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(74) Agents: ESMOND, Robert, W. et al.; Sterne, Kessler,
Goldstein & Fox, P.L.L.C., 1100 New York Avenue, N.W.,
Washington, DC 20005 (US).

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(71) Applicant: INVITROGEN CORPORATION [US/US];
1600 Faraday Avenue, Carlsbad, CA 92008 (US).

(71) Applicants and

(72) Inventors: BENNETT, Robert, P. [US/US]; 1269 Rain-
bow Ridge Lane, Encinitas, CA 92024 (US). WELCH,
Peter, J. [US/US]; 4689 E. Talmadge Drive, San Diego,
CA 92116 (US). HARWOOD, Steven [US/US]; 5672
Stetson Place, Oceanside, CA 92057 (US). MADDEN,
Knut [US/US]; 2705 Glasgow Drive, Carlsbad, CA 92008
(US). FRIMPONG, Kenneth [GH/US]; 4152 Camino
Ticino, San Diego, CA 92122 (US). FRANKE, Kenneth,
E. [US/US]; 9934 Shelburne Terrace, #308, Gaithersburg,
MD 20878 (US).

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(54) Title: VIRAL VECTORS CONTAINING RECOMBINATION SITES

(57) Abstract: The present invention provides compositions and methods for the construction of nucleic acids comprising all or portion of a viral genome. Nucleic acid molecules of the invention may be constructed to contain multiple recombination and/or topoisomerase recognition sites. The compositions include vectors having multiple recombination sites with unique specificity that contain all or a portion of a viral genome. The methods permit the insertion of a sequence of interest into a viral genome using recombinational and/or topoisomerase-mediated cloning. The present invention also provides methods of constructing recombinant virus, methods of expressing polypeptides, and methods of expressing fusion polypeptides.

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VIRAL VECTORS CONTAINING RECOMBINATION SITES

BACKGROUND OF THE INVENTION

Field of the Invention

[0001] The present invention relates to the fields of biotechnology and molecular biology. In particular, the present invention relates to nucleic acids comprising multiple recombination sites and comprising all or a portion of a viral genome as well as viruses and/or plasmids containing multiple recombination sites and their uses.

Related Art

[0002] Recombinant viruses are currently used in wide variety of applications. Viruses may be used for medical applications, for example, in gene therapy applications and/or as vaccines. Viruses may also be used in biotechnology applications, for example, as vectors to clone nucleic acids of interests and/or to produce proteins. Examples of recombinant viruses that have been used include, but are not limited to, herpes viruses (see, for example, U.S. patent no. 5,672,344, issued to Kelly, *et al.*), pox viruses such as vaccinia virus (see, for example, Moss, *et al.*, 1997, in Current Protocols in Molecular Biology, Chapters 16.15-16.18, John Wiley & Sons), papilloma viruses (see, for example, U.S. patent no. 6,342,224, issued to Bruck, *et al.*), retroviruses (see, for example U.S. patent no. 6,300,118, issued to Chavez, *et al.*), adenoviruses (see, for example, U.S. patent no. 6,261,807, issued to Crouzet, *et al.*), adeno-associated viruses (AAV, see for example, U.S. patent no. 5,252,479, issued to Srivastava), and coxsackie viruses (see, for example, U.S. patent no. 6,323,024).

[0003] When the viral nucleic acid is not infectious—for example, pox viruses—construction of recombinant viruses may involve *in vivo* homologous recombination in a virus-infected cell between the viral genome and concomitantly transfected plasmid bearing a sequence of interest flanked by viral sequences. When the viral nucleic acid is infectious—for example, adenovirus—a modified viral nucleic acid may be prepared and transfected

into a host cell. Either methodology requires the preparation of a nucleic acid molecule containing a sequence of interest and some or all of the viral sequence. The preparation of this nucleic acid molecule may be a time-consuming, laborious process.

[0004] Adenoviruses are non-enveloped viruses with a 36 kb DNA genome that encodes more than 30 proteins. At the ends of the genome are inverted terminal repeats (ITRs) of approximately 100-150 base pairs. A sequence of approximately 300 base pairs located next to the 5'-ITR is required for packaging of the genome into the viral capsid. The genome as packaged in the virion has terminal proteins covalently attached to the ends of the linear genome.

[0005] The genes encoded by the adenoviral genome are divided into early and late genes depending upon the timing of their expression relative to the replication of the viral DNA. The early genes are expressed from four regions of the adenoviral genome termed E1-E4 and are transcribed prior to onset of DNA replication. Multiple genes are transcribed from each region. Portions of the adenoviral genome may be deleted without affecting the infectivity of the deleted virus. The genes transcribed from regions E1, E2, and E4 are essential for viral replication while those from the E3 region may be deleted without affecting replication. The genes from the essential regions can be supplied in trans to allow the propagation of a defective virus. For example, deletion of the E1 region of the adenoviral genome results in a virus that is replication defective. Viruses deleted in this region are grown on 293 cells that express the viral E1 genes from the genome of the cell.

[0006] In addition to permitting the construction of a safer, replication-defective viruses, deletion and complementation in trans of portions of the adenoviral genome and/or deletion of non-essential regions make space in the adenoviral genome for the insertion of heterologous DNA sequences. The packaging of viral DNA into a viral particle is size restricted with an upper limit of approximately 38 kb of DNA. In order to maximize the amount of heterologous DNA that may be inserted and packaged, viruses have been constructed that lack all of the viral genome except the ITRs and packaging sequence (see, U.S. patent no. 6,228,646). All of the viral functions necessary

for replication and packaging are provided in trans from a defective helper virus that is deleted in the packaging signal.

[0007] Recombinant adenoviruses have been used as a gene transfer vectors both *in vitro* and *in vivo*. Their principal attractions as a gene transfer vector are their ability to infect a wide variety of cells including dividing and non-dividing cells and their ability to be grown in cell culture to high titers. A number of systems to insert heterologous DNA into the adenoviral genome have been developed. The adenoviral genome has been inserted into a yeast artificial chromosome (YAC, see Ketner, *et al.*, *PNAS* 91:6186-90, 1994). Mutations may be introduced into the genome by transfecting a mutation-containing plasmid into a yeast cell that contains the adenoviral YAC. Homologous recombination between the YAC and the plasmid introduces the mutation into the adenoviral genome. The adenoviral genome can be removed from the YAC by restriction digest and the genome released by restriction digest is infectious when transfected into host cells. A similar system using two plasmids has been developed in *E. coli* (see Crouzet, *et al.*, *PNAS* 94:1414-1419, 1997, and U.S. patent no. 6,261,807). In this system, the adenoviral genome is introduced into a *inc-P* derived replicon. Mutations are introduced by homologous recombination with a plasmid containing a *ColE1* origin of replication. The ITRs in the *inc-P* plasmid are flanked by a restriction site not present in the rest of the viral genome, thus, infectious DNA can be liberated from the plasmid by restriction digest.

[0008] A number of viruses containing recombination site sequences and/or encoding recombinases have been prepared. For example, the Cre recombinase has been expressed from recombinant adenovirus and used to excise fragments from a mouse genome that were flanked with lox sites (see, Wang, *et al.*, *PNAS* 93:3932-3936, 1996). U.S. patent no. 6,156,497 describes a system for constructing adenoviral genomes utilizing a first nucleic acid having an ITR, packaging signal, DNA of interest, and recombination site and a second nucleic acid having a recombination site and an ITR to which is bound a terminal protein. In the presence of recombinase, the two molecules are joined to form an infectious viral DNA.

[0009] Baculoviruses are large, enveloped viruses that infect arthropods. Baculoviral genomes are double-stranded DNA molecules of approximately

130 kbp in length. Baculoviruses have gained widespread use as systems in which to express proteins, particularly proteins from eukaryotic organisms (e.g., mammals), as the insect cells used to culture the virus may more closely mimic the post-translational modifications (e.g., glycosylation, acylation, etc.) of the native organism.

[0010] Numerous expression systems utilizing recombinant baculoviruses have been developed. General methods for constructing recombinant baculoviruses for expression of heterologous proteins may be found in Piwnicka-Worms, *et al.*, (1997) *Expression of Proteins in Insect Cells Using Baculovirus Vectors*, in *Current Protocols in Molecular Biology*, Chapter 16, pp. 16.9.1 to 16.11.12, Ausubel, *et al.* Eds., John Wiley & Sons, Inc. Other expression systems are known, for example, United States patent number 6,255,060, issued to Clark, *et al.* discloses a baculoviral expression system for expressing nucleotide sequences that include a tag. United States patent number 5,244,805, issued to Miller, discloses a baculoviral expression system that utilizes a modified promoter not naturally found in baculoviruses. United States patent number 5,169,784, issued to Summers, *et al.* discloses a baculoviral expression system that utilizes dual promoters (e.g., a baculoviral early promoter and a baculoviral late promoter). United States patent number 5,162,222, issued to Guarino, *et al.* discloses a baculoviral expression system that can be used to create stable cells lines or infectious viruses expressing heterologous proteins from a baculoviral immediate-early promoter (*i.e.*, IEN). United States patent number 5,155,037, issued to Summers, *et al.* discloses a baculoviral expression system that utilizes insect cell secretion signal to improve efficiency of processing and secretion of heterologous genes. United States patent number 5,077,214, issued to Guarino, *et al.* discloses the use of baculoviral early gene promoters to construct stable cell lines expression heterologous genes. United States patent number 4,879,239, issued to Smith, *et al.* discloses a baculoviral expression system that utilizes the baculoviral polyhedrin promoter to control the expression of heterologous genes.

[0011] Various methods of constructing recombinant baculoviruses have been used. A frequently used method involves transfecting baculoviral DNA and a plasmid containing baculoviral sequences flanking a heterologous sequence. Homologous recombination between the plasmid and the baculoviral genome

results in a recombinant baculovirus containing the heterologous sequences. This results in a mixed population of recombinant and non-recombinant viruses. Recombinant baculoviruses may be isolated from non-recombinant by plaque purification. Viruses produced in this fashion may require several rounds of plaque purification to obtain a pure strain. Methods to reduce the background of non-recombinant viruses produced by homologous recombination methods have been developed. For example, a linearized baculoviral genome containing a lethal deletion, BaculoGold™, is commercially available from BD Biosciences, San Jose, CA. The lethal deletion is rescued by homologous recombination with plasmids containing baculoviral sequences from the polyhedrin locus.

[0012] Methods utilizing direct insertion of foreign sequences into a baculoviral genome are also known. For example, Peakman, *et al.* (*Nucleic Acids Res* 20(3):495-500, 1992) disclose the construction of baculoviruses having a *lox* site in the genome. Heterologous sequences may be moved into the genome by *in vitro* site-specific recombination between a plasmid having a *lox* site and the baculoviral genome in the presence of Cre recombinase. United States patent number 5,348,886, issued to Lee, *et al.* discloses a baculoviral expression system that utilizes a bacmid (a hybrid molecule comprising a baculoviral genome and a prokaryotic origin of replication and selectable marker) containing a recombination site for Tn7 transposon. Prokaryotic cells carrying the bacmid are transformed with a plasmid having a Tn7 recombination site and with a plasmid expressing the activities necessary to catalyze recombination between the Tn7 sites. Heterologous sequences present on the plasmid are introduced into the bacmid by site-specific recombination between the Tn7 sites. The recombinant bacmid may be purified from the prokaryotic host and introduced into insect cells to initiate an infection. Recombinant viruses carrying the heterologous sequence are produced by the cells transfected with the bacmid.

[0013] The family *Retroviridae* contains three subfamilies: 1) *oncovirinae*; 2) *spumavirinae*; and 3) *lentivirinae*. Retroviruses (*e.g.*, lentiviruses) are viruses having an RNA genome that replicate through a DNA intermediate. A retroviral particle contains two copies of the RNA genome and viral replication enzymes in a RNA-protein viral core. The core is surrounded by a

viral envelop made up of virally encoded glycoproteins and host cell membrane. In the early steps of infection, retroviruses deliver the RNA-protein complex into the cytoplasm of the target cell. The RNA is reverse transcribed into double-stranded cDNA and a pre-integration complex containing the cDNA and the viral factors necessary to integrate the cDNA into the target cell genome is formed. The complex migrates to the nucleus of the target cell and the cDNA is integrated into the genome of the target cell. As a consequence of this integration, the DNA corresponding to the viral genome (and any heterologous sequences contained in the viral genome) is replicated and passed on to daughter cells. This makes it possible to permanently introduce heterologous sequences into cells.

[0014] A wide variety of retroviruses are known, for example, leukemia viruses such as a Moloney Murine Leukemia Virus (MMLV) and immunodeficiency viruses such as the Human Immunodeficiency Virus (HIV). Representative examples of retroviruses include, but are not limited to, the Gibbon Ape Leukemia virus (GALV), Avian Sarcoma-Leukosis Virus (ASLV), which includes but is not limited to Rous Sarcoma Virus (RSV), Avian Myeloblastosis Virus (AMV), Avian Erythroblastosis Virus (AEV) Helper Virus, Avian Myelocytomatosis Virus, Avian Reticuloendotheliosis Virus, Avian Sarcoma Virus, Rous Associated Virus (RAV), and Myeloblastosis Associated Virus (MAV).

[0015] Retroviruses have found widespread use as gene therapy vectors. To reduce the risk of transmission of the gene therapy vector, gene therapy vectors have been developed that have modifications that prevent the production of replication competent viruses once introduced into a target cell. For example, United States patent number 5,741,486 issued to Pathak, *et al.* describes retroviral vectors comprising direct repeats flanking a sequence that is desired to be deleted (*e.g.*, a *cis*-acting packing signal) upon reverse transcription in a host cell. Deletion of the packing signal prevents packaging of the recombinant viral genome into retroviral particles, thus preventing spread of retroviral vectors to non-target cells in the event of infection with replication competent viruses. United States patent numbers 5,686,279, 5,834,256, 5,858, 740, 5,994,136, 6,013, 516, 6,051, 427, 6,165,782, and 6,218,187 describe a retroviral packaging system for preparing high titer

stocks of recombinant retroviruses. Plasmids encoding the retroviral functions required to package a recombinant retroviral genome are provided *in trans*. The packaged recombinant retroviral genomes may be harvested and used to infect a desired target cell.

[0016] The family *Herpesviridae* contains three subfamilies 1) *alphaherpesvirinae*, containing among others human herpesvirus 1; 2) *betaherpesvirinae*, containing the cytomegaloviruses; and 3) *gammaherpesvirinae*. Herpesviruses are enveloped DNA viruses. Herpesviruses form particles that are approximately spherical in shape and that contain one molecule of linear dsDNA and approximately 20 structural proteins. Numerous herpesviruses have been isolated from a wide variety of hosts. For example, United Patent No. 6,121,043 issued to Cochran, *et al.* describes recombinant herpesvirus of turkeys comprising a foreign DNA inserted into a non-essential region of the herpesvirus of turkeys genome; United States Patent No. 6,410,311 issued to Cochran, *et al.* describes recombinant feline herpesvirus comprising a foreign DNA inserted into a region corresponding to a 3.0 kb EcoRI-SalI fragment of a feline herpesvirus genome, United States Patent No. 6,379,967 issued to Meredith, *et al.*, describes herpesvirus saimiri, (HVS; a lymphotropic virus of squirrel monkeys) as a viral vector; and United States Patent No. 6,086,902 issued to Zamb, *et al.* describes recombinant bovine herpesvirus type 1 vaccines.

[0017] Herpesviruses have been used as vectors to deliver exogenous nucleic acid material to a host cell. In addition to the examples above, United States Patent No. 4,859,587, issued to Roizman describes recombinant herpes simplex viruses, vaccines and methods, United States Patent No. 5,998,208 issued to Fraefel, *et al.*, describes a helper virus-free herpesvirus vector packaging system, United States Patent No. 6,342,229 issued to O'Hare, *et al.*, describes herpesvirus particles comprising fusion protein and their preparation and use and United States Patent 6,319,703 issued to Speck describes recombinant virus vectors that include a double mutant herpesvirus such as an herpes simplex virus-1 (HSV-1) mutant lacking the essential glycoprotein gH gene and having a mutation impairing the function of the gene product VP16.

[0018] RNA viruses, such as those of the families *Flaviviridae* and *Togaviridae* have also been used to deliver exogenous nucleic acids to target

cells. For example, members of the genus *alphavirus* in the family *Togaviridae* have been engineered for the high level expression of heterologous RNAs and polypeptides (Frolov *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 93: 11371-11377 (1996)). Alphaviruses are positive stranded RNA viruses. A single genomic RNA molecule is packaged in the virion. RNA replication occurs by synthesis of a full-length minus strand RNA intermediate that is used as a template for synthesis of positive strand genomic RNA as well for synthesis of a positive strand sub-genomic RNA initiated from an internal promoter. The sub-genomic RNA can accumulate to very high levels in infected cells making alphaviruses attractive as transient expression systems. Examples of alphaviruses are Sindbis virus and Semliki Forest Virus. Kunjin virus is an example of a flavivirus. Sub-genomic replicons of Kunjin virus have been engineered to express heterologous polypeptides (Khromykh and Westaway, *J. Virol.* 71: 1497-1505 (1997)). The genomic RNA of both flaviviruses and togaviruses are infectious; transfection of the naked genomic RNA results in production of infective virus particles.

[0019] Methods for constructing recombinant viruses are typically laborious and time consuming. There remains a need in the art for materials and methods for the rapid and precise and rapid construction of recombinant viruses containing a nucleic acid region of interest. This need and others are met by the present invention.

BRIEF SUMMARY OF THE INVENTION

[0020] The present invention provides, in part, a nucleic acid molecule comprising all or a portion of a viral genome (*e.g.*, an adenovirus genome, a baculovirus genome, a herpesvirus genome, a pox virus genome, an adeno-associated virus genome, a retrovirus genome, a flavivirus genome, a togavirus genome, an alphavirus genome, an RNA virus genome, etc.). Nucleic acid molecules of the invention may further comprise at least two recombination sites (*e.g.*, three, four, five, six, seven, eight, nine, ten, etc.) that, in most instances, do not recombine with each other. In particular embodiments, the viral genome may be an adenoviral genome, a baculoviral genome, a retroviral genome (*e.g.*, a lentiviral genome), an RNA virus genome or a herpesvirus genome. In some embodiments, the viral genome is not an

adenoviral genome, is not a baculoviral genome, is not a retroviral genome (*e.g.*, a lentiviral genome), and/or is not a herpesvirus genome. In some embodiments, the viral genome is not from a virus that infects prokaryotic organisms. In some embodiments, one or more of the two or more recombination sites is not a *lox* site. In some embodiments, nucleic acid molecules comprising one or more sequences of interest are combined with nucleic acid molecules comprising all or a portion of a viral genome using a recombination system that does not use a recombination system derived from a transposon (*e.g.*, Tn7). In some embodiments, nucleic acid molecules of the invention may not contain a *lox* site.

[0021] Optionally, nucleic acid molecules of the invention may comprise one or more features that confer desired characteristics on the nucleic acid molecules. Examples of features include, but are not limited to, promoters, viral terminal repeats (*e.g.*, long terminal repeats (LTRs)), splice sites (*e.g.*, 5'-splice donor sites and/or 3'-splice acceptor sites), packaging signals, nucleic acid sequences responsive to one or more viral proteins (*e.g.*, rev response element (RRE)), recognition sites (*e.g.*, restriction enzyme recognition sites), recombination sites, sequences encoding marker proteins or polypeptides (*e.g.*, antibiotic resistance enzymes, toxic proteins, etc.), sequences encoding epitopes recognizable by an antibody (*e.g.*, V5 epitope), origins of replication (which may function in prokaryotic and/or eukaryotic cells), intervening sequences (*e.g.*, β -globin intron), internal ribosome entry sequences (IRES), and polyadenylation signals (*e.g.*, SV40 polyadenylation signal). Additional examples of such nucleic acid molecules include those which contain at least (1) one or more (*e.g.*, one, two, three, four, five, six, seven, eight, nine, etc.) component of one or more of the vectors represented in FIGs. 1, 2, 4, 5, 6, 7, 8, 9, 10, 15, 18, 20, 22, 34, 36, 37, 49, 57, 58, 59, 60, 69, 70, 71 or 72; or (2) one or more components of such vectors which confer the same or similar feature upon a nucleic acid molecule. As a specific example, a nucleic acid molecule of the invention may be a vector which comprises, in addition to recombination sites, at least one blasticidin resistance marker (*see, e.g.*, FIG. 22), at least one GP64 promoter (*see, e.g.*, FIG. 22), at least one RSV promoter (*see, e.g.*, FIG. 36A), at least one β -globin intron (*see, e.g.*, FIG. 37A), at least one ampicillin resistance marker (*see, e.g.*, FIG. 37A), and at

least one bacterial origin of replication (*see, e.g.*, FIG. 37A). In most instances, the combinations of components selected for inclusion in a nucleic acid molecule will be designed to provide activities intended for a particular use. For example, a vector which is capable of expressing a nucleic acid insert in more than one type of eukaryotic cells (*e.g.*, human cells and insect cells) and is replicable in prokaryotic cells (*e.g.*, *E. coli* cells) may be desired. Thus, the components which are selected for inclusion in nucleic acid molecules of the invention will typically be determined by the particular use for which it is designed. The invention further includes methods for making and using such nucleic acid molecules as described, for example, elsewhere herein.

[0022] Viruses produced using nucleic acids of the present invention may be used as viral vectors (*e.g.*, viruses containing at least one heterologous sequence), for example, to deliver exogenous sequences to cells or organisms. The present invention also contemplates compositions comprising nucleic acids and/or viruses of the invention, as well as methods of making and using such nucleic acids, viruses, and compositions.

[0023] Viral genomes that may be used with the present invention (*e.g.*, retroviral genomes, adenoviral genomes, herpesvirus genomes, genomes of RNA viruses, and/or baculoviral genomes) may be wild type or may contain one or more mutations, insertions and/or deletions. In some embodiments, viral genomes for use in the practice of the present invention may be adenoviral genomes containing one or more deletions. Deleted adenoviral genomes may be deleted in one or more regions of the genome. Regions of the adenoviral genome that may be deleted, include, but are not limited to, the E1 and E3 regions.

[0024] Adenoviral genomes for use in the present invention may be infectious. In some embodiments, an adenoviral genome may be infectious when introduced into cells expressing one or more adenoviral proteins (*e.g.*, the E1 proteins as in 293 cells). In some embodiments, a viral genome used in the invention is an Ad5 viral genome.

[0025] Baculoviral genomes that may be used in the practice of the present invention may be entire genomes or may contain one or more deletions, for example, at the polyhedrin locus. Suitable genomes include those from any virus in the family *Baculoviridae*. Suitable viral genomes include, but are not

limited to, those from occluded baculoviruses (e.g., nuclear polyhedrosis viruses (NPV) such as *Autographa californica* nuclear polyhedrosis virus (AcMNPV), *Choristoneura fumiferana* MNPV (CfMNPV), *Mamestra brassicae* MNPV (MbMNPV), *Orgyia pseudotsugata* MNPV (OpMNPV), *Bombyx mori* S Nuclear Polyhedrosis Virus (BmNPV), *Heliothis zea* SNPV (HzSnpv), and *Trichoplusia ni* SNPV (TnSnpv) and granulosis viruses (GV) (e.g., *Plodia interpunctella* granulosis virus (PiGV), *Trichoplusia ni* granulosis virus (TnGV), *Pieris brassicae* granulosis virus (PbGV), *Artogeia rapae* granulosis virus (ArGV), and *Cydia pomonella* granulosis virus (CpGV)). Suitable genomes also include, but are not limited to, those from non-occluded baculoviruses (NOB) (e.g., *Heliothis zea* NOB (HzNOB), *Oryctes rhinoceros* virus), etc.

[0026] In some embodiments, viral genomes for use in the practice of the present invention may be retroviral genomes containing one or more deletions. Deleted retroviral genomes may be deleted in one or more regions of the genome. Regions of the retroviral genome that may be deleted, include, but are not limited to, the gag, pol, env, and rev regions. In some embodiments, a retroviral genome may be deleted of all retroviral sequences except the 5'-LTR, 3'-LTR and packaging signal (Ψ). In some embodiments, retroviral genomes of the present invention may comprise one or more heterologous sequences (e.g., sequences derived from another organism such as another virus). In a particular embodiment, nucleic acid molecules of the invention may comprise a deleted retroviral genome and may also comprise one or more heterologous sequences that may be promoter sequences. In some embodiments, nucleic acid molecules of the invention may comprise a deleted retroviral genome and may further comprise the CMV promoter.

[0027] In some embodiments, nucleic acid molecules of the present invention may be in the form of plasmids and/or bacmids comprising one or more origins of replication and, optionally, one or more selectable markers. In certain embodiments, nucleic acid molecules of the invention (e.g., plasmids and/or bacmids) may comprise one or more recognition sequences (e.g., recombination sequences, topoisomerase sequences, restriction enzyme sequences, etc.), which may be recognized by the same or different enzymes. For example, in some embodiments, plasmids comprising all or a portion of

the viral genome may comprise one or more recombination sites that may not recombine with each other. In certain embodiments, nucleic acid molecules of the invention (*e.g.*, plasmids and/or bacmids) may comprise restriction enzyme recognition sequences, which may be recognized by the same or different restriction endonucleases, arranged such that digestion with one or more restriction enzymes that recognize the recognition sequences produces a linear molecule comprising the viral genome. In some embodiments, digestion with a restriction enzyme may remove a portion of plasmid and/or bacmid. For example, in some embodiments, plasmids comprising all or a portion of the adenoviral genome may be digested so as to remove the origin of replication and, optionally, the selectable marker from the plasmid. In another example, a nucleic acid molecule comprising all or a portion of a baculoviral genome may be digested with a restriction enzyme that linearizes the baculoviral genome, for example, by cleaving the nucleic acid molecule at a recognition site located between two recombination sites (see Fig. 20). In embodiments of this type, the baculoviral genome may be re-circularized by recombination with a second nucleic acid molecule having recombination sites that are capable of recombining with those in the nucleic acid molecule comprising all or a portion of the baculoviral genome. In particular embodiments, the restriction enzyme recognition sites may be recognized by two different restriction enzymes. Thus, the invention includes methods for selecting recombinant nucleic acid molecules (*e.g.*, recombinant baculoviral vectors). The method may comprise recombining a first nucleic acid molecule, which may be linearized, with a second nucleic acid molecule to produce a circularized molecule that is capable of replicating when introduced into a suitable host cell. The method may also comprise selecting against re-circularized first nucleic acid molecule that did not undergo recombination with the second nucleic acid molecule. In some embodiments, the first nucleic acid molecule may be a linearized baculoviral genome.

[0028] A nucleic acid sequence of interest may be inserted into the nucleic acid molecule of the invention using recombinational cloning techniques. In some embodiments, a nucleic acid molecule of the invention may comprise a heterologous promoter (*e.g.*, the CMV promoter) and one or more recombination sites arranged such that a nucleic acid sequence of interest can

be inserted into the nucleic acid molecule of the invention by recombination with one or more of the recombination sites and, after insertion, the nucleic acid sequence of interest may be operably linked to the heterologous promoter. In some embodiments, a nucleic acid molecule of the invention may have a heterologous promoter located adjacent to two recombination sites that do not recombine with each other. A nucleic acid sequence of interest can be inserted into the nucleic acid molecule of the invention between the two recombination sites and may then be operably linked to the heterologous promoter.

[0029] Any nucleic acid sequence of interest may be placed between the recombination sites present in the nucleic acids of the present invention. For example, the nucleic acid sequence between the recombination sites may encode one or more polypeptides of interest. The viral vectors of the present invention may be used to express libraries of sequences, for example, genomic libraries or cDNA libraries. A sequence of interest may be a sequence coding for a polypeptide or may be a sequence that does not encode a polypeptide. Examples of sequences of interest that do not encode a polypeptide include, but are not limited to, sequences encoding tRNA sequences (*e.g.*, suppressor tRNA sequences), sequences encoding ribozyme sequences, promoter sequences, enhancer sequences, repressor sequences and the like. In some embodiments, the sequence of interest may encode one or more polypeptides and may further comprise one or more stop codons in the sequence. In some embodiments, the nucleic acid between the recombination sites comprises at least one selectable marker. In some embodiments, the sequence of interest comprises a sequence encoding at least one suppressor tRNA and/or at least one aminoacyl-tRNA synthetase.

[0030] In some embodiments, the present invention provides nucleic acid molecules comprising all or a portion of more than one viral genome. For example, a nucleic acid molecule of the invention may comprise all or a portion of a first viral genome (*e.g.*, a retroviral genome) and all or a portion of one or more additional viral genomes (*e.g.*, an adenoviral genome, a baculoviral genome, a herpesvirus genome, a pox virus genome, an RNA virus genome, etc). In some embodiments, the nucleic acid molecules of the invention may comprise nucleic acid sequences from more than one virus. Nucleic acid molecules of this type may comprise viral sequences that permit

the replication of the nucleic acid in more than one type of organism (*e.g.*, mammalian cells and insect cells) and may also include sequences capable of functioning as transcriptional regulatory sequences (*e.g.*, promoters, enhancers, etc.) that function in more than one cell type. For example, one viral sequence may function as a promoter in one cell type (*e.g.*, mammalian) while another viral sequence may function as a promoter in another cell type (*e.g.*, insect).

[0031] In another aspect, the present invention provides a method of constructing a nucleic acid molecule comprising all or a portion of one or more viral genomes (*e.g.*, a recombinant virus such as a viral vector). In some embodiments, methods of the invention may comprise providing at least a first nucleic acid molecule comprising all or a portion of at least one viral genome and at least a first and a second recombination site that do not recombine with each other. Methods of the invention may also entail contacting at least a first nucleic acid molecule with at least a second nucleic acid molecule comprising at least one sequence of interest flanked by at least a third and a fourth recombination site under conditions causing recombination between the first and third recombination site and between the second and fourth recombination site. In some embodiments, the viral genome may be an adenoviral genome, for example, an Ad5 adenoviral genome. In some embodiments, the viral genome may be a baculoviral genome, for example, an *Autographa californica* multiple nuclear polyhedrosis virus (AcMNPV) genome. In some embodiments, the viral genome may be a retroviral genome (*e.g.*, a lentiviral genome).

[0032] In some embodiments, a first nucleic acid molecule comprising all or a portion of a viral genome for use in the methods of the invention may be a plasmid that may comprise an origin of replication and a selectable marker. The first nucleic acid molecule may, optionally, contain two restriction enzyme recognition sequences, which may be for the same or different restriction enzymes, arranged such that digestion with the appropriate restriction enzyme or restriction enzymes produces a linear molecule comprising the viral genome (*e.g.*, adenoviral genome) and lacking the origin of replication and/or the selectable marker.

[0033] In some embodiments, the first nucleic acid molecule may comprise at least a first and a second recombination site, which may or may not recombine with each other, and the portion of the first nucleic acid molecule between the first and second recombination sites may comprise a sequence encoding at least one selectable marker. In some embodiments, a second nucleic acid molecule, which may or may not comprise viral sequences, may comprise at least a third and a fourth recombination site and a sequence of interest between the third and fourth recombination site. The sequence of interest may be any sequence, for example, a sequence encoding a polypeptide or a sequence of a functional RNA (*e.g.*, a suppressor tRNA sequence). In some embodiments, the first and second nucleic acid molecules may be contacted with one or more recombination proteins such that the sequence of interest is transferred to the first nucleic acid molecule resulting in a first nucleic acid molecule comprising all or a portion of a viral genome and further comprising at least one sequence of interest (*e.g.*, a polypeptide coding region, a tRNA coding sequence etc.). The present invention also contemplates compositions comprising a nucleic acid molecule comprising all or a portion of a viral genome and further comprising at least one sequence of interest, as well as methods of making and using such nucleic acids and compositions. In some embodiments, the sequence of interest may be a tRNA coding sequence.

[0034] In some embodiments, a first nucleic acid molecule comprising all or a portion of a viral genome for use in the methods of the invention may be a bacmid that may comprise an origin of replication and a selectable marker. The first nucleic acid molecule may, optionally, contain a restriction enzyme recognition sequence, located such that digestion with the appropriate restriction enzyme produces a linear molecule comprising the viral genome (*e.g.*, baculoviral genome). In some embodiments, the first nucleic acid molecule may comprise at least a first and a second recombination site, which may or may not recombine with each other, and the recognition site for the restriction enzyme may be located between the recombination sites. Optionally, the portion of the first nucleic acid molecule between the first and second recombination sites may comprise a sequence encoding at least one selectable marker. In some embodiments, a second nucleic acid molecule, which may or may not comprise viral sequences, may comprise at least a third

and a fourth recombination site and the sequence between the third and fourth recombination site comprises a sequence of a functional RNA (e.g., a suppressor tRNA sequence). In some embodiments, the first and second nucleic acid molecules may be contacted with one or more recombination proteins such that the functional sequence (e.g., a sequence encoding a suppressor tRNA sequence) is transferred to the first nucleic acid molecule resulting in the first nucleic acid molecule re-circularizing and further comprising at least one functional sequence (e.g., a sequence encoding a tRNA). The present invention also contemplates compositions comprising a nucleic acid molecule comprising all or a portion of a viral genome and further comprising at least one functional sequence, as well as methods of making and using such nucleic acids and compositions.

[0035] The present invention also provides, in part, materials and methods for joining or combining two or more (e.g., two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, seventy-five, one hundred, two hundred, etc.) nucleic acid segments and/or nucleic acid molecules by a recombination reaction between recombination sites—at least one of which is present on each molecule and/or segment—in order to construct a nucleic acid molecule comprising all or a portion of a viral genome (e.g., a retroviral genome, an adenoviral genome and/or a baculoviral genome). In embodiments of this type, one or more nucleic acid segments and/or nucleic acid molecules may comprise viral nucleic acid sequences. Such recombination reactions to join multiple nucleic acid segments and/or nucleic acid molecules according to the invention may be conducted *in vivo* (e.g., within a cell, tissue, organ or organism) or *in vitro* (e.g., cell-free systems). The invention also relates to hosts and host cells comprising the viral vectors and/or nucleic acid molecules of the invention. The invention also relates to kits for carrying out methods of the invention, and to compositions for carrying out methods of the invention, as well as to compositions used in and made while carrying out the methods of the invention.

[0036] Nucleic acid molecules prepared by methods of the invention may be used for any purpose known to those skilled in the art. For example, nucleic acid molecules of the invention may be used to express proteins or peptides encoded by these nucleic acid molecules and may also be used to create novel

fusion proteins by expressing different nucleic acid sequences linked by the methods of the invention. Nucleic acids of the invention may also be used to produce RNA molecules that are not translated into polypeptides or proteins, for example, tRNAs, anti-sense molecules, interfering RNA and/or ribozymes.

[0037] Nucleic acid molecules of the invention may be used as part of a system to generate replication-defective viral particles. For example, nucleic acid molecules of the invention may be packaged into a viral particle using techniques known in the art. Packaging may be accomplished by providing requisite packaging activities *in trans*, for example, on a different nucleic acid molecule and/or in the genome of a cell. In a particular example, nucleic acid molecules of the invention may be used to construct a replication-defective lentivirus. In a particular embodiment, nucleic acid molecules of the invention may comprise lentiviral long terminal repeats and packaging signal and other activities required to package the nucleic acid molecule of the invention may be provided *in trans*, for example, may be expressed from one or more plasmids.

[0038] In some aspects, methods of the present invention may comprise introducing a nucleic acid molecule of the invention into a cell or population of cells and detecting the presence or absence of the nucleic acid molecule. Such detection may be accomplished, for example, by detecting the presence or absence of one or more selectable marker present on the nucleic acid molecule. Optionally, a selectable marker may be a nucleic acid sequence encoding a polypeptide having β -lactamase activity. Detection may be accomplished by contacting a cell or population of cells with a fluorogenic substrate for β -lactamase activity and detecting fluorescence of the cell or population of cells. In a specific embodiment, the fluorogenic substrate may be CCF2/AM and fluorescence may be detected by illuminating the cell with light having a wavelength of 405 nm and detecting fluorescence at a wavelength of approximately 450 nm and at a wavelength of approximately 520 nm. Methods may also comprise comparing the amount of fluorescence observed at 450 nm and 520 nm, for example, by determining a ratio between the observed fluorescence amounts. Methods may also comprise physically separating cells having a desired nucleic acid molecule by fluorescent activated cell sorting (FACS).

[0039] The present invention provides methods for infecting, transfecting, transducing and/or otherwise introducing the nucleic acid molecules of the invention into host cells and, optionally, expressing one or more sequences of interest present on the nucleic acid molecule of the invention. Suitable host cells may be dividing or non-dividing cells. In a particular embodiment, host cells using in connection with the methods of the invention are non-dividing cells. For example, one or more nucleic acid molecule of the invention may be introduced into one or more non-dividing cells. One or more of the nucleic acid molecules may comprise a sequence of interest that may encode a polypeptide or an untranslated RNA. The methods of the invention may result in the production in the non-dividing cells of a polypeptide or untranslated RNA encoded by the sequence of interest. Nucleic acid molecules of the invention for use in the expression of a sequence of interest in a non-dividing cell may comprise one or more sequences from one or more viruses, for example, from an adenovirus and/or a lentivirus. A nucleic acid molecule of the invention for expression of a sequence of interest in a non-dividing cell may comprise one or more adenoviral sequences. A nucleic acid molecule of the invention for expression of a sequence of interest in a non-dividing cell may comprise one or more lentiviral sequences.

[0040] Recombination sites for use in the methods and/or compositions of the invention may be any recognition sequence on a nucleic acid molecule that participates in a recombination reaction mediated or catalyzed by one or more recombination proteins. In those embodiments of the present invention utilizing more than one (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.) recombination sites, such recombination sites may be the same or different and may recombine with each other or may not recombine or not substantially recombine with each other. Recombination sites contemplated by the invention also include mutants, derivatives or variants of wild-type or naturally occurring recombination sites. Desired modifications can also be made to the recombination sites to include changes to the nucleotide sequence of the recombination site that cause desired sequence changes to the transcription product (*e.g.*, mRNA, tRNA, ribozyme, etc.) and/or desired amino acid changes in the translation product (*e.g.*,

polypeptide or protein) when transcription occurs across the modified recombination site.

[0041] Preferred recombination sites used in accordance with the invention include att sites, frt sites, dif sites, psi sites, cer sites, and lox sites or mutants, derivatives and variants thereof (or combinations thereof). Recombination sites contemplated by the invention also include portions of such recombination sites. Depending on the recombination site specificity used, the invention allows directional linking of nucleic acid molecules to provide desired orientations of the linked molecules or non-directional linking to produce random orientations of the linked molecules.

[0042] In certain embodiments, recombination proteins used in the practice of the invention comprise one or more proteins selected from the group consisting of Cre, Int, IHF, Xis, Flp, Fis, Hin, Gin, Cin, Tn3 resolvase, TndX, XerC, XerD, and Φ C31. In specific embodiments, the recombination sites comprise one or more recombination sites selected from the group consisting of lox sites; psi sites; dif sites; cer sites; frt sites; att sites; and mutants, variants, and derivatives of these recombination sites that retain the ability to undergo recombination.

[0043] In a specific aspect, the invention provides nucleic acid molecules and/or viral vectors that permit controlled expression of fusion polypeptides by suppression of one or more stop codons. According to the invention, a nucleic acid molecule, which may be any nucleic acid molecule, for example, a plasmid and/or a nucleic acid molecule comprising all or a portion of a viral genome and/or a viral vector produced by the methods of the invention, may comprise a sequence of interest that may comprise one or more stop codons (*e.g.*, TAG, TAA, and/or TGA) that may be suppressed. In embodiments of this type, mRNA is transcribed from the nucleic acid molecule. The transcribed mRNA molecule comprises at least a first coding sequence corresponding to the sequence of interest and at least one additional sequence containing a second coding region separated from the first coding sequence by a stop codon. Suppression of the stop codon allows expression of both the first and second coding sequences in a single polypeptide molecule. The nucleic acid sequence corresponding to the additional sequence may be contained on the sequence of interest or may be contained in a recombination

site or on the nucleic acid molecule. One or more suppressor tRNA molecules may be provided, for example, from any nucleic acid molecule such as a plasmid, a nucleic acid molecule comprising all or a portion of a viral genome and/or a viral vector of the invention.

[0044] Some embodiments of the present invention allow selective or controlled fusion protein expression by varying the suppression of selected stop codons. For example, a nucleic acid molecule, which may be a viral vector of the invention, may comprise three coding regions of interest separated by regions comprising stop codons. One or more of the coding regions of interest may be flanked by recombination sites. By suppressing the stop codon between the first and second coding regions a fusion polypeptide may be produced comprising amino acids encoded by the first and second coding region but not containing the amino acids encoded by the third region. Thus, use of different stop codons and variable control of suppression allows production of various fusion proteins or portions thereof encoded by all or different portions of the nucleic acid sequence of interest. In some embodiments, one or more of the coding regions in the sequence of interest may encode a polypeptide that comprises a sequence (preferably an N-terminal and/or a C-terminal tag sequence) encoding all or a portion of one or more of the following: the Fc portion of an immunoglobulin, an antibody, a β -glucuronidase, a β -lactamase, a β -galactosidase, a fluorescent protein (*e.g.*, green fluorescent protein, yellow fluorescent protein, red fluorescent protein, cyan fluorescent protein, etc.), a transcription activation domain, a protein or domain involved in translation, protein localization tag, a protein stabilization or destabilization sequence, a protein interaction domains, a binding domain for DNA, a protein substrate, a purification tag (*e.g.*, an epitope tag, maltose binding protein, a six histidine tag, glutathione S-transferase, etc.), and an epitope tag.

[0045] In one aspect, a stop codon may be included anywhere within the sequence of interest or within a recombination site contained by nucleic acid molecules, which may be nucleic acid molecules comprising all or a portion of a viral genome. Preferably, stop codons are located at or near the termini of the sequence of interest, although stop codons may be included internally within the sequence. In another aspect, the sequence of interest may comprise

the coding sequence of all or a portion of a target gene or open reading frame (ORF) of interest wherein the coding sequence is followed by a stop codon. The stop codon may then be followed by a recombination site allowing joining the sequence of interest to another nucleic acid molecule, which may be a nucleic acid molecule comprising all or a portion of a viral genome. After joining the sequence of interest with the nucleic acid molecule to form a recombinant nucleic acid molecule, the stop codon may be optionally suppressed by a suppressor tRNA molecule. In some embodiments of this type, one or more genes coding for one or more suppressor tRNA molecules (that may be the same or different) may be provided on the same nucleic acid molecule, or on another nucleic acid molecule. One or more genes coding for one or more suppressor tRNA molecules (that may be the same or different) may be provided on a different nucleic acid molecule, for example, a viral genome, a plasmid, a bacmid, a cosmid, a BAC, a YAC, a chromosome of the host cell into which the nucleic acid molecule of the invention is inserted, or any other nucleic acid molecule known to those skilled in the art. In some embodiments, one or more sequences encoding suppressor tRNAs may be provided on a nucleic acid molecule comprising all or a portion of a viral genome. In some embodiments, more than one copy (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc. copies) of the gene encoding the suppressor tRNA may be provided. In some embodiments, the transcription of the suppressor tRNA may be under the control of a regulatable (*e.g.*, inducible or repressible) promoter. In other embodiments, the transcription of the suppressor tRNA may be under the control of a constitutive promoter. When more than one gene encoding a suppressor tRNA is provided, the genes may be the same or different and may be expressed from the same or different promoters.

[0046] The sequence of interest may comprise a ORF of interest that may be provided with translation initiation signals (*e.g.*, Shine-Delgarno sequences, Kozak sequences and/or IRES sequences) in order to permit the expression of a polypeptide from the ORF with a native N-terminus when the stop codon is not suppressed. Further, the sequence of interest may be constructed by recombinational cloning of two or more different sequences resulting in recombination sites within the sequence of interest. Recombination sites that

reside between nucleic acid segments that encode components of fusion proteins may be designed either to not encode stop codons or to not encode stop codons in the fusion protein reading frame. A sequence of interest encoding a polypeptide may also be provided with a stop codon (*e.g.*, a suppressible stop codon) at the 3' end of the coding sequence. Similarly, when a fusion protein is produced from multiple nucleic acid segments (*e.g.*, three, four, five, six, eight, ten, etc. segments), nucleic acids sequences that encode stop codons can be omitted between each nucleic acid segment and/or nucleic acids that encodes a stop codon can be positioned at the 3' end of one or more of the segments and/or at the 3' end of the 3'-most segment of the fusion protein coding region.

[0047] In some embodiments, a tag sequence may be provided at both the N- and C-termini of the gene of interest. Optionally, the tag sequence at the N-terminus may be provided with a stop codon and an ORF of interest may be provided with a stop codon and the tag at the C-terminus may be provided with a stop codon. The stop codons may be the same or different.

[0048] In some embodiments, the stop codon of the N-terminal tag is different from the stop codon of the ORF of interest. In embodiments of this type, suppressor tRNAs corresponding to one or both of the stop codons may be provided. When both are provided, each of the suppressor tRNAs may be independently provided on the same vector (*e.g.*, plasmid, virus, etc.), on a different viral vector or other vector, or in the host cell genome. The suppressor tRNAs need not both be provided in the same way, for example, one may be provided on the vector contain the gene of interest while the other may be provided in the host cell genome.

[0049] Depending on the location of the expression signals (*e.g.*, promoters), suppression of the stop codon(s) during expression allows production of a fusion peptide having the tag sequence at the N- and/or C-terminus of the expressed protein. By not suppressing the stop codon(s), expression of the sequence of interest without the N- and/or C-terminal tag sequence may be accomplished. Thus, the invention allows through recombination efficient construction of vectors (*e.g.*, viral vectors) containing one or more ORFs (*e.g.*, one, two, three, four, five, six, ten, or more ORFs) or other sequence of interest (*e.g.*, untranslated sequences such as RNAi, tRNAs, ribozymes, etc.)

for controlled expression of fusion proteins depending on the need. Those skilled in the art will appreciate that suppression is not 100% effective. Thus, under suppressing conditions a mixture of polypeptides is produced, the mixture comprising polypeptides that terminate at the stop codon and polypeptides that contain amino acid sequences encoded after the stop codon. For example, in the case discussed above where three coding regions are separated by two stop codons, under conditions designed to suppress both stop codons, a mixture containing various amounts of the polypeptide encoded by the first coding region plus a polypeptide encoded by the first and the second coding regions and a polypeptide containing amino acids of all three coding regions might be produced.

[0050] The present invention provides methods of making stable cell lines and cell lines made by the methods of the invention. Stable cell lines may incorporate one or more sequences of interest that may be incorporated into the genome of the cell or may be maintained extra-chromasomally. Optionally, a sequence of interest may include one or more stop codons, one or more of which may be located at or near the 3' end of a coding sequence present in the sequence of interest. A stable cell line of the invention may be contacted with one or more nucleic acid molecules comprising all or a portion of a viral genome under conditions causing suppression of one or more of the stop codons present in the sequence of interest. A nucleic acid molecule comprising all or a portion of a viral genome may also comprise one or more copies (*e.g.*, two, three, four, five, six, seven, eight, nine, ten, fifteen, twenty, twenty five, etc.) of a sequence that produces a suppressor tRNA. In the absence of the nucleic acid molecule expressing a suppressor tRNA, for example, a nucleic acid molecule comprising all or a portion of a viral genome and comprising one or more sequence encoding a suppressor tRNA, a stable cell line of the invention may express a polypeptide encoded by a sequence of interest such that the polypeptide has a native primary structure. In the presence of a suppressor expressing nucleic acid molecule, for example, a nucleic acid molecule comprising all or a portion of a viral genome and comprising one or more sequence encoding a suppressor tRNA, a stable cell line of the invention may express a fusion protein incorporating the polypeptide encoded by the sequence of interest and some additional peptide

sequence. A stable cell line of the invention may also comprise a suppressor tRNA encoding sequence in the genome of the cell, which sequence may be under the control of a promoter that is inducible (*e.g.*, inducible by a nucleic acid molecule comprising all or a portion of a viral genome or a polypeptide encoded by such a nucleic acid molecule). Thus, contacting the cell with a nucleic acid molecule comprising all or a portion of a viral genome may result in production of a suppressor tRNA and suppression of one or more stop codons present in a sequence of interest.

[0051] The sequences of interest to be incorporated in the viral vectors and/or nucleic acids molecules of the invention may comprise at least one open reading frame (ORF) (*e.g.*, one, two, three, four, five, seven, ten, twelve, or fifteen ORFs). Such sequences may also comprise functional sequences (*e.g.*, primer binding sites, transcriptional or translation sites or signals), termination sites (*e.g.*, stop codons that may be optionally suppressed), origins of replication, and the like, and often will comprise sequences that regulate gene expression including transcriptional regulatory sequences and sequences that function as internal ribosome entry sites (IRES). Often, either the sequence of interest and/or the portions of the nucleic acid comprising the viral genome adjacent to the sequence of interest comprise sequences that function as a promoter. Either or both the sequence of interest and/or nucleic acid comprising all or a part of a viral genome may also comprise transcription termination sequences, selectable markers, restriction enzyme recognition sites, and the like.

[0052] In some embodiments, nucleic acid molecules of the invention comprising all or a portion of a viral genome may comprise two copies of the same selectable marker, each copy flanked by two recombination sites. In other embodiments, these molecules may comprise two different selectable markers each flanked by two recombination sites. In some embodiments, one or more of these selectable markers may be a negative selectable marker (*e.g.*, *ccdB*, *kicB*, Herpes simplex thymidine kinase, cytosine deaminase, etc.).

[0053] In one aspect, the present invention provides a composition comprising a recombinant viral vector which encodes one or more suppressor tRNAs. Such compositions may comprise any number of additional components, for example, cells, media, buffers, proteins, lipids, and the like. In some

embodiments, the viral vector may be an adenovirus. A viral vector may encode one or more suppressor tRNAs that recognize one of the stop codons selected from TAG, TGA, or TAA. In some embodiments, the viral vector encodes a plurality of suppressor tRNAs, for example, eight suppressor tRNAs that recognize the stop codon TAG.

[0054] In some embodiments, the present invention provides compositions comprising a nucleic acid molecule comprising all or a portion of at least one viral genome and further comprising at least two recombination sites that do not substantially recombine with each other ; and a polypeptide. Any polypeptide may be included in compositions of this type, for example, the polypeptide may be a viral envelop polypeptide. A composition of this type may be in the form of a particle comprising the nucleic acid molecule and the polypeptide. All or a portion of any viral genome may be included on the nucleic acid molecule, for example, the viral genomes may be a lentiviral genome, for example an HIV genome (such as HIV-1). A polypeptide suitable for compositions of this type is vesicular stomatitis virus G-protein.

[0055] In another aspect, the present invention provides host cells comprising a first nucleic acid sequence encoding a fusion polypeptide, wherein the sequence comprises at least a first coding region, and a second coding region separated by a sequence comprising a stop codon, and a second nucleic acid sequence comprising one or more suppressor tRNAs that suppresses the stop codon. In some embodiments, at least one of the first and/or second nucleic acid sequence is present on a nucleic acid molecule comprising all or a portion of at least one viral genome (*e.g.*, an adenoviral genome). In some embodiments, the one or more suppressor tRNAs are expressed from a nucleic acid molecule comprising all or a portion of at least one viral genome (*e.g.*, an adenoviral genome). A nucleic acid molecule may encode one or more suppressor tRNAs that recognizes one of the stop codons selected from TAG, TGA, or TAA. In some embodiments, the nucleic acid molecule may encode a plurality of suppressor tRNAs. In some embodiments, the nucleic acid molecule may encode eight suppressor tRNAs that recognize the stop codon TAG and may comprise all or a portion of an adenoviral genome.

[0056] In one aspect, the present invention provides a host cell comprising a nucleic acid molecule comprising all or a portion of at least one viral genome

and further comprising at least two recombination sites that do not substantially recombine with each other. In some embodiments, at least one of the viral genomes may be a lentiviral genome (*e.g.*, an HIV genome). In some aspects, a nucleic acid molecule may be stably integrated into the genome of the host cell. In some embodiments, at least one of the viral genomes may be an RNA virus genome (*e.g.*, of the family *Togaviridae* or *Flaviviridae* such as an alphavirus, a Sindbis virus and a Kunjin virus).

[0057] In one aspect, the present invention provides a method of expressing a polypeptide. Such methods may comprise contacting a cell with a nucleic acid molecule comprising a sequence encoding the polypeptide operably linked to a promoter and a repressor sequence, wherein the nucleic acid molecule comprises all or a portion of a viral genome, contacting the cell with a nucleic acid molecule encoding a protein that binds to the repressor sequence; and incubating the cell under conditions sufficient to express the polypeptide. In embodiments of this type, the viral genome may be a lentiviral genome (*e.g.*, an HIV). In some aspects, the repressor sequence may be the tetracycline operator sequence and the protein may be the tetracycline repressor protein and conditions sufficient to express the polypeptide comprise incubating the cell in the presence of a compound that reduces binding of the protein to the repressor sequence (*e.g.*, tetracycline).

[0058] In another aspect, the present invention provides a method of expressing a polypeptide, comprising contacting a cell with a nucleic acid molecule comprising a sequence encoding the polypeptide operably linked to a promoter and a repressor sequence, wherein the nucleic acid molecule comprises all or a portion of a viral genome and wherein the cell express a protein that binds to the repressor sequence; and incubating the cell under conditions sufficient to express the polypeptide. In embodiments of this type, the viral genome may be a lentiviral genome (*e.g.*, an HIV). In some aspects, the repressor sequence may be the tetracycline operator sequence and the protein may be the tetracycline repressor protein and conditions sufficient to express the polypeptide comprise incubating the cell in the presence of a compound that reduces binding of the protein to the repressor sequence (*e.g.*, tetracycline).

[0059] The present invention also relates to kits for carrying out methods of the invention, and particularly for use in creating recombinant viral vectors and/or nucleic acids molecules of the invention. Kits of the invention may also comprise further components for further manipulating nucleic acids and/or viral vectors produced by methods of the invention. Kits of the invention may comprise one or more nucleic acid molecules comprising all or a portion of a viral genome. Such kits may optionally comprise one or more additional components selected from the group consisting of one or more host cells (*e.g.*, two, three, four, five etc.), one or more reagents for introducing (*e.g.*, by transfection or transformation) molecules or compounds into one or more host cells, one or more nucleotides, one or more polymerases and/or reverse transcriptases (*e.g.*, two, three, four, five, etc.), one or more suitable buffers (*e.g.*, two, three, four, five, etc.), one or more primers (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.), one or more populations of molecules for creating combinatorial libraries (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.) and one or more combinatorial libraries (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.). Kits of the invention may also contain directions or protocols for carrying out one or more methods of the invention.

[0060] In another aspect the invention provides kits for joining, deleting, or replacing nucleic acid segments in the viral vectors and/or nucleic acids molecules of the invention, these kits comprising at least one component selected from the group consisting of (1) one or more recombination proteins; (2) one or more compositions comprising one or more recombination proteins; (3) at least one nucleic acid molecule comprising one or more recombination sites (preferably a vector having at least two different recombination specificities); (4) one or more nucleic acid molecules comprising all or a portion of a viral genome and one or more recombination sites; (5) one or more enzymes having ligase activity; (6) one or more enzymes having polymerase activity; (7) one or more enzymes having reverse transcriptase activity; (9) one or more enzymes having restriction endonuclease activity; (10) one or more primers; (11) one or more nucleic acid libraries; (12) one or more reagents for introducing macromolecules into cells; (13) one or more buffers; (14) one or more detergents or solutions containing detergents; (15)

one or more nucleotides; (16) one or more terminating agents; (17) one or more transfection reagents; (18) one or more host cells; (19) one or more topoisomerases; (20) one or more nucleic acid molecules to which at least one topoisomerases is bound; (21) one or more nucleic acid molecules comprising at least one topoisomerases recognition sequence; and (22) instructions for using kit components.

[0061] Further, kits of the invention may contain one or more recombination proteins. Any recombination protein known to those skilled in the art may be provided in the kits of the invention. Examples of suitable recombination proteins include, but are not limited to, Cre, Int, IHF, Xis, Flp, Fis, Hin, Gin, Cin, Tn3 resolvase, Φ C31, TndX, XerC, and XerD.

[0062] In addition, kits of the invention may contain one or more nucleic acids having more than one recombination site (*e.g.*, one or more recombination sites with different recombination specificities such as att sites with different seven base pair overlap regions). In specific embodiments, kits of the invention contain compositions comprising one or more recombination proteins capable of catalyzing recombination between recombination sites, *e.g.*, between att sites. In related embodiments, these compositions comprise one or more recombination proteins capable of catalyzing attB x attP (BP) reactions, attL x attR (LR) reactions, or both BP and LR reactions.

[0063] The invention also relates to compositions for carrying out methods of the invention and to compositions created while carrying out methods of the invention. In particular, the invention includes recombinant viral vectors prepared by methods of the invention, methods for preparing host cells that contain these viral vectors, host cells prepared by these methods, and methods employing these host cells for producing products (*e.g.*, RNA, protein, etc.) encoded by these viral vectors, and products encoded by these viral vectors (*e.g.*, RNA, protein, etc.).

[0064] Compositions, methods and kits of the invention may be prepared and carried out using a phage-lambda site-specific recombination system, such as with the GATEWAY™ Recombinational Cloning System available from Invitrogen Corporation, Carlsbad, CA. The GATEWAY™ Technology Instruction Manual (catalog number 12539-011, version C, Invitrogen

Corporation, Carlsbad, CA) describes in more detail this system and is incorporated herein by reference in its entirety.

[0065] Other embodiments of the invention will be apparent to one or ordinary skill in the art in light of what is known in the art, in light of the following drawings and description of the invention, and in light of the claims.

BRIEF DESCRIPTION OF THE FIGURES

[0066] Fig. 1 is a schematic representation of the basic recombinational cloning reaction.

[0067] Fig. 2 is a schematic representation of the use of the present invention to clone two nucleic acid segments by performing an LR recombination reaction.

[0068] Figs. 3A to 3D illustrate various embodiments of compositions and methods of the invention for generating a covalently linked double-stranded recombinant nucleic acid molecule. Topoisomerase is shown as a solid circle, and is either attached to a terminus of a substrate nucleic acid molecule or is released following a linking reaction. As illustrated, the substrate nucleic acid molecules have 5' overhangs, although they similarly can have 3' overhangs or can be blunt ended. In addition, while the illustrated nucleic acid molecules are shown having the topoisomerases bound thereto (topoisomerase-charged), one or more of the termini shown as having a topoisomerase bound thereto also can be represented as having a topoisomerase recognition site, in which case the joining reaction would further require addition of one or more site specific topoisomerases, as appropriate.

[0069] Fig. 3A shows a first nucleic acid molecule having a topoisomerase linked to each of the 5' terminus and 3' terminus of one end, and further shows linkage of the first nucleic acid molecule to a second nucleic acid molecule.

[0070] Fig. 3B shows a first nucleic acid molecule having a topoisomerase bound to the 3' terminus of one end, and a second nucleic acid molecule having a topoisomerase bound to the 3' terminus of one end, and further shows a covalently linked double-stranded recombinant nucleic acid molecule

generated due to contacting the ends containing the topoisomerase-charged substrate nucleic acid molecules.

[0071] Fig. 3C shows a first nucleic acid molecule having a topoisomerase bound to the 5' terminus of one end, and a second nucleic acid molecule having a topoisomerase bound to the 5' terminus of one end, and further shows a covalently linked double-stranded recombinant nucleic acid molecule generated due to contacting the ends containing the topoisomerase-charged substrate nucleic acid molecules.

[0072] Fig. 3D shows a nucleic acid molecule having a topoisomerase linked to each of the 5' terminus and 3' terminus of both ends, and further shows linkage of the topoisomerase-charged nucleic acid molecule to two nucleic acid molecules, one at each end. The topoisomerases at each of the 5' termini and/or at each of the 3' termini can be the same or different.

[0073] Fig. 4 is a schematic representation of one embodiment of the invention.

[0074] Figs. 5A-5F are schematic representation of exemplary vectors of the invention. Fig. 5A depicts a vector that contains two different DNA inserts, the transcription of which is driven in different directions by promoters (*e.g.*, polyhedrin, p10, T7, CMV, MMTV, metalothionine, RSV, SV40, hGH promoters). Depending on the type of transcripts which are to be produced, either of DNA-A and/or DNA-B may be in an orientation which results in the production of either sense or anti-sense RNA.

[0075] Fig. 5B is a schematic representation of an exemplary vector of the invention which contains one DNA insert, the transcription of which may proceed in either direction (or both directions) driven by two promoters which may be the same or different. Thus, RNA produced by transcription driven by one promoter will be sense RNA and RNA produced by transcription driven by the other promoter will be anti-sense RNA. RNA can be produced from both promoters, for example, to make small interfering RNA (siRNA).

[0076] Fig. 5C is a schematic representation of an exemplary vector of the invention which contains two different DNA inserts having the same nucleotide sequence (*i.e.*, DNA-A), the transcription of which are driven in different directions by two separate promoters, which may be the same or different. In this example, RNA produced by transcription driven by one

promoter will be sense RNA and RNA produced by transcription driven by the other promoter will be anti-sense RNA.

[0077] Fig. 5D is a schematic representation of an exemplary vector of the invention that contains two DNA inserts having the same nucleotide sequence (*i.e.*, DNA-A) in opposite orientations, the transcription of which is driven by one promoter (*e.g.*, CMV promoter). A transcription termination signal is not present between the two copies of DNA-A and the DNA-A inserts. Transcription of one segment produces a sense RNA and of the other produces an anti-sense RNA. The RNA produced from this vector will undergo intramolecular hybridization and, thus, will form a double-stranded molecule with a hairpin turn.

[0078] Figs. 5E and 5F are schematic representations of two exemplary vectors of the invention, each of which contains a DNA insert having the same nucleotide sequence (*i.e.*, DNA-A). Transcription of these inserts results in the production of sense and anti-sense RNA which may then hybridize to form double stranded RNA molecules.

[0079] Fig. 6 is a plasmid map of pAd/CMV/V5-DEST.

[0080] Fig. 7 is a plasmid map of pAd-GW-TO/tRNA.

[0081] Fig. 8 is a plasmid map of pAdenoTAG tRNA.

[0082] Fig. 9 is a plasmid map of pAd/PL-DEST.

[0083] Fig. 10 is a plasmid map of pAd/CMV/V5-GW/lacZ.

[0084] Fig. 11 shows the recombination region of pAd/CMV/V5-DEST.

[0085] Fig. 12 shows the recombination region of pAd/PL-DEST.

[0086] Fig. 13 shows a schematic representation of producing an exemplary adenoviral vector produced as described in Example 4.

[0087] Figs. 14A-C show the cytopathic effect (CPE) in 293A cells transfected with Pac I-digested pAd/CMV/V5-GW/lacZ plasmid as described in Example 4. Fig. 14A shows 293A cells at days 4-6 post-transfection. At this early stage, cells producing adenovirus first appear as patches of rounding, dying cells. Fig. 14B shows 293A cells at day 6-8 post-transfection. As the infection proceeds, cells containing viral particles lyse and infect neighboring cells. A plaque begins to form. Fig. 14C shows cells at day 8-10 post-transfection. At this late stage, infected neighboring cells lyse, forming a plaque that is clearly visible.

- [0088] Fig. 15 is a plasmid map of pIB/V5-His-DEST.
- [0089] Fig. 16 provides the nucleotide sequence of the *OpIE2* promoter.
- [0090] Fig. 17 shows the recombination region of pIB/V5-His-DEST.
- [0091] Fig. 18 is a plasmid map of pIB/V5-His-GW/lacZ.
- [0092] Fig. 19A shows a schematic representation of the BaculoDirect™ V5-His Dest cassette. Fig. 19B shows a schematic representation of the BaculoDirect™ Mel/V5-His Dest cassette.
- [0093] Fig. 20 shows a schematic representation of the genome of a baculovirus of the invention and an entry clone to introduce a gene of interest into the baculoviral genome.
- [0094] Fig. 21 shows a schematic representation of the topoisomerase mediate insertion of the gp64 promoter into pIB/V5-His.
- [0095] Fig. 22 is a plasmid map of pIB/V5-His/gp64/DEST.
- [0096] Fig. 23 is a bar graph showing the results of a transient transfection assay.
- [0097] Fig. 24 is a Western blot showing protein expression levels of stably transfected cells and transiently transfected cells.
- [0098] Figs. 25A and 25B are Western blots showing protein expression levels of stably transfected cells.
- [0099] Fig. 26 is a bar graph showing the results of a lacZ transfection assay.
- [0100] Fig. 27A shows a schematic representation of the construction of BaculoDirect™ vector. Fig. 27B shows a schematic representation of an LR reaction between the BaculoDirect™ vector and an entry clone containing a gene of interest.
- [0101] Fig. 28 shows a schematic representation of a high throughput cloning protocol using the baculoviruses of the present invention.
- [0102] Fig. 29 shows the results of a comparison of the use of circular virus DNA and linear virus DNA in the initial LR clonase reaction.
- [0103] Fig. 30 shows the results obtained in the presence of ganciclovir selection.
- [0104] Fig. 31 shows the results of a Western blot of various polypeptides expressed using BaculoDirect™.

- [0105] Fig. 32 shows a comparison of the titers of recombinant baculoviruses obtained using various techniques. Virus titer was obtained using the TCID₅₀ technique (upper panel) and by plaque assay (lower panel).
- [0106] Fig. 33 shows a comparison of the cumulative time required to prepare a viral stock using Bac to Bac™ and BaculoDirect™.
- [0107] Fig. 34 shows a schematic representation of plasmid pVL1393 GST p10 stop.
- [0108] Fig. 35 shows a schematic representation of a method of making a nucleic acid molecule comprising all or a portion of a lentiviral genome.
- [0109] Fig. 36 shows a schematic representation of plasmids for use in the present invention. Fig. 36A shows a schematic representation pLenti6/V5-DEST. Fig. 36B shows a schematic representation of pLenti6/V5-D-TOPO®. Figure 36C shows a plasmid map of pLenti4/V5-DEST. Figure 36D shows a plasmid map of pLenti6/UbC/V5-DEST.
- [0110] Fig. 37 shows a schematic representation of plasmids for use in the present invention. Fig. 37A shows a schematic representation pLP1. Fig. 37B shows a schematic representation of pLP2. Fig. 37C shows a schematic representation of pLP/VSVG.
- [0111] Fig. 38 shows the results of an experiment in which two LR reactions were performed with either pLenti6/V5-DEST alone or pLenti6/V5-DEST plus pENTR/CAT and 3 µl of each was transformed into TOP10 cells. 100 µl of the transformations were plated on regular LB-amp plates (no Bsd) or LB-amp containing 50 µg/ml blasticidin. Fig. 38A is photograph shown the observed colony morphologies. Figure 38B shows the results in tabular form.
- [0112] Figs. 39A and 39B show the results of a Western blot with anti-lacZ antibody (Fig. 39A) and anti-V5-antibody (Fig. 39B).
- [0113] Fig. 40 shows in tabular form the titers of lentiviral stocks prepared with inserts of varying size.
- [0114] Figs. 41A, 41B, and 41C show the expression of marker genes using the lentiviral expression system. Fig. 41A shows the expression of lacZ using the GATEWAY™ adapted lentiviral system. Figs. 41B and 41C show the expression of GFP using the topoisomerase adapted lentiviral system.
- [0115] Figs. 42A and 42B show Western blots of the expression of various genes using the lentiviral expression system described herein. Fig. 42A shows

the expression of lacZ, CAT and GFP. Fig. 42B shows the expression of PKC and GFP.

[0116] Figs. 43A and 43B show the results of varying the multiplicity of infection on the observed expression level of lacZ using the lentiviral expression system of the invention. Fig. 43A shows photographs of cells stained to detect β -galactosidase activity. Fig. 43B is a graph of β -galactosidase activity as a function of MOI.

[0117] Figs. 44A and 44B show the results of transduction of various cell types with lentiviral vectors prepared according to the methods of the invention. Fig. 44A is a bar graph of β -galactosidase activity observed in various actively growing or G1/S arrested cell types. Fig. 44B provides photographs of contacted-inhibited primary foreskin cells transduced with lentiviral vectors and stained to detect lacZ activity.

[0118] Figs. 45A and 45B show long term expression of genes from cells transduced with the nucleic acid molecules of the invention. Fig. 45A shows photographs of transduced cells stained for β -galactosidase activity after 10 days. Fig. 45B shows photographs of transduced cells stained for β -galactosidase activity after 6 weeks.

[0119] Fig. 46A shows the recombination region of pLenti6/V5-DEST. Figure 46B shows the recombination region of the expression clone resulting from pLenti6/UbC/V5-DEST x entry clone. Figure 46C shows the complete sequence of the UbC promoter.

[0120] Fig. 47 is a schematic representation of directional topoisomerase cloning according to the invention.

[0121] Fig. 48 shows the cloning region of pLenti6/V5-D-TOPO®.

[0122] Fig. 49 shows a plasmid map of pCMVSPORT6TA_g.neo.

[0123] Fig. 50 shows a schematic representation of the Tag-On-Demand™ method described in Example 14. A coding sequence of interest (GOI) is cloned with a TAG stop codon into an expression vector such that it is operably linked to a promoter (as an example, the CMV promoter is indicated in the figure). If its native stop codon is not TAG, it must be changed to TAG to be compatible with this particular method although by changing the anticodon on the suppressor tRNA molecule any stop codon can be used. Downstream of, and in frame with, the GOI is an epitope tag to be fused to the

C-terminus of the protein of interest (*e.g.*, V5, GFP, etc.). Under normal expression conditions (*i.e.*, in the absence of tRNA suppressor (-tRNA^{TAG}), native protein is expressed. In the presence of the tRNA suppressor (+tRNA), the TAG stop codon is translated as a serine in this example, and translation continues along to produce a tagged protein. The expression vector contains at least one non-TAG stop codon (*e.g.*, TAA or TGA) downstream of the C-terminal epitope tag to terminate translation of the fusion protein.

[0124] Figs. 51A-B shows western blots from plasmid tRNA suppression using the V5 epitope and GFP Tag-On-Demand™ method described in Example 14. Fig. 51A shows the western blots of CHO cells that were co-transfected with one of three reporters: pcDNA3.2/V5-GW/CAT^{TAA}, -GW/CAT^{TAG} or -GW/CAT^{TGA} in the presence or absence of its cognate tRNA suppressor: pUC12-tRNA^{TAA}, pUC12-tRNA^{TAG} or pUC12-tRNA^{TGA}, as indicated. Forty-eight hours post transfection, 20 µg of cell lysate was analyzed by either anti-V5 or anti-CAT western blotting as indicated. A control transfection of pcDNA3.1/CAT was also included in each experiment (CAT lane). Fig. 51B is the western blot of 293FT cells that were co-transfected with one of three reporters: pcDNA6.2/GFP-GW/CAT^{TAA}, -GW/CAT^{TAG} or -GW/CAT^{TGA} and one of the tRNA suppressors: pUC12-tRNA^{TAA}, pUC12-tRNA^{TAG} or pUC12-tRNA^{TGA}, as indicated. Forty-eight hours post transfection, 20 µg of cell lysate was analyzed by anti-CAT western blotting as indicated. A control transfection of pcDNA3.1/CAT was also included in each experiment (CAT lane).

[0125] Fig. 52 shows the stop codon specificity of tRNA suppression using plasmid tRNA suppression. CHO cells were co-transfected with pcDNA3.1/lacZ-stop^{TAG}-GFP and one of each of the three tRNA suppressors: pUC12-tRNA^{TAA}, pUC12-tRNA^{TAG} and pUC12-tRNA^{TGA}. Forty-eight hours post-transfection, brightfield (upper panes) and fluorescent (lower panels) photographs were taken.

[0126] Fig. 53 shows the expression of the gene of interest after adenovirus delivery of the monomer vs. octamer tRNA^{TAG} construct. COS-7 cells were transduced with crude lysates of Adeno-tRNA^{TAG} (monomer) or Adeno-tRNA^{TAG} (octamer) at an MOI of 50 for 6 hours, followed by an overnight transfection with pcDNA3.1/lacZ-stop^{TAG}-GFP. 72 hours post-transduction,

fluorescent photographs (upper panels) and anti-lacZ western blotting (lower panel) were performed. Lane 1: mock, Lane 2: co-transfection of pUC12-tRNA^{TAG} and reporter vector (positive control), Lane 3: Adeno-tRNA^{TAG} (monomer), Lane 4: Adeno-tRNA8^{TAG} (octamer).

[0127] Fig. 54 shows the expression of the indicated pENTR-ORF clone. Three pENTR-ORF clones were taken from the Invitrogen Corporation, Carlsbad, CA human ORF collection and LxR crossed into either pcDNA6.2/GFP-DEST or pcDNA6.2/V5-DEST to create expression vectors. COS-7 cells were transduced with Ad-tRNA8^{TAG} (MOI 50) followed by transfection with the ORF expression vectors. Twenty-four hours post transfection, fluorescent photographs were taken (upper panels). V5-western blotting was performed on RIPA lysates following co-transfection of COS-7 cells with the ORF expression clone and the pUC12-tRNA^{TAG} (lower panel). ORF6 expresses a protein similar to CGI-130, ORF7 expresses a splicing factor and ORF12 expresses a truncated c-myc p64 protein. "lacZ" refers to pcDNA3.1/lacZ-stop^{TAG}-V5 and "GFP-V5" refers to constitutive GFP expression from pcDNA5/GFP.

[0128] Figs. 55A and 55B shows western blots from cells transduced with adenovirus-tRNA^{TAG} for the suppression of either transient or stable target genes. Fig. 55A shows a western blot of the tRNA suppression of a stably-expressed target gene. FlpIn-CHO cells stably expressing a single copy of pcDNA6/FRT/lacZ-stop^{TAG}-GFP were transduced with Adeno-tRNA8^{TAG} at various MOIs. 48 hours post-transduction, cell lysates were analyzed by anti-lacZ western blotting and percent suppression was determined by densitometry. The additional band present in the "stable GOI" western blot (indicated by *) is the endogenous lacZeo fusion protein present in the Flp-In CHO cell line. Fig. 55B shows a western blot of the tRNA suppression of a transiently-expressed target gene. COS-7 cells were transiently transfected with the plasmid pcDNA3.1/lacZ-stop^{TAG}-GFP following transduction with CsCl purified Adeno-tRNA8^{TAG} at various MOIs. 48 hours post-transduction, cell lysates were analyzed by anti-lacZ western blotting and percent suppression was determined by densitometry.

[0129] Fig. 56 shows the use of the Tag-On-Demand™ method in five mammalian cell lines. BHK-21, CHO-S, COS-7, HeLa and HT1080 cells

were transduced with CsCl purified Adeno-tRNA⁸^{TAG} at an MOI of 50 followed by a transfection with pcDNA3.1/lacZ-stop^{TAG}-GFP. Brightfield (upper panels) and fluorescent (lower panels) photographs were taken 48 hours post transduction.

- [0130] Figure 57 is a plasmid map of pcDNATM6.2/V5-DEST.
- [0131] Figure 58 is a plasmid map of pcDNATM6.2/GFP-DEST.
- [0132] Figure 59 is a plasmid map of pcDNATM6.2/V5-GW/p64^{TAG}.
- [0133] Figure 60 is a plasmid map of pcDNATM6.2/GFP-GW-p64^{TAG}.
- [0134] Figures 61A and 61B provide the sequences of the recombination regions of vectors pcDNATM6.2/V5-DEST and pcDNATM6.2/GFP-DEST, respectively.
- [0135] Figure 62 provides a schematic representation of a method of using an adenovirus of the invention to produce C-terminal fusion proteins in a transient transfection experiment.
- [0136] Figure 63 provides a schematic representation of a method of using an adenovirus of the invention to produce C-terminal fusion proteins in a stable cell line containing an expression construct.
- [0137] Figure 64 shows fluorescent micrographs of GFP-fusion proteins made using the present invention.
- [0138] Figure 65 shows a schematic of the use of a fluorogenic substrate to assay β -lactamase activity according to one aspect of the invention.
- [0139] Figure 66 shows a comparison of sequential (left column) versus simultaneous (right column) transduction/transfection.
- [0140] Figure 67 shows Western blots showing the effects of various lipid/DNA ratios and MOI in a simultaneous transduction/transfection method (upper panels) and a sequential transduction/transfection method (lower panels).
- [0141] Figure 68 is a Western blot showing the results of an experiment in which COS-7 cells were transduced with an adenovirus expressing suppressor tRNA molecules at various MOIs and simultaneously transfected with the pcDNATM6.2/GFP-GW/p64^{TAG} plasmid.
- [0142] Figure 69 is a vector map of pLenti6/TR, a nucleic acid molecule of the invention that can be used to generate blasticidin resistant mammalian cells that stably express the tetracycline repressor, TetR.

- [0143] Figure 70 is a vector map of pLenti4/TO/V5-DEST, a nucleic acid molecule of the invention.
- [0144] Figure 71 is a vector map of pLenti6/V5.
- [0145] Figure 72 is a vector map of pLenti3/V5-TREx.
- [0146] Figure 73 shows a schematic representation of a method of attaching a topoisomerase to a nucleic acid molecule of the invention.

DETAILED DESCRIPTION OF THE INVENTION

Definitions

- [0147] In the description that follows, a number of terms used in recombinant nucleic acid technology are utilized extensively. In order to provide a clear and more consistent understanding of the specification and claims, including the scope to be given such terms, the following definitions are provided.
- [0148] **Gene:** As used herein, the term "gene" refers to a nucleic acid that contains information necessary for expression of a polypeptide, protein, or untranslated RNA (*e.g.*, rRNA, tRNA, anti-sense RNA). When the gene encodes a protein, it includes the promoter and the structural gene open reading frame sequence (ORF), as well as other sequences involved in expression of the protein. When the gene encodes an untranslated RNA, it includes the promoter and the nucleic acid that encodes the untranslated RNA.
- [0149] **Structural Gene:** As used herein, the phrase "structural gene" refers to a nucleic acid that is transcribed into messenger RNA that is then translated into a sequence of amino acids characteristic of a specific polypeptide.
- [0150] **Host:** As used herein, the term "host" refers to any prokaryotic or eukaryotic (*e.g.*, mammalian, insect, yeast, plant, avian, animal, etc.) organism that is a recipient of a replicable expression vector, cloning vector or any nucleic acid molecule. The nucleic acid molecule may contain, but is not limited to, a sequence of interest, a transcriptional regulatory sequence (such as a promoter, enhancer, repressor, and the like) and/or an origin of replication. As used herein, the terms "host," "host cell," "recombinant host" and "recombinant host cell" may be used interchangeably. For examples of

such hosts, see Sambrook, *et al.*, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York.

- [0151] Transcriptional Regulatory Sequence: As used herein, the phrase "transcriptional regulatory sequence" refers to a functional stretch of nucleotides contained on a nucleic acid molecule, in any configuration or geometry, that act to regulate the transcription of (1) one or more structural genes (*e.g.*, two, three, four, five, seven, ten, etc.) into messenger RNA or (2) one or more genes into untranslated RNA. Examples of transcriptional regulatory sequences include, but are not limited to, promoters, enhancers, repressors, operators (*e.g.*, the tet operator), and the like.
- [0152] Promoter: As used herein, a promoter is an example of a transcriptional regulatory sequence, and is specifically a nucleic acid generally described as the 5'-region of a gene located proximal to the start codon or nucleic acid that encodes untranslated RNA. The transcription of an adjacent nucleic acid segment is initiated at or near the promoter. A repressible promoter's rate of transcription decreases in response to a repressing agent. An inducible promoter's rate of transcription increases in response to an inducing agent. A constitutive promoter's rate of transcription is not specifically regulated, though it can vary under the influence of general metabolic conditions.
- [0153] Target Nucleic Acid Molecule: As used herein, the phrase "target nucleic acid molecule" refers to a nucleic acid segment of interest, preferably nucleic acid that is to be acted upon using the compounds and methods of the present invention. Such target nucleic acid molecules may contain one or more (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.) genes or one or more portions of genes.
- [0154] Insert Donor: As used herein, the phrase "Insert Donor" refers to one of the two parental nucleic acid molecules (*e.g.*, RNA or DNA) of the present invention that carries the an insert (see Fig. 1). The Insert Donor molecule comprises the insert flanked on both sides with recombination sites. The Insert Donor can be linear or circular. In one embodiment of the invention, the Insert Donor is a circular nucleic acid molecule, optionally supercoiled, and further comprises a cloning vector sequence outside of the recombination signals. When a population of inserts or population of nucleic acid segments

are used to make the Insert Donor, a population of Insert Donors result and may be used in accordance with the invention. An Insert Donor may be referred to as an Entry Clone.

[0155] Insert: As used herein, the term "insert" refers to a desired nucleic acid segment that is a part of a larger nucleic acid molecule. In many instances, the insert will be introduced into the larger nucleic acid molecule. For example, the nucleic acid segments labeled ccdB, DNA-A, and DNA-B in Fig. 2, are nucleic acid inserts with respect to the larger nucleic acid molecule shown therein. In most instances, the insert will be flanked by recombination sites, topoisomerase sites and/or other recognition sequences (*e.g.*, at least one recognition sequence will be located at each end). In certain embodiments, however, the insert will only contain a recognition sequence on one end.

[0156] Product: As used herein, the term "Product" refers to one the desired daughter molecules comprising the A and D sequences that is produced after the second recombination event during the recombinational cloning process (see Fig. 1). The Product contains the nucleic acid that was to be cloned or subcloned. In accordance with the invention, when a population of Insert Donors are used, the resulting population of Product molecules will contain all or a portion of the population of Inserts of the Insert Donors and preferably will contain a representative population of the original molecules of the Insert Donors.

[0157] Byproduct: As used herein, the term "Byproduct" refers to a daughter molecule (a new clone produced after the second recombination event during the recombinational cloning process) lacking the segment that is desired to be cloned or subcloned.

[0158] Cointegrate: As used herein, the term "Cointegrate" refers to at least one recombination intermediate nucleic acid molecule of the present invention that contains both parental (starting) molecules. Cointegrates may be linear or circular. RNA and polypeptides may be expressed from cointegrates using an appropriate host cell strain, for example *E. coli* DB3.1 (particularly *E. coli* LIBRARY EFFICIENCY® DB3.1™ Competent Cells), and selecting for both selection markers found on the cointegrate molecule.

[0159] Recognition Sequence: As used herein, the phrase "recognition sequence" or "recognition site" refers to a particular sequence to which a

protein, chemical compound, DNA, or RNA molecule (*e.g.*, restriction endonuclease, a modification methylase, topoisomerases, or a recombinase) recognizes and binds. In the present invention, a recognition sequence may refer to a recombination site or topoisomerases site. For example, the recognition sequence for Cre recombinase is loxP which is a 34 base pair sequence comprising two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence (see Fig. 1 of Sauer, B., *Current Opinion in Biotechnology* 5:521-527 (1994)). Other examples of recognition sequences are the attB, attP, attL, and attR sequences, which are recognized by the recombinase enzyme λ Integrase. attB is an approximately 25 base pair sequence containing two 9 base pair core-type Int binding sites and a 7 base pair overlap region. attP is an approximately 240 base pair sequence containing core-type Int binding sites and arm-type Int binding sites as well as sites for auxiliary proteins integration host factor (IHF), FIS and excisionase (Xis) (see Landy, *Current Opinion in Biotechnology* 3:699-707 (1993)). Such sites may also be engineered according to the present invention to enhance production of products in the methods of the invention. For example, when such engineered sites lack the P1 or H1 domains to make the recombination reactions irreversible (*e.g.*, attR or attP), such sites may be designated attR' or attP' to show that the domains of these sites have been modified in some way.

[0160] **Recombination Proteins:** As used herein, the phrase "recombination proteins" includes excisive or integrative proteins, enzymes, co-factors or associated proteins that are involved in recombination reactions involving one or more recombination sites (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.), which may be wild-type proteins (see Landy, *Current Opinion in Biotechnology* 3:699-707 (1993)), or mutants, derivatives (*e.g.*, fusion proteins containing the recombination protein sequences or fragments thereof), fragments, and variants thereof. Examples of recombination proteins include Cre, Int, IHF, Xis, Flp, Fis, Hin, Gin, Φ C31, Cin, Tn3 resolvase, TndX, XerC, XerD, TnpX, Hjc, SpCCE1, and ParA.

[0161] **Recombinases:** As used herein, the term "recombinases" is used to refer to the protein that catalyzes strand cleavage and re-ligation in a recombination reaction. Site-specific recombinases are proteins that are

present in many organisms (*e.g.*, viruses and bacteria) and have been characterized as having both endonuclease and ligase properties. These recombinases (along with associated proteins in some cases) recognize specific sequences of bases in a nucleic acid molecule and exchange the nucleic acid segments flanking those sequences. The recombinases and associated proteins are collectively referred to as "recombination proteins" (see, *e.g.*, Landy, A., *Current Opinion in Biotechnology* 3:699-707 (1993)).

[0162] Numerous recombination systems from various organisms have been described. See, *e.g.*, Hoess, *et al.*, *Nucleic Acids Research* 14(6):2287 (1986); Abremski, *et al.*, *J. Biol. Chem.* 261(1):391 (1986); Campbell, *J. Bacteriol.* 174(23):7495 (1992); Qian, *et al.*, *J. Biol. Chem.* 267(11):7794 (1992); Araki, *et al.*, *J. Mol. Biol.* 225(1):25 (1992); Maeser and Kahnmann, *Mol. Gen. Genet.* 230:170-176 (1991); Esposito, *et al.*, *Nucl. Acids Res.* 25(18):3605 (1997). Many of these belong to the integrase family of recombinases (Argos, *et al.*, *EMBO J.* 5:433-440 (1986); Voznyanov, *et al.*, *Nucl. Acids Res.* 27:930 (1999)). Perhaps the best studied of these are the Integrase/att system from bacteriophage λ (Landy, A. *Current Opinions in Genetics and Devel.* 3:699-707 (1993)), the Cre/loxP system from bacteriophage P1 (Hoess and Abremski (1990) In *Nucleic Acids and Molecular Biology*, vol. 4. Eds.: Eckstein and Lilley, Berlin-Heidelberg: Springer-Verlag; pp. 90-109), and the FLP/FRT system from the *Saccharomyces cerevisiae* 2 μ circle plasmid (Broach, *et al.*, *Cell* 29:227-234 (1982)).

[0163] **Recombination Site:** A used herein, the phrase "recombination site" refers to a recognition sequence on a nucleic acid molecule that participates in an integration/recombination reaction by recombination proteins. Recombination sites are discrete sections or segments of nucleic acid on the participating nucleic acid molecules that are recognized and bound by a site-specific recombination protein during the initial stages of integration or recombination. For example, the recombination site for Cre recombinase is loxP, which is a 34 base pair sequence comprised of two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence (see Fig. 1 of Sauer, B., *Curr. Opin. Biotech.* 5:521-527 (1994)). Other examples of recombination sites include the attB, attP, attL, and attR sequences described in United States provisional patent applications

60/136,744, filed May 28, 1999, and 60/188,000, filed March 9, 2000, and in co-pending United States patent applications 09/517,466 and 09/732,91—all of which are specifically incorporated herein by reference—and mutants, fragments, variants and derivatives thereof, which are recognized by the recombination protein λ Int and by the auxiliary proteins integration host factor (IHF), FIS and excisionase (Xis) (see Landy, Curr. Opin. Biotech. 3:699-707 (1993)).

[0164] Recombination sites may be added to molecules by any number of known methods. For example, recombination sites can be added to nucleic acid molecules by blunt end ligation, PCR performed with fully or partially random primers, or inserting the nucleic acid molecules into a vector using a restriction site flanked by recombination sites.

[0165] Topoisomerase recognition site. As used herein, the term "topoisomerase recognition site" or "topoisomerase site" means a defined nucleotide sequence that is recognized and bound by a site specific topoisomerase. For example, the nucleotide sequence 5'-(C/T)CCTT-3' is a topoisomerase recognition site that is bound specifically by most poxvirus topoisomerases, including vaccinia virus DNA topoisomerase I, which then can cleave the strand after the 3'-most thymidine of the recognition site to produce a nucleotide sequence comprising 5'-(C/T)CCTT-PO₄-TOPO, *i.e.*, a complex of the topoisomerase covalently bound to the 3' phosphate through a tyrosine residue in the topoisomerase (see Shuman, *J. Biol. Chem.* 266:11372-11379, 1991; Sekiguchi and Shuman, *Nucl. Acids Res.* 22:5360-5365, 1994; each of which is incorporated herein by reference; see, also, U.S. Pat. No. 5,766,891; PCT/US95/16099; and PCT/US98/12372 also incorporated herein by reference). In comparison, the nucleotide sequence 5'-GCAACTT-3' is the topoisomerase recognition site for type IA *E. coli* topoisomerase III.

[0166] Recombinational Cloning: As used herein, the phrase "recombinational cloning" refers to a method, such as that described in U.S. Patent Nos. 5,888,732; 6,143,557; 6,171,861; 6,270,969; and 6,277,608 (the contents of which are fully incorporated herein by reference), whereby segments of nucleic acid molecules or populations of such molecules are exchanged, inserted, replaced, substituted or modified, *in vitro* or *in vivo*. Preferably, such cloning method is an *in vitro* method.

[0167] Cloning systems that utilize recombination at defined recombination sites have been previously described in U.S. patent no. 5,888,732, U.S. patent no. 6,143,557, U.S. patent no. 6,171,861, U.S. patent no. 6,270,969, and U.S. patent no. 6,277,608, and in pending United States application no. 09/517,466 filed March 2, 2000, and in published United States application no. 2002 0007051-A1, all assigned to the Invitrogen Corporation, Carlsbad, CA, the disclosures of which are specifically incorporated herein in their entirety. In brief, the GATEWAY™ Cloning System described in these patents and applications utilizes vectors that contain at least one recombination site to clone desired nucleic acid molecules *in vivo* or *in vitro*. In some embodiments, the system utilizes vectors that contain at least two different site-specific recombination sites that may be based on the bacteriophage lambda system (*e.g.*, att1 and att2) that are mutated from the wild-type (att0) sites. Each mutated site has a unique specificity for its cognate partner att site (*i.e.*, its binding partner recombination site) of the same type (for example attB1 with attP1, or attL1 with attR1) and will not cross-react with recombination sites of the other mutant type or with the wild-type att0 site. Different site specificities allow directional cloning or linkage of desired molecules thus providing desired orientation of the cloned molecules. Nucleic acid fragments flanked by recombination sites are cloned and subcloned using the GATEWAY™ system by replacing a selectable marker (for example, ccdB) flanked by att sites on the recipient plasmid molecule, sometimes termed the Destination Vector. Desired clones are then selected by transformation of a ccdB sensitive host strain and positive selection for a marker on the recipient molecule. Similar strategies for negative selection (*e.g.*, use of toxic genes) can be used in other organisms such as thymidine kinase (TK) in mammals and insects.

[0168] Mutating specific residues in the core region of the att site can generate a large number of different att sites. As with the att1 and att2 sites utilized in GATEWAY™, each additional mutation potentially creates a novel att site with unique specificity that will recombine only with its cognate partner att site bearing the same mutation and will not cross-react with any other mutant or wild-type att site. Novel mutated att sites (*e.g.*, attB 1-10, attP 1-10, attR 1-10 and attL 1-10) are described in previous patent application serial number

09/517,466, filed March 2, 2000, which is specifically incorporated herein by reference. Other recombination sites having unique specificity (*i.e.*, a first site will recombine with its corresponding site and will not recombine or not substantially recombine with a second site having a different specificity) may be used to practice the present invention. Examples of suitable recombination sites include, but are not limited to, loxP sites; loxP site mutants, variants or derivatives such as loxP511 (see U.S. Patent No. 5,851,808); frt sites; frt site mutants, variants or derivatives; dif sites; dif site mutants, variants or derivatives; psi sites; psi site mutants, variants or derivatives; cer sites; and cer site mutants, variants or derivatives.

[0169] Repression Cassette: As used herein, the phrase "repression cassette" refers to a nucleic acid segment that contains a repressor or a selectable marker present in the subcloning vector.

[0170] Selectable Marker: As used herein, the phrase "selectable marker" refers to a nucleic acid segment that allows one to select for or against a molecule (*e.g.*, a replicon) or a cell that contains it and/or permits identification of a cell or organism that contains or does not contain the nucleic acid segment. Frequently, selection and/or identification occur under particular conditions and do not occur under other conditions.

[0171] Markers can encode an activity, such as, but not limited to, production of RNA, peptide, or protein, or can provide a binding site for RNA, peptides, proteins, inorganic and organic compounds or compositions and the like. Examples of selectable markers include but are not limited to: (1) nucleic acid segments that encode products that provide resistance against otherwise toxic compounds (*e.g.*, antibiotics); (2) nucleic acid segments that encode products that are otherwise lacking in the recipient cell (*e.g.*, tRNA genes, auxotrophic markers); (3) nucleic acid segments that encode products that suppress the activity of a gene product; (4) nucleic acid segments that encode products that can be readily identified (*e.g.*, phenotypic markers such as β -lactamase, β -galactosidase, green fluorescent protein (GFP), yellow fluorescent protein (YFP), red fluorescent protein (RFP), cyan fluorescent protein (CFP), and cell surface proteins); (5) nucleic acid segments that bind products that are otherwise detrimental to cell survival and/or function; (6) nucleic acid segments that otherwise inhibit the activity of any of the nucleic acid segments

described in Nos. 1-5 above (*e.g.*, antisense oligonucleotides); (7) nucleic acid segments that bind products that modify a substrate (*e.g.*, restriction endonucleases); (8) nucleic acid segments that can be used to isolate or identify a desired molecule (*e.g.*, specific protein binding sites); (9) nucleic acid segments that encode a specific nucleotide sequence that can be otherwise non-functional (*e.g.*, for PCR amplification of subpopulations of molecules); (10) nucleic acid segments that, when absent, directly or indirectly confer resistance or sensitivity to particular compounds; and/or (11) nucleic acid segments that encode products that either are toxic (*e.g.*, Diphtheria toxin) or convert a relatively non-toxic compound to a toxic compound (*e.g.*, Herpes simplex thymidine kinase, cytosine deaminase) in recipient cells; (12) nucleic acid segments that inhibit replication, partition or heritability of nucleic acid molecules that contain them; and/or (13) nucleic acid segments that encode conditional replication functions, *e.g.*, replication in certain hosts or host cell strains or under certain environmental conditions (*e.g.*, temperature, nutritional conditions, etc.).

[0172] Selection and/or identification may be accomplished using techniques well known in the art. For example, a selectable marker may confer resistance to an otherwise toxic compound and selection may be accomplished by contacting a population of host cells with the toxic compound under conditions in which only those host cells containing the selectable marker are viable. In another example, a selectable marker may confer sensitivity to an otherwise benign compound and selection may be accomplished by contacting a population of host cells with the benign compound under conditions in which only those host cells that do not contain the selectable marker are viable. A selectable marker may make it possible to identify host cells containing or not containing the marker by selection of appropriate conditions. In one aspect, a selectable marker may enable visual screening of host cells to determine the presence or absence of the marker. For example, a selectable marker may alter the color and/or fluorescence characteristics of a cell containing it. This alteration may occur in the presence of one or more compounds, for example, as a result of an interaction between a polypeptide encoded by the selectable marker and the compound (*e.g.*, an enzymatic reaction using the compound as a substrate). Such alterations in visual

characteristics can be used to physically separate the cells containing the selectable marker from those not contain it by, for example, fluorescent activated cell sorting (FACS).

[0173] Multiple selectable markers may be simultaneously used to distinguish various populations of cells. For example, a nucleic acid molecule of the invention may have multiple selectable markers, one or more of which may be removed from the nucleic acid molecule by a suitable reaction (*e.g.*, a recombination reaction). After the reaction, the nucleic acid molecules may be introduced into a host cell population and those host cells comprising nucleic acid molecules having all of the selectable markers may be distinguished from host cells comprising nucleic acid molecules in which one or more selectable markers have been removed (*e.g.*, by the recombination reaction). For example, a nucleic acid molecule of the invention may have a blasticidin resistance marker outside a pair of recombination sites and a β -lactamase encoding selectable marker inside the recombination sites. After a recombination reaction and introduction of the reaction mixture into a cell population, cells comprising any nucleic acid molecule can be selected for by contacting the cell population with blasticidin. Those cell comprising a nucleic acid molecule that has undergone a recombination reaction can be distinguished from those containing an unreacted nucleic acid molecules by contacting the cell population with a fluorogenic β -lactamase substrate as described below and observing the fluorescence of the cell population. Optionally, the desired cells can be physically separated from undesirable cells, for example, by FACS.

[0174] In a specific embodiment of the invention, a selectable marker may be a nucleic acid sequence encoding a polypeptide having an enzymatic activity (*e.g.*, β -lactamase activity). Assays for β -lactamase activity are known in the art. United States Patent nos. 5,955,604, issued to Tsien, *et al.* September 21, 1999, 5,741,657 issued to Tsien, *et al.*, April 21, 1998, 6,031,094, issued to Tsien, *et al.*, February 29, 2000, 6,291,162, issued to Tsien, *et al.*, September 18, 2001, and 6,472,205, issued to Tsien, *et al.* October 29, 2002, disclose the use of β -lactamase as a reporter gene and fluorogenic substrates for use in detecting β -lactamase activity and are specifically incorporated herein by reference. In one embodiment of the invention, a selectable marker may be a

nucleic acid sequence encoding a polypeptide having β -lactamase activity and desired host cells may be identified by assaying the host cells for β -lactamase activity.

[0175] A β -lactamase catalyzes the hydrolysis of a β -lactam ring. Those skilled in the art will appreciate that the sequences of a number of polypeptides having β -lactamase activity are known. In addition to the specific β -lactamases disclosed in the Tsien, *et al.* patents listed above, any polypeptide having β -lactamase activity is suitable for use in the present invention.

[0176] β -lactamases are classified based on amino acid and nucleotide sequence (Ambler, R. P., *Phil. Trans. R. Soc. Lond.* [Ser.B.] 289: 321-331 (1980)) into classes A-D. Class A β -lactamases possess a serine in the active site and have an approximate weight of 29 kD. This class contains the plasmid-mediated TEM β -lactamases such as the RTEM enzyme of pBR322. Class B β -lactamases have an active-site zinc bound to a cysteine residue. Class C enzymes have an active site serine and a molecular weight of approximately 39 kD, but have no amino acid homology to the class A enzymes. Class D enzymes also contain an active site serine. Representative examples of each class are provided below with the accession number at which the sequence of the enzyme may be obtained in the indicated database.

Class A β -lactamases

<i>Bacteroides fragilis</i> CS30	L13472	GenBank
<i>Bacteroides uniformis</i> WAL-7088	P30898	SWISS-PROT
PER-1, <i>P. aeruginosa</i> RNL-1	P37321	SWISS-PROT
<i>Bacteroides vulgatus</i> CLA341	P30899	SWISS-PROT
OHIO-1, <i>Enterobacter cloacae</i>	P18251	SWISS-PROT
SHV-1, <i>K. pneumoniae</i>	P23982	SWISS-PROT
LEN-1, <i>K. pneumoniae</i> LEN-1	P05192	SWISS-PROT
TEM-1, <i>E. coli</i>	P00810	SWISS-PROT
<i>Proteus mirabilis</i> GN179	P30897	SWISS-PROT
PSE-4, <i>P. aeruginosa</i> Dalglish	P16897	SWISS-PROT
<i>Rhodopseudomonas capsulatus</i> SP108	P14171	SWISS-PROT
NMC, <i>E. cloacae</i> NOR-1	P52663	SWISS-PROT
Sme-1, <i>Serratia marcescens</i> S6	P52682	SWISS-PROT
OXY-2, <i>Klebsiella oxytoca</i> D488	P23954	SWISS-PROT
<i>K. oxytoca</i> E23004/SL781/SL7811	P22391	SWISS-PROT
<i>S. typhimurium</i> CAS-5	X92507	GenBank
MEN-1, <i>E. coli</i> MEN	P28585	SWISS-PROT
<i>Serratia fonticola</i> CUV	P80545	SWISS-PROT
<i>Citrobacter diversus</i> ULA27	P22390	SWISS-PROT

<i>Proteus vulgaris</i> 5E78-1	P52664	SWISS-PROT
<i>Burkholderia cepacia</i> 249	U85041	GenBank
<i>Yersinia enterocolitica</i> serotype O:3/Y-56	Q01166	SWISS-PROT
<i>M. tuberculosis</i> H37RV	Q10670	SWISS-PROT
<i>S. clavuligerus</i> NRRL 3585	Z54190	GenBank
III, <i>Bacillus cereus</i> 569/H	P06548	SWISS-PROT
<i>B. licheniformis</i> 749/C	P00808	SWISS-PROT
I, <i>Bacillus mycoides</i> NI10R	P28018	SWISS-PROT
I, <i>B. cereus</i> 569/H/9	P00809	SWISS-PROT
I, <i>B. cereus</i> 5/B	P10424	SWISS-PROT
<i>B. subtilis</i> 168/6GM	P39824	SWISS-PROT
2, <i>Streptomyces cacaoi</i> DSM40057	P14560	SWISS-PROT
<i>Streptomyces badius</i> DSM40139	P35391	SWISS-PROT
<i>Actinomadura</i> sp. strain R39	X53650	GenBank
<i>Nocardia lactamdurans</i> LC411	Q06316	SWISS-PROT
<i>S. cacaoi</i> KCC S0352	Q03680	SWISS-PROT
ROB-1, <i>H. influenzae</i> F990/LNPB51/ serotype A1	P33949	SWISS-PROT
<i>Streptomyces fradiae</i> DSM40063	P35392	SWISS-PROT
<i>Streptomyces lavendulae</i> DSM2014	P35393	SWISS-PROT
<i>Streptomyces albus</i> G	P14559	SWISS-PROT
<i>S. lavendulae</i> KCCS0263	D12693	GenBank
<i>Streptomyces aureofaciens</i>	P10509	SWISS-PROT
<i>Streptomyces cellulosae</i> KCCS0127	Q06650	SWISS-PROT
<i>Mycobacterium fortuitum</i>	L25634	GenBank
<i>S. aureus</i> PC1/SK456/NCTC9789	P00807	SWISS-PROT
BRO-1, <i>Moraxella catarrhalis</i> ATCC 53879	Z54181 Q59514	GenBank; SWISS-PROT
Class B β -lactamase		
II, <i>B. cereus</i> 569/H	P04190	SWISS-PROT
II, <i>Bacillus</i> sp. 170	P10425	SWISS-PROT
II, <i>B. cereus</i> 5/B/6	P14488	SWISS-PROT
<i>Chryseobacterium meningosepticum</i> CCUG4310	X96858	GenBank
IMP-1, <i>S. marcescens</i> AK9373/TN9106	P52699	SWISS-PROT
<i>B. fragilis</i> TAL3636/TAL2480	P25910	SWISS-PROT
<i>Aeromonas hydrophila</i> AE036	P26918	SWISS-PROT
L1, <i>Xanthomonas maltophilia</i> IID 1275	P52700	SWISS-PROT
Class C β -lactamase		
<i>Citrobacter freundii</i> OS60/GN346	P05193	SWISS-PROT
<i>E. coli</i> K-12/MG1655	P00811	SWISS-PROT
P99, <i>E. cloacae</i> P99/Q908R/MHN1	P05364	SWISS-PROT
<i>Y. enterocolitica</i> IP97/serotype O:5B	P45460	SWISS-PROT
<i>Morganella morganii</i> SLM01	Y10283	GenBank
<i>A. sobria</i> 163a	X80277	GenBank
FOX-3, <i>K. oxytoca</i> 1731	Y11068	GenBank
<i>K. pneumoniae</i> NU2936	D13304	GenBank
<i>P. aeruginosa</i> PAO1	P24735	SWISS-PROT
<i>S. marcescens</i> SR50	P18539	SWISS-PROT

<i>Psychrobacter immobilis</i> A5	X83586	GenBank
Class D β -lactamases		
OXA-18, <i>Pseudomonas aeruginosa</i> Mus	U85514	GenBank
OXA-9, <i>Klebsiella pneumoniae</i>	P22070	SWISS-PROT
<i>Aeromonas sobria</i> AER 14	X80276	GenBank
OXA-1, <i>Escherichia coli</i> K10-35	P13661	SWISS-PROT
OXA-7, <i>E. coli</i> 7181	P35695	SWISS-PROT
OXA-11, <i>P. aeruginosa</i> ABD	Q06778	SWISS-PROT
OXA-5, <i>P. aeruginosa</i> 76072601	Q00982	SWISS-PROT
LCR-1, <i>P. aeruginosa</i> 2293E	Q00983	SWISS-PROT
OXA-2, <i>Salmonella typhimurium</i> type 1A	P05191	SWISS-PROT

[0177] For additional β -lactamases and a more detailed description of substrate specificities, consult Bush *et al.* (1995) *Antimicrob. Agents Chemother.* 39:1211-1233. Those skilled in the art will appreciate that the polypeptides having β -lactamase activity disclosed herein may be altered by for example, mutating, deleting, and/or adding one or more amino acids and may still be used in the practice of the invention so long as the polypeptide retains detectable β -lactamase activity. An example of a suitably altered polypeptide having β -lactamase activity is one from which a signal peptide sequence has been deleted and/or altered such that the polypeptide is retained in the cytosol of prokaryotic and/or eukaryotic cells. The amino acid sequence of one such polypeptide is provided in Table 30.

[0178] As described in the above-referenced United States patents, host cells to be assayed may be contacted with a fluorogenic substrate for β -lactamase activity. In the presence of β -lactamase, the substrate is cleaved and the fluorescence emission spectrum of the substrate is altered. As an example, un-cleaved substrate may fluoresce green (*i.e.*, have an emission maxima at approximately 520 nm) when excited with light having a wavelength of 405 nm and the cleaved substrate may fluoresce blue (*i.e.*, have an emission maxima at approximately 447 nm). By determining the ratio of green fluorescence intensity to blue fluorescence intensity it is possible to determine the amount of β -lactamase produced and from that, to calculate what % of the cells express β -lactamase. Kits for conducting a fluorescence-based β -lactamase assay are commercially available, for example, from PanVerra, LLC, Madison, WI, catalog number K1032.

[0179] Preferred β -lactam fluorogenic substrates for use in the present invention include those which comprise a fluorescence donor moiety and a fluorescence acceptor moiety linked to a cephalosporin backbone such that, upon hydrolysis of the β -lactam, the acceptor moiety is released from the molecule. Before the β -lactam is hydrolyzed, the donor and acceptor moiety are positioned such that efficient fluorescence resonance energy transfer (FRET) occurs. Upon excitation with light of a suitable wavelength, fluorescence from the acceptor moiety is observed. After hydrolysis of the β -lactam, the acceptor moiety is released from the molecule and the FRET is disrupted resulting in a change in the fluorescence emission spectrum. An example of a suitable fluorescence donor molecule is a coumarin or derivative thereof (*e.g.*, 6-chloro-7-hydroxycoumarin) and examples of suitable acceptor moieties include, but are not limited to, fluorescein, rhodol, or rhodamine or derivatives thereof. Examples of suitable substrates include CCF2 and the acetoxymethyl ester derivative thereof (CCF2/AM). Those skilled in the art will appreciate that CCF2/AM is membrane permeable and is converted to CCF2 inside a cell by the action of endogenous esterase enzymes. A schematic showing the result of hydrolysis of CCF2 by a β -lactamase is shown in Figure 65.

[0180] Selection Scheme: As used herein, the phrase "selection scheme" refers to any method that allows selection, enrichment, or identification of a desired nucleic acid molecules or host cells containing them (in particular Product or Product(s) from a mixture containing an Entry Clone or Vector, a Destination Vector, a Donor Vector, an Expression Clone or Vector, any intermediates (*e.g.*, a Cointegrate or a replicon), and/or Byproducts). In one aspect, selection schemes of the invention rely on one or more selectable markers. The selection schemes of one embodiment have at least two components that are either linked or unlinked during recombinational cloning. One component is a selectable marker. The other component controls the expression *in vitro* or *in vivo* of the selectable marker, or survival of the cell (or the nucleic acid molecule, *e.g.*, a replicon) harboring the plasmid carrying the selectable marker. Generally, this controlling element will be a repressor or inducer of the selectable marker, but other means for controlling expression or activity of the selectable marker can be used. Whether a repressor or

activator is used will depend on whether the marker is for a positive or negative selection, and the exact arrangement of the various nucleic acid segments, as will be readily apparent to those skilled in the art. In some preferred embodiments, the selection scheme results in selection of, or enrichment for, only one or more desired nucleic acid molecules (such as Products). As defined herein, selecting for a nucleic acid molecule includes (a) selecting or enriching for the presence of the desired nucleic acid molecule (referred to as a "positive selection scheme"), and (b) selecting or enriching against the presence of nucleic acid molecules that are not the desired nucleic acid molecule (referred to as a "negative selection scheme").

[0181] In one embodiment, the selection schemes (which can be carried out in reverse) will take one of three forms, which will be discussed in terms of Fig.

1. The first, exemplified herein with a selectable marker and a repressor therefore, selects for molecules having segment D and lacking segment C. The second selects against molecules having segment C and for molecules having segment D. Possible embodiments of the second form would have a nucleic acid segment carrying a gene toxic to cells into which the *in vitro* reaction products are to be introduced. A toxic gene can be a nucleic acid that is expressed as a toxic gene product (a toxic protein or RNA), or can be toxic in and of itself. (In the latter case, the toxic gene is understood to carry its classical definition of "heritable trait.")

[0182] Examples of such toxic gene products are well known in the art, and include, but are not limited to, restriction endonucleases (*e.g.*, DpnI, Nla3, etc.); apoptosis-related genes (*e.g.*, ASK1 or members of the bcl-2/ced-9 family); retroviral genes; including those of the human immunodeficiency virus (HIV); defensins such as NP-1; inverted repeats or paired palindromic nucleic acid sequences; bacteriophage lytic genes such as those from Φ X174 or bacteriophage T4; antibiotic sensitivity genes such as *rpsL*; antimicrobial sensitivity genes such as *pheS*; plasmid killer genes' eukaryotic transcriptional vector genes that produce a gene product toxic to bacteria, such as GATA-1; genes that kill hosts in the absence of a suppressing function, *e.g.*, *kicB*, *ccdB*, Φ X174 E (Liu, Q., *et al.*, *Curr. Biol.* 8:1300-1309 (1998)); and other genes that negatively affect replicon stability and/or replication. A toxic gene can alternatively be selectable *in vitro*, *e.g.*, a restriction site.

- [0183] Many genes coding for restriction endonucleases operably linked to inducible promoters are known, and may be used in the present invention (see, e.g., U.S. Patent Nos. 4,960,707 (DpnI and DpnII); 5,082,784 and 5,192,675 (KpnI); 5,147,800 (NgoAIII and NgoAI); 5,179,015 (FspI and HaeIII); 5,200,333 (HaeII and TaqI); 5,248,605 (HpaII); 5,312,746 (ClaI); 5,231,021 and 5,304,480 (XhoI and XhoII); 5,334,526 (AluI); 5,470,740 (NsiI); 5,534,428 (SstI/SacI); 5,202,248 (NcoI); 5,139,942 (NdeI); and 5,098,839 (PacI). (See also Wilson, G.G., *Nucl. Acids Res.* 19:2539-2566 (1991); and Lunnen, K.D., *et al.*, *Gene* 74:25-32 (1988)).
- [0184] In the second form, segment D carries a selectable marker. The toxic gene would eliminate transformants harboring the Vector Donor, Cointegrate, and Byproduct molecules, while the selectable marker can be used to select for cells containing the Product and against cells harboring only the Insert Donor.
- [0185] The third form selects for cells that have both segments A and D in cis on the same molecule, but not for cells that have both segments in trans on different molecules. This could be embodied by a selectable marker that is split into two inactive fragments, one each on segments A and D.
- [0186] The fragments are so arranged relative to the recombination sites that when the segments are brought together by the recombination event, they reconstitute a functional selectable marker. For example, the recombinational event can link a promoter with a structural nucleic acid molecule (e.g., a gene), can link two fragments of a structural nucleic acid molecule, or can link nucleic acid molecules that encode a heterodimeric gene product needed for survival, or can link portions of a replicon.
- [0187] Site-Specific Recombinase: As used herein, the phrase "site-specific recombinase" refers to a type of recombinase that typically has at least the following four activities (or combinations thereof): (1) recognition of specific nucleic acid sequences; (2) cleavage of said sequence or sequences; (3) topoisomerase activity involved in strand exchange; and (4) ligase activity to reseal the cleaved strands of nucleic acid (see Sauer, B., *Current Opinions in Biotechnology* 5:521-527 (1994)). Conservative site-specific recombination is distinguished from homologous recombination and transposition by a high degree of sequence specificity for both partners. The strand exchange mechanism involves the cleavage and rejoining of specific nucleic acid

sequences in the absence of DNA synthesis (Landy, A. (1989) *Ann. Rev. Biochem.* 58:913-949).

[0188] Suppressor tRNAs. A tRNA molecule that results in the incorporation of an amino acid in a polypeptide in a position corresponding to a stop codon in the mRNA being translated.

[0189] Homologous Recombination: As used herein, the phrase "homologous recombination" refers to the process in which nucleic acid molecules with similar nucleotide sequences associate and exchange nucleotide strands. A nucleotide sequence of a first nucleic acid molecule that is effective for engaging in homologous recombination at a predefined position of a second nucleic acid molecule will therefore have a nucleotide sequence that facilitates the exchange of nucleotide strands between the first nucleic acid molecule and a defined position of the second nucleic acid molecule. Thus, the first nucleic acid will generally have a nucleotide sequence that is sufficiently complementary to a portion of the second nucleic acid molecule to promote nucleotide base pairing.

[0190] Homologous recombination requires homologous sequences in the two recombining partner nucleic acids but does not require any specific sequences. As indicated above, site-specific recombination that occurs, for example, at recombination sites such as att sites, is not considered to be "homologous recombination," as the phrase is used herein.

[0191] Vector: As used herein, the term "vector" refers to a nucleic acid molecule (preferably DNA) that provides a useful biological or biochemical property to an insert. A vector may be a nucleic acid molecule comprising all or a portion of a viral genome. Examples include plasmids, phages, autonomously replicating sequences (ARS), centromeres, and other sequences that are able to replicate or be replicated *in vitro* or in a host cell, or to convey a desired nucleic acid segment to a desired location within a host cell. A vector can have one or more recognition sites (*e.g.*, two, three, four, five, seven, ten, etc. recombination sites, restriction sites, and/or topoisomerases sites) at which the sequences can be manipulated in a determinable fashion without loss of an essential biological function of the vector, and into which a nucleic acid fragment can be spliced in order to bring about its replication and cloning. Vectors can further provide primer sites (*e.g.*, for PCR),

transcriptional and/or translational initiation and/or regulation sites, recombinational signals, replicons, selectable markers, etc. Clearly, methods of inserting a desired nucleic acid fragment that do not require the use of recombination, transpositions or restriction enzymes (such as, but not limited to, uracil N-glycosylase (UDG) cloning of PCR fragments (U.S. Patent No. 5,334,575 and 5,888,795, both of which are entirely incorporated herein by reference), T:A cloning, and the like) can also be applied to clone a fragment into a cloning vector to be used according to the present invention. The cloning vector can further contain one or more selectable markers (*e.g.*, two, three, four, five, seven, ten, etc.) suitable for use in the identification of cells transformed with the cloning vector.

[0192] Subcloning Vector: As used herein, the phrase "subcloning vector" refers to a cloning vector comprising a circular or linear nucleic acid molecule that includes, preferably, an appropriate replicon. In the present invention, the subcloning vector (segment D in Fig. 1) can also contain functional and/or regulatory elements that are desired to be incorporated into the final product to act upon or with the cloned nucleic acid insert (segment A in Fig. 1). The subcloning vector can also contain a selectable marker (preferably DNA).

[0193] Vector Donor: As used herein, the phrase "Vector Donor" refers to one of the two parental nucleic acid molecules (*e.g.*, RNA or DNA) of the present invention that carries the nucleic acid segments comprising the nucleic acid vector that is to become part of the desired Product. The Vector Donor comprises a subcloning vector D (or it can be called the cloning vector if the Insert Donor does not already contain a cloning vector) and a segment C flanked by recombination sites (see Fig. 1). Segments C and/or D can contain elements that contribute to selection for the desired Product daughter molecule, as described above for selection schemes. The recombination signals can be the same or different, and can be acted upon by the same or different recombinases. In addition, the Vector Donor can be linear or circular. A Vector Donor may be referred to as a Destination Vector.

[0194] Primer: As used herein, the term "primer" refers to a single stranded or double stranded oligonucleotide that is extended by covalent bonding of nucleotide monomers during amplification or polymerization of a nucleic acid molecule (*e.g.*, a DNA molecule). In one aspect, the primer may be a

sequencing primer (for example, a universal sequencing primer). In another aspect, the primer may comprise a recombination site or portion thereof.

[0195] Adapter: As used herein, the term "adapter" refers to an oligonucleotide or nucleic acid fragment or segment (preferably DNA) that comprises one or more recombination sites (or portions of such recombination sites) that can be added to a circular or linear Insert Donor molecule as well as to other nucleic acid molecules described herein. When using portions of recombination sites, the missing portion may be provided by the Insert Donor molecule. Such adapters may be added at any location within a circular or linear molecule, although the adapters are preferably added at or near one or both termini of a linear molecule. Preferably, adapters are positioned to be located on both sides (flanking) a particular nucleic acid molecule of interest. In accordance with the invention, adapters may be added to nucleic acid molecules of interest by standard recombinant techniques (*e.g.*, restriction digest and ligation). For example, adapters may be added to a circular molecule by first digesting the molecule with an appropriate restriction enzyme, adding the adapter at the cleavage site and reforming the circular molecule that contains the adapter(s) at the site of cleavage. In other aspects, adapters may be added by homologous recombination, by integration of RNA molecules, and the like. Alternatively, adapters may be ligated directly to one or more and preferably both termini of a linear molecule thereby resulting in linear molecule(s) having adapters at one or both termini. In one aspect of the invention, adapters may be added to a population of linear molecules, (*e.g.*, a cDNA library or genomic DNA that has been cleaved or digested) to form a population of linear molecules containing adapters at one and preferably both termini of all or substantial portion of said population.

[0196] Adapter-Primer: As used herein, the phrase "adapter-primer" refers to a primer molecule that comprises one or more recombination sites (or portions of such recombination sites) that can be added to a circular or to a linear nucleic acid molecule described herein. When using portions of recombination sites, the missing portion may be provided by a nucleic acid molecule (*e.g.*, an adapter) of the invention. Such adapter-primers may be added at any location within a circular or linear molecule, although the adapter-primers are preferably added at or near one or both termini of a linear

molecule. Such adapter-primers may be used to add one or more recombination sites or portions thereof to circular or linear nucleic acid molecules in a variety of contexts and by a variety of techniques, including but not limited to amplification (*e.g.*, PCR), ligation (*e.g.*, enzymatic or chemical/synthetic ligation), recombination (*e.g.*, homologous or non-homologous (illegitimate) recombination) and the like.

[0197] Template: As used herein, the term "template" refers to a double stranded or single stranded nucleic acid molecule that is to be amplified, synthesized or sequenced. In the case of a double-stranded DNA molecule, denaturation of its strands to form a first and a second strand is preferably performed before these molecules may be amplified, synthesized or sequenced, or the double stranded molecule may be used directly as a template. For single stranded templates, a primer complementary to at least a portion of the template hybridizes under appropriate conditions and one or more polypeptides having polymerase activity (*e.g.*, two, three, four, five, or seven DNA polymerases and/or reverse transcriptases) may then synthesize a molecule complementary to all or a portion of the template. Alternatively, for double stranded templates, one or more transcriptional regulatory sequences (*e.g.*, two, three, four, five, seven or more promoters) may be used in combination with one or more polymerases to make nucleic acid molecules complementary to all or a portion of the template. The newly synthesized molecule, according to the invention, may be of equal or shorter length compared to the original template. Mismatch incorporation or strand slippage during the synthesis or extension of the newly synthesized molecule may result in one or a number of mismatched base pairs. Thus, the synthesized molecule need not be exactly complementary to the template. Additionally, a population of nucleic acid templates may be used during synthesis or amplification to produce a population of nucleic acid molecules typically representative of the original template population.

[0198] Incorporating: As used herein, the term "incorporating" means becoming a part of a nucleic acid (*e.g.*, DNA) molecule or primer.

[0199] Library: As used herein, the term "library" refers to a collection of nucleic acid molecules (circular or linear). In one embodiment, a library may comprise a plurality of nucleic acid molecules (*e.g.*, two, three, four, five,

seven, ten, twelve, fifteen, twenty, thirty, fifty, one hundred, two hundred, five hundred one thousand, five thousand, or more), that may or may not be from a common source organism, organ, tissue, or cell. In another embodiment, a library is representative of all or a portion or a significant portion of the nucleic acid content of an organism (a "genomic" library), or a set of nucleic acid molecules representative of all or a portion or a significant portion of the expressed nucleic acid molecules (a cDNA library or segments derived therefrom) in a cell, tissue, organ or organism. A library may also comprise nucleic acid molecules having random sequences made by de novo synthesis, mutagenesis of one or more nucleic acid molecules, and the like. Such libraries may or may not be contained in one or more vectors (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.).

[0200] Amplification: As used herein, the term "amplification" refers to any *in vitro* method for increasing the number of copies of a nucleic acid molecule with the use of one or more polypeptides having polymerase activity (*e.g.*, one, two, three, four or more nucleic acid polymerases or reverse transcriptases). Nucleic acid amplification results in the incorporation of nucleotides into a DNA and/or RNA molecule or primer thereby forming a new nucleic acid molecule complementary to a template. The formed nucleic acid molecule and its template can be used as templates to synthesize additional nucleic acid molecules. As used herein, one amplification reaction may consist of many rounds of nucleic acid replication. DNA amplification reactions include, for example, polymerase chain reaction (PCR). One PCR reaction may consist of 5 to 100 cycles of denaturation and synthesis of a DNA molecule.

[0201] Nucleotide: As used herein, the term "nucleotide" refers to a base-sugar-phosphate combination. Nucleotides are monomeric units of a nucleic acid molecule (DNA and RNA). The term nucleotide includes ribonucleoside triphosphates ATP, UTP, CTG, GTP and deoxyribonucleoside triphosphates such as dATP, dCTP, dITP, dUTP, dGTP, dTTP, or derivatives thereof. Such derivatives include, for example, [α -S]dATP, 7-deaza-dGTP and 7-deaza-dATP. The term nucleotide as used herein also refers to dideoxyribonucleoside triphosphates (ddNTPs) and their derivatives. Illustrated examples of dideoxyribonucleoside triphosphates include, but are

not limited to, ddATP, ddCTP, ddGTP, ddITP, and ddTTP. According to the present invention, a "nucleotide" may be unlabeled or detectably labeled by well known techniques. Detectable labels include, for example, radioactive isotopes, fluorescent labels, chemiluminescent labels, bioluminescent labels and enzyme labels.

[0202] Nucleic Acid Molecule: As used herein, the phrase "nucleic acid molecule" refers to a sequence of contiguous nucleotides (riboNTPs, dNTPs, ddNTPs, or combinations thereof) of any length. A nucleic acid molecule may encode a full-length polypeptide or a fragment of any length thereof, or may be non-coding. As used herein, the terms "nucleic acid molecule" and "polynucleotide" may be used interchangeably and include both RNA and DNA.

[0203] Oligonucleotide: As used herein, the term "oligonucleotide" refers to a synthetic or natural molecule comprising a covalently linked sequence of nucleotides that are joined by a phosphodiester bond between the 3' position of the pentose of one nucleotide and the 5' position of the pentose of the adjacent nucleotide.

[0204] Polypeptide: As used herein, the term "polypeptide" refers to a sequence of contiguous amino acids of any length. The terms "peptide," "oligopeptide," or "protein" may be used interchangeably herein with the term "polypeptide."

[0205] Hybridization: As used herein, the terms "hybridization" and "hybridizing" refer to base pairing of two complementary single-stranded nucleic acid molecules (RNA and/or DNA) to give a double stranded molecule. As used herein, two nucleic acid molecules may hybridize, although the base pairing is not completely complementary. Accordingly, mismatched bases do not prevent hybridization of two nucleic acid molecules provided that appropriate conditions, well known in the art, are used. In some aspects, hybridization is said to be under "stringent conditions." By "stringent conditions," as the phrase is used herein, is meant overnight incubation at 42°C in a solution comprising: 50% formamide, 5x SSC (750 mM NaCl, 75m M trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 µg/ml denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1 x SSC at about 65°C.

[0206] Transduce: As used herein, "transduce" and "transduction" refer to a process of introducing a virus into a cell type that does not support replication of the virus and does not result in the production of infectious viral progeny. In contrast, "infect" or "infection" are used to indicate introduction of a virus into a cell type that supports replication and results in the production of infectious viral progeny.

[0207] Other terms used in the fields of recombinant nucleic acid technology and molecular and cell biology as used herein will be generally understood by one of ordinary skill in the applicable arts.

Overview

[0208] The present invention relates to methods, compositions and kits for the recombinational joining of two or more segments or nucleic acid molecules to produce a nucleic acid molecule comprising all or a portion of a viral genome, for example, a recombinant viral vector. Further, the present invention relates to methods, compositions and kits for the topoisomerase-mediated joining of two or more segments or nucleic acid molecules to produce a nucleic acid molecule comprising all or a portion of a viral genome, for example, a recombinant viral vector. The present invention also relates to methods, compositions and kits for the joining by other means (*e.g.*, ligase) of two or more segments or nucleic acid molecules to produce a nucleic acid molecule comprising all or a portion of a viral genome, for example, a recombinant viral vector. The invention also includes methods for preparing such nucleic acid molecules, as well as compositions comprising such nucleic acid molecules. The present invention also contemplates methods for using these molecules to generate host cells, methods of using these molecules to produce polypeptide and/or RNA expression products.

[0209] In one embodiment, at least two nucleic acid segments, each comprising at least one recombination site, are contacted with suitable recombination proteins to effect the joining of all or a portion of the two molecules, depending on the position in the molecules of the recombination sites that undergo recombination. Each individual nucleic acid segment may comprise a variety of sequences including, but not limited to viral sequences,

sequences suitable for use as primer binding sites (*e.g.*, sequences for which a primer such as a sequencing primer or amplification primer may hybridize to initiate nucleic acid synthesis, amplification or sequencing), transcription or translation signals or regulatory sequences such as promoters and/or enhancers, ribosomal binding sites, Kozak sequences, start codons, termination signals such as stop codons, origins of replication, recombination sites (or portions thereof), selectable markers, and genes or portions of genes to create protein fusions (*e.g.*, N-terminal or C-terminal) such as GST, GUS, GFP, YFP, CFP, maltose binding protein, 6 histidines (HIS6), epitopes, haptens and the like and combinations thereof. The vectors used for cloning such segments may also comprise these functional sequences (*e.g.*, promoters, primer sites, etc.).

[0210] After joining of the segments, the product molecule will often contain at least sufficient viral sequences to permit the packaging of the product molecule in a viral particle. In the case where the viral sequences are adenoviral sequences, the product molecule may contain a left ITR, a packaging sequence and a right ITR, and/or sufficient other sequences to result in a molecule of appropriate size for packaging. In some embodiments, the product molecule comprises sufficient viral sequences to be an infectious viral genome when introduced into a permissive host cell. In some embodiments, a recombinant adenoviral vector produced by the methods of the invention may comprise a left ITR, a packaging sequence a first recombination site, a sequence of interest, a second recombination site, and additional adenoviral sequences including a right ITR. In the case where the viral sequences are retroviral sequences, the product molecule may contain a 5'-LTR, a 3'-LTR and a packaging sequence (Ψ), and/or sufficient other sequences to result in a molecule of appropriate size for packaging. In some embodiments, the product molecule comprises sufficient retroviral sequences to integrate into the genome of host cell into which it is introduced but not enough viral sequences to produce an infectious virus in the host cell. In some embodiments, a recombinant retroviral vector produced by the methods of the invention may be a plasmid comprising a 5'-LTR, a packaging sequence a first recombination site, a sequence of interest, and a second recombination site, and additional retroviral sequences including a 3'-LTR.

Recombination Sites

[0211] Recombination sites for use in the invention may be any nucleic acid that can serve as a substrate in a recombination reaction. Such recombination sites may be wild-type or naturally occurring recombination sites, or modified, variant, derivative, or mutant recombination sites. Examples of recombination sites for use in the invention include, but are not limited to, phage-lambda recombination sites (such as attP, attB, attL, and attR and mutants or derivatives thereof) and recombination sites from other bacteriophages such as phi80, P22, P2, 186, P4 and P1 (including lox sites such as loxP and loxP511).

[0212] In some embodiments, recombination sites that may be used in the practice of the invention include recombination sites that undergo recombination with compatible recombination sites in the presence of one or more recombination proteins active in the phage lambda recombination system, for example, one or more of Int, IHF, FIS, and/or Xis. The invention also contemplates nucleic acid molecules comprising such recombination sites and compositions comprising such nucleic acid molecules. Preferred recombination proteins and mutant, modified, variant, or derivative recombination sites for use in the invention include those described in U.S. Patent Nos. 5,888,732, 6,143,557, 6,171,861, 6,270,969, and 6,277,608 and in U.S. application no. 09/438,358 (filed November 12, 1999), based upon United States provisional application no. 60/108,324 (filed November 13, 1998). Mutated att sites (*e.g.*, attB 1-10, attP 1-10, attR 1-10 and attL 1-10) are described in United States application numbers 09/517,466, filed March 2, 2000, and 09/732,914, filed December 11, 2000 (published as 2002 0007051-A1) the disclosures of which are specifically incorporated herein by reference in their entirety. Other suitable recombination sites and proteins are those associated with the GATEWAY™ Cloning Technology available from Invitrogen Corporation, Carlsbad, CA, and described in the product literature of the GATEWAY™ Cloning Technology, the entire disclosures of all of which are specifically incorporated herein by reference in their entireties.

[0213] Sites that may be used in the present invention include att sites. The 15 bp core region of the wildtype att site (GCTTTTTTAT ACTAA (SEQ ID NO:)), which is identical in all wildtype att sites, may be mutated in one or

more positions. Other att sites that specifically recombine with other att sites can be constructed by altering nucleotides in and near the 7 base pair overlap region, bases 6-12 of the core region. Thus, recombination sites suitable for use in the methods, molecules, compositions, and vectors of the invention include, but are not limited to, those with insertions, deletions or substitutions of one, two, three, four, or more nucleotide bases within the 15 base pair core region (see U.S. Application Nos. 08/663,002, filed June 7, 1996 (now U.S. Patent No. 5,888,732) and 09/177,387, filed October 23, 1998, which describes the core region in further detail, and the disclosures of which are incorporated herein by reference in their entireties). Recombination sites suitable for use in the methods, compositions, and vectors of the invention also include those with insertions, deletions or substitutions of one, two, three, four, or more nucleotide bases within the 15 base pair core region that are at least 50% identical, at least 55% identical, at least 60% identical, at least 65% identical, at least 70% identical, at least 75% identical, at least 80% identical, at least 85% identical, at least 90% identical, or at least 95% identical to this 15 base pair core region.

[0214] As a practical matter, whether any particular nucleic acid molecule is at least 50%, 60%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to, for instance, a given recombination site nucleotide sequence or portion thereof can be determined conventionally using known computer programs such as DNAsis software (Hitachi Software, San Bruno, California) for initial sequence alignment followed by ESEE version 3.0 DNA/protein sequence software (cabot@trog.mbb.sfu.ca) for multiple sequence alignments. Alternatively, such determinations may be accomplished using the BESTFIT program (Wisconsin Sequence Analysis Package, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711), which employs a local homology algorithm (Smith and Waterman, *Advances in Applied Mathematics* 2: 482-489 (1981)) to find the best segment of homology between two sequences. When using DNAsis, ESEE, BESTFIT or any other sequence alignment program to determine whether a particular sequence is, for instance, 95% identical to a reference sequence according to the present invention, the parameters are set such that the percentage of identity is calculated over the full length of the reference nucleotide sequence and that

gaps in homology of up to 5% of the total number of nucleotides in the reference sequence are allowed. Computer programs such as those discussed above may also be used to determine percent identity and homology between two proteins at the amino acid level.

[0215] Analogously, the core regions in attB1, attP1, attL1 and attR1 are identical to one another, as are the core regions in attB2, attP2, attL2 and attR2. Nucleic acid molecules suitable for use with the invention also include those comprising insertions, deletions or substitutions of one, two, three, four, or more nucleotides within the seven base pair overlap region (TTTATAC, bases 6-12 in the core region). The overlap region is defined by the cut sites for the integrase protein and is the region where strand exchange takes place. Examples of such mutants, fragments, variants and derivatives include, but are not limited to, nucleic acid molecules in which (1) the thymine at position 1 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; (2) the thymine at position 2 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; (3) the thymine at position 3 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; (4) the adenine at position 4 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or thymine; (5) the thymine at position 5 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; (6) the adenine at position 6 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or thymine; and (7) the cytosine at position 7 of the seven bp overlap region has been deleted or substituted with a guanine, thymine, or adenine; or any combination of one or more (e.g., two, three, four, five, etc.) such deletions and/or substitutions within this seven bp overlap region. The nucleotide sequences of representative seven base pair core regions are set out below.

[0216] Altered att sites have been constructed that demonstrate that (1) substitutions made within the first three positions of the seven base pair overlap (TTTATAC) strongly affect the specificity of recombination, (2) substitutions made in the last four positions (TTTATAC) only partially alter recombination specificity, and (3) nucleotide substitutions outside of the seven bp overlap, but elsewhere within the 15 base pair core region, do not affect

specificity of recombination but do influence the efficiency of recombination. Thus, nucleic acid molecules and methods of the invention include those comprising or employing one, two, three, four, five, six, eight, ten, or more recombination sites which affect recombination specificity, particularly one or more (*e.g.*, one, two, three, four, five, six, eight, ten, twenty, thirty, forty, fifty, etc.) different recombination sites that may correspond substantially to the seven base pair overlap within the 15 base pair core region, having one or more mutations that affect recombination specificity. Particularly preferred such molecules may comprise a consensus sequence such as NNNATAC wherein "N" refers to any nucleotide (*i.e.*, may be A, G, T/U or C). Preferably, if one of the first three nucleotides in the consensus sequence is a T/U, then at least one of the other two of the first three nucleotides is not a T/U.

[0217] The core sequence of each att site (attB, attP, attL and attR) can be divided into functional units consisting of integrase binding sites, integrase cleavage sites and sequences that determine specificity. Specificity determinants are defined by the first three positions following the integrase top strand cleavage site. These three positions are shown with underlining in the following reference sequence: CAACTTTTTTATAC AAAGTTG (SEQ ID NO:). Modification of these three positions (64 possible combinations) can be used to generate att sites that recombine with high specificity with other att sites having the same sequence for the first three nucleotides of the seven base pair overlap region. The possible combinations of first three nucleotides of the overlap region are shown in Table 1.

[0218]

Table 1. Modifications of the First Three Nucleotides of the <i>att</i> Site Seven Base Pair Overlap Region that Alter Recombination Specificity.			
AAA	CAA	GAA	TAA
AAC	CAC	GAC	TAC
AAG	CAG	GAG	TAG
AAT	CAT	GAT	TAT
ACA	CCA	GCA	TCA
ACC	CCC	GCC	TCC
ACG	CCG	GCG	TCG
ACT	CCT	GCT	TCT
AGA	CGA	GGA	TGA
AGC	CGC	GGC	TGC
AGG	CGG	GGG	TGG
AGT	CGT	GGT	TGT
ATA	CTA	GTA	TTA
ATC	CTC	GTC	TTC
ATG	CTG	GTG	TTG
ATT	CTT	GTT	TTT

[0219] Representative examples of seven base pair *att* site overlap regions suitable for in methods, compositions and vectors of the invention are shown in Table 2. The invention further includes nucleic acid molecules comprising one or more (e.g., one, two, three, four, five, six, eight, ten, twenty, thirty, forty, fifty, etc.) nucleotides sequences set out in Table 2. Thus, for example, in one aspect, the invention provides nucleic acid molecules comprising the nucleotide sequence GAAATAC, GATATAC, ACAATAC, or TGCATAC.

Table 2. Representative Examples of Seven Base Pair <i>att</i> Site Overlap Regions Suitable for use in the recombination sites of the Invention.			
AAAATAC	CAAATAC	GAAATAC	TAAATAC
AACATAC	CACATAC	GACATAC	TACATAC
AAGATAC	CAGATAC	GAGATAC	TAGATAC
AATATAC	CATATAC	GATATAC	TATATAC
ACAATAC	CCAATAC	GCAATAC	TCAATAC
ACCATAC	CCCATAC	GCCATAC	TCCATAC
ACGATAC	CCGATAC	GCGATAC	TCGATAC
ACTATAC	CCTATAC	GCTATAC	TCTATAC
AGAATAC	CGAATAC	GGAATAC	TGAATAC
AGCATAC	CGCATAC	GGCATAC	TGCATAC
AGGATAC	CGGATAC	GGGATAC	TGGATAC
AGTATAC	CGTATAC	GGTATAC	TGTATAC
ATAATAC	CTAATAC	GTAATAC	TTAATAC
ATCATAC	CTCATAC	GTCATAC	TTCATAC
ATGATAC	CTGATAC	GTGATAC	TTGATAC
ATTATAC	CTTATAC	GTTATAC	TTTATAC

[0220] As noted above, alterations of nucleotides located 3' to the three base pair region discussed above can also affect recombination specificity. For example, alterations within the last four positions of the seven base pair overlap can also affect recombination specificity.

[0221] For example, mutated att sites that may be used in the practice of the present invention include attB1 (AGCCTGCTTT TTTGTACAAA CTTGT (SEQ ID NO:)), attP1 (TACAGGTCAC TAATACCATC TAAGTAGTTG ATTCATAGTG ACTGGATATG TTGTGTTTCA CAGTATTATG TAGTCTGTTT TTTATGCAAA ATCTAATTCA ATATATTGAT ATTTATATCA TTTTACGTTT CTCGTTTCAGC TTTTTTGTAC AAAGTTGGCA TTATAAAAAA GCATTGCTCA TCAATTTGTT GCAACGAACA GGTCACATC AGTCAAAATA AAATCATTAT TTG (SEQ ID NO:)), attL1 (CAAATAATGA TTTTATTTTG ACTGATAGTG ACCTGTTCGT TGCAACAAAT TGATAAGCAA TGCTTTTTTA TAATGCCAAC TTTGTACAAA AAAGCAGGCT (SEQ ID NO:)), and attR1 (ACAAGTTTGT ACAAAAAAGC TGAACGAGAA ACGTAAAATG ATATAAATAT CAATATATTA AATTAGATTT TGCATAAAAA ACAGACTACA TAATACTGTA AAACACAACA TATCCAGTCA CTATG (SEQ ID NO:)). Table 3 provides the sequences of the regions surrounding the core region for the wild type att sites (attB0, P0, R0, and L0) as well as a variety of other suitable recombination sites. Those skilled in the art will appreciate that the remainder of the site may be the same as the corresponding site (B, P, L, or R) listed above.

Table 3. Nucleotide sequences of att sites.		
attB0	AGCCTGCTTT TTTATACTAA CTTGAGC	(SEQ ID NO:)
attP0	G TTCAGCTTT TTTATACTAA GTTGGCA	(SEQ ID NO:)
attL0	AGCCTGCTTT TTTATACTAA GTTGGCA	(SEQ ID NO:)
attR0	G TTCAGCTTT TTTATACTAA CTTGAGC	(SEQ ID NO:)
attB1	AGCCTGCTTT TTTGTACAAA CTTGT	(SEQ ID NO:)
attP1	G TTCAGCTTT TTTGTACAAA GTTGGCA	(SEQ ID NO:)
attL1	AGCCTGCTTT TTTGTACAAA GTTGGCA	(SEQ ID NO:)
attR1	G TTCAGCTTT TTTGTACAAA CTTGT	(SEQ ID NO:)

Table 3. Nucleotide sequences of att sites.		
attB2	ACCCAGCTTT CTTGTACAAA GTGGT	(SEQ ID NO:)
attP2	G TTCAGCTTT CTTGTACAAA GTTGGCA	(SEQ ID NO:)
attL2	ACCCAGCTTT CTTGTACAAA GTTGGCA	(SEQ ID NO:)
attR2	G TTCAGCTTT CTTGTACAAA GTGGT	(SEQ ID NO:)
attB5	CAACTTTATT ATACAAAGTT GT	(SEQ ID NO:)
attP5	G TTCAACTTT ATTATACAAA GTTGGCA	(SEQ ID NO:)
attL5	CAACTTTATT ATACAAAGTT GGCA	(SEQ ID NO:)
attR5	G TTCAACTTT ATTATACAAA GTTGT	(SEQ ID NO:)
attB11	CAACTTTTCT ATACAAAGTT GT	(SEQ ID NO:)
attP11	G TTCAACTTT TCTATACAAA GTTGGCA	(SEQ ID NO:)
attL11	CAACTTTTCT ATACAAAGTT GGCA	(SEQ ID NO:)
attR11	G TTCAACTTT TCTATACAAA GTTGT	(SEQ ID NO:)
attB17	CAACTTTTGT ATACAAAGTT GT	(SEQ ID NO:)
attP17	G TTCAACTTT TGTATACAAA GTTGGCA	(SEQ ID NO:)
attL17	CAACTTTTGT ATACAAAGTT GGCA	(SEQ ID NO:)
attR17	G TTCAACTTT TGTATACAAA GTTGT	(SEQ ID NO:)
attB19	CAACTTTTTC GTACAAAGTT GT	(SEQ ID NO:)
attP19	G TTCAACTTT TTCGTACAAA GTTGGCA	(SEQ ID NO:)
attL19	CAACTTTTTC GTACAAAGTT GGCA	(SEQ ID NO:)
attR19	G TTCAACTTT TTCGTACAAA GTTGT	(SEQ ID NO:)
attB20	CAACTTTTGT GTACAAAGTT GT	(SEQ ID NO:)
attP20	G TTCAACTTT TTGGTACAAA GTTGGCA	(SEQ ID NO:)
attL20	CAACTTTTGT GTACAAAGTT GGCA	(SEQ ID NO:)
attR20	G TTCAACTTT TTGGTACAAA GTTGT	(SEQ ID NO:)
attB21	CAACTTTTTA ATACAAAGTT GT	(SEQ ID NO:)
attP21	G TTCAACTTT TTAATACAAA GTTGGCA	(SEQ ID NO:)
attL21	CAACTTTTTA ATACAAAGTT GGCA	(SEQ ID NO:)

Table 3. Nucleotide sequences of att sites.

attR21	GTTCAACTTT TTAATACAAA GTTGT	(SEQ ID NO:)
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[0222] Other recombination sites having unique specificity (*i.e.*, a first site will recombine with its corresponding site and will not substantially recombine with a second site having a different specificity) are known to those skilled in the art and may be used to practice the present invention.

Corresponding recombination proteins for these systems may be used in accordance with the invention with the indicated recombination sites. Other systems providing recombination sites and recombination proteins for use in the invention include the FLP/FRT system from *Saccharomyces cerevisiae*, the resolvase family (*e.g.*, $\gamma\delta$, TndX, TnpX, Tn3 resolvase, Hin, Hjc, Gin, SpCCE1, ParA, and Cin), and IS231 and other *Bacillus thuringiensis* transposable elements. Other suitable recombination systems for use in the present invention include the XerC and XerD recombinases and the psi, dif and cer recombination sites in *E. coli*. Other suitable recombination sites may be found in United States patent no. 5,851,808 issued to Elledge and Liu which is specifically incorporated herein by reference.

[0223] The materials and methods of the invention may further encompass the use of "single use" recombination sites which undergo recombination one time and then either undergo recombination with low frequency (*e.g.*, have at least five fold, at least ten fold, at least fifty fold, at least one hundred fold, or at least one thousand fold lower recombination activity in subsequent recombination reactions) or are essentially incapable of undergoing recombination. The invention also provides methods for making and using nucleic acid molecules which contain such single use recombination sites and molecules which contain these sites. Examples of methods which can be used to generate and identify such single use recombination sites are set out below. Further examples of methods which can be used to generate and identify such single use recombination sites are set out in PCT/US00/21623, published as WO 01/11058, which claims priority to United States provisional patent application 60/147,892, filed August 9, 1999, both of which are specifically incorporated herein by reference.

[0224] The att system core integrase binding site comprises an interrupted seven base pair inverted repeat having the following nucleotide sequence:

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caactttnnnnnnnaaagttg (SEQ ID NO:39),

as well as variations thereof which can comprise either perfect or imperfect repeats.

[0225] The repeat elements can be subdivided into two distal and/or proximal "domains" composed of caac/gttg segments (underlined), which are distal to the central undefined sequence (the nucleotides of which are represented by the letter "n"), and tt/aaa segments, which are proximal to the central undefined sequence.

[0226] Alterations in the sequence composition of the distal and/or proximal domains on one or both sides of the central undefined region can affect the outcome of a recombination reaction. The scope and scale of the effect is a function of the specific alterations made, as well as the particular recombinational event (*e.g.*, LR vs. BP reactions).

[0227] For example, it is believed that an attB site altered to have the following nucleotide sequence:

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caactttnnnnnnnaaacaag (SEQ ID NO:40),

will functionally interact with a cognate attP and generate attL and attR. However, whichever of the latter two recombination sites acquires the segment containing "caag" (located on the left side of the sequence shown above) will be rendered non-functional to subsequent recombination events. The above is only one of many possible alterations in the core integrase binding sequence which can render att sites non-functional after engaging in a single recombination event. Thus, single use recombination sites may be prepared by altering nucleotides in the seven base pair inverted repeat regions which abut seven base pair overlap regions of att sites. This region is represented schematically as:

CAAC TTT [Seven Base Pair Overlap Region] AAA GTTG.

[0228] In generating single use recombination sites, one, two, three, four or more of nucleotides of the sequences CAAC TTT or AAAGTTG (*i.e.*, the seven base pair inverted repeat regions) may be substituted with other nucleotides or deleted altogether. These seven base pair inverted repeat

regions represent complementary sequences with respect to each other. Thus, alterations may be made in either seven base pair inverted repeat region in order to generate single use recombination sites. Further, when DNA is double stranded and one seven base pair inverted repeat region is present, the other seven base pair inverted repeat region will also be present on the other strand.

[0229] Using the sequence CAACTTT for illustration, examples of seven base pair inverted repeat regions which can form single use recombination sites include, but are not limited to, nucleic acid molecules in which (1) the cytosine at position 1 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, adenine, or thymine; (2) the adenine at position 2 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, cytosine, or thymine; (3) the adenine at position 3 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, cytosine, or thymine; (4) the cytosine at position 4 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, adenine, or thymine; (5) the thymine at position 5 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, cytosine, or adenine; (6) the thymine at position 6 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, cytosine, or adenine; and (7) the thymine at position 7 of the seven base pair inverted repeat region has been deleted or substituted with a guanine, cytosine, or adenine; or any combination of one, two, three, four, or more such deletions and/or substitutions within this seven base pair region. Representative examples of nucleotide sequences of the above described seven base pair inverted repeat regions are set out below in Table 4.

Table 4. Representative examples of nucleotide sequences of seven base pair inverted repeat regions.

aagaaaa	aagagcg	aagagaa	aagatat
ccgccac	ccgcctc	ccgcaca	ccgcttt
ggtggga	ggtgctc	ggtgata	ggtgtat
ttctttg	ttctctc	ttctgaa	ttctttt
aatacac	aatagcg	aataaca	aatatat
cctcgga	cctcccg	cctcaca	cctcttt
ggcgaaa	ggcgccg	ggcggaa	ggcgtat
ttgtcac	ttgtgcg	ttgtaca	ttgtttt
acaagga	acaaccg	acaataa	acaattt
caccttg	caccaga	caccgaa	cacctat
gaggcac	gaggcg	gaggaca	gaggttt
tattgga	tattaga	tattaca	tatttat
agaaaaa	agaaaga	agaagaa	agaattt
cgcccac	cgccctc	cgccaca	cgctttt
gcgggga	gcgggcg	gcggata	gcggtat
tcttttg	tcttcg	tcttgaa	tcttttt
ataacac	ataactc	ataaaca	ataattt
ctccaaa	ctccgcg	ctccata	ctcctat
gtgggga	gtggccg	gtgggaa	gtggtat
tgttttg	tgttctc	tgttaca	tgttttt

[0230] Representative examples of nucleotide sequences which form single use recombination sites may also be prepared by combining a nucleotide sequence set out in Table 5, Section 1, with a nucleotide sequence set out in Table 5, Section 2. Single use recombination sites may also be prepared by the insertion of one or more (*e.g.*, one, two, three, four, five six, seven, etc.) nucleotides internally within these regions.

Table 5. Representative examples of nucleotide sequences which form single use recombination sites.

Section 1 (CAAG)				Section 2 (TTT)		
aaaa	cccc	gggg	tttt	aaa	cca	ttc
aaac	ccca	ggga	ttta	aac	cac	ttg
aaag	ccct	gggc	tttc	aag	cgc	tat
aaat	cccg	gggt	tttg	aat	ctc	tct
aaca	ccac	ggag	ttat	aca	ggg	tgt
aaga	ccgc	ggtg	ttct	aga	gga	
aata	cctc	ggcg	ttgt	ata	ggc	
acaa	cacc	gagg	tatt	caa	ggt	
agaa	cgcc	gcgg	tctt	gaa	gag	
ataa	ctcc	gtgg	tggt	taa	gcg	
caaa	accc	aggg	atth	ccc	gtg	
gaaa	gccc	CGG	ctth	ccg	ttt	
taaa	tccc	tggg	gtth	cct	tta	

[0231] In most instances where one seeks to prevent recombination events with respect to a particular nucleic acid segment, the altered sequence will be

located proximally to the nucleic acid segment. Using the following schematic for illustration:

= 5' Nucleic Acid Segment 3' = caac ttt (Seven Base Pair Overlap Region)

AAA GTTG,

the lower case nucleotide sequence which represent a seven base pair inverted repeat region (*i.e.*, caac ttt) will generally have a sequence altered by insertion, deletion, and/or substitution to form a single use recombination site when one seeks to prevent recombination at the 3' end (*i.e.*, proximal end with respect to the nucleic acid segment) of the nucleic acid segment shown. Thus, a single recombination reaction can be used, for example, to integrate the nucleic acid segments into another nucleic acid molecule, then the recombination site becomes effectively non-functional, preventing the site from engaging in further recombination reactions. Similarly, single use recombination sites can be position at both ends of a nucleic acid segment so that the nucleic acid segment can be integrated into another nucleic acid molecule, or circularized, and will remain integrated, or circularized even in the presence of recombinases.

[0232] A number of methods may be used to screen potential single use recombination sites for functional activity (*e.g.*, undergo one recombination event followed by the failure to undergo subsequent recombination events). For example, with respect to the screening of recombination sites to identify those which become non-functional after a single recombination event, a first recombination reaction may be performed to generate a plasmid in which a negative selection marker is linked to one or more potentially defective recombination sites. The plasmid may then be reacted with another nucleic acid molecule which comprises a positive selection marker similarly linked to recombination sites. Thus, this selection system is designed such that molecules which recombine are susceptible to negative selection and molecules which do not recombine may be selected for by positive selection. Using such a system, one may then directly select for desired single use core site mutants.

[0233] As one skilled in the art would recognize, any number of screening assays may be designed which achieve the same results as those described above. In many instances, these assays will be designed so that an initial

recombination event takes place and then recombination sites which are unable to engage in subsequent recombination events are identified or molecules which contain such recombination sites are selected for. A related screening assay would result in selection against nucleic acid molecule which have undergone a second recombination event. Further, as noted above, screening assays can be designed where there is selection against molecules which have engaged in subsequent recombination events and selection for those which have not engaged in subsequent recombination events.

[0234] Single use recombination sites are especially useful for either decreasing the frequency of or preventing recombination when either large number of nucleic acid segments are attached to each other or multiple recombination reactions are performed. Thus, the invention further includes nucleic acid molecules which contain single use recombination sites, as well as methods for performing recombination using these sites.

[0235] Recombination sites used with the invention may also have embedded functions or properties. An embedded functionality is a function or property conferred by a nucleotide sequence in a recombination site that is not directly associated with recombination efficiency or specificity. For example, recombination sites may contain protein coding sequences (*e.g.*, intein coding sequences), intron/exon splice sites, origins of replication, and/or stop codons. Further, recombination sites that have more than one (*e.g.*, two, three, four, five, etc.) embedded functions or properties may also be prepared.

[0236] In some instances it will be advantageous to remove either RNA corresponding to recombination sites from RNA transcripts or amino acid residues encoded by recombination sites from polypeptides translated from such RNAs. Removal of such sequences can be performed in several ways and can occur at either the RNA or protein level. One instance where it may be advantageous to remove RNA transcribed from a recombination site will be when constructing a fusion polypeptide between a polypeptide of interest and a coding sequence present on the vector. The presence of an intervening recombination site between the ORF of the polypeptide of interest and the vector coding sequences may result in the recombination site (1) contributing codons to the mRNA that result in the inclusion of additional amino acid residues in the expression product, (2) contributing a stop codon to the mRNA

that prevents the production of the desired fusion protein, and/or (3) shifting the reading frame of the mRNA such that the two protein are not fused "in-frame."

[0237] In one aspect, the invention provides methods for removing nucleotide sequences encoded by recombination sites from RNA molecules. One example of such a method employs the use of intron/exon splice sites to remove RNA encoded by recombination sites from RNA transcripts. Nucleotide sequences that encode intron/exon splice sites may be fully or partially embedded in the recombination sites used in the present invention and/or may encoded by adjacent nucleic acid sequence. Sequences to be excised from RNA molecules may be flanked by splice sites that are appropriately located in the sequence of interest and/or on the vector. For example, one intron/exon splice site may be encoded by a recombination site and another intron/exon splice site may be encoded by other nucleotide sequences (e.g., nucleic acid sequences of the vector or a nucleic acid of interest). Nucleic acid splicing is well known to those skilled in the art and is discussed in the following publications: R. Reed, *Curr. Opin. Genet. Devel.* 6:215-220 (1996); S. Mount, *Nucl. Acids. Res.* 10:459-472, (1982); P. Sharp, *Cell* 77:805-815, (1994); K. Nelson and M. Green, *Genes and Devel.* 23:319-329 (1988); and T. Cooper and W. Mattox, *Am. J. Hum. Genet.* 61:259-266 (1997).

[0238] Splice sites can be suitably positioned in a number of locations. For example, a Destination Vector designed to express an inserted ORF with an N-terminal fusion—for example, with a detectable marker—the first splice site could be encoded by vector sequences located 3' to the detectable marker coding sequences and the second splice site could be partially embedded in the recombination site that separates the detectable marker coding sequences from the coding sequences of the ORF. Further, the second splice site either could abut the 3' end of the recombination site or could be positioned a short distance (e.g., 2, 4, 8, 10, 20 nucleotides) 3' to the recombination site. In addition, depending on the length of the recombination site, the second splice site could be fully embedded in the recombination site.

[0239] A modification of the method described above involves the connection of multiple nucleic acid segments that, upon expression, results in the

production of a fusion protein. In one specific example, one nucleic acid segment encodes detectable marker—for example, GFP—and another nucleic acid segment that encodes an ORF of interest. Each of these segments is flanked by recombination sites. In addition, the nucleic acid segments that encodes the detectable marker contains an intron/exon splice site near its 3' terminus and the nucleic acid segments that contains the ORF of interest also contains an intron/exon splice site near its 5' terminus. Upon recombination, the nucleic acid segment that encodes the detectable marker is positioned 5' to the nucleic acid segment that encodes the ORF of interest. Further, these two nucleic acid segments are separated by a recombination site that is flanked by intron/exon splice sites. Excision of the intervening recombination site thus occurs after transcription of the fusion mRNA. Thus, in one aspect, the invention is directed to methods for removing RNA transcribed from recombination sites from transcripts generated from nucleic acids described herein.

[0240] Splice sites may introduced into nucleic acid molecules to be used in the present invention in a variety of ways. One method that could be used to introduce intron/exon splice sites into nucleic acid segments is PCR. For example, primers could be used to generate nucleic acid segments corresponding to an ORF of interest and containing both a recombination site and an intron/exon splice site.

[0241] The above methods can also be used to remove RNA corresponding to recombination sites when the nucleic acid segment that is recombined with another nucleic acid segment encodes RNA that is not produced in a translatable format. One example of such an instance is where a nucleic acid segment is inserted into a vector in a manner that results in the production of antisense RNA. As discussed below, this antisense RNA may be fused, for example, with RNA that encodes a ribozyme. Thus, the invention also provides methods for removing RNA corresponding to recombination sites from such molecules.

[0242] The invention further provides methods for removing amino acid sequences encoded by recombination sites from protein expression products by protein splicing. Nucleotide sequences that encode protein splice sites may be fully or partially embedded in the recombination sites that encode amino

acid sequences excised from proteins or protein splice sites may be encoded by adjacent nucleotide sequences. Similarly, one protein splice site may be encoded by a recombination site and another protein splice sites may be encoded by other nucleotide sequences (e.g., nucleic acid sequences of the vector or a nucleic acid of interest).

[0243] It has been shown that protein splicing can occur by excision of an intein from a protein molecule and ligation of flanking segments (see, e.g., Derbyshire, *et al.*, *Proc. Natl. Acad. Sci. (USA)* 95:1356-1357 (1998)). In brief, inteins are amino acid segments that are post-translationally excised from proteins by a self-catalytic splicing process. A considerable number of intein consensus sequences have been identified (see, e.g., Perler, *Nucleic Acids Res.* 27:346-347 (1999)).

[0244] Similar to intron/exon splicing, N- and C-terminal intein motifs have been shown to be involved in protein splicing. Thus, the invention further provides compositions and methods for removing amino acid residues encoded by recombination sites from protein expression products by protein splicing. In particular, this aspect of the invention is related to the positioning of nucleic acid sequences that encode intein splice sites on both the 5' and 3' end of recombination sites positioned between two coding regions. Thus, when the protein expression product is incubated under suitable conditions, amino acid residues encoded these recombination sites will be excised.

[0245] Protein splicing may be used to remove all or part of the amino acid sequences encoded by recombination sites. Nucleic acid sequence that encode inteins may be fully or partially embedded in recombination sites or may adjacent to such sites. In certain circumstances, it may be desirable to remove considerable numbers of amino acid residues beyond the N- and/or C-terminal ends of amino acid sequences encoded by recombination sites. In such instances, intein coding sequence may be located a distance (e.g., 30, 50, 75, 100, etc. nucleotides) 5' and/or 3' to the recombination site.

[0246] While conditions suitable for intein excision will vary with the particular intein, as well as the protein that contains this intein, Chong *et al.*, *Gene* 192:271-281 (1997), have demonstrated that a modified *Saccharomyces cerevisiae* intein, referred to as Sce VMA intein, can be induced to undergo self-cleavage by a number of agents including 1,4-dithiothreitol (DTT), β -

mercaptoethanol, and cysteine. For example, intein excision/splicing can be induced by incubation in the presence of 30 mM DTT, at 4°C for 16 hours.

Topoisomerase cloning

[0247] The present invention also relates to methods of using one or more topoisomerases to generate a recombinant nucleic acid molecules of the invention (*e.g.*, molecules comprising all or a portion of a viral genome such as a viral vector) comprising two or more nucleotide sequences, any one or more of which may comprise, for example, all or a portion of a viral genome. Topoisomerases may be used in combination with recombinational cloning techniques described above. For example, a topoisomerase-mediated reaction may be used to attach one or more recombination sites to one or more nucleic acid segments. The segments may then be further manipulated and combined using, for example, recombinational cloning techniques.

[0248] In one aspect, the present invention provides methods for linking a first and at least a second nucleic acid segment (either or both of which may contain viral sequences and/or sequences of interest) with at least one (*e.g.*, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, etc.) topoisomerase (*e.g.*, a type IA, type IB, and/or type II topoisomerase) such that either one or both strands of the linked segments are covalently joined at the site where the segments are linked.

[0249] A method for generating a double stranded recombinant nucleic acid molecule covalently linked in one strand can be performed by contacting a first nucleic acid molecule which has a site-specific topoisomerase recognition site (*e.g.*, a type IA or a type II topoisomerase recognition site), or a cleavage product thereof, at a 5' or 3' terminus, with a second (or other) nucleic acid molecule, and optionally, a topoisomerase (*e.g.*, a type IA, type IB, and/or type II topoisomerase), such that the second nucleotide sequence can be covalently attached to the first nucleotide sequence. As disclosed herein, the methods of the invention can be performed using any number of nucleotide sequences, typically nucleic acid molecules wherein at least one of the nucleotide sequences has a site-specific topoisomerase recognition site (*e.g.*, a type IA, type IB or type II topoisomerase), or cleavage product thereof, at one or both 5' and/or 3' termini.

[0250] In some embodiments, two double-stranded nucleic acid molecules can be joined into a one larger molecule such that each strand of the larger molecule is covalently joined (*e.g.*, the larger molecule has no nicks). With reference to Fig. 3, a first double-stranded nucleic acid molecule having a topoisomerase linked to each of the 5' terminus and 3' terminus of one end may be contacted with a second nucleic acid under conditions causing the linkage of both strands of the first nucleic acid molecule to both strands of the second nucleic acid molecule (Fig. 3A). The end of the first nucleic acid molecules to which the topoisomerases are attached may have either a 5'-overhang, 3'-overhang or be blunt ended. The end of the second nucleic acid molecule to be joined to the first nucleic acid molecule may have the same type of end as the topoisomerase-linked end of the first nucleic acid molecule. The end of the second molecule that is not to be joined may have a different end if directional joining of the segments is desired and may have the same type of end if directionality is not required.

[0251] In another embodiment, a first nucleic acid molecule having a topoisomerase bound to the 3' terminus of one end, and a second nucleic acid molecule having a topoisomerase bound to the 3' terminus of one end may be joined using the methods of the invention (Fig. 3B). A covalently linked double-stranded recombinant nucleic acid molecule is generated by contacting the ends containing the topoisomerase-charged substrate nucleic acid molecules.

[0252] Figure 3C shows a first nucleic acid molecule having a topoisomerase bound to the 5' terminus of one end, and a second nucleic acid molecule having a topoisomerase bound to the 5' terminus of one end, and further shows the production of a covalently linked double-stranded recombinant nucleic acid molecule generated by contacting the ends containing the topoisomerase-charged substrate nucleic acid molecules.

[0253] Figure 3D shows a nucleic acid molecule having a topoisomerase linked to each of the 5' terminus and 3' terminus of both ends, and further shows linkage of the topoisomerase-charged nucleic acid molecule to two nucleic acid molecules, one at each end. The topoisomerases at each of the 5' termini and/or at each of the 3' termini can be the same or different. Those skilled in the art will appreciate that nicked molecules (*e.g.*, covalently joined

in only one strand) may be produced by omitting one of the topoisomerases from the any one of the methods described above for Figs. 3A-3D.

[0254] A method for generating a double stranded recombinant nucleic acid molecule covalently linked in both strands can be performed, for example, by contacting a first nucleic acid molecule having a first end and a second end, wherein, at the first end or second end or both ends, the first nucleic acid molecule has a topoisomerase recognition site (or cleavage product thereof) at or near the 5' or 3' terminus; at least a second nucleic acid molecule having a first end and a second end, wherein, at the first end or second end or both ends, the at least second double stranded nucleotide sequence has a topoisomerase recognition site (or cleavage product thereof) at or near a 5' or 3' terminus; and at least one site specific topoisomerase (*e.g.*, a type IA and/or a type IB topoisomerase), under conditions such that all components are in contact and the topoisomerase can effect its activity. A covalently linked double stranded recombinant nucleic acid generated according to a method of this aspect of the invention is characterized, in part, in that it does not contain a nick in either strand at the position where the nucleic acid molecules are joined. In one embodiment, the method is performed by contacting a first nucleic acid molecule and a second (or other) nucleic acid molecule, each of which has a topoisomerase recognition site in addition to viral sequences an/or sequences of interest, or a cleavage product thereof, at the 3' termini or at the 5' termini of two ends to be covalently linked. In another embodiment, the method is performed by contacting a first nucleic acid molecule having a topoisomerase recognition site, or cleavage product thereof, at the 5' terminus and the 3' terminus of at least one end, and a second (or other) nucleic acid molecule having a 3' hydroxyl group and a 5' hydroxyl group at the end to be linked to the end of the first nucleic acid molecule containing the recognition sites. As disclosed herein, the methods can be performed using any number of nucleic acid molecules having various combinations of termini and ends.

[0255] Topoisomerases are categorized as type I, including type IA and type IB topoisomerases, which cleave a single strand of a double stranded nucleic acid molecule, and type II topoisomerases (gyrases), which cleave both strands of a nucleic acid molecule. Type IA and IB topoisomerases cleave one strand of a nucleic acid molecule. Cleavage of a nucleic acid molecule by type IA

topoisomerases generates a 5' phosphate and a 3' hydroxyl at the cleavage site, with the type IA topoisomerase covalently binding to the 5' terminus of a cleaved strand. In comparison, cleavage of a nucleic acid molecule by type IB topoisomerases generates a 3' phosphate and a 5' hydroxyl at the cleavage site, with the type IB topoisomerase covalently binding to the 3' terminus of a cleaved strand. As disclosed herein, type I and type II topoisomerases, as well as catalytic domains and mutant forms thereof, are useful for generating double stranded recombinant nucleic acid molecules covalently linked in both strands according to a method of the invention.

[0256] Type IA topoisomerases include *E. coli* topoisomerase I, *E. coli* topoisomerase III, eukaryotic topoisomerase II, archeal reverse gyrase, yeast topoisomerase III, *Drosophila* topoisomerase III, human topoisomerase III, *Streptococcus pneumoniae* topoisomerase III, and the like, including other type IA topoisomerases (see Berger, *Biochim. Biophys. Acta* 1400:3-18, 1998; DiGate and Marians, *J. Biol. Chem.* 264:17924-17930, 1989; Kim and Wang, *J. Biol. Chem.* 267:17178-17185, 1992; Wilson, *et al.*, *J. Biol. Chem.* 275:1533-1540, 2000; Hanai, *et al.*, *Proc. Natl. Acad. Sci., USA* 93:3653-3657, 1996, U.S. Pat. No. 6,277,620, each of which is incorporated herein by reference). *E. coli* topoisomerase III, which is a type IA topoisomerase that recognizes, binds to and cleaves the sequence 5'-GCAACTT-3', can be particularly useful in a method of the invention (Zhang, *et al.*, *J. Biol. Chem.* 270:23700-23705, 1995, which is incorporated herein by reference). A homolog, the traE protein of plasmid RP4, has been described by Li, *et al.*, *J. Biol. Chem.* 272:19582-19587 (1997) and can also be used in the practice of the invention. A DNA-protein adduct is formed with the enzyme covalently binding to the 5'-thymidine residue, with cleavage occurring between the two thymidine residues.

[0257] Type IB topoisomerases include the nuclear type I topoisomerases present in all eukaryotic cells and those encoded by vaccinia and other cellular poxviruses (see Cheng, *et al.*, *Cell* 92:841-850, 1998, which is incorporated herein by reference). The eukaryotic type IB topoisomerases are exemplified by those expressed in yeast, *Drosophila* and mammalian cells, including human cells (see Caron and Wang, *Adv. Pharmacol.* 29B:271-297, 1994; Gupta, *et al.*, *Biochim. Biophys. Acta* 1262:1-14, 1995, each of which is

incorporated herein by reference; see, also, Berger, *supra*, 1998). Viral type IB topoisomerases are exemplified by those produced by the vertebrate poxviruses (vaccinia, Shope fibroma virus, ORF virus, fowlpox virus, and molluscum contagiosum virus), and the insect poxvirus (*Amsacta moorei* entomopoxvirus) (see Shuman, *Biochim. Biophys. Acta* 1400:321-337, 1998; Petersen, *et al.*, *Virology* 230:197-206, 1997; Shuman and Prescott, *Proc. Natl. Acad. Sci., USA* 84:7478-7482, 1987; Shuman, *J. Biol. Chem.* 269:32678-32684, 1994; U.S. Pat. No. 5,766,891; PCT/US95/16099; PCT/US98/12372, each of which is incorporated herein by reference; see, also, Cheng, *et al.*, *supra*, 1998).

[0258] Type II topoisomerases include, for example, bacterial gyrase, bacterial DNA topoisomerase IV, eukaryotic DNA topoisomerase II, and T-even phage encoded DNA topoisomerases (Roca and Wang, *Cell* 71:833-840, 1992; Wang, *J. Biol. Chem.* 266:6659-6662, 1991, each of which is incorporated herein by reference; Berger, *supra*, 1998;). Like the type IB topoisomerases, the type II topoisomerases have both cleaving and ligating activities. In addition, like type IB topoisomerase, substrate nucleic acid molecules can be prepared such that the type II topoisomerase can form a covalent linkage to one strand at a cleavage site. For example, calf thymus type II topoisomerase can cleave a substrate nucleic acid molecule containing a 5' recessed topoisomerase recognition site positioned three nucleotides from the 5' end, resulting in dissociation of the three nucleotide sequence 5' to the cleavage site and covalent binding the of the topoisomerase to the 5' terminus of the nucleic acid molecule (Andersen, *et al.*, *supra*, 1991). Furthermore, upon contacting such a type II topoisomerase charged nucleic acid molecule with a second nucleotide sequence containing a 3' hydroxyl group, the type II topoisomerase can ligate the sequences together, and then is released from the recombinant nucleic acid molecule. As such, type II topoisomerases also are useful for performing methods of the invention.

[0259] The various topoisomerases exhibit a range of sequence specificity. For example, type II topoisomerases can bind to a variety of sequences, but cleave at a highly specific recognition site (see Andersen, *et al.*, *J. Biol. Chem.* 266:9203-9210, 1991, which is incorporated herein by reference;). In comparison, the type IB topoisomerases include site specific topoisomerases,

which bind to and cleave a specific nucleotide sequence ("topoisomerase recognition site"). Upon cleavage of a nucleic acid molecule by a topoisomerase, for example, a type IB topoisomerase, the energy of the phosphodiester bond is conserved via the formation of a phosphotyrosyl linkage between a specific tyrosine residue in the topoisomerase and the 3' nucleotide of the topoisomerase recognition site. Where the topoisomerase cleavage site is near the 3' terminus of the nucleic acid molecule, the downstream sequence (3' to the cleavage site) can dissociate, leaving a nucleic acid molecule having the topoisomerase covalently bound to the newly generated 3' end.

[0260] With reference to Fig. 4, a combination of restriction digestion/ligation and recombinational cloning may be used to construct nucleic acid molecules of the invention. A nucleic acid molecule (*e.g.*, a plasmid) having at least one recognition site (*e.g.*, recombination site) (RS₁) and at least one restriction enzyme site (RE) may be constructed. A molecule of this type may comprise a tag sequence, optionally located adjacent to the restriction enzyme site. The molecule may be digested with a restriction enzyme resulting in a linear molecule. The resultant linear molecule may be contacted with a second nucleic acid molecule comprising at least one recombination site and having an end compatible with the restriction digested end of the linear first nucleic acid molecule. In the presence of ligase and the appropriate recombination proteins, the second nucleic acid molecule is covalently coupled to the first nucleic acid molecule replacing the portion of the first nucleic acid molecule between the recombination site and the restriction enzyme site. Those skilled in the art will appreciate that one or more topoisomerases may be used in place of or in combination with the restriction enzyme digestion and/or ligation reactions. Thus, the invention contemplates linear molecules, which may be charged at one end with one or more topoisomerases, containing at least one recombination site. The invention also contemplates compositions comprising such molecules, reaction mixtures comprising such molecules, and methods of making and using such molecules.

Suppressor tRNAs

[0261] Mutant tRNA molecules that recognize what are ordinarily stop codons suppress the termination of translation of an mRNA molecule and are termed suppressor tRNAs. Three codons are used by both eukaryotes and prokaryotes to signal the end of gene. When transcribed into mRNA, the codons have the following sequences: UAG (amber), UGA (opal) and UAA (ochre). Under most circumstances, the cell does not contain any tRNA molecules that recognize these codons. Thus, when a ribosome translating an mRNA reaches one of these codons, the ribosome stalls and falls off the RNA, terminating translation of the mRNA. The release of the ribosome from the mRNA is mediated by specific factors (see S. Mottagui-Tabar, *Nucleic Acids Research* 26(11), 2789, 1998). A gene with an in-frame stop codon (TAA, TAG, or TGA) will ordinarily encode a protein with a native carboxy terminus. However, suppressor tRNAs can result in the insertion of amino acids and continuation of translation past stop codons.

[0262] A number of such suppressor tRNAs have been found. Examples include, but are not limited to, the supE, supP, supD, supF and supZ suppressors, which suppress the termination of translation of the amber stop codon, supB, glT, supL, supN, supC and supM suppressors, which suppress the function of the ochre stop codon and glyT, trpT and Su-9 suppressors, which suppress the function of the opal stop codon. In general, suppressor tRNAs contain one or more mutations in the anti-codon loop of the tRNA that allows the tRNA to base pair with a codon that ordinarily functions as a stop codon. The mutant tRNA is charged with its cognate amino acid residue and the cognate amino acid residue is inserted into the translating polypeptide when the stop codon is encountered. For a more detailed discussion of suppressor tRNAs, the reader may consult Eggertsson, *et al.*, (1988) *Microbiological Review* 52(3):354-374, and Engleberg-Kukla, *et al.* (1996) in *Escherichia coli and Salmonella Cellular and Molecular Biology*, Chapter 60, pps 909-921, Neidhardt, *et al.* eds., ASM Press, Washington, DC.

[0263] Mutations that enhance the efficiency of termination suppressors, *i.e.*, increase the read through of the stop codon, have been identified. These include, but are not limited to, mutations in the uar gene (also known as the

prfA gene), mutations in the ups gene, mutations in the sueA, sueB and sueC genes, mutations in the rpsD (ramA) and rpsE (spcA) genes and mutations in the rplL gene.

[0264] Under ordinary circumstances, host cells would not be expected to be healthy if suppression of stop codons is too efficient. This is because of the thousands or tens of thousands of genes in a genome, a significant fraction will naturally have one of the three stop codons; complete read-through of these would result in a large number of aberrant proteins containing additional amino acids at their carboxy termini. If some level of suppressing tRNA is present, there is a race between the incorporation of the amino acid and the release of the ribosome. Higher levels of tRNA may lead to more read-through although other factors, such as the codon context, can influence the efficiency of suppression.

[0265] Organisms ordinarily have multiple genes for tRNAs. Combined with the redundancy of the genetic code (multiple codons for many of the amino acids), mutation of one tRNA gene to a suppressor tRNA status does not lead to high levels of suppression. The TAA stop codon is the strongest, and most difficult to suppress. The TGA is the weakest, and naturally (in *E. coli*) leaks to the extent of 3%. The TAG (amber) codon is relatively tight, with a read-through of ~1% without suppression. In addition, the amber codon can be suppressed with efficiencies on the order of 50% with naturally occurring suppressor mutants. Suppression in some organisms (*e.g.*, *E. coli*) may be enhanced when the nucleotide following the stop codon is an adenosine. Thus, the present invention contemplates nucleic acid molecules having a stop codon followed by an adenosine (*e.g.*, having the sequence TAGA, TAAA, and/or TGAA).

[0266] Suppression has been studied for decades in bacteria and bacteriophages. In addition, suppression is known in yeast, flies, plants and other eukaryotic cells including mammalian cells. For example, Capone, *et al.* (*Molecular and Cellular Biology* 6(9):3059-3067, 1986) demonstrated that suppressor tRNAs derived from mammalian tRNAs could be used to suppress a stop codon in mammalian cells. A copy of the *E. coli* chloramphenicol acetyltransferase (cat) gene having a stop codon in place of the codon for serine 27 was transfected into mammalian cells along with a gene encoding a

human serine tRNA that had been mutated to form an amber, ochre, or opal suppressor derivative of the gene. Successful expression of the cat gene was observed. An inducible mammalian amber suppressor has been used to suppress a mutation in the replicase gene of polio virus and cell lines expressing the suppressor were successfully used to propagate the mutated virus (Sedivy, *et al.*, *Cell* 50: 379-389 (1987)). The context effects on the efficiency of suppression of stop codons by suppressor tRNAs has been shown to be different in mammalian cells as compared to *E. coli* (Phillips-Jones, *et al.*, *Molecular and Cellular Biology* 15(12): 6593-6600 (1995), Martin, *et al.*, *Biochemical Society Transactions* 21: (1993)) Since some human diseases are caused by nonsense mutations in essential genes, the potential of suppression for gene therapy has long been recognized (see Temple, *et al.*, *Nature* 296(5857):537-40 (1982)). The suppression of single and double nonsense mutations introduced into the diphtheria toxin A-gene has been used as the basis of a binary system for toxin gene therapy (Robinson, *et al.*, *Human Gene Therapy* 6:137-143 (1995)).

Use of Suppressor tRNAs to Conditionally Express Fusion Proteins

[0267] Because the methods used to create the nucleic acids of the invention are site specific, the orientation and/or reading frame of a nucleic acid sequence on a first nucleic acid molecule can be controlled with respect to the orientation and/or reading frame of a sequence on a second nucleic acid molecule when all or a portion of the molecules are joined in a recombination and/or topoisomerase-mediated reaction. This control makes the construction of fusions between sequences present on different nucleic acid molecules a simple matter.

[0268] In general terms, an open reading frame may be expressed in four forms: native at both amino and carboxy termini, modified at either end, or modified at both ends. A nucleic acid sequence of interest comprising an ORF of interest may include the N-terminal methionine ATG codon, and a stop codon at the carboxy end, of the ORF, thus ATG - ORF - stop. Frequently, the nucleic acid molecule comprising the sequence of interest will include translation initiation sequences, *tis*, that may be located upstream of the ATG that allow expression of the gene, thus *tis* - ATG - ORF - stop. Constructs of

this sort allow expression of an ORF as a protein that contains the same amino and carboxy amino acids as in the native, uncloned, protein. When such a construct is fused in-frame with an amino-terminal protein tag, *e.g.*, GST, the tag will have its own tis, thus tis - ATG - tag - tis - ATG - ORF - stop, and the bases comprising the tis of the ORF will be translated into amino acids between the tag and the ORF. In addition, some level of translation initiation may be expected in the interior of the mRNA (*i.e.*, at the ORF's ATG and not the tag's ATG) resulting in a certain amount of native protein expression contaminating the desired protein.

- [0269] DNA (lower case): tis1 - atg - tag - tis2 - atg - orf - stop
- [0270] RNA (lower case, italics): tis1 - atg - tag - tis2 - atg - orf - stop
- [0271] Protein (upper case): ATG - TAG - TIS2 - ATG - ORF (tis1 and stop are not translated) + contaminating ATG - ORF (translation of ORF beginning at tis2).
- [0272] Using the methods disclosed herein, one skilled in the art can construct a vector containing a tag adjacent to a recombination site permitting the in frame fusion of a tag to the C- and/or N-terminus of the ORF of interest.
- [0273] Given the ability to rapidly create a number of clones in a variety of vectors, there is a need in the art to maximize the number of ways a single cloned ORF can be expressed without the need to manipulate the construct itself. The present invention meets this need by providing materials and methods for the controlled expression of a C- and/or N-terminal fusion to a target ORF using one or more suppressor tRNAs to suppress the termination of translation at a stop codon. Thus, the present invention provides materials and methods in which a gene construct is prepared flanked with recombination sites.
- [0274] The construct may be prepared with a sequence coding for a stop codon preferably at the C-terminus of the ORF encoding the protein of interest. In some embodiments, a stop codon can be located adjacent to the ORF, for example, within the recombination site flanking the gene or at or near the 3' end of the sequence of interest before a recombination site. The target gene construct can be transferred through recombination to various vectors that can provide various C-terminal or N-terminal tags (*e.g.*, GFP, GST, His Tag, GUS, etc.) to the ORF of interest. When the stop codon is

located at the carboxy terminus of the ORF, expression of the ORF with a “native” carboxy end amino acid sequence occurs under non-suppressing conditions (*i.e.*, when the suppressor tRNA is not expressed) while expression of the ORF as a carboxy fusion protein occurs under suppressing conditions. Those skilled in the art will recognize that any suppressors and any codons could be used in the practice of the present invention. Suppressors may insert any amino acid at the position corresponding to the stop codon, for example, Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Lys, Met, Phe, Pro, Ser, Thr, Trp, Tyr, or Val may be inserted. In some embodiments, serine may be inserted.

[0275] In some embodiments, the gene coding for the suppressing tRNA may be incorporated into the vector from which the target ORF is to be expressed. In other embodiments, the gene for the suppressor tRNA may be in the genome of the host cell. In still other embodiments, the gene for the suppressor may be located on a separate viral vector or other vector—*i.e.*, plasmid—and provided *in trans*. In embodiments of this type, the vector containing the suppressor gene may be a recombinant adenoviral vector and cells may be co-infected with a viral vector expressing a sequence of interest and a viral vector expressing a suppressor tRNA.

[0276] More than one copy of a suppressor tRNA may be provided in all of the embodiments described herein. For example, a host cell may be provided that contains multiple copies of a gene encoding the suppressor tRNA. Alternatively, multiple gene copies of the suppressor tRNA under the same or different promoters may be provided in the same vector background as the target ORF of interest. In some embodiments, multiple copies of a suppressor tRNA may be provided in a different vector than the one containing the target ORF of interest. In other embodiments, one or more copies of the suppressor tRNA gene may be provided on the vector containing the ORF for the protein of interest and/or on another vector and/or in the genome of the host cell or in combinations of the above. When more than one copy of a suppressor tRNA gene is provided, the genes may be expressed from the same or different promoters that may be the same or different as the promoter used to express the ORF encoding the protein of interest.

[0277] In some embodiments, two or more different suppressor tRNA genes may be provided. In embodiments of this type one or more of the individual suppressors may be provided in multiple copies and the number of copies of a particular suppressor tRNA gene may be the same or different as the number of copies of another suppressor tRNA gene. Each suppressor tRNA gene, independently of any other suppressor tRNA gene, may be provided on the vector used to express the ORF of interest and/or on a different vector and/or in the genome of the host cell. A given tRNA gene may be provided in more than one place in some embodiments. For example, a copy of the suppressor tRNA may be provided on the vector containing the ORF of interest while one or more additional copies may be provided on an additional vector and/or in the genome of the host cell. When more than one copy of a suppressor tRNA gene is provided, the genes may be expressed from the same or different promoters that may be the same or different as the promoter used to express the ORF encoding the protein of interest and may be the same or different as a promoter used to express a different tRNA gene.

[0278] In some embodiments of the present invention, the target ORF of interest and the gene expressing the suppressor tRNA may be controlled by the same promoter. In other embodiments, the target ORF of interest may be expressed from a different promoter than the suppressor tRNA. Those skilled in the art will appreciate that, under certain circumstances, it may be desirable to control the expression of the suppressor tRNA and/or the target ORF of interest using a regulatable promoter. For example, either the target ORF of interest and/or the gene expressing the suppressor tRNA may be controlled by a promoter such as the lac promoter or derivatives thereof such as the tac promoter. In some embodiments, both the target ORF of interest and the suppressor tRNA gene are expressed from the T7 RNA polymerase promoter and, optionally, are expressed as part of one RNA molecule. In embodiments of this type, the portion of the RNA corresponding to the suppressor tRNA is processed from the originally transcribed RNA molecule by cellular factors.

[0279] In some embodiments, the expression of the suppressor tRNA gene may be under the control of a different promoter from that of the ORF of interest. In some embodiments, it may be possible to express the suppressor gene before the expression of the target ORF. This would allow levels of

suppressor to build up to a high level, before they are needed to allow expression of a fusion protein by suppression of a the stop codon. For example, in embodiments of the invention where the suppressor gene is controlled by a promoter inducible with IPTG, the target ORF is controlled by the T7 RNA polymerase promoter and the expression of the T7 RNA polymerase is controlled by a promoter inducible with an inducing signal other than IPTG, *e.g.*, NaCl, one could turn on expression of the suppressor tRNA gene with IPTG prior to the induction of the T7 RNA polymerase gene and subsequent expression of the ORF of interest. In some embodiments, the expression of the suppressor tRNA might be induced about 15 minutes to about one hour before the induction of the T7 RNA polymerase gene. In one embodiment, the expression of the suppressor tRNA may be induced from about 15 minutes to about 30 minutes before induction of the T7 RNA polymerase gene. In some embodiments, the expression of the T7 RNA polymerase gene is under the control of an inducible promoter.

[0280] In additional embodiments, the expression of the target ORF of interest and the suppressor tRNA can be arranged in the form of a feedback loop. For example, the target ORF of interest may be placed under the control of the T7 RNA polymerase promoter while the suppressor gene is under the control of both the T7 promoter and the lac promoter. The T7 RNA polymerase gene itself is also under the control of both the T7 promoter and the lac promoter. In addition, the T7 RNA polymerase gene has an amber stop mutation replacing a normal tyrosine codon, *e.g.*, the 28th codon (out of 883). No active T7 RNA polymerase can be made before levels of suppressor are high enough to give significant suppression. Then expression of the polymerase rapidly rises, because the T7 polymerase expresses the suppressor gene as well as itself. In other preferred embodiments, only the suppressor gene is expressed from the T7 RNA polymerase promoter. Embodiments of this type would give a high level of suppressor without producing an excess amount of T7 RNA polymerase. In other preferred embodiments, the T7 RNA polymerase gene has more than one amber stop mutation. This will require higher levels of suppressor before active T7 RNA polymerase is produced.

[0281] In some embodiments of the present invention it may be desirable to have more than one stop codon suppressible by more than one suppressor

tRNA. A recombinant viral vector may be constructed so as to permit the regulatable expression of N- and/or C-terminal fusions of a protein of interest from the same construct. A viral vector may comprise a first tag sequence expressed from a promoter and may include a first stop codon in the same reading frame as the tag. The stop codon may be located anywhere in the tag sequence and is preferably located at or near the C-terminal of the tag sequence. The stop codon may also be located in a recombination site or in an internal ribosome entry sequence (IRES). The viral vector may also include a sequence of interest preferably comprising a ORF of interest that includes a second stop codon. The first tag and the ORF of interest are preferably in the same reading frame although inclusion of a sequence that causes frame shifting to bring the first tag into the same reading frame as the ORF of interest is within the scope of the present invention. The second stop codon is preferably in the same reading frame as the ORF of interest and is preferably located at or near the end of the coding sequence for the ORF. The second stop codon may optionally be located within a recombination site located 3' to the sequence of interest. The construct may also include a second tag sequence in the same reading frame as the ORF of interest and the second tag sequence may optionally include a third stop codon in the same reading frame as the second tag. A transcription terminator and/or a polyadenylation sequence may be included in the construct after the coding sequence of the second tag. The first, second and third stop codons may be the same or different. In some embodiments, all three stop codons are different. In embodiments where the first and the second stop codons are different, the same construct may be used to express an N-terminal fusion, a C-terminal fusion and the native protein by varying the expression of the appropriate suppressor tRNA. For example, to express the native protein, no suppressor tRNAs are expressed and protein translation is controlled by an appropriately located IRES. When an N-terminal fusion is desired, a suppressor tRNA that suppresses the first stop codon is expressed while a suppressor tRNA that suppresses the second stop codon is expressed in order to produce a C-terminal fusion. In some instances it may be desirable to express a doubly tagged protein of interest in which case suppressor tRNAs that suppress both the first and the second stop codons may be expressed.

Construction and Uses Nucleic Acid Molecules of the Invention.

[0282] As discussed below in more detail, in one aspect, the invention provides a modular system for constructing viruses, *e.g.*, viral vectors, having particular functions or activities. The present invention also includes methods for preparing viruses, *e.g.*, viral vectors, containing more than one nucleic acid insert (*e.g.*, two, three, four, five, six, eight, ten, twelve, fifteen, twenty, thirty, forty, fifty, etc. inserts). In one general embodiment of the invention, viral vectors and/or nucleic acids molecules of the invention are prepared as follows. Nucleic acid molecules that are to ultimately be incorporated into the viral vector are obtained (*e.g.*, purchased, prepared by PCR or by the preparation of cDNA using reverse transcriptase). Suitable recombination sites are either incorporated into the 5' and/or 3' ends of the nucleic acid molecules during synthesis or added later. A nucleic acid comprising all or a portion of a viral genome and the nucleic acid to be incorporated are combined in the presence of one or more recombination proteins in order to construct the desired viral vector.

[0283] In some embodiments of the invention nucleic acid molecules of the invention may be combined using various combinations of techniques known in the art. When a first nucleic acid molecule is to be joined with a second nucleic acid molecule, the ends of the molecules may be joined using the same or different techniques. For example, one end of a first nucleic acid molecule to be joined with a second nucleic acid molecule may comprise one type of recognition site (*e.g.*, a topoisomerase site) and the other end may comprise a different type of site (*e.g.*, a recombination site or a restriction enzyme site). In various embodiments, a nucleic acid molecule may have a restriction enzyme site on one end and a topoisomerase site on the other end, a restriction enzyme site on one end and a recombination site on the other end, or a topoisomerase site on one end and a recombination site on the other end. Those skilled in the art will appreciate that a ligase and/or topoisomerase may be used to link an end having a restriction site with another nucleic acid molecule. When topoisomerase is used to join two nucleic acid molecules, either or both strands may be covalently joined. Figure 3 shows examples of the covalent joining of both strands.

[0284] To construct a modular viral vector, one or more nucleic acid segments comprising one or more recombination sites and also comprising a viral sequence may be prepared. In some embodiments, multiple segments, each having at least one recombination site and some having viral sequences (e.g., baculoviral or adenoviral sequences) may be constructed and combined to produce a nucleic acid molecule of the invention. For example, a nucleic acid segment comprising an adenoviral ITR and a recombination site may be prepared. Further, a plurality of nucleic acid segments, each comprising a different portion of the adenoviral genome flanked by recombination sites, may be prepared. In some embodiments, the entire genome of an adenovirus is prepared in segments flanked by recombination sites. Such segments may be combined with one or more additional segments comprising additional sequences of interest such that, after combining, a nucleic acid comprising all or a portion of an adenoviral genome and comprising a sequence of interest is formed.

[0285] Segments of an adenoviral genome may be prepared from different serotypes of adenovirus, for example, Ad5, Ad3, Ad10, etc., and viral vectors having a mixed serotype, (e.g., some determinants of Ad5 and some of Ad10) may be prepared. It may be desirable to vary the most immunogenic portions of the viruses in situations where multiple administrations of viral vectors are contemplated.

[0286] Each segment of the adenoviral genome may comprise one or more regions of the genome, for example, left ITR, right ITR, packaging signal, E1, E2, E3, E4, and/or one or more late regions. In some embodiments, a segment may comprise the entire adenoviral genome except one region that is on a different segment. For example, an entire adenoviral genome except for the packaging signal may be prepared on one segment and the packaging signal may be prepared on a different segment. The two segments may be combined (e.g., using recombinational cloning) to produce a viral vector of the invention. Likewise, an entire adenoviral genome may be prepared that lacks one or more of the following elements: left ITR, E1, E2, E3, E4, or right ITR. The lacking element may be prepared on a separate segment and the two segments may be combined to produce a viral vector. One or more sequences of interest may be incorporated into either segment prior to combining the

segments in order to produce an adenoviral vector containing one or more sequences of interest. More than one viral region may be prepared on a segment, for example, the left ITR, packaging signal, and E3 region may be prepared on one segment with the remainder of the adenoviral functions necessary to prepare a viral vector present on one or more other segments. Sequences of interest may be present on any one of the segments.

[0287] Typically, the nucleic acid molecules may be dissolved in an aqueous buffer and added to the reaction mixture. One suitable set of conditions is 4 μ l CLONASE™ enzyme mixture (*e.g.*, Invitrogen Corporation, Cat. Nos. 11791-019 and 11789-013), 4 μ l 5X reaction buffer and nucleic acid and water to a final volume of 20 μ l. This will typically result in the inclusion of about 200 ng of Int and about 80 ng of IHF in a 20 μ l BP reaction and about 150 ng Int, about 25 ng IHF and about 30 ng Xis in a 20 μ l LR reaction.

[0288] Proteins for conducting an LR reaction may be stored in a suitable buffer, for example, LR Storage Buffer, which may comprise about 50 mM Tris at about pH 7.5, about 50 mM NaCl, about 0.25 mM EDTA, about 2.5 mM Spermidine, and about 0.2 mg/ml BSA. When stored, proteins for an LR reaction may be stored at a concentration of about 37.5 ng/ μ l INT, 10 ng/ μ l IHF and 15 ng/ μ l XIS. Proteins for conducting a BP reaction may be stored in a suitable buffer, for example, BP Storage Buffer, which may comprise about 25 mM Tris at about pH 7.5, about 22 mM NaCl, about 5 mM EDTA, about 5 mM Spermidine, about 1 mg/ml BSA, and about 0.0025% Triton X-100. When stored, proteins for an BP reaction may be stored at a concentration of about 37.5 ng/ μ l INT and 20 ng/ μ l IHF. One skilled in the art will recognize that enzymatic activity may vary in different preparations of enzymes. The amounts suggested above may be modified to adjust for the amount of activity in any specific preparation of enzymes.

[0289] A suitable 5X reaction buffer for conducting recombination reactions may comprise 100 mM Tris pH 7.5, 88 mM NaCl, 20 mM EDTA, 20 mM Spermidine, and 4 mg/ml BSA. Thus, in a recombination reaction, the final buffer concentrations may be 20 mM Tris pH 7.5, 17.6 mM NaCl, 4 mM EDTA, 4 mM Spermidine, and 0.8 mg/ml BSA. Those skilled in the art will appreciate that the final reaction mixture may incorporate additional components added with the reagents used to prepare the mixture, for example,

a BP reaction may include 0.005% Triton X-100 incorporated from the BP Clonase™.

[0290] In some preferred embodiments, particularly those in which *attL* sites are to be recombined with *attR* sites, the final reaction mixture may include about 50 mM Tris HCl, pH 7.5, about 1 mM EDTA, about 1 mg/ml BSA, about 75 mM NaCl and about 7.5 mM spermidine in addition to recombination enzymes and the nucleic acids to be combined. In other preferred embodiments, particularly those in which an *attB* site is to be recombined with an *attP* site, the final reaction mixture may include about 25 mM Tris HCl, pH 7.5, about 5 mM EDTA, about 1 mg/ml bovine serum albumin (BSA), about 22 mM NaCl, and about 5 mM spermidine.

[0291] In some preferred embodiments, particularly those in which *attL* sites are to be recombined with *attR* sites, the final reaction mixture may include about 40 mM Tris HCl, pH 7.5, about 1 mM EDTA, about 1 mg/ml BSA, about 64 mM NaCl and about 8 mM spermidine in addition to recombination enzymes and the nucleic acids to be combined. One of skill in the art will appreciate that the reaction conditions may be varied somewhat without departing from the invention. For example, the pH of the reaction may be varied from about 7.0 to about 8.0; the concentration of buffer may be varied from about 25 mM to about 100 mM; the concentration of EDTA may be varied from about 0.5 mM to about 2 mM; the concentration of NaCl may be varied from about 25 mM to about 150 mM; and the concentration of BSA may be varied from 0.5 mg/ml to about 5 mg/ml. In other preferred embodiments, particularly those in which an *attB* site is to be recombined with an *attP* site, the final reaction mixture may include about 25 mM Tris HCl, pH 7.5, about 5 mM EDTA, about 1 mg/ml bovine serum albumin (BSA), about 22 mM NaCl, about 5 mM spermidine and about 0.005% detergent (*e.g.*, Triton X-100).

[0292] The invention also includes viral vectors, in addition to adenoviral vectors (*e.g.*, baculoviral vectors), which contain either all or, part of one or more viral genome. Using vectors comprising baculoviral nucleic acid for purposes of illustration, vectors of the invention include those which comprise one or more element (*e.g.*, one or more functional element) of a baculoviral genome, as well as vectors which comprise one or more element (*e.g.*,

promoters, transcription terminators, polyA signals or sequences, ribosome binding sites, enhancers, ORFs or portions thereof, etc.) of one or more other viral genomes. Typically, these vectors will include one or more recombination site, as described elsewhere herein.

[0293] One specific example of a vector of the invention is shown schematically in Fig. 13. This vector contains three separate baculoviral elements. More specifically, the vector shown in Fig. 13 comprises the IE2 gene promoter and IE2 gene polyA region of *Orgyia pseudotsugata*. The vector also includes the GP64 promoter of *Autographa californica*. Thus, nucleic acid molecules of the invention include vectors which contain one or more elements (*e.g.*, an element described herein) derived from one or more viral genome (*e.g.*, adenoviral genome, baculoviral genome, etc.). Further, these elements may be from the same or different viruses.

[0294] The invention further includes nucleic acid molecules which comprise modified elements of viral genomes. These modified elements may be defined and/or described within the scope of the invention in any number of ways. Examples of such ways include (1) function (*e.g.*, a property conferred upon a nucleic acid which contains the element), (2) % sequence identity, and (3) % homology or sequence identity of expression products, as well as combinations of these ways. Percent homology or sequence identity will typically be determined with reference to the nucleotide or amino acid sequence of another nucleic acid or polypeptide.

[0295] As indicated above, viral elements and modified viral elements suitable for use with the invention may be described by their ability to confer one or more functional properties on nucleic acid molecules which contain them. Using the GP64 promoter as an example, this promoter is an inducible promoter which exhibits low level basal constitutive activity. In other words, in the absence of induction, the GP64 promoter allows for low level of transcription when operably linked to a nucleic acid segment. Functional properties are also associated with other viral elements, such as origins of replication, polyA tail sequences, packaging signals, LTRs, etc. Further, depending on the particular element, functional activity can be assessed at either the level of the vector (*e.g.*, the DNA or RNA level), a transcription product (*e.g.*, the RNA level), and/or a translation product (*e.g.*, the

polypeptide level). Thus, the invention further includes nucleic acid molecules which comprise modified viral elements which retain all or some of the functions of the viral elements from which they are derived (*e.g.*, the “wild-type” viral element). In many instances, a modified element will retain at least one functional property of the element from which they are derived. In particular embodiments, the modified element will (1) have at least one additional property not associated with the element from which it was derived, (2) be deficient in at least one property associated with the element from which it was derived, and/or (3) have increased or decreased activity with respect to at least one property associated with the element from which it was derived.

[0296] As also indicated above, modified elements (*e.g.*, modified viral elements) contained in nucleic acid molecules of the invention may be described by their structural similarity to elements from which they are derived. For example, modified elements may be at least 50% identical, at least 55% identical, at least 60% identical, at least 65% identical, at least 70% identical, at least 75% identical, at least 80% identical, at least 85% identical, at least 90% identical, or at least 95% identical at the nucleic acid level to the nucleic acid molecules from which they are derived. Modified elements may also be defined by having sufficient structural similarity to the nucleic acid molecules from which they are derived (*e.g.*, an element the nucleotide sequence of which is set out elsewhere herein) so that the two nucleic acids will hybridized. Often, these molecules will hybridized to each other under stringent hybridization conditions. In many instances, these modified elements will retain at least one property associated from the elements from which they are derived.

[0297] When modified elements of a viral genome encode a polypeptide expression product, the polypeptide may be at least 50% identical or homologous, at least 55% identical or homologous, at least 60% identical or homologous, at least 65% identical or homologous, at least 70% identical or homologous, at least 75% identical or homologous, at least 80% identical or homologous, at least 85% identical or homologous, at least 90% identical or homologous, or at least 95% identical or homologous at the amino acid level to the amino acid sequences of the polypeptide which is expressed from the

nucleic acid from which the modified elements is derived. Typically, polypeptide expression products of modified elements will retain at least one functional property of polypeptides which are expressed from nucleic acids from which the modified elements are derived. In particular embodiments, the polypeptide expression product of a modified element will (1) have at least one additional property not associated with the polypeptide expression product from which the element from which it was derived, (2) be deficient in at least one property associated with the polypeptide expression product from which the element from which it was derived, and/or (3) have increased or decreased activity with respect to at least one property associated with the polypeptide expression product from which the element from which it was derived.

[0298] One example of a vector of the invention is a vector which contains the GP64 promoter of *Autographa californica* operably linked to a heterologous nucleic acid. In particular embodiments, the GP64 promoter has all or part of the nucleotide sequence set out in Table 12 beginning at nucleotide 3364. The invention further include nucleic acid molecules which comprise modified forms of the GP64 promoter. These modified forms of the GP64 promoter include deleted forms of the promoter which comprise at least 30 nucleotides, at least 40 nucleotides, at least 50 nucleotides, at least 60 nucleotides, at least 70 nucleotides, at least 80 nucleotides, at least 90 nucleotides, or at least 95 nucleotides.

[0299] As indicated above, vectors of the invention may comprise all or part of a viral genome. For example, vectors of the invention may comprise at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 95%, or 100% of a viral genome used to prepare the vector. For example, a baculoviral vector which contains about 50% of the used to prepare it may contain about 66 kb of baculoviral nucleic acid.

[0300] It is not necessary that all viral functions required for replication be contained on a segment and be included in the final nucleic acid molecule comprising all or a portion of the viral genome. One or more required functions may be provided *in trans*. For example, a required function may be incorporated into the genome of a cell line and still provide the function.

Viruses lacking the function could be prepared in the cell line expressing the function. These viruses could only replicate in the cell line expressing the function and, thus, would be replication-deficient in any other cell line. Any required function could be used in this fashion, for example, the adenovirus E2 and/or E4 functions (*see, Weinberg, et al., Proc. Ntl. Acad. Sci. USA 80:5383, 5386, 1983*).

[0301] Segments prepared as above may be linear fragments (*e.g.*, PCR fragments) or segments may be part of larger nucleic acid molecule (*e.g.*, a plasmid). The segments may be combined to form a viral vector of the invention. When the segments are combined, the resultant adenoviral vector may be a linear molecule, for example, by combining linear segments using recombination cloning. A linear viral vector may be introduced (*e.g.*, by transfection, electroporation, etc.) into an appropriate host cell and packaged virus may be isolated as described elsewhere herein. Alternatively, a viral vector may be prepared as part of a circular molecule (*e.g.*, a plasmid) and the viral vector may be released from the circular molecule (*e.g.*, by restriction digest) and introduced into an appropriate host cell and packaged virus isolated.

[0302] When one seeks to prepare or construct a viral vector containing multiple nucleic acid inserts, these inserts can be inserted into a viral vector in either one reaction mixture or a series of reaction mixtures. For example, multiple nucleic acid segments can be linked end to end and inserted into a viral vector using reactions performed, for example, in a single reaction mixture. The nucleic acid segments in this reaction mixture can be designed so that recombination sites on their 5' and 3' ends result in their insertion into a nucleic acid comprising all or a portion of a viral genome in a specific order and a specific 5' to 3' orientation. Alternatively, nucleic acid segments can be designed so that they are inserted into a nucleic acid comprising all or a portion of a viral genome without regard to order, orientation (*i.e.*, 5' to 3' orientation), the number of inserts, and/or the number of duplicate inserts.

[0303] Methods of the invention can also be used to prepare viral vectors that, upon expression of a sequence of interest contained in the viral vector, produce one or more polypeptides having one or more desired property, function, or activity (*e.g.*, an enzymatic activity, the ability to bind a nucleic

acid, etc.). For example, a polypeptide having one or more enzymatic activities might be expressed from the viral vectors of the present invention. Viral vectors of this type might be used, for example, in a gene therapy protocol to replace a missing enzymatic activity. Polypeptides produced from the viral vectors of the present invention may have other desirable characteristics, for example, a polypeptide may comprise one or more antigenic determinants. Expression of such a polypeptide may result in an immune response specific for the expressed polypeptide. Such a viral vector may be used, for example, as an immunotherapeutic, for example, a vaccine.

[0304] Methods of the invention can also be used to prepare viral vectors that, upon expression of a sequence of interest contained in the viral vector, produce one or more un-translated RNA molecules, for example, ribozymes, antisense molecules, RNAi and the like. Such a viral vector might be used, for example, to modulate (*e.g.*, inhibit) the expression of one more RNA or polypeptide molecules produced by a host organism. Such a vector might be used, for example, to inhibit the expression of a disease associated RNA or polypeptide.

[0305] Methods of the invention can also be used to prepare viral vectors that, upon expression of a sequence of interest contained in the viral vector, produce fusion proteins having more than one property, function, or activity. Further, the expression product can be produced in such a manner as to facilitate its export from the cell. For example, these expression products can be fusion proteins that contain a signal peptide that results in export of the protein from the cell. One application where cell export may be desirable is where the proteins that are to be exported are enzymes that interact with extracellular substrates.

[0306] In a specific embodiment, the invention further provides methods for introducing viral vectors and/or nucleic acids molecules of the invention into animals (*e.g.*, humans) and animal cells (*e.g.*, human cells), as part of a gene therapy protocol. Viral vectors of the present invention may be designed such that compositions comprising the vectors are free of viral vectors that are replication competent in the target cell. Thus, in some embodiments, viral vectors of the present invention are replication restricted, *i.e.*, can replicate in a

permissive cell type, *e.g.*, 293 cells, and cannot replicate in a target cell type, *e.g.*, patient cells.

[0307] Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid molecule. In many embodiments of the invention, nucleic acid molecules of the invention will encoded one or more proteins (*e.g.*, one or more fusion proteins) that mediate at least one therapeutic effect. Thus, the invention provide nucleic acid molecules and methods for use in gene therapy.

[0308] Viral vectors and/or nucleic acids molecules of the invention can be used to prepare gene therapy vectors designed to replace genes that reside in the genome of a cell, to delete such genes, or to insert a heterologous gene or groups of genes. When viral vectors and/or nucleic acids molecules of the invention function to delete or replace a gene or genes, the gene or genes being deleted or replaced may lead to the expression of either a "normal" phenotype or an aberrant phenotype. One example of an aberrant phenotype is the disease cystic fibrosis. Further, the gene therapy vectors may be either stably maintained (*e.g.*, integrate into cellular nucleic acid by homologous or site specific recombination) or non-stably maintained in cells.

[0309] Further, viral vectors and/or nucleic acids molecules of the invention may be used to suppress "abnormal" phenotypes or complement or supplement "normal" phenotypes that result from the expression of endogenous genes. One example of a viral vector of the invention designed to suppress an abnormal phenotype would be where an expression product of the viral vector has dominant/negative activity. An example of a viral vector of the invention designed to supplement a normal phenotype would be where introduction of the viral vector effectively results in the amplification of a gene resident in the cell.

[0310] In some embodiments, viral vectors and/or nucleic acids of the present invention may be used to prevent or inhibit the expression of one or more genes in an organism, for example, by homology-dependent gene silencing (HDGS, see, for example, Bernstein, *et al.*, *RNA* 7:1509-21 (2001), and Bass, *Cell* 101:235-238 (2000)). The genes expression of which is to be inhibited, *i.e.*, silenced, may be endogenous to the organism or may be exogenous to the organism.

[0311] Viral vectors and/or nucleic acid molecules of the invention may be prepared to generate interfering RNAs (RNAi). RNAi is double-stranded RNA that results in degradation of specific mRNAs, and can also be used to lower or eliminate gene expression. Viral vectors and/or nucleic acid molecules of the invention may be engineered, for example, to produce dsRNA molecules by, for example, engineering the viral vectors and/or nucleic acid molecules to have a sequence that, when transcribed, folds back upon itself to generate a hairpin molecule containing a double-stranded portion. One strand of the double-stranded portion may correspond to all or a portion of the sense strand of the mRNA transcribed from the gene to be silenced while the other strand of the double-stranded portion may correspond to all or a portion of the antisense strand. Other methods of producing a double-stranded RNA molecule may be used, for example, a viral vector and/or nucleic acid molecules may be engineered to have a first sequence that, when transcribed, corresponds to all or a portion of the sense strand of the mRNA transcribed from the gene to be silenced and a second sequence that, when transcribed, corresponds to all or portion of an antisense strand (*i.e.*, the reverse complement) of the mRNA transcribed from the gene to be silenced. This may be accomplished by putting the first and the second sequence on the same strand of the viral vector each under the control of its own promoter. Alternatively, two promoters may be positioned on opposite strands of the viral vector such that expression from each promoter results in transcription of one strand of the double-stranded RNA. In some embodiments, it may be desirable to have the first sequence on one viral vector or nucleic acid molecule and the second sequence on a second viral vector or nucleic acid molecule and to introduce both vectors or molecules into a cell containing the gene to be silenced. In other embodiments, a viral vector or nucleic acid molecule containing only the antisense strand may be introduced and the mRNA transcribed from the gene to be silenced may serve as the other strand of the double-stranded RNA. In some embodiments, a dsRNA to be used to silence a gene may have one or more regions of homology to a gene to be silenced. Regions of homology may be from about 20 bp to about 5 kbp in length, 20 bp to about 4 kbp in length, 20 bp to about 3 kbp in length, 20 bp to about 2.5 kbp in length, from about 20 bp to about 2 kbp in length, 20 bp to

about 1.5 kbp in length, from about 20 bp to about 1 kbp in length, 20 bp to about 750 bp in length, from about 20 bp to about 500 bp in length, 20 bp to about 400 bp in length, 20 bp to about 300 bp in length, 20 bp to about 250 bp in length, from about 20 bp to about 200 bp in length, from about 20 bp to about 150 bp in length, from about 20 bp to about 100 bp in length, from about 20 bp to about 90 bp in length, from about 20 bp to about 80 bp in length, from about 20 bp to about 70 bp in length, from about 20 bp to about 60 bp in length, from about 20 bp to about 50 bp in length, from about 20 bp to about 40 bp in length, from about 20 bp to about 30 bp in length, from about 20 bp to about 25 bp in length, from about 15 bp to about 25 bp in length, from about 17 bp to about 25 bp in length, from about 19 bp to about 25 bp in length, from about 15 bp to about 23 bp, from about 17 bp to about 23 bp, from about 19 bp to about 23 bp in length, from about 15 bp to about 21 bp, from about 17 bp to about 21 bp, or from about 19 bp to about 21 bp in length.

[0312] As discussed above, a hairpin containing molecule having a double-stranded region may be used as RNAi. The length of the double stranded region may be from about 20 bp to about 2.5 kbp in length, from about 20 bp to about 2 kbp in length, 20 bp to about 1.5 kbp in length, from about 20 bp to about 1 kbp in length, 20 bp to about 750 bp in length, from about 20 bp to about 500 bp in length, 20 bp to about 400 bp in length, 20 bp to about 300 bp in length, 20 bp to about 250 bp in length, from about 20 bp to about 200 bp in length, from about 20 bp to about 150 bp in length, from about 20 bp to about 100 bp in length, 20 bp to about 90 bp in length, 20 bp to about 80 bp in length, 20 bp to about 70 bp in length, 20 bp to about 60 bp in length, 20 bp to about 50 bp in length, 20 bp to about 40 bp in length, 20 bp to about 30 bp in length, from about 20 bp to about 25 bp in length, from about 15 bp to about 25 bp in length, from about 17 bp to about 25 bp in length, from about 19 bp to about 25 bp in length, from about 15 bp to about 23 bp, from about 17 bp to about 23 bp, from about 19 bp to about 23 bp in length, from about 15 bp to about 21 bp, from about 17 bp to about 21 bp, or from about 19 bp to about 21 bp in length. The non-base-paired portion of the hairpin (*i.e.*, loop) can be of any length that permits the two regions of homology that make up the double-stranded portion of the hairpin to fold back upon one another.

[0313] Any suitable promoter may be used to control the production of RNA from the nucleic acid molecules of the invention. Promoters may be those recognized by any polymerase enzyme. For example, promoters may be promoters for RNA polymerase II or RNA polymerase III (*e.g.*, a U6 promoter, an H1 promoter, etc.). Other suitable promoters include, but are not limited to, T7 promoter, cytomegalovirus (CMV) promoter, mouse mammary tumor virus (MMTV) promoter, metallothionine, RSV (Rous sarcoma virus) long terminal repeat, SV40 promoter, human growth hormone (hGH) promoter. Other suitable promoters are known to those skilled in the art and are within the scope of the present invention.

[0314] One example of a construct designed to produce RNAi is shown in Figure 5B. In this construct, a DNA segment is inserted into a vector such that RNA corresponding to both strands are produced as two separate transcripts. Another example of a construct designed to produce RNAi is shown in Figure 5C. In this construct, two copies of a DNA segment are inserted into a vector such that RNA corresponding to both strands are again produced. Yet another example of a construct designed to produce RNAi is shown in Figure 5D. In this construct, two copies of a DNA segment are inserted into a vector such that RNA corresponding to both strands are produced as a single transcript. The exemplary vector system shown in shown in Figures 5E and 5F comprises two vectors, each of which contain copies of the same DNA segment. Expression of one of these DNA segments results in the production of sense RNA while expression of the other results in the production of an anti-sense RNA. RNA strands produced from vectors represented in Figures 5B-5F will thus have complementary nucleotide sequences and will generally hybridize either to each or intramolecularly under physiological conditions.

[0315] Nucleic acid segments designed to produce RNAi, such as the vectors represented in Figures 5B-5F, need not correspond to the full-length gene or open reading frame. For example, when the nucleic acid segment corresponds to an ORF, the segment may only correspond to part of the ORF (*e.g.*, 50 nucleotides at the 5' or 3' end of the ORF). Further, while Figures 5B-5F show vectors designed to produce RNAi, nucleic acid segments may also perform the same function in other forms (*e.g.*, when inserted into the chromosome of a host cell).

[0316] Gene silencing methods involving the use of compounds such as RNAi and antisense RNA, for examples, are particularly useful for identifying gene functions. More specifically, gene silencing methods can be used to reduce or prevent the expression of one or more genes in a cell or organism. Phenotypic manifestations associated with the selective inhibition of gene functions can then be used to assign role to the "silenced" gene or genes. As an example, Chuang, *et al.*, *Proc. Natl. Acad. Sci. (USA)* 97:4985-4990 (2000), have demonstrated that *in vivo* production of RNAi can alter gene activity in *Arabidopsis thaliana*. Thus, the invention provides methods for regulating expression of nucleic acid molecules in cells and tissues comprising the expression of RNAi and antisense RNA. The invention further provides methods for preparing nucleic acid molecules which can be used to produce RNA corresponding to one or both strands of a DNA molecule.

[0317] Further, viral vectors and/or nucleic acids molecules of the invention may be used to insert into cells nucleic acid segments that encode expression products involved in each step of particular biological pathways (*e.g.*, biosynthesis of amino acids such as lysine, threonine, etc.) or expression products involved in one or a few steps of such pathways. These nucleic acid molecules can be designed to, in effect, amplify genes encoding expression products in such pathways, insert genes into cells that encode expression products involved in pathways not normally found in the cells, or to replace one or more genes involved one or more steps of particular biological pathways in cells. Thus, gene therapy vectors of the invention may contain nucleic acid that results in the production one or more products (*e.g.*, one, two, three, four, five, eight, ten, fifteen, etc.). Such vectors, especially those that lead to the production of more than one product, will be particularly useful for the treatment of diseases and/or conditions that result from the expression and/or lack of expression of more than one gene or for the treatment of more than one diseases and/or conditions.

[0318] Thus, in related aspects, the invention provides gene therapy vectors that express one or more expression products (*e.g.*, one or more fusion proteins), methods for producing such vectors, methods for performing gene therapy using vectors of the invention, expression products of such vector

(e.g., encoded RNA and/or proteins), and host cells that contain vectors of the invention.

- [0319] For general reviews of the methods of gene therapy, see Goldspiel, *et al.*, *Clinical Pharmacy* 12:488-505 (1993); Wu and Wu, *Biotherapy* 3:87-95 (1991); Tolstoshev, *Ann. Rev. Pharmacol. Toxicol.* 32:573-596 (1993); Mulligan, *Science* 260:926-932 (1993); and Morgan and Anderson, *Ann. Rev. Biochem.* 62:191-217 (1993); May, *TIBTECH* 11(5):155-215 (1993)).

Methods commonly known in the art of recombinant DNA technology that can be used are described in Ausubel, *et al.* (eds.), 1993, *Current Protocols in Molecular Biology*, John Wiley & Sons, NY; and Kriegler, 1990, *Gene Transfer and Expression, A Laboratory Manual*, Stockton Press, NY.

- [0320] Delivery of the viral vectors and/or nucleic acids molecules of the invention into a patient may be either direct, in which case the patient is directly exposed to the nucleic acids and/or viral vectors of the invention, or indirect, in which case, cells are first transfected/transduced with the nucleic acid/viral vector *in vitro*, then transplanted into the patient. These two approaches are known, respectively, as *in vivo* or *ex vivo* gene therapy.

- [0321] In another specific embodiment, viral vectors that contain nucleic acid sequences encoding an antibody or other antigen-binding protein of the invention are used. The nucleic acid sequences encoding the antibody to be used in gene therapy are cloned into one or more viral vectors, which facilitates delivery of the gene into a patient.

- [0322] Adenoviruses are examples of viruses that can be used to prepare viral vectors that can be used in gene therapy. Adenoviral vectors are especially attractive vehicles for delivering genes to respiratory epithelia and the use of such vectors are included within the scope of the invention. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviral vectors have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, *Current Opinion in Genetics and Development* 3:499-503 (1993) present a review of adenovirus-based gene therapy. Bout, *et al.*, *Human Gene Therapy* 5:3-10 (1994) demonstrated the use of adenoviral vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of

adenoviral vectors in gene therapy can be found in Rosenfeld, *et al.*, *Science* 252:431-434 (1991); Rosenfeld, *et al.*, *Cell* 68:143- 155 (1992); Mastrangeli, *et al.*, *J. Clin. Invest.* 91:225-234 (1993); PCT Publication Nos. WO 94/12649 and WO 96/17053; U.S. Patent No. 5,998,205; and Wang, *et al.*, *Gene Therapy* 2:775-783 (1995), the disclosures of all of which are incorporated herein by reference in their entireties. In a one embodiment, adenoviral vectors are used for *in vivo* gene therapy.

[0323] Another approach to gene therapy involves transferring a gene to cells in tissue culture, for example, by infection with a viral vector of the present invention. The viral vector may contain a sequence encoding a therapeutic polypeptide or nucleic acid (*i.e.*, antisense molecule) and may further include a sequence encoding a selectable marker. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

[0324] In this embodiment, the viral vector is introduced into a cell prior to administration *in vivo* of the resulting recombinant cell. The resulting recombinant cells can be delivered to a patient by various methods known in the art. Recombinant blood cells (*e.g.*, hematopoietic stem or progenitor cells) will generally be administered intravenously. The amount of cells envisioned for use depends on the desired effect, patient state, etc., and can be determined by one skilled in the art.

[0325] Cells into which a viral vector can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as T-lymphocytes, B-lymphocytes, monocytes, macrophages, neutrophils, eosinophils, megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells (*e.g.*, as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, etc.).

[0326] In a certain embodiment, the cell used for gene therapy is autologous to the patient.

[0327] In an embodiment in which recombinant cells are used in gene therapy, viral vectors containing nucleic acids encoding an antibody or other antigen-binding protein are introduced into the cells such that they are expressible by

the cells and/or their progeny, and the recombinant cells are then administered *in vivo* for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells that can be isolated and maintained *in vitro* can potentially be used in accordance with this embodiment of the present invention (see, *e.g.*, PCT Publication WO 94/08598, dated April 28, 1994; Stemple and Anderson, *Cell* 71:973-985 (1992); Rheinwald, *Meth. Cell Bio.* 21A:229 (1980); and Pittelkow and Scott, *Mayo Clinic Proc.* 61:771 (1986)).

[0328] In a specific embodiment, viral vectors and/or nucleic acids molecules of the invention comprise nucleic acid sequences to be introduced for purposes of gene therapy under the control of an inducible promoter operably linked to the coding region, such that expression of the nucleic acid sequences is controllable by controlling the presence or absence of the appropriate inducer of transcription.

[0329] The viral vectors and/or nucleic acids molecules of the invention can also be used to produce transgenic organisms (*e.g.*, animals). Animals of any species, including, but not limited to, mice, rats, rabbits, hamsters, guinea pigs, pigs, micro-pigs, goats, sheep, cows and non-human primates (*e.g.*, baboons, monkeys, and chimpanzees) may be used to generate transgenic animals. Viruses capable of infecting the desired cell type are known to those skilled in the art and viral vectors based on these viruses may be used in the methods of the invention.

[0330] The present invention provides for transgenic organisms that carry the viral vectors and/or nucleic acids molecules of the invention or nucleic acid sequences provided by the viral vectors and/or nucleic acids molecules of the invention in all their cells, as well as organisms that carry these viral vectors or sequences in some, but not all, of their cells, *i.e.*, mosaic organisms or chimeric. The viral vectors and/or nucleic acids molecules of the invention may be integrated as a single copy or as multiple copies. The viral vectors and/or nucleic acids molecules of the invention may also be selectively introduced into and activated in a particular cell type by following, for example, the teaching of Lasko, *et al.* (Lasko, *et al.*, *Proc. Natl. Acad. Sci. USA* 89:6232-6236 (1992)). The regulatory sequences required for such a cell-type specific activation will depend upon the particular cell type of

interest, and will be apparent to those of skill in the art. When it is desired that the sequences of interest contained in the viral vectors and/or nucleic acids molecules of the invention be integrated into the chromosomal site of the endogenous gene, this will normally be done by gene targeting. Briefly, when such a technique is to be utilized, viral vectors containing some nucleotide sequences homologous to the endogenous gene are designed for the purpose of integrating, via homologous recombination with chromosomal sequences, into and disrupting the function of the nucleotide sequence of the endogenous gene. Viral vectors and/or nucleic acids molecules of the invention may also be selectively introduced into a particular cell type, thus inactivating the endogenous gene in only that cell type, by following, for example, the teaching of Gu, *et al.* (Gu, *et al.*, *Science* 265:103-106 (1994)). The regulatory sequences required for such a cell-type specific inactivation will depend upon the particular cell type of interest, and will be apparent to those of skill in the art. The contents of each of the documents recited in this paragraph is herein incorporated by reference in its entirety.

[0331] Once transgenic organisms have been generated, the expression of the recombinant gene may be assayed utilizing standard techniques. Initial screening may be accomplished by Southern blot analysis or PCR techniques to analyze organism tissues to verify that integration of nucleic acid molecules of the invention has taken place. The level of mRNA expression of nucleic acid sequences introduced by the viral vectors and/or nucleic acids molecules of the invention in the tissues of the transgenic organisms may also be assessed using techniques including, but not limited to, Northern blot analysis of tissue samples obtained from the organism, in situ hybridization analysis, and reverse transcriptase-PCR (RT-PCR). Samples of tissue that express the inserted sequences may also be evaluated immunocytochemically or immunohistochemically using antibodies specific for the expression product of these nucleic acid molecules.

[0332] Once the founder organisms are produced, they may be bred, inbred, outbred, or crossbred to produce colonies of the particular organism. Examples of such breeding strategies include, but are not limited to: outbreeding of founder organisms with more than one integration site in order to establish separate lines; inbreeding of separate lines in order to produce

compound transgenic organisms that express sequences of interest at higher levels because of the effects of additive expression of each copy of nucleic acid molecules of the invention; crossing of heterozygous transgenic organisms to produce organisms homozygous for a given integration site in order to both augment expression and eliminate the need for screening of organisms by DNA analysis; crossing of separate homozygous lines to produce compound heterozygous or homozygous lines; and breeding to place the nucleic acid molecules of the invention on a distinct background that is appropriate for an experimental model of interest.

[0333] Transgenic and "knock-out" organisms of the invention have uses that include, but are not limited to, model systems (*e.g.*, animal model systems) useful in elaborating the biological function of expression products of sequences of interest, studying conditions and/or disorders associated with aberrant expression of expression products of sequences of interest, and in screening for compounds effective in ameliorating such conditions and/or disorders.

[0334] As one skilled in the art would recognize, in many instances when viral vectors containing sequences of interest are introduced into metazoan organisms, it will be desirable to operably link the sequences that encode expression products to tissue-specific transcriptional regulatory sequences (*e.g.*, tissue-specific promoters) where production of the expression product is desired. Such promoters can be used to facilitate production of these expression products in desired tissues. A considerable number of tissue-specific promoters are known in the art.

Host Cells

[0335] The invention also relates to host cells comprising one or more of the viral vectors and/or nucleic acids molecules of the invention containing one or more sequences of interest (*e.g.*, two, three, four, five, seven, ten, twelve, fifteen, twenty, thirty, fifty, etc.), particularly those viral vectors described in detail herein. Representative host cells that may be used according to this aspect of the invention include, but are not limited to, bacterial cells, yeast cells, plant cells and animal cells. Preferred bacterial host cells include *Escherichia* spp. cells (particularly *E. coli* cells and most particularly *E. coli*

strains DH10B, Stbl2, DH5 α , DB3, DB3.1 (preferably *E. coli* LIBRARY EFFICIENCY \circledR DB3.1TM Competent Cells; Invitrogen Corporation, Carlsbad, CA), DB4, DB5, JDP682 and *ccdA*-over (see U.S. Application No. 09/518,188, filed March 2, 2000, and U.S. provisional Application No. 60/475,004, filed June 3, 2003, by Louis Leong *et al.*, entitled "Cells Resistant to Toxic Genes and Uses Thereof," the disclosures of which are incorporated by reference herein in their entireties); a DB3 cell (deposit number NRRL B-30097), a DB3.1 cell (deposit number NRRL B-30098), a DB4 cell (deposit number NRRL B-30106), a DB5 cell (deposit number NRRL B-30107), a JDP682 cell (deposit number NRRL B-30667), a *ccdA*-over cell (deposit number NRRL B-30668), or a mutant or derivative thereof; *Bacillus* spp. cells (particularly *B. subtilis* and *B. megaterium* cells), *Streptomyces* spp. cells, *Erwinia* spp. cells, *Klebsiella* spp. cells, *Serratia* spp. cells (particularly *S. marcessans* cells), *Pseudomonas* spp. cells (particularly *P. aeruginosa* cells), and *Salmonella* spp. cells (particularly *S. typhimurium* and *S. typhi* cells). Preferred animal host cells include insect cells (most particularly *Drosophila melanogaster* cells, *Spodoptera frugiperda* Sf9 and Sf21 cells and *Trichoplusia* High-Five cells), nematode cells (particularly *C. elegans* cells), avian cells, amphibian cells (particularly *Xenopus laevis* cells), reptilian cells, and mammalian cells (most particularly NIH3T3, 293, CHO, COS, VERO, BHK and human cells). Preferred yeast host cells include *Saccharomyces cerevisiae* cells and *Pichia pastoris* cells. These and other suitable host cells are available commercially, for example, from Invitrogen Corporation, (Carlsbad, CA), American Type Culture Collection (Manassas, Virginia), and Agricultural Research Culture Collection (NRRL; Peoria, Illinois).

[0336] Nucleic acid molecules to be used in the present invention may comprise one or more origins of replication (ORIs), and/or one or more selectable markers. In some embodiments, molecules may comprise two or more ORIs at least two of which are capable of functioning in different organisms (*e.g.*, one in prokaryotes and one in eukaryotes). For example, a nucleic acid may have an ORI that functions in one or more prokaryotes (*e.g.*, *E. coli*, *Bacillus*, etc.) and another that functions in one or more eukaryotes (*e.g.*, yeast, insect, mammalian cells, etc.). Selectable markers may likewise be included in nucleic acid molecules of the invention to allow selection in

different organisms. For example, a nucleic acid molecule may comprise multiple selectable markers, one or more of which functions in prokaryotes and one or more of which functions in eukaryotes.

[0337] Methods for introducing the viral vectors and/or nucleic acids molecules of the invention into the host cells described herein, to produce host cells comprising one or more of the viral vectors and/or nucleic acids molecules of the invention, will be familiar to those of ordinary skill in the art. For instance, the nucleic acid molecules and/or viral vectors of the invention may be introduced into host cells using well known techniques of infection, transduction, electroporation, transfection, and transformation. The nucleic acid molecules and/or viral vectors of the invention may be introduced alone or in conjunction with other nucleic acid molecules and/or vectors and/or proteins, peptides or RNAs. Alternatively, the nucleic acid molecules and/or viral vectors of the invention may be introduced into host cells as a precipitate, such as a calcium phosphate precipitate, or in a complex with a lipid. Electroporation also may be used to introduce the nucleic acid molecules and/or viral vectors of the invention into a host. Likewise, such molecules may be introduced into chemically competent cells such as *E. coli*. If the vector is a virus, it may be packaged *in vitro* or introduced into a packaging cell and the packaged virus may be transduced into cells. Thus nucleic acid molecules of the invention may contain and/or encode one or more packaging signal (*e.g.*, viral packaging signals that direct the packaging of viral nucleic acid molecules). Hence, a wide variety of techniques suitable for introducing the nucleic acid molecules and/or vectors of the invention into cells in accordance with this aspect of the invention are well known and routine to those of skill in the art. Such techniques are reviewed at length, for example, in Sambrook, J., *et al.*, *Molecular Cloning, a Laboratory Manual*, 2nd Ed., Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press, pp. 16.30-16.55 (1989), Watson, J.D., *et al.*, *Recombinant DNA*, 2nd Ed., New York: W.H. Freeman and Co., pp. 213-234 (1992), and Winnacker, E.-L., *From Genes to Clones*, New York: VCH Publishers (1987), which are illustrative of the many laboratory manuals that detail these techniques and which are incorporated by reference herein in their entireties for their relevant disclosures.

Kits

[0338] In another aspect, the invention provides kits that may be used in conjunction with methods the invention. Kits according to this aspect of the invention may comprise one or more containers, which may contain one or more components selected from the group consisting of one or more nucleic acid molecules (*e.g.*, one or more nucleic acid molecules comprising one or more viral sequences and /or one or more recombination sites) and/or viral vectors of the invention, one or more primers, the molecules and/or compounds of the invention, one or more polymerases, one or more reverse transcriptases, one or more recombination proteins (or other enzymes for carrying out the methods of the invention), one or more ligases, one or more buffers, one or more detergents, one or more restriction endonucleases, one or more nucleotides, one or more terminating agents (*e.g.*, ddNTPs), one or more transfection reagents, pyrophosphatase, and the like.

[0339] A wide variety of nucleic acid molecules and/or viral vectors of the invention can be used with the invention. Further, due to the modularity of the invention, these nucleic acid molecules can be combined in wide range of ways. Examples of nucleic acid molecules that can be supplied in kits of the invention include those that contain promoters, signal peptides, enhancers, repressors, selection markers, transcription signals, translation signals, primer hybridization sites (*e.g.*, for sequencing or PCR), recombination sites, restriction sites and polylinkers, sites that suppress the termination of translation in the presence of a suppressor tRNA, suppressor tRNA coding sequences, sequences that encode domains and/or regions (*e.g.*, 6 His tag) for the preparation of fusion proteins, origins of replication, telomeres, centromeres, and the like. Similarly, libraries can be supplied in kits of the invention. These libraries may be in the form of replicable nucleic acid molecules or they may comprise nucleic acid molecules that are not associated with an origin of replication. As one skilled in the art would recognize, the nucleic acid molecules of libraries, as well as other nucleic acid molecules that are not associated with an origin of replication, either could be inserted into other nucleic acid molecules that have an origin of replication or would be an expendable kit components.

[0340] Further, in some embodiments, libraries supplied in kits of the invention may comprise two components: (1) the nucleic acid molecules of these libraries and (2) 5' and/or 3' recombination sites. In some embodiments, when the nucleic acid molecules of a library are supplied with 5' and/or 3' recombination sites, it will be possible to insert these molecules into nucleic acid molecules comprising all or a portion of a viral genome, which also may be supplied as a kit component, using recombination reactions. In other embodiments, recombination sites can be attached to the nucleic acid molecules of the libraries before use (*e.g.*, by the use of a ligase, which may also be supplied with the kit). In such cases, nucleic acid molecules that contain recombination sites or primers that can be used to generate recombination sites may be supplied with the kits.

[0341] Nucleic acid molecules comprising all or a portion of a viral genome to be supplied in kits of the invention can vary greatly. In some instances, these molecules will contain an origin of replication, at least one selectable marker, and at least one recombination site. For example, molecules supplied in kits of the invention can have four separate recombination sites that allow for insertion of sequence of interest at two different locations of a nucleic acid molecule, for example, as shown in Fig. 2. Other attributes of vectors supplied in kits of the invention are described elsewhere herein.

[0342] In some embodiments, the kits of the invention may comprise a plurality of containers, each container comprising one or more nucleic acid segments comprising viral sequences and/or one or more recombination sites and/or topoisomerase recognition sites. Segments may be provided with recombination sites such that a series of segments (*e.g.*, two, three, four, five, six, seven, eight, nine, ten, etc.) may be combined in order to construct a viral vector or other nucleic acid molecule of the present invention. Segments may be combined in reactions involving two or more segments (*e.g.*, three, four, five, six, seven, eight, nine, ten, etc.). Each individual segment may be, independently of any other segment, from about 100 bp to about 35 kb in length, or from about 100 bp to about 20 kb in length, or from about 100 bp to about 10 kb in length, or from about 100 bp to about 5 kb in length, or from about 100 bp to about 2.5 kb in length, or from about 100 bp to about 1 kb in length, or from about 100 bp to about 500 bp in length. The present invention

also contemplates methods for assembling and using such segments, nucleic acid molecules assembled by such methods, and compositions comprising such nucleic acid molecules.

[0343] Segments may be prepared so as to contain viral transcription units. For example, when an adenoviral vector is to be prepared, one segment may comprise, in addition to one or more recombination sites and/or one or more topoisomerase recognition sites, sequences corresponding to the E1 region, the E2 region, the E3 region, and/or the E4 region. Other segments may comprise sequences corresponding to one or more late transcription units and/or viral inverted terminal repeats. Segments comprising nucleic acid sequences of interest may be prepared so as to construct a viral vector or other nucleic acid molecule in which one or more viral nucleic acid sequences, present in a wild-type virus, are not present in the viral vector. Segments comprising a nucleic acid sequence of interest may be prepared and inserted into a viral vector in place of one or more segments comprising viral sequences. In some embodiments, sequences that are present in a wild-type virus but not present in the viral vectors of the invention are those that are not required for replication in cultured cells. For example, a segment comprising a nucleic acid sequence of interest may be used to construct an adenoviral vector wherein the nucleic acid sequence of interest replaces one or more of the E1 region and/or the E3 region. Where necessary (e.g., in the case of the E1 functions) viral functions required to support replication of the viral vector may be supplied in trans (e.g., from the genome of the host cell). Segments may be prepared to construct viral vectors wherein a nucleic acid sequence of interest is placed in the viral genome in a position known to be tolerant of nucleic acid insertions, for example, upstream of the E4 region.

[0344] A kit of the present invention may comprise a container containing a nucleic acid molecule comprising all or a portion of a viral genome and comprising two recombination sites that do not recombine with each other. The recombination sites may flank a selectable marker that allows selection for or against the presence of the nucleic acid molecule in a host cell or identification of a host cell containing or not containing the nucleic acid. A nucleic acid molecule to be included in a kit may comprise more than two recombination sites, for example, a nucleic acid molecule may comprise

multiple pairs of recombination sites (e.g., two, three, four, five, six, seven, eight, nine, ten, etc.) where members of a pair of recombination sites do not recombine or substantially recombine with each other. In some embodiments, members of one pair of recombination sites do not recombine with members of another pair present in the same nucleic acid molecule.

[0345] Kits of the invention may comprise containers containing one or more recombination proteins. Suitable recombination proteins have been disclosed above and include, but are not limited to, Cre, Int, IHF, Xis, Flp, Fis, Hin, Gin, Cin, Tn3 resolvase, Φ C31, TndX, XerC, and XerD.

[0346] Kits of the invention may also comprise one or more topoisomerase proteins and/or one or more nucleic acids comprising one or more topoisomerase recognition sequence. Suitable topoisomerases include Type IA topoisomerases, Type IB topoisomerases and/or Type II topoisomerases. Suitable topoisomerases include, but are not limited to, poxvirus topoisomerases, including vaccinia virus DNA topoisomerase I, *E. coli* topoisomerase III, *E. coli* topoisomerase I, topoisomerase III, eukaryotic topoisomerase II, archeal reverse gyrase, yeast topoisomerase III, *Drosophila* topoisomerase III, human topoisomerase III, *Streptococcus pneumoniae* topoisomerase III, bacterial gyrase, bacterial DNA topoisomerase IV, eukaryotic DNA topoisomerase II, and T-even phage encoded DNA topoisomerases, and the like. Suitable recognition sequences have been described above.

[0347] In use, a nucleic acid molecule comprising all or a portion of a viral genome provided in a kit of the invention may be combined with a nucleic acid molecule comprising a sequence of interest using recombinational cloning. The nucleic acid molecule comprising all or a portion of a viral genome may be provided, for example, with two recombination sites that do not recombine with each other. The nucleic acid molecule comprising a sequence of interest may also be provided with two recombination sites, each of which is capable of recombining with one of the two sites present on the a nucleic acid molecule comprising all or a portion of a viral genome. In the presence of the appropriate recombination proteins, the nucleic acid molecule reacts with the nucleic acid molecule comprising all or a portion of a viral genome in order to form a recombinant nucleic acid molecule containing the

sequence of interest and all or a portion of a viral genome. When the nucleic acid molecule comprising all or a portion of a viral genome comprises multiple pairs of recombination sites, multiple nucleic acid molecules comprising sequences of interest, which may be the same or different, may be combined with the nucleic acid molecule comprising all or a portion of a viral genome in order to form a nucleic acid molecule comprising all or a portion of a viral genome and also comprises multiple sequence of interest.

[0348] Kits of the invention can also be supplied with primers. These primers will generally be designed to anneal to molecules having specific nucleotide sequences. For example, these primers can be designed for use in PCR to amplify a particular nucleic acid molecule. Further, primers supplied with kits of the invention can be sequencing primers designed to hybridize to vector sequences. Thus, such primers will generally be supplied as part of a kit for sequencing nucleic acid molecules that have been inserted into a vector.

[0349] One or more buffers (*e.g.*, one, two, three, four, five, eight, ten, fifteen) may be supplied in kits of the invention. These buffers may be supplied at a working concentrations or may be supplied in concentrated form and then diluted to the working concentrations. These buffers will often contain salt, metal ions, co-factors, metal ion chelating agents, etc. for the enhancement of activities of the stabilization of either the buffer itself or molecules in the buffer. Further, these buffers may be supplied in dried or aqueous forms. When buffers are supplied in a dried form, they will generally be dissolved in water prior to use.

[0350] Kits of the invention may contain virtually any combination of the components set out above or described elsewhere herein. As one skilled in the art would recognize, the components supplied with kits of the invention will vary with the intended use for the kits. Thus, kits may be designed to perform various functions set out in this application and the components of such kits will vary accordingly.

[0351] Kits of the invention may comprise one or more pages of written instructions for carrying out the methods of the invention. For example, instructions may comprise methods steps necessary to carry out recombinational cloning of an ORF provided with recombination sites and a

vector also comprising recombination sites and optionally further comprising one or more functional sequences.

[0352] It will be understood by one of ordinary skill in the relevant arts that other suitable modifications and adaptations to the methods and applications described herein are readily apparent from the description of the invention contained herein in view of information known to the ordinarily skilled artisan, and may be made without departing from the scope of the invention or any embodiment thereof. Having now described the present invention in detail, the same will be more clearly understood by reference to the following examples, which are included herewith for purposes of illustration only and are not intended to be limiting of the invention.

[0353] The entire disclosures of U.S. Appl. No. 08/486,139, (now abandoned), filed June 7, 1995, U.S. Appl. No. 08/663,002, filed June 7, 1996 (now U.S. Patent No. 5,888,732), U.S. Appl. No. 09/233,492, filed January 20, 1999, (now U.S. Patent No. 6,270,969), U.S. Appl. No. 09/233,493, filed January 20, 1999, (now U.S. Patent No. 6,143,557), U.S. Appl. No. 09/005,476, filed January 12, 1998, (now U.S. Patent No. 6,171,861), U.S. Appl. No. 09/432,085 filed November 2, 1999, U.S. Appl. No. 09/498,074 filed February 4, 2000, U.S. Appl. No. 60/065,930, filed October 24, 1997, U.S. Appl. No. 09/177,387, filed October 23, 1998, U.S. Appl. No. 09/296,280, filed April 22, 1999, (now U.S. Patent No. 6,277,608), U.S. Appl. No. 09/296,281, filed April 22, 1999, (now abandoned), U.S. Appl. No. 09/648,790, filed August 28, 2000, U.S. Appl. No. 09/732,914 (published as US 2002 0007051), filed December 11, 2000, U.S. Appl. No. 09/855,797, filed May 16, 2001, U.S. Appl. No. 09/907,719, filed July 19, 2001, U.S. Appl. No. 09/907,900, filed July 19, 2001, U.S. Appl. No. 09/985,448, filed November 2, 2001, U.S. Appl. No. 60/108,324, filed November 13, 1998, U.S. Appl. No. 09/438,358, filed November 12, 1999, U.S. Appl. No. 60/161,403, filed October 25, 1999, U.S. Appl. No. 09/695,065, filed October 25, 2000, U.S. Appl. No. 09/984,239, filed October 29, 2001, U.S. Appl. No. 60/122,389, filed March 2, 1999, U.S. Appl. No. 60/126,049, filed March 23, 1999, U.S. Appl. No. 60/136,744, filed May 28, 1999, U.S. Appl. No. 09/517,466, filed March 2, 2000, U.S. Appl. No. 60/122,392, filed March 2, 1999, U.S. Appl. No. 09/518,188, filed March 2, 2000, U.S. Appl. No. 60/169,983, filed December 10, 1999, U.S. Appl. No.

60/188,000, filed March 9, 2000, U.S. Appl. No. 09/732,914, filed December 11, 2001, U.S. Appl. No. 60/284,528, filed April 19, 2001, U.S. Appl. No. 60/291,973, filed May 21, 2001, U.S. Appl. No. 60/318,902, filed September 14, 2001, U.S. Appl. No. 60/333,124, filed November 27, 2001, and U.S. Appl. No. 10/005,876, filed December 7, 2001, are herein incorporated by reference.

EXAMPLES

[0354] The present invention provides an extremely versatile method for the modular construction of nucleic acids and production of polypeptides. Both insert nucleic acid segments and the vector can contain sequences selected so as to confer desired characteristics on the product molecules. In some embodiments, in addition to the insert, one or more of the portions of the nucleic acid comprising all or a portion of a viral genome adjacent to the insert, can contain one or more selected sequences. The selected sequences might encode ribozymes, epitope tags, structural domains, selectable markers, internal ribosome entry sequences, promoters, enhancers, recombination sites and the like.

[0355] In some embodiments, more than one sequence of interest may be incorporated in a nucleic acid molecule comprising all or a portion of a viral genome. The incorporated sequences of interest may be adjacent to one another or may be separated by a portion of the nucleic acid molecule comprising all or a portion of a viral genome. When separated, the portion of the nucleic acid molecule separating the sequences of interest may comprise one or more selectable markers flanked by a reactive pair of recombination sites in addition to containing the recombination sites used to insert the nucleic acid segments. The portion of the nucleic acid molecule separating the sequences of interest may also comprise viral sequences and/or other sequences conferring a desired characteristic on the nucleic acid molecule and/or sequences of interest.

[0356] A sequence of interest may be a sequence of any type. For example, the sequence may encode one or more polypeptides and/or may contain one or more un-translated regions. Sequences of interest may be transcribed and translated into polypeptides or may be transcribed and not translated into

polypeptides, for example, anti-sense molecules, ribozymes, and RNAi. Sequences of interest may or may not comprise a stop codon. Sequences comprising a stop codon may or may not comprise additional sequences 3' to the stop codon that may be in frame with sequences 5' to the stop codon. In some embodiments, stop codons may be suppressed in order to produce a fusion polypeptide.

[0357] Throughout this disclosure, the term gene of interest (GOI) may be used for the sake of convenience. This should not be construed as limiting the present invention to nucleic acid sequences comprising genes. Any nucleic acid sequence of interest can be inserted into a vector of the invention using materials and methods described herein.

EXAMPLE 1

Preparation of a viral vector of the invention.

[0358] Fig. 6 is a plasmid map of the pAd/CMV/V5-DEST vector, one example of a nucleic acid comprising all or a portion of a viral genome according to the present invention. The nucleotide sequence of the plasmid is provided in Table 6 (SEQ ID NO:). The plasmid contains the first 458 nucleotides of Ad5, including the left ITR and packaging sequence, followed the cytomegalovirus promoter (CMV) and the T7 promoter. The promoters are followed by a sequence containing selectable markers flanked by recombination sites attR1 and attR2. Any other suitable pair of recombination sites might be employed as long as they are selected so as not to recombine with each other. After the attR2 site, the V5 epitope coding sequence is followed by stop codons in all three reading frames and the herpes virus thymidine kinase polyadenylation signal. This is followed by the nucleotides from position 3513 to the right end of the adenoviral genome including the right ITR. After the adenoviral sequences, are plasmid sequences including a plasmid origin of replication followed by the ampicillin resistance gene. The plasmid sequences are flanked by PacI restriction enzyme recognition sites. Thus, after replacement of the replaceable sequence with a sequence of interest flanked by attL1 and attL2 in a recombination reaction, an infectious viral genome can be prepared by digestion of the recombination reaction

product with PacI to remove the plasmid sequences. In this particular embodiment, the viral genome is an adenoviral genome deleted in the E1 and E3 regions. The E1 function must be supplied *in trans* in order for the virus to replicate, for example, from the host cell as in 293 cells. The gene products of the E3 region are not required for replication.

[0359] In order to prepare a viral vector according to the present invention, a particular sequence of interest may be prepared with recombination sites compatible to those in the pAd/CMV/V5-DEST vector. This may be accomplished using standard techniques, for example, by amplifying a sequences of interest with primers comprising the appropriate recombination site sequences. If a PCR product contains the appropriate recombination site sequences, it may be used directly in a recombination reaction. Optionally, a PCR product or other nucleic acid comprising the sequence of interest may be cloned into a GATEWAY™ entry vector. This can be accomplished using any conventional technique, for example, by a) traditional restriction fragment ligation, b) TOPO-mediated cloning of the nucleic acid comprising the sequence of interest into pENTR-dTOPO, or c) GATEWAY™ clonase reaction between PCR-amplified sequence of interest (*e.g.*, gene of interest (GOI)) containing flanking attB sites with pDONR DNA. Any of these three methods will result in the sequence of interest being inserted into an entry vector. Using the terminology of the GATEWAY™ Technology, the resultant vector would be designated pENTR-GOI for an entry vector comprising a gene of interest (GOI). This should not be construed as limiting the sequences of interest to those encoding genes; any sequence of interest may be inserted into a pENTR vector in this fashion. In this example, this results in the sequence of interest being flanked by attL1 and attL2 recombination sites.

[0360] In an *in vitro* GATEWAY™ LR reaction, the pENTR-GOI vector may be combined with pAd-CMV-DEST. The reaction may be incubated for an appropriate period of time, for example, 1 hour at room temperature. This reaction moves the sequence of interest into the adenoviral vector, pAd-CMV-DEST.

[0361] The adenoviral vector containing a sequence of interest is used to transform competent bacteria (*i.e.*, DH5 α , TOP10, HB101, etc.). All or a portion of the LR reaction mixture is used to transform competent bacteria and

the transformed bacteria are plated on LB-ampicillin bacterial plates and incubated overnight at 37°C.

[0362] Several bacterial colonies—2-4 is usually sufficient— may be picked and used to inoculate overnight cultures in LB-ampicillin liquid medium and grown overnight at 37°C.

[0363] Plasmid DNA is prepared from the cultures using conventional techniques and analyzed for the presence of the sequence of interest, for example, by restriction enzyme digests or PCR.

[0364] To prepare a larger quantity of viral vector, 2 to 5 micrograms of destination vector comprising the sequence of interest may be digested with *PacI* restriction enzyme to expose the adenoviral ITRs (immediately adjacent to the *PacI* sites on the 5' and 3' ends of the adenoviral genome). The digested DNA may be purified using any conventional technique, for example, phenol/chloroform extraction followed by ethanol precipitation, or use of a commercially available kit for this purpose.

[0365] The digested DNA is used to transfect an appropriate host cell, for example, 293 cells. The day before transfection, 6 well plates with 5×10^5 293 cells per well may be prepared. On the day of transfection, 2 micrograms of DNA is used to transfect the cells in each well. Transfection may be accomplished using standard techniques using, for example, calcium phosphate, lipids, electroporation, etc. Preferred methods of transfection include those utilizing cationic lipids or mixtures of cationic and neutral lipids. Suitable transfection reagents are commercially available, for example, from Invitrogen Corporation, Carlsbad, Ca. One suitable lipid formulation is Lipofectamine™ 2000.

[0366] The day after transfection, the transfection media may be removed and replaced with fresh media. The next day, the transfected cells may be trypsinized and transferred. The cells from one well are used to seed a 100 mm dish. The cells are grown in the 100 mm dish for 7-10 days. The media is replaced with fresh media every 2-3 days. At about 9 days post transfection, "plaques" may be observed forming in the monolayer of 293 cells. Plaques will appear as cleared areas when viewed by the naked eye. Under the microscope, plaques will be fringed with rounded, lysing cells. This is

referred to as cytopathic effect (CPE). The media should be replaced with fresh media every 2 days until most of the cells are demonstrating CPE.

[0367] Harvest the plate by squirting off the cells using the growth media and transfer the cells and media to a 15 ml tube. Freeze/thaw the tube 3 times by alternating -80°C and 37°C . This releases the viral particles from the cells. Centrifuge the tube to remove the unwanted cellular debris (3000 rpm x 10 minutes). Remove the supernatant and transfer it to a fresh tube. This material now contains recombinant adenoviral vector containing the sequence of interest. This can be used directly in experiments to deliver the sequence of interest.

[0368] To increase the titer of the viral vector, the viral vector may be amplified, for example, by applying a small amount (typically 100 microliters) of the initial viral vector to a fresh plate of 293 cells (typically 5×10^6 293 cells in a 100 mm dish). Infection of the cells occurs within the first couple hours and three days later CPE is observed throughout the plate. Viral vector is harvested as described above.

[0369] Viral vector produced in this way (called "crude viral lysates", or CVLs) is typically high titer ($>10^9$ infectious virus/ml) and can be used directly for most applications. To determine the exact titer of the CVL (or of any adenoviral stock), 293 cells are plated at 1×10^6 cells per well in 6-well plates. The next day, each well is transduced with 1 ml media containing ten-fold serial dilutions of CVL ranging from 10^{-5} to 10^{-10} . After overnight incubation, the media is removed and the cell monolayers are overlaid with 2 ml of fresh media containing 0.4% Ultrapure agarose. This semi-solid medium prevents viral vector from diffusing throughout the plate and keeps individual plaques distinct. After 7 to 10 days, distinct plaques will be visible to the naked eye. MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide) can be used to stain the wells to aid in plaque visualization. Plaques are counted, and that number is multiplied by the dilution factor to obtain the titer of infectious viral vector present in the original CVL. If higher titer viral vector is required, the viral vector in the CVLs can be concentrated and purified using a number of different approaches including: cesium chloride density ultracentrifugation, HPLC, or commercially available columns

designed for virus purification (*e.g.* Virapur). These methods typically result in titers of $>10^{11}$ infectious virus/ml.

EXAMPLE 2

Use of Suppressor tRNAs to Generate Fusion Polypeptides

- [0370] Detection of expressed polypeptides is often facilitated by the use of epitope tags (*e.g.* V5 or myc) or detectable markers (*e.g.*, β -lactamase, β -galactosidase, β -glucuronidase, GFP, etc.). This is especially useful if there is no specific antibody available for the polypeptide of interest. However, addition of epitope tags and/or fusion to a detectable marker may adversely affect polypeptide activity, structure, or its interaction with other molecules. One common approach to this problem is to clone the gene of interest twice: with and without the tag.
- [0371] The present invention provides materials and methods to express a polypeptide with and without a tag or marker from the same genetic construct. This is accomplished using mammalian suppressor tRNAs that specifically recognize and decode one of the three stop codons (Ochre, Amber, and Opal) and result in the insertion of an amino acid at the position coded for by the stop codon. The suppressor tRNAs may insert any amino acid into the position coded for by the stop codon. In the specific embodiments described below, the amino acid serine was inserted; however, any amino acid desired can be inserted by preparing and expressing the appropriate suppressor tRNA according to the present invention.
- [0372] Expression plasmids encoding a reporter gene with all three possible stop codons in frame with C-terminal tags were constructed. Following delivery of suppressor tRNAs in trans, the stop codons between the gene and the epitope tag were suppressed, allowing translation of the 3' sequences.
- [0373] Plasmids encoding each suppressor tRNA were co-transfected with the corresponding expression plasmid to test the efficiency of suppression. Suppression of TAA and TAG were approximately 50% to 60% efficient, while TGA was only 30%. Changing the nucleotide following the TGA stop codon from an adenine to a cytosine improved suppression to about 70%.

[0374] A recombinant adenoviral vector was constructed that expresses a suppressor tRNA. A map of a plasmid containing the adenoviral construct pAd-GW-TO/tRNA in which a suppressor tRNA is under the control of a tetracycline-inducible CMV promoter is shown in Fig. 7. The nucleotide sequence of pAd-GW-TO/tRNA is provided in Table 7 (SEQ ID NO:). An additional adenoviral construct expressing a suppressor tRNA is pAdenoTAG tRNA shown in Fig. 8. The nucleotide sequence of pAdenoTAG tRNA is provided in Table 8. Table 9 provides the nucleotide sequence of a Sau3A fragment that may be used to construct suppressor tRNA containing nucleic acid molecules of the invention (*e.g.*, pAdenoTag tRNA.) A transcription terminator is located at bases 600 to 606 of the fragment, the sequence corresponding to the suppressor tRNA is located at bases 512 to 593 of the fragment, the anti-codon is located at bases 545 to 547, and the tetracycline operator sequence is located at bases 474 to 511. The suppressor tRNA produced from this sequence suppresses the amber stop codon UAG. Those skilled in the art will appreciate that it is possible to prepare suppressors for opal and ochre stop codons by mutating the bases in the anti-codon to make the anti-codon the reverse complement of the stop codon. *i.e.*, TCA for the opal stop codon and TTA for the ochre stop codon. Other anti-codons may be used, for example, those employing other bases in the wobble position. Constructing a suitable sequence from which to produce a desired suppressor tRNA (*e.g.*, by introducing one or more point mutations in a sequence) is routine in the art.

[0375] The plasmid may be digested with PacI to generate an infectious adenoviral genome. The viral vector expressing the suppressor tRNA may be used in conjunction with any vector comprising a sequence with a stop codon to be suppressed. In some embodiments, a viral vector expressing a suppressor tRNA and a viral vector comprising a sequence of interest may be used to co-infect a cell and produce a fusion polypeptide. A fusion polypeptide may be encoded entirely by the sequence of interest, for example, the sequence may have one open reading frame (ORF) separated from another ORF by a stop codon. Alternatively, one ORF may be present on the sequence of interest and one or more additional ORFs may be present on the viral vector. Co-infection with a suppressor-expressing viral vector an expression

vector will result in the expression of a fusion polypeptide; infection without the suppressor-expressing viral vector will produce a native polypeptide. Thus, the suppression technology allows expression of tagged and untagged polypeptides using a single expression vector.

EXAMPLE 3

Detailed materials and method for construction of adenoviral vectors and kits.

[0376] Kits of the invention may comprise one or more sets of instructions for carrying out the methods of the invention. For example, the instructions may related to the propagation of cells used in the methods of the invention and/or to conducting individual reactions that are part of the methods. In a one embodiments, kits of the invention may comprise instructions for growth and maintenance of cell used in methods of the invention (*e.g.*, the 293A cell line manual, catalog no. R705-07 version B, Invitrogen Corporation, Carlsbad, CA) and a manual for the preparation of the viral vectors of the invention (*e.g.*, the ViraPower™ Adenoviral Expression System manual, catalog no. K4930-00, version A, Invitrogen Corporation, Carlsbad, CA).

[0377] In one embodiment, a kit of the invention may comprise the necessary reagents and instructions to prepare a viral vector according to the invention. Such a kit may comprise one or more components selected from the group consisting of : the ViraPower™ Adenoviral GATEWAY™ Expression Kit, ViraPower™ Adenoviral Promoterless GATEWAY™ Expression Kit, pAd/CMV/V5-DEST™ GATEWAY™ Vector Pack, or pAd/PL-DEST™ GATEWAY™ Vector Pack all available from Invitrogen Corporation, Carlsbad, CA.

[0378] A plasmid map of pAd/PL-DEST™ is provided in Figure 9 and the sequence of the plasmid is provided in Table 10.

[0379] A kit may also comprise one or more control reagents. For example, a kit may comprise an adenoviral vector comprising a detectable marker that may be used as a control for transfection of cells and infection of cells. One suitable control reagent is pAd/CMV/V5-GW/*lacZ* control. A map of the pAd/CMV/V5-GW/*lacZ* plasmid is provide as Fig. 10 and the nucleotide sequence of the plasmid is provided as Table 11.

[0380] Kits of the invention may comprise one or more additional products (*e.g.*, accessory products). Such products include, but are not limited to, reagents and materials for purifying nucleic acids (*e.g.*, plasmid purification), host cells for propagating plasmids and/or viruses (*e.g.*, *E. coli* and 293 cells), transfection reagents (*e.g.*, lipids), reagents for assaying control vector expression (*e.g.*, β -lactamase assay reagents, β -galactosidase assay reagents, antibodies to β -galactosidase), recombination polypeptides, and antibiotics for selection of transformed cells. The contents of one suitable kit include, ViraPower™ Adenoviral GATEWAY™ Expression Kit, ViraPower™ Adenoviral Promoterless GATEWAY™ Expression Kit, 293A Cell Line, GATEWAY™ LR Clonase™ Enzyme Mix, Library Efficiency® DB3.1™ Competent Cells, One Shot® TOP10 Chemically Competent *E. coli*, S.N.A.P.™ MidiPrep Kit, Lipofectamine™ 2000, β -gal Antiserum, and Ampicillin all available from Invitrogen Corporation, Carlsbad, CA.

[0381] A polypeptide encoded by a sequence of interest may be expressed as a fusion polypeptide with a detectable epitope. For example, a polypeptide expressed from pAd/CMV/V5-DEST™ (Fig. 6), can be detected with an antibody to the V5 epitope. Antibodies to the detectable epitope may be labeled, for example, horseradish peroxidase (HRP) or alkaline phosphatase (AP) may be conjugated to the antibody to allow one-step detection using chemiluminescent or colorimetric detection methods. A fluorescent label, (*e.g.*, FITC) may be conjugated to the antibody to allow one-step detection in immunofluorescence experiments. Thus, kits of the invention may comprise one or more antibodies to one or more detectable epitopes. Antibodies to detectable epitopes may be labeled. Suitable antibodies include, but are not limited to, an anti-V5 antibody, an anti-V5-HRP antibody, an anti-V5-AP antibody, and/or an anti-V5-FITC antibody.

[0382] Examples of nucleic acid molecules of the invention include pAd/CMV/V5-DEST™ (36.7 kb) and pAd/PL-DEST™ (34.9 kb), which are destination vectors adapted for use with recombinational cloning (*e.g.*, GATEWAY™ Technology), and are designed to allow high-level, transient expression of recombinant fusion polypeptides in dividing and non-dividing mammalian cells, for example, using ViraPower™ Adenoviral Expression

System, catalog nos. K4930-00 and K4940-00 available from Invitrogen Corporation, Carlsbad, CA.

[0383] A choice of vectors permits the construction of an adenovirus expressing a sequence of interest. Each vector provides different features that may be useful under different circumstances. For example, the pAd/CMV/V5-DEST™ vector contains the CMV promoter that provides high-level, constitutive expression of the sequence of interest and the C-terminal V5 epitope for detection of recombinant polypeptide using anti-V5 antibodies. The pAd/PL-DEST™ vector has no promoter allowing expression of a sequence of interest from any desired promoter that may be operably linked to the sequence of interest, optionally, prior to insertion in the viral vectors of the invention. Additionally, the pAd/PL-DEST™ vector has no 3' sequences allowing addition of a C-terminal epitope tag (if desired) and a polyadenylation signal of choice.

[0384] The pAd/CMV/V5-DEST™ vector (36686 bp) contains the following features.

Feature	Benefit
Human adenovirus type 5 sequences (corresponds to wild-type 1-458 and 3513-35935 sequence) Note: The E1 and E3 regions are deleted.	Encodes all elements (except E1 and E3 polypeptides) required to produce replication-incompetent adenovirus (Russell, (2000) <i>J. Gen. Virol.</i> 81, 2573-2604.) including: Left and right ITRs Encapsidation signal for packaging E2 and E4 regions Late genes
pAd forward priming site CMV promoter	Permits sequencing of the insert. Permits high-level expression of the gene of interest
T7 promoter/priming site	Allows <i>in vitro</i> transcription in the sense orientation and sequencing through the insert.
<i>attR1</i> and <i>attR2</i> sites	Bacteriophage λ-derived DNA recombination sequences that permit recombinational cloning of the gene of interest from a GATEWAY™ entry clone.
<i>ccdB</i> gene Chloramphenicol resistance gene (Cm ^R)	Permits negative selection of the plasmid. Allows counterselection of the plasmid.
V5 epitope	Allows detection of the recombinant fusion polypeptide by the Anti-V5 Antibodies

Feature	Benefit
Herpes Simplex Virus thymidine kinase (TK)	Permits efficient transcription termination and polyadenylation of mRNA
°polyadenylation signal	
pAd reverse priming site	Allows sequencing of the insert in the anti-sense orientation.
pUC origin	Permits high-copy replication and maintenance in <i>E. coli</i> .
<i>bla</i> promoter	Allows expression of the ampicillin resistance gene.
Ampicillin resistance gene (β -lactamase)	Allows selection of the plasmid in <i>E. coli</i> .
<i>Pac</i> I restriction sites (positions 34610 and 36684)	Permits exposure of the left and right ITRs required for viral replication and packaging.

[0385] The pAd/PL-DEST™ vector (34864 bp) contains the following features.

Feature	Benefit
Human adenovirus type 5 sequences (corresponds to wild-type 1-458 and 3513-35935 sequence) Note: The E1 and E3 regions are deleted.	Encodes all elements (except E1 and E3 proteins) required to produce replication-incompetent adenovirus (Russell, 2000) including: Left and right ITRs Encapsidation signal for packaging E2 and E4 regions Late genes
pAd forward priming site <i>attR1</i> and <i>attR2</i> sites	Permits sequencing of the insert. Bacteriophage λ -derived DNA recombination sequences that permit recombinational cloning of the DNA sequence of interest from a GATEWAY™ entry clone (Landy, 1989, <i>Annu. Rev. Biochem.</i> 58, 913-949.).
Chloramphenicol resistance gene (Cm^R) <i>ccdB</i> gene	Allows counterselection of the plasmid.
pAd reverse priming site	Permits negative selection of the plasmid.
pUC origin	Allows sequencing of the insert in the anti-sense orientation.
<i>bla</i> promoter	Permits high-copy replication and maintenance in <i>E. coli</i> .
Ampicillin resistance gene (β -lactamase)	Allows expression of the ampicillin resistance gene.
<i>Pac</i> I restriction sites (positions 32788 and 34862)	Allows selection of the plasmid in <i>E. coli</i> .
	Permits exposure of the left and right ITRs required for viral replication and packaging.

[0386] The pAd/CMV/V5-DEST™ and pAd/PL-DEST™ vectors contain the following features: human adenovirus type 5 sequences (Ad 1-458), upstream of the *attR1* site, containing the “Left” Inverted Terminal Repeat (L-ITR) and the encapsidation signal sequence required for viral packaging; human cytomegalovirus (CMV) immediate early promoter for high-level constitutive expression of the gene of interest in a wide range of mammalian cells (in pAd/CMV/V5-DEST™ only; (Andersson, *et al.*, 1989, *J. Biol. Chem.* 264, 8222-8229; Boshart, *et al.*, 1985, *Cell* 41, 521-530; Nelson, *et al.*, 1987, *Molec. Cell. Biol.* 7, 4125-4129); two recombination sites, *attR1* and *attR2* for recombinational cloning of the DNA sequence of interest from an entry clone; chloramphenicol resistance gene (Cm^R) located between the two *attR* sites for counterselection; the *ccdB* gene located between the *attR* sites for negative selection; C-terminal V5 epitope for detection of the recombinant polypeptide of interest (in pAd/CMV/V5-DEST™ only); (Southern, *et al.*, 1991, *J. Gen. Virol.* 72, 1551-1557); human adenovirus type 5 sequences (Ad 3513-35935) containing genes and elements (*e.g.* E2 and E4 regions, late genes, and “Right” ITR) required for proper packaging and production of adenovirus (Hitt, *et al.*, (1999) In *The Development of Human Gene Therapy*, T. Friedmann, ed. (Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press), pp. 61-86.; Russell, (2000)); ampicillin resistance gene for selection in *E. coli*; and the pUC origin for high-copy replication and maintenance of the plasmid in *E. coli*. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the destination vector containing *attP* sites flanking the *ccdB* and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. It has recently been found that the use of spectinomycin selection instead of chloramphenicol selection results in an increase in the number of colonies obtained on selection plates, indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene.

[0387] The plasmid, pAd/CMV/V5-GW/*lacZ*, is included and may be used as a positive expression control in the mammalian cell line of choice.

pAd/CMV/V5-GW/*lacZ* (Fig. 10) is a 37567 bp vector expressing β -galactosidase, and was generated using the GATEWAY™ LR recombination reaction between an entry clone containing the *lacZ* gene and pAd/CMV/V5-DEST™. β -galactosidase is expressed as a C-terminal V5 fusion polypeptide with a molecular weight of approximately 120 kDa.

[0388] Nucleic acid molecules of the invention may be constructed using any technique known to those skilled in the art, for example recombinational cloning (*e.g.*, using GATEWAY™). GATEWAY™ is a universal cloning technology that takes advantage of the site-specific recombination properties of bacteriophage lambda (Landy, 1989) to provide a rapid and highly efficient way to move a DNA sequence of interest into multiple vector systems. To express a sequence of interest in mammalian cells using the GATEWAY™ Technology the following method may be used. First, a sequence of interest may be cloned into a GATEWAY™ entry vector of choice to create an entry clone. If pAd-DEST™ is used, a promoter of choice and a polyadenylation signal may be operably attached to the sequence of interest. Next, a recombination reaction (*e.g.*, an LR reaction) may be performed to generate an expression clone by transferring the sequence of interest into a GATEWAY™ destination vector (*e.g.* pAd/CMV/V5-DEST™ or pAd-DEST™). An expression clone may then be used to generate viral vector using the ViraPower™ Adenoviral Expression System.

[0389] For more information about the GATEWAY™ Technology, generating an entry clone, and performing the LR recombination reaction, refer to the GATEWAY™ Technology manual.

[0390] Materials and methods of the invention (*e.g.*, The ViraPower™ Adenoviral Expression System) facilitate highly efficient, *in vitro* or *in vivo* delivery of a target gene to dividing and non-dividing mammalian cells using a replication-incompetent adenovirus. The System utilizes GATEWAY™-adapted destination vectors to allow highly efficient and rapid creation of adenoviral vectors that circumvent the need for traditional, homologous recombination and the use of *recA*⁺ bacteria to produce adenovirus. To express a sequence of interest in mammalian cells using the ViraPower™ Adenoviral Expression System the following method may be used. First, an expression clone in pAd/CMV/V5-DEST™ or pAd-DEST™ may be created (*e.g.*, using

GATEWAY™ Technology or other suitable methodology). Next, the expression clone may be digested with *Pac* I to expose the viral inverted terminal repeats (ITRs). The digested expression clone may be introduced into suitable host cells (e.g., 293 or 293A cells) to produce adenovirus. The adenovirus may be amplified by infecting additional cells and allowing the virus to replicate. The virus may be used to transduce a suitable cell line (e.g., a mammalian cell line of choice). The transduced cell line may be assayed for expression of the sequence of interest using any suitable means.

[0391] The pAd/CMV/V5-DEST™ and pAd/PL-DEST™ vectors may be linear or may be supercoiled plasmids. Each destination vector may be supplied as 6 µg of plasmid, lyophilized in TE, pH 8.0. To use, resuspend the destination plasmid in 40 µl of sterile water to a final concentration of 150 ng/µl.

[0392] It may be desirable to propagate and maintain the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ vectors. One suitable method is to use Library Efficiency® DB3.1™ Competent Cells (Invitrogen Corporation, Carlsbad, CA) for transformation. The DB3.1™ *E. coli* strain is resistant to CcdB effects and can support the propagation of plasmids containing the *ccdB* gene. To maintain integrity of the vector, select for transformants in media containing 50-100 µg/ml ampicillin and 15-30 µg/ml chloramphenicol. General *E. coli* cloning strains including TOP10 or DH5α are not recommended for propagation and maintenance as these strains are sensitive to CcdB effects.

[0393] To recombine a sequence of interest into pAd/CMV/V5-DEST™ or pAd-DEST™, the sequence of interest should be cloned into an entry clone. Many entry vectors including pENTR/D-TOPO® are available from Invitrogen Corporation, Carlsbad, CA to facilitate generation of entry clones.

[0394] pAd/CMV/V5-DEST™ is a C-terminal fusion vector; however, this vector may be used to express native polypeptides or C-terminal fusion polypeptides. A sequence of interest encoding a polypeptide of interest must contain an ATG initiation codon in the context of a Kozak consensus sequence for proper initiation of translation in mammalian cells (Kozak, M. (1987). *Nucleic Acids Res.* 15, 8125-8148. Kozak, M. (1991). *J. Cell Biology* 115, 887-903. Kozak, M. (1990). *Proc. Natl. Acad. Sci. USA* 87, 8301-8305.). An example of a Kozak consensus sequence is (G/A)NNATGG (SEQ ID NO:). The ATG initiation codon is underlined. Note that other sequences are

possible, but the G or A at position -3 and the G at position +4 are the most critical for function (shown in bold).

[0395] If it is desired to include the V5 epitope tag, a sequence of interest in the entry clone should not contain a stop codon. In addition, the sequence encoding the polypeptide should be in frame with the V5 epitope tag after recombination. To express a native polypeptide (*e.g.*, without a tag sequence) from a sequence of interest, the sequence of interest must contain a stop codon in the entry clone. The C-terminal peptide containing the V5 epitope and the *attB2* site will add approximately 4.3 kDa to the size of a polypeptide expressed from a sequence of interest.

[0396] pAd/PL-DEST[™] allows generation of an adenovirus that contains a sequence of interest whose expression is controlled by a promoter of choice. To facilitate proper expression of a sequence of interest from pAd/PL-DEST[™], an entry clone containing the following should be generated: 1) a promoter of choice to control expression of the sequence of interest in mammalian cells; 2). the sequence of interest; 3) a stop codon; and 4) a polyadenylation signal sequence of choice for proper transcription termination and polyadenylation of mRNA. To express a polypeptide from a sequence of interest, the ORF of the polypeptide should contain an ATG initiation codon in the context of a Kozak consensus sequence for proper initiation of translation in mammalian cells (Kozak, 1987; Kozak, 1991; Kozak, 1990). If desired, an N-terminal and/or C-terminal fusion tag sequence may be included.

[0397] In some embodiments, an entry clone contains *attL* sites flanking the sequence of interest. Sequences of interest in an entry clone are transferred to the destination vector backbone by mixing the DNAs with the GATEWAY[™] LR Clonase[™] Enzyme Mix, Invitrogen Corporation, Carlsbad, CA. The resulting LR recombination reaction is then transformed into *E. coli* (*e.g.* TOP10 or DH5 α [™]-T1^R) and the expression clone selected using ampicillin. Recombination between the *attR* sites on the destination vector and the *attL* sites on the entry clone replaces the chloramphenicol (Cm^R) gene and the *ccdB* gene with the sequence of interest and results in the formation of *attB* sites in the expression clone.

[0398] The *ccdB* gene mutates at a very low frequency, resulting in a very low number of false positives. True expression clones will be ampicillin- and

blasticidin-resistant and chloramphenicol-sensitive. Transformants containing a plasmid with a mutated *ccdB* gene will be ampicillin-, blasticidin-, and chloramphenicol-resistant. To check a putative expression clone, test for growth on LB plates containing 30 µg/ml chloramphenicol. A true expression clone should not grow in the presence of chloramphenicol.

[0399] The recombination region of the expression clone resulting from pAd/CMV/V5-DEST™ x entry clone is shown in Fig. 8. Shaded regions correspond to those DNA sequences transferred from the entry clone into the pAd/CMV/V5-DEST™ vector by recombination. Non-shaded regions are derived from the pAd/CMV/V5-DEST™ vector. Bases 1414 and 3657 of the pAd/CMV/V5-DEST™ sequence are marked. The recombination region of the expression clone resulting from pAd/PL-DEST™ x entry clone is shown IN Fig. 9. Shaded regions correspond to those DNA sequences transferred from the entry clone into the pAd/PL-DEST™ vector by recombination. Non-shaded regions are derived from the pAd/PL-DEST™ vector. Bases 519 and 2202 of the pAd/PL-DEST™ sequence are marked.

[0400] To confirm that a sequence of interest is in the correct orientation and in frame with a fusion tag (if present), an expression construct may be sequenced. The following primer binding may be used to sequence an expression construct. Refer to the Figs. 8 and 9 for the location of the primer binding sites. The pAd/CMV/V5-DEST™ vector contains the T7 promoter/priming site 5'-TAATACGACTCACTATAGGG-3' (SEQ ID NO:) and the V5 (C-term) reverse priming site 5'-ACCGAGGAGAGGGTTAGGGAT-3' (SEQ ID NO:). The pAd/PL-DEST™ vector contains the pAd forward priming site 5'-GACTTTGACCGTTTACGTGGAGAC-3' (SEQ ID NO:) and the pAd reverse priming site 5'-CCTTAAGCCACGCCCACACATTTC-3' (SEQ ID NO:).

[0401] Once purified plasmid DNA of a pAd/CMV/V5-DEST™ or pAd/PL-DEST™ expression construct has been obtained, the vector may be used in ViraPower™ Adenoviral Expression System (Invitrogen Corporation, Carlsbad, CA) by digesting with *Pac* I. The *Pac* I-digested vector is used to produce an adenoviral stock, which after amplification, may then be used to

transduce a mammalian cell line of choice to express the sequence of interest or a polypeptide encoded by the sequence of interest.

[0402] Once a pAd/CMV/V5-DEST™ and/ or a pAd/PL-DEST™ expression clone has been constructed, purified plasmid DNA may be prepared. Suitable purification methods include the S.N.A.P.™ MidiPrep Kit (Invitrogen Corporation, Carlsbad, CA) and CsCl gradient centrifugation. To verify the integrity of an expression construct after plasmid preparation, the plasmid may be analyzed by restriction digests.

[0403] Before transfecting an expression clone into 293A cells, the left and right viral ITRs on the vector should be exposed to allow proper viral replication and packaging. Both pAd/CMV/V5-DEST™ and pAd/PL-DEST™ vectors contain *Pac* I restriction sites. Digestion of the vector with *Pac* I allows exposure of the left and right viral ITRs and removal of the bacterial sequences (*i.e.* pUC origin and ampicillin resistance gene). The sequence of interest must not contain any *Pac* I restriction sites.

[0404] Digest at least 5 µg of purified plasmid DNA of pAd/CMV/V5-DEST™ or pAd/PL-DEST™ expression construct with *Pac* I using commercially available *Pac* I enzyme. Follow the manufacturer's instructions. Purify the digested plasmid DNA using phenol/chloroform extraction followed by ethanol precipitation or a DNA purification kit (*e.g.* S.N.A.P. MiniPrep™ Kit, catalog no. K19001, Invitrogen Corporation, Carlsbad, CA). Gel purification is not required.

[0405] Resuspend or elute the purified plasmid, as appropriate in sterile water or TE Buffer, pH 8.0 to a final concentration of 0.1-3.0 µg/µl.

[0406] To express a gene of interest from pAd/CMV/V5-DEST™ or pAd/PL-DEST™ using Invitrogen's ViraPower™ Adenoviral Expression System, the following reagents are required: 1) a host cell (*e.g.*, 293 or 293A cell lines); and 2) a transfection reagent (*e.g.*, Lipofectamine™ 2000 Reagent, catalog no. 11668019, Invitrogen Corporation, Carlsbad, CA). The 293A cell line is a subclone of the 293 cell line and supplies the E1 proteins required for production of replication-competent adenovirus and exhibits a flattened morphology to enhance visualization of plaques.

[0407] pAd/CMV/V5-GW/*lacZ* is included with the each kit for use as a positive control for expression in the ViraPower™ Adenoviral Expression

System. In pAd/CMV/V5-GW/*lacZ*, β -galactosidase is expressed as a C-terminally tagged fusion polypeptide that may be easily detected by western blot or functional assay. To propagate and maintain the plasmid: resuspend the vector in 10 μ l of sterile water to prepare a 1 μ g/ μ l stock solution. Use the stock solution to transform a *recA*, *endA* *E. coli* strain like TOP10, DH5 α [™]-T1^R, or equivalent. Use 10 ng of plasmid for transformation. Select transformants on LB agar plates containing 50-100 μ g/ml ampicillin. Prepare a glycerol stock of a transformant containing plasmid for long-term storage.

EXAMPLE 4

Exemplary instruction manual for kits of the invention.

[0408] Provided for in the methods of the present invention is a kit containing a viral system for high-level, transient expression in dividing and non-dividing mammalian cells. One nonlimiting example of such a kit is the ViraPower[™] Adenoviral Expression System, Invitrogen catalog nos. K4930-00 and K4940-00, Version A, July 15, 2002, 25-0543, as described in this example.

[0409] The ViraPower[™] Adenoviral Expression Kits include the following components. For a detailed description of the contents of each component, see below.

Components	Catalog No. K4930-00	Catalog No. K4940-00
pAd/CMV/V5-DEST [™] GATEWAY [™] Vector	✓	
pAd/PL-DEST [™] GATEWAY [™] Vector		✓
293A Cell Line	✓	✓

[0410] The ViraPower[™] Adenoviral Expression Kits are shipped as described below. Upon receipt, store each component as detailed below.

Item	Shipping	Storage
pAd-DEST [™] GATEWAY [™] Vector	Blue ice	-20°C
293A Cell Line	Dry ice	Liquid nitrogen

- [0411] Each ViraPower™ Adenoviral Expression Kit includes a destination vector (pAd/CMV/V5-DEST™ or pAd/PL-DEST™) for cloning a DNA sequence of interest and a corresponding expression control vector. For information about the vectors see, for example, the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual, catalog nos. V493-20 and 494-20, version B, Invitrogen Corporation, Carlsbad, CA.
- [0412] Methods of the invention may be practiced using any suitable cell line (e.g., 293A Cell Line, catalog no. R-705-07, Invitrogen Corporation, Carlsbad, CA).
- [0413] A number of reagents that are commercially available may be used in conjunction with the methods of the invention. For example, the following reagents may be obtained from Invitrogen Corporation, Carlsbad, CA.

Item	Catalog no.
pAd/CMV/V5-DEST™ GATEWAY™ Vector	V493-20
pAd/PL-DEST™ GATEWAY™ Vector	V494-20
293A Cell Line	R705-07
Lipofectamine™ 2000	11668-027
	11668-019
Opti-MEM® I Reduced Serum Medium	31985-062
	31985-062
Phosphate-Buffered Saline (PBS), pH 7.4	10010-023
	10010-031
S.N.A.P.™ MidiPrep Kit	K1910-01

- [0414] The ViraPower™ Adenoviral Expression System allows creation of a replication-incompetent adenovirus that can be used to deliver and express a gene of interest in either dividing or non-dividing mammalian cells. The major components of the ViraPower™ Adenoviral Expression System include: a choice of GATEWAY™-adapted adenoviral vectors that allow highly efficient generation of a recombinant adenovirus containing the gene of interest under the control of the human cytomegalovirus (CMV) immediate-early enhancer/promoter (pAd/CMV/V5-DEST™) or a promoter of choice (pAd/PL-DEST™); a optimized cell line, 293A, which allows production and subsequent, titering of the recombinant adenovirus; and a control expression plasmid containing the lacZ gene which, when packaged into virions and transduced into a mammalian cell line, expresses β -galactosidase. For more information about the adenoviral vectors, the corresponding positive control vector containing the lacZ gene, and GATEWAY™ Technology, refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vectors manual.

This manual is supplied with each ViraPower™ Adenoviral Expression Kit, but may also be obtained by contacting Invitrogen Corporation, Carlsbad, CA.

[0415] Use of the ViraPower™ Adenoviral Expression System to facilitate DNA virus-based expression of the gene of interest provides the following advantages: uses GATEWAY™ Technology to allow highly efficient, rapid cloning of a gene of interest into a full-length adenoviral vector, bypassing the need for a shuttle vector and inefficient homologous recombination in human or bacterial cells; allows generation of high titer adenoviral stocks (*i.e.*, 1×10^9 pfu/ml in crude preparations and 1×10^{11} pfu/ml in concentrated preparations); efficiently delivers the gene of interest to actively dividing and non-dividing mammalian cells in culture or *in vivo*; generates adenoviral constructs with such a high degree of efficiency and accuracy that the system is amenable for use in high-throughput applications or library transfer procedures; and allows production of a replication-incompetent virus that enhances the biosafety of the system and its use as a gene delivery vehicle.

[0416] This example provides an overview of the ViraPower™ Adenoviral Expression System and provides instructions and guidelines to: transfect the pAd/CMV/V5-DEST™ or pAd/PL-DEST™ expression construct into the 293A Cell Line to produce an adenoviral stock; amplify the adenoviral stock; titer the adenoviral stock; use the amplified adenoviral stock to transduce any mammalian cell line of choice; and assay for transient expression of any polynucleotide of interest or recombinant polypeptide. This expression may be used to express, for example, a polypeptide, a protein, or an untranslated RNA, *e.g.*, tRNA, all of which are encompassed by the term “gene of interest” as used herein.

[0417] For details and instructions to generate an expression construct using pAd/CMV/V5-DEST™ or pAd/PL-DEST™, refer to the pAd/CMV/V5-DEST™ or pAd/PL-DEST™ GATEWAY™ Vector manual. For instructions to culture and maintain the 293A producer cell line, refer to the 293A Cell Line manual. These manuals are supplied with the ViraPower™ Adenoviral Expression Kits, and are also available from Invitrogen Corporation, Carlsbad, CA.

[0418] The ViraPower™ Adenoviral Expression System facilitates highly efficient, *in vitro* or *in vivo* delivery of a target gene to dividing and non-

dividing mammalian cells using a replication-incompetent adenovirus. Based on the second-generation vectors developed by Bett, A.J., *et al.*, *Proc. Natl. Acad. Sci. USA* 91:8802-8806 (1994), the ViraPower™ Adenoviral Expression System takes advantage of the GATEWAY™ Technology to simplify and greatly enhance the efficiency of generating high-titer, recombinant adenovirus.

[0419] The first major component of the system described in this example is an E1 and E3-deleted, pAd-DEST™-based expression vector into which the gene of interest will be cloned. Expression of the gene of interest is controlled by the human cytomegalovirus (CMV) promoter (in pAd/CMV/V5-DEST™) or the promoter of choice (in pAd/PL-DEST™). The vector also contains the elements required to allow packaging of the expression construct into virions (*e.g.*, 5' and 3' ITRs, encapsidation signal, adenoviral late genes). For more information about the pAd-DEST™ expression vectors, refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual, available from Invitrogen Corporation, Carlsbad, CA.

[0420] The second major component of the system is an optimized 293A Cell Line that will be used to facilitate initial production, amplification, and titering of replication-incompetent adenovirus. The 293A cells contain a stably integrated copy of E1 that supplies the E1 proteins (E1a and E1b) in trans that are required to generate adenovirus. For more information about the 293A Cell Line, refer to the 293A Cell Line manual, available from Invitrogen Corporation, Carlsbad, CA. The pAd-DEST™ vector containing the gene of interest is transfected into 293A cells to produce a replication-incompetent adenovirus. The crude adenoviral stock is used to infect 293A cells to produce an amplified adenoviral stock. Once the adenoviral stock is amplified and titered, this high-titer stock may be used to transduce the recombinant adenovirus into the mammalian cell line of choice for expression of the recombinant polypeptide of interest.

[0421] Adenovirus enters target cells by binding to the Coxsackie/Adenovirus Receptor (CAR). After binding to the CAR, the adenovirus is internalized via integrin-mediated endocytosis followed by active transport to the nucleus. Once in the nucleus, the early events are initiated (*e.g.*, transcription and translation of E1 proteins), followed by expression of the adenoviral late genes

and viral replication. Expression of the late genes is dependent upon E1. In the ViraPower™ Adenoviral Expression System, E1 is supplied by the 293A producer cells. The viral life cycle spans approximately 3 days. For more information about the adenovirus life cycle and adenovirus biology, refer to the following references as well as published reviews: Bergelson, J. M., *et al.* *Science* 275:1320-1323 (1997); Hitt, M.M., *et al.*, "Structure and Genetic Organization of Adenovirus Vectors," in *The Development of Human Gene Therapy*, Friedmann, T., ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1999), pp. 61-86.

- [0422] After adenovirus is transduced into the target cell and is transported to the nucleus, it does not integrate into the host genome. Therefore, expression of the gene of interest is typically detectable within 24 hours after transduction and is transient, only persisting for as long as the viral genome is present. Additional information regarding the use of adenoviral vectors and host cells may be obtained from the following references: Bett, A.J., *et al.*, *Proc. Natl. Acad. Sci. USA* 91:8802-8806 (1994); Chen, H.H., *et al.*, *Hum. Gene Ther.* 10:365-373 (1999); Ciccarone, V., *et al.*, *Focus* 21:54-55 (1999); Dion, L.D., *et al.*, *J. Virol. Methods* 56:99-107 (1996); Engelhardt, J.F., *et al.*, *Nature Genetics* 4:27-34 (1993); Fallaux, F.J., *et al.*, *Hum. Gene Ther.* 9:1909-1917 (1998); Fallaux, F.J., *et al.*, *Hum. Gene Ther.* 7:215-222 (1996); Fan, X., *et al.*, *Hum. Gene Ther.* 11:1313-1327 (2000); Graham, F.L., *et al.*, *J. Gen. Virol.* 36:59-74 (1977); Hitt, M.M., *et al.*, "Structure and Genetic Organization of Adenovirus Vectors," in *The Development of Human Gene Therapy*, Friedmann, T., ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1999), pp. 61-86; Kozarsky, K.F., and Wilson, J.M., *Curr. Opin. Genet. Dev.* 3:499-503 (1993); Krougliak, V., and Graham, F.L., *Hum. Gene Ther.* 6:1575-1586 (1995); Lochmuller, H., *et al.*, *Hum. Gene Ther.* 5:1485-1491 (1994); Navarro, V., *et al.*, *Gene Ther.* 6:1884-1892 (1999); Russell, W.C., *J. Gen. Virol.* 81:2573-2604 (2000); Wang, I.I., and Huang, I.I., *Drug Discovery Today* 5:10-16 (2000); Wivel, N.A., "Adenoviral Vectors," in *The Development of Human Gene Therapy*, Friedmann, T., ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1999), pp. 87-110; and Zhang, W. W., *et al.*, *BioTechniques* 18:444-447 (1995).

- [0423] Viral infection is referred to in some procedures in this example, and viral transduction in other procedures. These terms are defined below.
- [0424] Infection: Applies to situations where viral replication occurs and infectious viral progeny are generated. Only cell lines that stably express E1 may be infected.
- [0425] Transduction: Applies to situations where no viral replication occurs and no infectious viral progeny are generated. Mammalian cell lines that do not express E1 are transduced. In this case, an adenovirus is used as a gene delivery vehicle.
- [0426] The ViraPower™ Adenoviral Expression System is suitable for *in vivo* gene delivery applications. Many groups have successfully used adenoviral vectors to express a target gene in a multitude of tissues including skeletal muscle, lung, heart, and brain. For more information about target genes that have been successfully expressed *in vivo* using adenoviral-based vectors, refer to the publications, *supra*.
- [0427] The ViraPower™ Adenoviral Expression System includes the following safety features. The entire E1 region is deleted in the pAd/CMV/V5-DEST™ or pAd/PL-DEST™ expression vectors. Expression of the E1 proteins is required for the expression of the other viral genes (*e.g.*, late genes), and thus viral replication only occurs in cells that express E1. Adenovirus produced from the pAd/CMV/V5-DEST™ or pAd/PL-DEST™ expression vectors is replication-incompetent in any mammalian cells that do not express the E1a and E1b proteins. Adenovirus does not integrate into the host genome upon transduction. Because the virus is replication-incompetent, the presence of the viral genome is transient and will eventually be diluted out as cell division occurs. For more information regarding adenoviral transduction and expression, see the publications listed *supra*.
- [0428] Despite the presence of the safety features discussed above, the adenovirus produced with this system may still pose some biohazardous risk since it can transduce primary human cells. For this reason, adenoviral stocks generated using this system be handled as Biosafety Level 2 (BL-2) organisms and strictly all published guidelines for BL-2 should be followed. Furthermore, extra caution should be taken when creating adenovirus carrying potential harmful or toxic genes (*e.g.*, activated oncogenes) or when producing

large-scale preparations of virus. For more information about the BL-2 guidelines and adenovirus handling, refer to the document, "Biosafety in Microbiological and Biomedical Laboratories," 4th Edition, published by the Centers for Disease Control (CDC). This document may be downloaded from the CDC Web site.

[0429] The genomic copy of E1 in all 293 cell lines contains homologous regions of overlap with the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ vectors. In rare instances, it is possible for homologous recombination to occur between the E1 genomic region in 293 cells and the viral DNA, causing the gene of interest to be replaced with the E1 region, and resulting in generation of a "wild-type," replication-competent adenovirus (RCA). This event is most likely to occur during large-scale preparation or amplification of virus, and the growth advantages of the RCA allow it to quickly overtake cultures of recombinant adenovirus. To reduce the likelihood of propagating RCA-contaminated adenoviral stocks, caution should be used when handling all viral preparations, which is considered to be BL-2 material. Routine screening for the presence of wild-type RCA contamination after large-scale viral preparations should be performed. Suitable methods to screen for RCA contamination include PCR screening or supernatant rescue assays. If RCA contamination occurs, plaque purification may be performed to re-isolate the recombinant adenovirus of interest. As an alternative, E1-containing producer cell lines such as 911 or PER.C6 which contain no regions of homologous overlap with the adenoviral vectors may be used to help reduce the incidence of RCA generation. For more information regarding RCA, see the publications listed *supra*, in particular Lochmuller, *et al.* (1994) and Zhang *et al.* (1995).

[0430] Figure 13 describes the general steps required to express the gene of interest using the ViraPower™ Adenoviral Expression System. For instructions to generate an adenovirus expression clone using pAd/CMV/V5-DEST™ or pAd/PL-DEST™, refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual, available from Invitrogen Corporation, Carlsbad, CA.

[0431] First, the adenovirus expression clone containing the gene of interest is generated and digested with *Pac* I to expose the ITRs according to the

methods described herein or by published methods, *e.g.*, the pAd/PL-DEST™ and pAd/CMV/V5-DEST™ manuals, from Invitrogen Corporation, Carlsbad, CA. Next, the 293A producer cell line is transfected with the adenovirus expression clone. The cells are harvested and lysed to produce a crude viral lysate. The adenovirus may be amplified by infecting 293A producer cells with the crude viral lysate, and the resulting viral stock is titered. The viral stock is used to infect a mammalian cell line of interest, which is then assayed for expression of the gene of interest.

[0432] The ViraPower™ Adenoviral Expression System is designed to create an adenovirus to deliver and transiently express a gene of interest in mammalian cells. Although the system has been designed to express any recombinant polypeptide of interest in the simplest, most direct fashion, use of the system is geared towards those users who are familiar with the biology of DNA viruses and adenoviral vectors and possess a working knowledge of viral and tissue culture techniques. For more information about these topics, refer to the following published reviews: Adenovirus biology: see Russell, W. C. *J. Gen. Virol.* 81:2573-2604 (2000). Adenoviral vectors: see Hitt, M.M., *et al.*, "Structure and Genetic Organization of Adenovirus Vectors," in *The Development of Human Gene Therapy*, Friedmann, T., ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1999), pp. 61-86, and Wivel, N.A., "Adenoviral Vectors," in *The Development of Human Gene Therapy*, Friedmann, T., ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1999), pp. 87-110. Adenovirus applications: see Wang, I.I., and Huang, I.I., *Drug Discovery Today* 5:10-16 (2000).

[0433] An expression clone may be created containing a DNA sequence of interest in pAd/CMV/V5-DEST™, which expresses the gene of interest under the control of the human CMV promoter, or in pAd/PL-DEST™, which is promoterless, thus allowing the insertion of a cassette containing the gene of interest under the control of any promoter. Refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual for further instructions. Once an expression clone has been created, any method of preparing purified plasmid DNA that is clean and free from phenol and sodium chloride may be used. Contaminants may kill the cells, and salt may interfere with lipid complexing, decreasing transfection efficiency. Suitable

methods of isolating plasmid DNA include, but are not limited to, the S.N.A.P.[™] MidiPrep Kit (Catalog No. K1910-01, Invitrogen Corporation, Carlsbad, CA) and cesium chloride gradient centrifugation.

[0434] Any 293-derived cell line or other cell line that expresses the E1 proteins may be used to produce adenovirus. One such cell lines particularly suited for use in the present invention is the human 293A Cell Line, included with the ViraPower[™] Adenoviral Expression kits to facilitate adenovirus production from the E1-deleted pAd-DEST[™] vectors. The 293A Cell Line, a subclone of the 293 cell line, supplies in *trans* the E1 proteins that are required for expression of adenoviral late genes, and thus viral replication. The cell line exhibits a flattened morphology, enabling easier visualization of plaques. For more information about how to culture and maintain 293A cells, refer to the 293A Cell Line manual, available from Invitrogen Corporation, Carlsbad, CA.

[0435] Once an expression clone, for example a pAd-DEST[™] expression clone, is created, the expression clone is transfected into a suitable host cell line (*e.g.*, 293A cells) to produce an adenoviral stock. The following section provides protocols and instructions to generate an adenoviral stock, using pAd-DEST[™] to illustrate the method of the present invention.

[0436] Before transfecting a pAd-DEST[™] expression clone into 293A cells, the left and right viral ITRs are exposed to allow proper viral replication and packaging. Each pAd-DEST[™] vector contains Pac I restriction sites (refer to the maps of each vector in the pAd/CMV/V5-DEST[™] and pAd/PL-DEST[™] manual for the location of the Pac I sites). Digestion of the vector with Pac I allows exposure of the left and right viral ITRs and removal of the bacterial sequences (*i.e.*, pUC origin and ampicillin resistance gene). The DNA sequence of interest should not contain any Pac I restriction sites. At least 5 mg of purified plasmid DNA of the pAd-DEST[™] expression construct is digested with *Pac* I (New England Biolabs, Catalog No. R0547S) according to the manufacturer's instructions. The digested plasmid DNA may be purified using phenol/chloroform extraction followed by ethanol precipitation or a DNA purification kit (*e.g.*, Invitrogen's S.N.A.P. MiniPrep[™] Kit; catalog No. K1900-01). Gel purification is not required. The purified plasmid is

resuspended or eluted, as appropriate, in sterile water or TE Buffer, pH 8.0 to a final concentration of 0.1-3.0 mg/ml.

[0437] The following materials are required before beginning: Pac I-digested pAd-DEST™ expression clone containing the DNA sequence of interest (0.1-3.0 mg/ml in sterile water or TE, pH 8.0); pAd/CMV/V5-GW/lacZ positive control vector (supplied with the kit; resuspended in sterile water to a concentration of 1 mg/ml); 293A cells cultured in the appropriate medium (see the 293A Cell Line manual for details); transfection reagent suitable for transfecting 293A cells (*e.g.*, Lipofectamine™ 2000); Opti-MEM® I Reduced Serum Medium (if using Lipofectamine™ 2000; pre-warmed); fetal bovine serum (FBS); sterile 6-well and 10 cm tissue culture plates; and sterile tissue culture supplies, *e.g.*, 15 ml sterile, capped, conical tubes, table-top centrifuge, water bath (set to 37 °C), and cryovials.

[0438] The pAd/CMV/V5-GW/lacZ plasmid is included with each ViraPower™ Adenoviral Expression kit as a positive control vector for expression. The positive control vector may be included in the transfection experiment to generate a control adenoviral stock that may be used to help optimize expression conditions in the mammalian cell line of interest. For more information about the positive control vector, refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual.

[0439] Any suitable transfection reagent may be used to introduce the pAd-DEST™ expression construct into 293A cells. Particularly suitable is the cationic lipid-based Lipofectamine™ 2000 Reagent available from Invitrogen. Using Lipofectamine™ 2000 to transfect 293A cells offers several advantages: provides the highest transfection efficiency in 293A cells; DNA-Lipofectamine™ 2000 complexes can be added directly to cells in culture medium in the presence of serum; and removal of complexes or medium change or addition following transfection are not required, although complexes can be removed after 4-6 hours without loss of activity. To facilitate optimal formation of DNA-Lipofectamine™ 2000 complexes, the Opti-MEM® I Reduced Serum Medium available from Invitrogen may be used. For more information about Opti-MEM® I, contact Invitrogen Corporation, Carlsbad, CA.

[0440] Provided below is one method by which adenoviral stocks may be produced in 293A cells using the following optimized transfection conditions below. The amount of adenovirus produced using these recommended conditions is approximately 10 ml of crude viral lysate with a titer ranging from 1×10^7 to 1×10^8 plaque-forming units (pfu)/ml. Lipofectamine™ 2000 is one suitable transfection reagent. Other transfection reagents are readily available and may be used according to the appropriate protocols.

Condition	Amount
Tissue culture plate size	6-well (one well per adenoviral construct)
Number of 293A cells to transfect	5×10^5 cells (see Note below)
Amount of Pac I-digested pAd-DEST™ expression plasmid	1 µg
Amount of Lipofectamine™ 2000	3 µl

[0441] 293A cells are plated 24 hours prior to transfection in complete medium, and should be healthy and 90-95% confluent on the day of transfection.

[0442] Provided herein is a method to transfect 293A cells using Lipofectamine™ 2000. One feature of the provided method is that cells may be kept in culture medium during transfection. A positive control and a negative control (no DNA, no Lipofectamine™ 2000) may be included the experiment to aid in evaluation of the results.

[0443] The day before transfection, the 293A cells are trypsinized and counted, then plated at 5×10^5 cells per well in a 6-well plate containing 2 ml of normal growth medium containing serum. On the day of transfection, the culture medium from the 293A cells is removed and replaced with 1.5 ml of normal growth medium containing serum (or Opti-MEM® I Medium containing serum). Antibiotics should not included.

[0444] The DNA-Lipofectamine™ 2000 complexes are prepared for each transfection sample as follows: 1 µg of Pac I-digested pAd-DEST™ expression plasmid DNA is diluted in 250 µl of Opti-MEM® I Medium without serum and mixed gently. The Lipofectamine™ 2000 reagent is mixed gently before use, then diluted 3 µl in 250 µl of Opti-MEM® I Medium without serum. The solution is mixed gently and incubated for 5 minutes at

room temperature. After the 5 minute incubation, the diluted DNA is combined with the diluted Lipofectamine™ 2000 and mixed gently. The solution is then incubated for 20 minutes at room temperature to allow the DNA-Lipofectamine™ 2000 complexes to form. The solution may appear cloudy, but this will not impede the transfection. The DNA-Lipofectamine™ 2000 complexes is added dropwise to each well and mixed gently by rocking the plate back and forth. The cells are incubated overnight at 37°C in a CO₂ incubator.

[0445] The next day, the medium containing the DNA-Lipofectamine™ 2000 complexes is removed and replaced with complete culture medium (*i.e.*, D-MEM containing 10% FBS, 2 mM L-glutamine, and 1% penicillin/streptomycin). 48 hours post transfection, the cells are trypsinized and transferred to a sterile 10 cm tissue culture plate containing 10 ml of complete culture medium. The recommended guidelines for working with BL-2 organisms should be followed throughout these procedures. The culture medium is replaced with fresh, complete culture medium every 2-3 days until visible regions of cytopathic effect (CPE) are observed (typically 7-10 days post-transfection). The infections proceed until approximately 80% CPE is observed (typically 10-13 days post-transfection). The recombinant adenovirus-containing cells are harvested by squirting cells off the plate with a 10 ml tissue culture pipette. The cells and media are transferred to a sterile, 15 ml, capped tube for lysing as described below.

[0446] In this example, Pac I-digested pAd/CMV/V5-GW/lacZ plasmid was transfected into 293A cells using the protocol described *supra*. Figures 14A-C show transfected cells as they undergo CPE.

[0447] Day 4-6 post-transfection (Figure 14A): at this early stage, cells producing adenovirus first appear as patches of rounding, dying cells.

[0448] Day 6-8 post-transfection (Figure 14B): as the infection proceeds, cells containing viral particles lyse and infect neighboring cells. A plaque begins to form.

[0449] Day 8-10 post-transfection (Figure 14C): at this late stage, infected neighboring cells lyse, forming a plaque that is clearly visible.

[0450] After the adenovirus-containing cells and media are harvested, several freeze/thaw cycles followed by centrifugation may be used to prepare a crude

viral lysate. The freeze/thaw cycles cause the cells to lyse and allow release of intracellular viral particles. The tube containing harvested transfected cells and media is placed at -80°C for 30 minute, then placed in a 37°C water bath for 15 minutes to thaw. The freezing and thawing steps are repeated twice. The cell lysate is centrifuged in a table-top centrifuge at 3000 rpm for 15 minutes at room temperature to pellet the cell debris. The supernatant containing viral particles, the viral stock, may be transferred to cryovials in 1 ml aliquots and stored at -80°C.

[0451] Once a crude viral stock is prepared, it may be amplified by infecting 293A cells as described below. This procedure is recommended to obtain the highest viral titers and optimal results in transduction studies. The titer of the crude viral stock may be determined, and this stock may be used to transduce the mammalian cells of interest to verify the functionality of the adenoviral construct in preliminary expression experiments.

[0452] The viral stocks are placed at -80°C for long-term storage. Because adenovirus is non-enveloped, viral stocks remain relatively stable and some freezing and thawing of the viral stocks is acceptable. Freezing and thawing viral stocks more than 10 times should be avoided as loss of viral titer can occur. When stored properly, viral stocks of an appropriate titer should be suitable for use for up to one year. After long-term storage, re-titering the viral stocks may be performed before use.

[0453] Once a crude viral stock is created, this stock may be used to infect 293A cells to generate a higher titer viral stock (*i.e.*, amplify the virus). The titer of the initial viral stock obtained from transfecting 293A cells generally ranges from 1×10^7 to 1×10^8 plaque-forming units (pfu)/ml. Amplification allows production of a viral stock with a titer ranging from 1×10^8 to 1×10^9 pfu/ml and is generally recommended. Guidelines and protocols are provided in this example to amplify the recombinant adenovirus using 293A cells plated in a 10 cm dish. Larger-scale amplification is possible. Other 293 cell lines or cell lines expressing the E1 proteins are also suitable.

[0454] The recommended Federal guidelines for working with BL-2 organisms should be followed for all work with infectious virus. All manipulations should be performed within a certified biosafety cabinet. Media containing virus should be treated with bleach. Used pipettes, pipette

tips, and other tissue culture supplies should be treated with bleach or disposed of as biohazardous waste. Gloves, a laboratory coat, and safety glasses or goggles should be worn when handling viral stocks and media containing virus.

[0455] Wild-type RCA contamination has not been observed in small-scale (*i.e.*, 3×10^6 293A cells plated in a 10 cm dish) adenoviral amplification using the protocol provided below. However, large-scale amplification of virus should be screened for wild-type RCA contamination. Even in large-scale preparations, contamination of adenoviral stocks with wild-type RCA is a rare event.

[0456] The following materials are required for amplifying the viral stock: crude adenoviral stock of the pAd-DEST™ construct; sterile 10 cm tissue culture plates; sterile, tissue culture supplies 15 ml sterile, capped, conical tubes; equipment and supplies such as table-top centrifuge, 37° C water bath, and cryovials.

[0457] A typical infection of 293A cells uses the following conditions:

Condition	Amount
Tissue culture plate size	10 cm (one per adenoviral construct)
Number of 293A cells to infect	3×10^6 cells
Amount of crude adenoviral stock to use	100 μ l

[0458] For infection, a 10 cm plate of 293A cells is infected with 100 μ l of untitered crude viral stock. Assuming a viral titer of 1×10^7 to 1×10^8 pfu/ml, this generally allows harvesting the desired number adenovirus-containing cells 2-3 days after infection. The volume of crude viral stock used to infect cells, may be varied proportionally according to the desired number of cells and/or amount of crude viral stock to as much as 1 ml of crude viral stock. If the titer of the crude viral stock is known, 293A cells are infected at a multiplicity of infection (MOI) = 3 to 5.

[0459] The procedure below may be used to amplify the adenoviral stock using 293A cells. The day before infection, the 293A cells are trypsinized and counted before plating them at 3×10^6 cells per 10 cm plate. Cells are plated in 10 ml of normal growth medium containing serum. On the day of infection, the cells are verified to be at 80-90% confluency before proceeding. The

desired amount of crude adenoviral stock (e.g., 100 μ l) is added to the cells. The plate is swirled gently to mix. The cells are incubated at 37°C in a CO₂ incubator and the infection is allowed to proceed until 80-90% of the cells have rounded up and are floating or lightly attached to the tissue culture dish (typically 2-3 days post-infection). This CPE indicates that cells are loaded with adenoviral particles. Using less than 100 μ l of crude viral stock or a lower titer stock for infection, may require a longer incubation to achieve CPE. The adenovirus-containing cells are harvested by squirting cells off the plate with a 10 ml tissue culture pipette. The cells and media are transferred to a sterile, 15 ml, capped tube which is then placed at -80°C for 30 minutes. The tube is removed and placed in a 37°C water bath for 15 minutes to thaw. The freezing and thawing steps are repeated twice. The cell lysate is centrifuged in a table-top centrifuge at 3000 rpm for 15 minutes at room temperature to pellet the cell debris. The supernatant containing viral particles is transferred to cryovials in 1 ml aliquots and may be stored at -80°C.

[0460] The amplification procedure is easily scalable to any size tissue culture dish or roller bottle. If it is desirable to scale up the amplification, the number of cells and amount of crude viral stock and medium used is increased in proportion to the difference in surface area of the culture vessel. A screen for the presence of wild-type RCA contamination in the amplified stock may be performed according to suitable screening protocols as described in published literature known to those skilled in the art.

[0461] Before proceeding to transduce the mammalian cell line of interest and express the polynucleotide of interest or recombinant polypeptide, determining the titer of the adenoviral stock may be useful. While this procedure is not required for some applications, it is necessary if the number of adenoviral particles introduced to each cell is to be controlled and to generate reproducible expression results. Guidelines and protocols are provided in this example.

[0462] To determine the titer of an adenoviral stock, 293A cells are plated in 6-well tissue culture plates. Ten-fold serial dilutions of the adenoviral stock are prepared, then used to infect 293A cells overnight. A plaque assay is performed by first overlaying the infected 293A cells with an agarose/plaquing

media solution then allowing 8-12 days for plaques to form. The cells are stained and the number of plaques are counted in each dilution

- [0463] A number of factors may influence viral titers. Titers generally decrease as the size of the insert increases. The size of the wild-type adenovirus type 5 genome is approximately 35.9 kb. Studies have demonstrated that recombinant adenovirus can efficiently package up to 108% of the wild-type virus size from E1 and E3-deleted vectors. Taking into account the size of the elements required for expression from each pAd-DEST™ vector, the DNA sequence or gene of interest should not exceed the size indicated below for efficient packaging.

Vector	Insert Size Limit
pAd/CMV/V5-DEST™	6.0 kbp
Ad/PL-DEST™	7.5 kb

- [0464] Other factors include the characteristics of the cell line used for titering and the age of the adenoviral stock. Viral titers may decrease with long-term storage at -80°C. If the adenoviral stock has been stored for 6 months to 1 year, re-titering the adenoviral stock may be performed prior to use in an expression experiment. The number of freeze/thaw cycles and storage of the adenoviral stock may also affect titer. A limited number of freeze/thaw cycles is acceptable, but viral titers may decrease with more than 10 freeze/thaw cycles. Adenoviral stocks may be aliquotted and stored at -80°C.

- [0465] The 293A cell line supplied with the kit is particularly suitable for use in titering the adenoviral stock, however other cell lines may be used. If another cell line is used, it should: express the E1 proteins, grow as an adherent cell line, be easy to handle, exhibit a doubling time in the range of 18-25 hours, and be non-migratory.

- [0466] The titer of an adenoviral construct may vary depending on which cell line is chosen. If more than one adenoviral construct is to be titered, all of the adenoviral constructs is preferably titered using the same mammalian cell line.

- [0467] To determine the titer of the adenoviral construct, the following materials are required: the pAd-DEST™ adenoviral stock (stored at -80°C until use); 293A Cell Line or other appropriate mammalian cell line of choice (see above); complete culture medium for the cell line; 6-well tissue culture

plates; 4% agarose (see Recipes; equilibrated to 65°C before use); plaquing media (normal growth medium containing 2% FBS; equilibrated to 37°C before use); and 5 mg/ml MTT solution or other appropriate reagent for staining (see Recipes; see below for alternatives).

- [0468] The vital dye, 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (thiazolyl blue (MTT)) is suitable for use as a staining reagent to help visualize plaques. Other vital stains including Neutral Red (Sigma-Aldrich, St. Louis, MO, catalog No. N7005) are suitable. To use Neutral Red, a 1% solution (100X stock solution) is prepared in water and stored at +4°C.
- [0469] The procedure presented herein is a method to determine the titer of the adenoviral stock using the 293A cell line or other appropriate cell line. Other suitable methods are available and known in the art. At least one 6-well plate is required for every adenoviral stock to be titered (six dilutions or one mock well and five dilutions). If an adenoviral stock of the pAd/CMV/V5-GW/lacZ positive expression control has been generated, titrating this stock may be done as well. The day before infection (Day 1), the cells are trypsinized and counted for plating at a density such that they will be 80-90% confluent at the time of infection. For example, 293A cells may be used to titer the adenoviral stock and 1×10^6 cells per well may be plated in each well of a 6-well plate. The cells are incubated at 37°C overnight.
- [0470] On the day of infection (Day 2), the adenoviral stock is thawed and diluted 10-fold serially to concentrations ranging from 10^{-4} to 10^{-9} . For each dilution, the adenoviral construct is diluted into complete culture medium to a final volume of 1 ml and mixed by gentle inversion. The culture medium is removed from the cells, and the dilutions are added to one well of cells (total volume = 1 ml). The plate is swirled gently to disperse the media, then incubated at 37°C overnight. The following day (Day 3), the media containing virus is removed and the cells are gently overlaid with 2 ml of Agarose Overlay solution per well.
- [0471] An agarose overlay solution (enough to overlay one 6-well plate at a time) may be prepared as follows. For one 6-well plate (2 ml overlay per well), 12 ml of pre-warmed (at 37°C) Plaquing Media and 1.2 ml of pre-warmed (at 65°C) 4% Agarose is gently mixed while avoiding the formation of bubbles. The overlay is applied to the cells by gently pipetting the overlay

down the side of each aspirated well while working quickly to prevent premature solidification. The 6-well plate is placed in a level tissue-culture hood at room temperature for 15 minutes or until the Agarose Overlay solidifies. The plate is returned to a 37°C humidified CO₂ incubator. 3-4 days following the initial overlay (Day 6-7), the cells are gently overlaid with an additional 1 ml of Agarose Overlay solution (prepared as before) per well. The Agarose Overlay is allowed to solidify before returning the plate to a 37°C humidified CO₂ incubator. The plates are monitored until plaques are visible (generally 8-12 days post-infection). For each well, the 5 mg/ml MTT solution (1/10 the volume of the Agarose Overlay) is layered gently on top of the solidified agar to stain. For example, if each well contains 3 ml of Agarose Overlay, 300 µl of 5 mg/ml MTT is used. The plates are incubated for 3 hours at 37°C. The plaques are counted to determine the titer of the adenoviral stock.

[0472] When titrating pAd/CMV/V5-DEST™ or pAd/PL-DEST™ adenoviral stocks using 293A cells, titers ranging from 1×10^8 to 1×10^9 pfu/ml are obtained. Adenoviral constructs with titers in this range are generally suitable for use in most applications. If the titer of the adenoviral stock is less than 1×10^7 pfu/ml, a new adenoviral stock may be produced to increase the titer. See the Troubleshooting section below for more tips and guidelines to optimize the viral yield.

[0473] For some applications, viral titers higher than 1×10^9 pfu/ml may be desired. It is possible to concentrate adenoviral stocks using a variety of methods (*e.g.*, CsCl purification; Engelhardt, J.F., *et al.*, *Nature Genetics* 4:27-34 (1993), without significantly affecting their transducibility. Use of these methods allows generation of adenoviral stocks with titers as high as 1×10^{11} pfu/ml.

[0474] Once an adenoviral stock with a suitable titer is generated, it may be used to transduce the adenoviral construct into the mammalian cell line of choice and assay for expression of the polynucleotide of interest. Guidelines illustrating one method of transduction are provided below, though it will be appreciated that many such methods are known in the art and may be used in the present invention.

- [0475] The pAd/CMV/V5-DEST™ or pAd/PL-DEST™ adenoviral construct is replication-incompetent and does not integrate into the host genome. Therefore, once transduced into the mammalian cells of choice, the gene of interest will be expressed only as long as the viral genome is present. The adenovirus terminal protein (TP) covalently binds to the ends of the viral DNA, and helps to stabilize the viral genome in the nucleus. In actively dividing cells, the adenovirus genome is gradually diluted out as cell division occurs, resulting in an overall decrease in transgene expression over time (generally to background levels within 1-2 weeks after transduction). In non-dividing cells (*e.g.*, quiescent CD34+ cells) or animal tissues (*e.g.*, skeletal muscle, neurons), transgene expression is more stable and can persist for as long as 6 months following transduction.
- [0476] In actively dividing cells (*i.e.*, doubling time of approximately 24 hours), transgene expression is generally detectable within 24 hours of transduction, with maximal expression observed at 48-96 hours (2-4 days) post transduction. Expression levels generally start to decline by 5 days after transduction. In cell lines that exhibit longer doubling times or non-dividing cell lines, high levels of transgene expression typically persist for a longer time. If transducing the adenoviral construct into the mammalian cell line for the first time, a time course of expression may be performed to determine the optimal conditions for expression of the gene of interest.
- [0477] To obtain optimal expression of the gene of interest, the adenoviral construct may be transduced into the mammalian cell line of choice using a suitable MOI. MOI is defined as the number of virus particles per cell and generally correlates with expression. Typically, expression levels increase linearly as the MOI increases.
- [0478] A number of factors can influence determination of an optimal MOI including the nature of the mammalian cell line to be used (*e.g.*, non-dividing vs. dividing cell type), its transduction efficiency, the application of interest, and the nature of the gene of interest. If transducing the adenoviral construct into the mammalian cell line of choice for the first time, using a range of MOIs (*e.g.*, 0, 0.5, 1, 2, 5, 10, 20, 50) to determine the MOI required to obtain optimal expression of the DNA or interest or recombinant polypeptide may be performed.

- [0479] In general, 80-90% of the cells in an actively dividing cell line (*e.g.*, HT1080) express a target gene when transduced at an MOI of ~1. Other cell types including non-dividing cells may transduce adenoviral constructs less efficiently. If transducing the adenoviral construct into a non-dividing cell type, the MOI may be increased to achieve optimal expression levels for the polynucleotide of interest or recombinant polypeptide.
- [0480] The pAd/CMV/V5-GW/lacZ control adenoviral construct may be used to determine the optimal MOI for the particular cell line and application. Once the Ad/CMV/V5-GW/lacZ adenovirus is transduced into the mammalian cell line of choice, the gene encoding β -galactosidase will be constitutively expressed and can be easily assayed (refer to the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual for details, available from Invitrogen Corporation, Carlsbad, CA).
- [0481] Viral supernatants are generated by lysing cells containing virus into spent media harvested from the 293A producer cells. Spent media lacks nutrients and may contain some toxic waste products. If a large volume of viral supernatant is used to transduce the mammalian cell line (*e.g.*, 1 ml of viral supernatant per well in a 6-well plate), growth characteristics or morphology of the target cells may be affected during transduction. These effects are generally alleviated after transduction when the media is replaced with fresh, complete media.
- [0482] The procedure described herein illustrates one method to transduce the mammalian cell line of choice with the adenoviral construct. Other methods suitable for use with the present invention are readily available for use by one skilled in the art. Mammalian cells of choice are plated in complete media. On the day of transduction (Day 1), the adenoviral stock is thawed, and the appropriate amount of virus is diluted (if necessary) into fresh complete medium. The culture medium is removed from the cells. The medium containing virus is mixed gently by pipetting and add to the cells. The plate is swirled gently to disperse the medium, then incubated at 37°C overnight. On the following day (Day 2), the medium containing virus is removed and replaced with fresh, complete culture medium. The cells are harvested (if needed) on the desired day (*e.g.*, 2 days post transduction) and assayed for expression of the polynucleotide of interest or recombinant polypeptide.

[0483] Any method of choice to detect the polynucleotide of interest or recombinant polypeptide of interest including functional analysis, immunofluorescence, northern blot, or western blot. If the gene of interest is cloned in frame with an epitope tag, the recombinant polypeptide of interest may be detected using an antibody to the epitope tag (see the pAd/CMV/V5-DEST™ and pAd/PL-DEST™ GATEWAY™ Vector manual for details, available from Invitrogen, Carlsbad, CA).

Troubleshooting

[0484] Below are listed some potential problems and possible solutions that may help troubleshoot the cotransfection and titering experiments.

Problem	Reason	Solution
Low viral titer	Low transfection efficiency: Incomplete Pac I digestion or digested DNA contaminated with phenol, ethanol, or salts Unhealthy 293A cells; cells exhibit low viability 293A cells plated too sparsely Plasmid DNA:transfection reagent ratio incorrect	Repeat the Pac I digestion. Make sure that the purified DNA is not contaminated with phenol, ethanol, or salts. Use healthy 293A cells; do not overgrow. Cells should be 90-95% confluent at the time of transfection. Optimize such that plasmid DNA (in µg):Lipofectamine™ 2000 (in µl) ratio ranges from 1:2 to 1:3. If using another transfection reagent, optimize according to the manufacturer's recommendations.
	Viral supernatant too dilute	Concentrate virus using CsCl purification (Engelhardt, J.F., et al., Nature Genetics 4:27-34 (1993) or any method of choice.
	Viral supernatant frozen and thawed multiple times	Do not freeze/thaw viral supernatant more than 10 times.
	Gene of interest is large	Viral titers generally decrease as the size of the insert increases; inserts larger than 6 kb (for pAd/CMV/V5-DEST™) and 7.5 kb (for pAd/PL-DEST™) are not recommended.
No plaques obtained upon titering	Gene of interest is toxic to cells	Generation of constructs containing activated oncogenes or potentially harmful genes is not recommended.
	Viral stocks stored incorrectly	Aliquot and store stocks at -80°C. Do not freeze/thaw more than 10 times.
	Incorrect titering cell line used	Use the 293A cell line or any cell line with the characteristics discussed.
	Agarose overlay incorrectly prepared	Make sure that the agarose is not too hot before addition to the cells; hot agarose will kill the cells.

Transducing Mammalian Cells

[0485] Below are listed some potential problems and possible solutions that may help troubleshoot the transduction and expression experiment.

Problem	Reason	Solution
Titer indeterminate; cells confluent	Viral supernatant not diluted sufficiently	Titer adenovirus using 10-fold serial dilutions ranging from 10^{-4} to 10^{-9} .
No expression	Viral stocks stored incorrectly	Aliquot and store stocks at -80°C . Do not freeze/thaw more than 10 times.
Poor expression	Gene of interest contains a Pac I site	Perform mutagenesis to change or remove the Pac I site.
	Poor transduction efficiency: Mammalian cells not healthy	Make sure that the cells are healthy before transduction.
	Non-dividing cell type used	Transduce the adenoviral construct into cells using a higher MOI.
	MOI too low	Transduce the adenoviral construct into cells using a higher MOI.
	Low viral titer	Amplify the adenoviral stock using the procedure.
Persistent toxicity in target cells	Adenoviral stock contaminated with RCA	Screen for RCA contamination (Dion, L.D., et al., J. Virol. Methods 56:99-107 (1996)). Prepare a new adenoviral stock or plaque purify to isolate recombinant adenovirus.
	Cells harvested too soon after transduction	Do not harvest cells until at least 24-48 hours after transduction.
	Cells harvested too long after transduction	For actively dividing cells, assay for maximal levels of recombinant polypeptide expression within 5 days of transduction.
	Gene of interest is toxic to cells	Generation of constructs containing activated oncogenes or potentially harmful genes is not recommended.
	Too much crude viral stock used	Reduce the amount crude viral stock used for transduction or dilute the crude viral stock.
Wild-type RCA contamination		Amplify the adenoviral stock.
		Concentrate the crude viral stock.
		Screen for RCA contamination (Dion, L.D., et al., J. Virol. Methods 56:99-107 (1996); Zhang, W. W., et al., BioTechniques 18:444-447 (1995). Plaque purify to isolate recombinant adenovirus or prepare a new adenoviral stock.

RECIPES

4% Agarose

- [0486] This procedure may be used to prepare a 4% Agarose solution.
- [0487] Materials Needed: Ultra Pure Agarose (Invitrogen, Catalog No. 15510-027) Deionized, sterile water.
- [0488] Protocol: Prepare a 4% stock solution in deionized, sterile water. Autoclave at 121°C for 20 minutes to sterilize. Equilibrate to 65°C in a water bath and use immediately or store at room temperature indefinitely. If the agarose solution is stored at room temperature, melting the agarose is required before use. To melt, microwave the agarose to melt, then equilibrate to 65°C in a water bath before use.

5 mg/ml MTT

- [0489] This procedure may be used to prepare a 5 mg/ml MTT solution.
- [0490] Materials Needed: 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium bromide; Thiazolyl blue (MTT; Sigma-Aldrich, St. Louis, MO, Catalog No. M2128). Phosphate-Buffered Saline (PBS; Invitrogen, Catalog No. 10010-023).
- [0491] Protocol: Prepare a 5 mg/ml stock solution in PBS. Filter-sterilize and dispense 5 ml aliquots into sterile, conical tubes. Store at +4°C for up to 6 months.

EXAMPLE 5

- [0492] The present invention provides materials and methods for the stable expression of heterologous polypeptides in cells (*e.g.*, insect cells). pIB/V5-His-DEST and pIB/V5-His-GW/*lacZ* are nucleic acid molecules of the invention that are commercially available from Invitrogen Corporation, Carlsbad, CA. Information concerning the construction and use of these vectors may be found in Catalog no. 12550-018 Version A, July 15, 2002, 25-0607, available from Invitrogen Corporation, Carlsbad, CA.
- [0493] Nucleic acid molecules of the invention may be used to express a polypeptide of interest as part of a fusion polypeptide. Numerous suitable

fusion partners are known to those in the art. For example a polypeptide of interest may be expressed as a fusion polypeptide containing the V5 epitope. Antibodies to detect the V5 epitope, a 14 amino acid epitope derived from the P and V proteins of the paramyxovirus, SV5 having the sequence GKPIPNLLGLDST (Southern, J.A., *et al.*, *J. Gen. Virol.* 72:1551-1557 (1991)) are commercially available from Invitrogen Corporation, Carlsbad, CA, for example, Anti-V5 Antibody catalog no. R960-25, Anti-V5-HRP Antibody catalog no. R961-25, and catalog no. Anti-V5-AP Antibody R962-25. A polypeptide of interest may be expressed as a fusion polypeptide with a polyhistidine sequence. Antibodies to detect a polyhistidine sequence are commercially available from Invitrogen Corporation, Carlsbad, CA. For example, Anti-His(C-term) Antibody catalog no. R930-25, Anti-His(C-term)-HRP Antibody catalog no. R931-25, and Anti-His(C-term)-AP Antibody R932-25, all of which detect a C-terminal polyhistidine (6xHis) tag and require the free carboxyl group for detection (*i.e.*, detect the sequence HHHHHH-COOH, see Lindner, P., *et al.*, *BioTechniques* 22:140-149 (1997)).

[0494] An open reading frame present on a sequence of interest may be cloned in frame with the C-terminal peptide containing the V5 epitope and the polyhistidine (6xHis) and Immobilized Metal Affinity Chromatography (IMAC) may be used to purify the recombinant fusion polypeptide. The ProBond™ Purification System as well as the Ni-NTA Purification System are available from Invitrogen Corporation, Carlsbad, CA.

Product	Catalog no.
ProBond™ Purification System	K850-01
ProBond™ Nickel-chelating Resin	R801-01
	R801-15
ProBond™ Purification System with Anti-His(C-term)-HRP Antibody	K853-01
ProBond™ Purification System with Anti-V5-HRP Antibody	K854-01
Purification Columns (10 ml polypropylene columns)	R640-50
Ni-NTA Purification System	K950-01
Ni-NTA Agarose	R901-01
	R901-15
Ni-NTA Purification System with Anti-His(C-term)-HRP Antibody	K953-01
Ni-NTA Purification System with Anti-V5-HRP Antibody	K954-01

[0495] pIB/V5-His-DEST is a 5.0 kb vector derived from pIB/V5-His and adapted for use with GATEWAY™ Technology. It is designed to allow transient

or stable expression of a sequence of interest, which may encode a polypeptide, in insect cell lines.

[0496] pIB/V5-His-DEST contains the following features:

Feature	Benefit
OpIE2 promoter	Allows constitutive expression of the gene of interest in lepidopteran insect cells (Theilmann, D.A., and Stewart, S., Virology 187:84-96 (1992))
attR1 and attR2 sites	Allows recombinational cloning of the gene of interest from an entry clone.
Chloramphenicol resistance gene (Cm ^R)	Allows counterselection of expression clones.
ccdB gene	Allows negative selection of expression clones.
V5 epitope	Allows detection of a recombinant polypeptide with the Anti-V5 Antibodies (Southern, J.A., et al., J. Gen. Virol. 72:1551-1557 (1991))
C-terminal polyhistidine tag	Allows purification of recombinant polypeptides on metal-chelating resin such as ProBond™ or Ni-NTA. Allows detection of the recombinant polypeptide by the Anti-His (C-term) Antibodies (Lindner, P., et al., BioTechniques 22:140-149 (1997))
OpIE2 polyadenylation sequence	Efficient transcription termination and polyadenylation of mRNA (Theilmann, D.A., and Stewart, S., Virology 187:84-96 (1992))
pUC origin	Allows high-copy number replication and growth in E. coli.
GP64 promoter	Allows constitutive expression of the blasticidin resistance gene in lepidopteran insect cells (Blissard, G.W., et al., Virology 190:783-793 (1992); Blissard, G.W., and Rohrmann, G.F., J. Virology 65:5820-5827 (1991))
EM7 promoter	Allows efficient expression of the blasticidin and ampicillin resistance genes in E. coli.
Blasticidin resistance gene (bsd)	Allows generation of stable insect cell lines (Kimura, M., et al., Biochim. Biophys. ACTA 1219:653-659 (1994))
Ampicillin resistance gene (bla)	Allows selection of transformants in E. coli Note: The native promoter has been removed. Transcription is assumed to start from the EM7 promoter.

A map of pIB/V5-His-DEST is provided in Figure 15 and the nucleotide sequence of the vector is provided in Table 12.

[0497] GATEWAY™ is a universal cloning technology that takes advantage of the site-specific recombination properties of bacteriophage lambda (Landy, 1989) to provide a rapid and highly efficient way to move a gene of interest into multiple vector systems. To express a sequence of interest using

GATEWAY™ Technology: clone the sequence of interest into a GATEWAY™ entry vector to create an entry clone; generate an expression clone by performing an LR recombination reaction between the entry clone and a GATEWAY™ destination vector (e.g. pIB/V5-His-DEST); and introduce the expression clone into insect cells for transient or stable expression.

[0498] Baculovirus immediate-early promoters utilize the host cell transcription machinery and do not require viral factors for activation. The *OpIE2* promoter is from the baculovirus *Orgyia pseudotsugata* multicapsid nuclear polyhedrosis virus (*OpMNPV*) and drives constitutive expression of the gene of interest in pIB/V5-His-DEST. The virus' natural host is the Douglas fir tussock moth; however, the promoter allows protein expression in *Lymantria dispar* (LD652Y), *Spodoptera frugiperda* cells (Sf9) (Hegedus, D.D., et al., *Gene* 207:241-249 (1998); Pfeifer, T.A., et al., *Gene* 188:183-190 (1997)), *Sf21* (Invitrogen), *Trichoplusia ni* (High Five™, Invitrogen Corporation, Carlsbad, CA), *Drosophila* (Kc1, S2) (Hegedus, D.D., et al., *Gene* 207:241-249 (1998); Pfeifer, T.A., et al., *Gene* 188:183-190 (1997)) and mosquito cell lines. The *OpIE2* promoter has been sequenced and analyzed. The sequence of the promoter is provided in Figure 16.

[0499] Although the *OpIE2* promoter provides relatively high levels of constitutive expression, some proteins may not be expressed at levels seen with baculovirus late promoters such as polyhedrin or very late promoters such as p10 (Jarvis, D.L., et al., *Protein Expression and Purification* 8:191-203 (1996)). Typical expression levels range from 1-2 µg/ml (human IL-6; Invitrogen) to 8-10 µg/ml (human melanotransferrin) (Hegedus, D.D., et al., *Protein Expression and Purification* 15:296-307 (1999)).

[0500] The *OpIE2* promoter has been analyzed by deletion analysis using a CAT reporter in both *Lymantria dispar* (LD652Y) and *Spodoptera frugiperda* (Sf9) cells. Expression in Sf9 cells was much higher than in LD652Y cells. Deletion analysis revealed that sequence up to -275 base pairs from the start of transcription is necessary for maximal expression (Theilmann, D.A., and Stewart, S., *Virology* 187:84-96 (1992)). Additional sequence beyond -275 may broaden the host range expression of this plasmid to other insect cell lines. In addition, an 18 bp element appears to be required for expression. This 18 bp element is repeated almost completely in three different locations and

partially at six other locations. These are marked in Fig. 16. Elimination of the three major 18 bp elements reduces expression to basal levels (Theilmann, D.A., and Stewart, S., *Virology* 187:84-96 (1992)). Primer extension experiments revealed that transcription initiates equally from either the C or the A indicated. These two transcriptional start sites are adjacent to a CAGT sequence motif that has been shown to be conserved in a number of early genes (Blissard, G.W., and Rohrmann, G.F., *Virology* 170:537-555 (1989)).

[0501] The *GP64* promoter regulates expression of the baculovirus major envelope glycoprotein gene (*GP64*) of the budded virion. Studies have shown that while the *GP64* promoter is stimulated by the transcriptional transactivator IE-1, low levels of activity still occur without transactivation (Blissard, G.W., *et al.*, *Virology* 190:783-793 (1992); Blissard, G.W., and Rohrmann, G.F., *J. Virology* 65:5820-5827 (1991)). Furthermore, deletion analysis has identified the specific region required for transcriptional initiation in the absence of IE-1 (Blissard, G.W., *et al.*, *Virology* 190:783-793 (1992); Blissard, G.W., and Rohrmann, G.F., *J. Virology* 65:5820-5827 (1991)).

[0502] pIB/V5-His-DEST contains a 100 bp region of the *Autographa californica* nuclear polyhedrosis virus (AcMNPV) *GP64* promoter which is sufficient for activation of the blasticidin resistance gene (*bsd*) in the absence of any baculovirus proteins. Using standard blasticidin concentrations (10-80 µg/ml), stable transfectants will only be selected if the *bsd* gene is expressed at suitable levels. Without wishing to be bound by theory, because of the minimal activity of the *GP64* promoter, it is likely that only stable transfectants containing pIB/V5-His-DEST integrated into the most transcriptionally active genomic loci will be selected. This allows generation of stable cell lines which will express higher levels of the protein of interest compared to cell lines expressing the *bsd* gene product from the *OpIE1* promoter, as in the parent pIB/V5-His vector.

[0503] Cell cultures of either *Sf9* (catalog no. B82501, Invitrogen Corporation, Carlsbad, CA), *Sf21* (catalog no. B82101, Invitrogen Corporation, Carlsbad, CA), or High Five™ cells (catalog no. B85502, Invitrogen Corporation, Carlsbad, CA) may be used in connection with the present invention and may be grown and stored using conventional techniques well known in the art (*e.g.*,

Baculoviral Expression Systems and Insect Cell Lines manual, February 27, 2002, Invitrogen Corporation, Carlsbad, CA).

[0504] The pIB/V5-His-DEST vector is supplied as a supercoiled plasmid. Linearization of this vector is not required to obtain optimal results for any downstream application. The vector may be resuspended at a concentration of 50-150 ng/ μ l in sterile water, pH 8.0. To propagate and maintain pIB/V5-His-DEST, Library Efficiency[®] DB3.1[™] Competent Cells (Invitrogen Corporation, Carlsbad, CA Catalog no. 11782-018) may be used. The DB3.1[™] *E. coli* strain is resistant to CcdB effects and can support the propagation of plasmids containing the *ccdB* gene. To maintain integrity of the vector, select for transformants in media containing 50-100 μ g/ml ampicillin and 15 μ g/ml chloramphenicol. The use of general *E. coli* cloning strains including TOP10 or DH5 α is not recommended for propagation and maintenance as these strains are sensitive to CcdB effects. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the pcDNA destination vector containing attP sites flanking the *ccdB* and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. It has recently been found that the use of spectinomycin selection instead of chloramphenicol selection results in an increase in the number of colonies obtained on selection plates, indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene.

[0505] To recombine a sequence of interest into pIB/V5-His-DEST, an entry clone containing the sequence of interest may be prepared. A commercially available kit (e.g., the pENTR Directional TOPO[®] Cloning Kit, Invitrogen Corporation, Carlsbad, CA Catalog no. K2400-20, version B) can be used. Other suitable entry vectors are available from Invitrogen Corporation, Carlsbad, CA. Detailed information on constructing an entry clone may be obtained from the manual provided with the specific entry vector. For detailed information on performing the LR recombination reaction, refer to the GATEWAY[™] Technology manual, Invitrogen Corporation, Carlsbad, CA.

[0506] A sequence of interest may contain a Kozak consensus sequence with an ATG initiation codon for proper initiation of translation (Kozak, M., *Nucleic Acids Res.* 15:8125-8148 (1987); Kozak, M., *J. Cell Biology* 115:887-903 (1991); Kozak, M., *Proc. Natl. Acad. Sci. USA* 87:8301-8305 (1990)). An example of a Kozak consensus sequence is provided below. Other sequences are possible, but the G or A at position -3 and the G at position +4 are the most critical for function (shown in bold). The ATG initiation codon is shown underlined.

(G/A)NNATGG

[0507] To include the V5 epitope and/or 6xHis tag encoded by the vector, the sequence of interest may not contain a stop codon. A coding sequence should also be designed to be in frame with the C-terminal epitope tag after recombination. To express a polypeptide with a native C-terminal (*i.e.*, without the V5 epitope and/or 6xHis tag), the sequence of interest should contain a stop codon in the entry clone.

[0508] Each entry clone contains *attL* sites flanking the sequence of interest. Sequences of interest in an entry clone may be transferred to the destination vector backbone by mixing the DNAs with the GATEWAY™ LR Clonase™ enzyme mix. The resulting LR recombination reaction may then be transformed into *E. coli* and the expression clone may be selected. In an embodiment, recombination between the *attR* sites on the destination vector and the *attL* sites on the entry clone replaces the *ccdB* gene and the chloramphenicol (Cm^R) gene with the sequence of interest and results in the formation of *attB* sites in the expression clone.

[0509] The LR Clonase™ reaction; subsequent transformation of a suitable *E. coli*, and selection for an expression clone may be performed using standard techniques such as those provide in the GATEWAY™ Technology manual.

[0510] The *ccdB* gene mutates at a very low frequency, resulting in a very low number of false positives. True expression clones will be ampicillin-resistant and chloramphenicol-sensitive. Transformants containing a plasmid with a mutated *ccdB* gene will be both ampicillin- and chloramphenicol-resistant. A putative expression clone can be tested by growth on LB plates containing 30

µg/ml chloramphenicol. A true expression clone will not grow in the presence of chloramphenicol.

[0511] The recombination region of the expression clone resulting from pIB/V5-His-DEST × entry clone is shown in Fig. 17. Shaded regions correspond to those DNA sequences transferred from the entry clone into pIB/V5-His-DEST by recombination. Non-shaded regions are derived from the pIB/V5-His-DEST vector. The underlined nucleotides flanking the shaded region correspond to bases 609 and 2292, respectively, of the pIB/V5-His-DEST vector sequence.

[0512] To confirm that a coding sequence on the sequence of interest is in frame with the C-terminal V5 epitope and polyhistidine tag, the expression construct may be sequenced, for example, using the OpIE2 Forward and Reverse primer sequences. Refer to Fig. 17 for the sequence and location of the primer binding sites.

[0513] Plasmid DNA for transfection into insect cells must be very clean and free from phenol and sodium chloride. Contaminants will kill the cells, and salt will interfere with lipid complexing, decreasing transfection efficiency. The expression construct plasmid may be prepared using standard techniques, for example, column chromatography(*e.g.*, the S.N.A.P.TM MiniPrep Kit Catalog no. K1900-01, Invitrogen Corporation, Carlsbad, CA). Typical yields of plasmid using this technique are 10-15 µg of plasmid DNA from 10-15 ml of bacterial culture. Plasmid can be used directly for transfection of insect cells.

[0514] One technique suitable to introduce the nucleic acid molecules of the invention into host cells is lipid-mediated transfection (*e.g.*, using Cellfectin® Reagent, catalog no. 10362010, Invitrogen Corporation, Carlsbad, CA). Other lipids may be substituted, although transfection conditions may have to be optimized. Expected Transfection Efficiency using Cellfectin® Reagent: 40-60% for Sf9 or Sf21 cells and 40-60% for High FiveTM cells. Other transfection methods (*e.g.*, calcium phosphate and electroporation (Mann and King, 1989)) may also be used with High FiveTM cells.

[0515] Controls may be included in the transfection reaction, for example, pIB/V5-His-GW/lacZ vector as a positive control for transfection and

expression and lipid only as a negative control DNA only to check for DNA contamination.

[0516] pIB/V5-His-GW/lacZ is provided as a positive control vector for transfection and expression (see Fig. 18 for a map). The vector allows expression of a C-terminally tagged β -galactosidase fusion polypeptide that may be detected by Western blot or functional assay. pIB/V5-His-GW/lacZ is a 6478 bp control vector containing the gene for β -galactosidase. pIB/V5-His-GW/lacZ was constructed using the GATEWAY™ LR recombination reaction between an entry clone containing the lacZ gene and pIB/V5-His-DEST. β -galactosidase is expressed as a fusion to the C-terminal tag. The molecular weight of the fusion polypeptide is approximately 120 kDa.

[0517] To propagate and maintain the plasmid: resuspend the vector in 10 μ l sterile water to prepare a 1 μ g/ μ l stock solution and use the stock solution to transform a recA, endA E. coli strain like TOP10, DH5a, JM109, or equivalent. Select transformants on LB agar plates containing 50-100 μ g/ml ampicillin. Optionally, a glycerol stock of a transformant containing plasmid may be prepared for long-term storage.

[0518] For each transfection, log-phase cells with greater than 95% viability may be used. A time course for expression of the sequence of interest may be performed. For example, expression of a polypeptide encoded by the sequence of interest may be assayed for at 2, 3, and 4 days post transfection. One or more 60 mm plate may be used for each time point. For Sf9, Sf21, or High Five™ cells, 1×10^6 cells may be seeded in appropriate serum-free medium in a 60 mm dish. Rock gently from side to side for 2 to 3 minutes to evenly distribute the cells. Cells may be 50 to 60% confluent.

[0519] Incubate the cells for at least 15 minutes without rocking to allow the cells to fully attach to the bottom of the dish to form a monolayer of cells.

[0520] Verify that the cells have attached by inspecting them under an inverted microscope.

[0521] Nucleic acid molecules of the invention may be introduced into host cells using standard techniques. A protocol for use of Cellfectin® Reagent is provided below. Other conditions for transfection may be empirically determined by one skilled in the art using routine experimentation. Preferably, a plasmid is not linearized prior to introduction into a host cell. Linearizing a

plasmid appears to decrease protein expression. The reason for this is not known.

[0522] A suitable transfection may employ: 1-10 μg of purified pIB/V5-His-DEST expression construct ($\sim 1 \mu\text{g}/\mu\text{l}$ in TE buffer); either log-phase *Sf9* or *Sf21* cells ($1.6\text{-}2.5 \times 10^6$ cells/ml, >95% viability) or log-phase High Five™ cells ($1.8\text{-}2.3 \times 10^6$ cells/ml, >95% viability), growing in serum-free medium (e.g., Grace's Medium without supplements; serum-free medium 60 mm tissue-culture dishes; 1.5 ml sterile microcentrifuge tubes; rocking platform only (NOT orbital); 27°C incubator; inverted microscope; paper towels and air-tight bags or containers; and 5 mM EDTA, pH 8.

[0523] Transfection may comprise mixing plasmid DNA and Cellfectin® in an appropriate medium and incubating with freshly seeded insect cells. The amount of cells, liposomes, and plasmid DNA described herein has been optimized for 60 mm culture plates. Other transfection conditions may be used with other size plates or flasks. Optimizing conditions for other volumes of transfection may be accomplished by one skilled in the art using routine experimentation. Serum-free medium (e.g., *Sf*-900 II SFM (catalog no. 1090207) to transfect *Sf9* or *Sf21* cells and Express Five® SFM (catalog no. 10486017) to transfect High Five™ cells, available from Invitrogen Corporation, Carlsbad, CA) can be used. Grace's Medium without supplements may also be used. The proteins in the FBS and supplements will interfere with the liposomes, causing the transfection efficiency to decrease.

[0524] To prepare each transfection mixture, a 1.5 ml microcentrifuge tube may be used. The following reagents may be added: 1 ml of Grace's Medium OR appropriate serum-free medium; 1-10 μl nucleic acid molecule of the invention (e.g., pIB/V5-His plasmid or construct) at a concentration of $\sim 1 \mu\text{g}/\mu\text{l}$ in TE, pH 8; 20 μl Cellfectin® Reagent (mixed well before use and always added last). The transfection mixture may be mixed gently for 10 seconds and incubated at room temperature for 15 minutes.

[0525] The medium covering the cells to be transfected should be removed without disrupting the monolayer. If the medium contained serum, wash the cells by carefully adding 2 ml of fresh Grace's Medium without supplements or FBS to remove trace amounts of serum that will decrease the efficiency of liposome transfection and remove the wash.

- [0526] The entire transfection mix described above may be added dropwise into the 60 mm dish. The drops may be evenly distributed over the monolayer. This method reduces the chances of disturbing the monolayer. Repeat for all transfections.
- [0527] The dishes may be incubated at room temperature for 4 hours on a side-to-side, rocking platform. A suitable speed for the platform is ~2 side to side motions per minute. Instead of a platform rocker, the dishes may be manually rocked periodically.
- [0528] Following the 4-hour incubation period, 1-2 ml of complete TNM-FH medium (Sf9 or Sf21 cells) or the appropriate serum-free medium (Sf9, Sf21, or High Five™ cells) may be added to each 60 mm dish. The dishes may be placed in a sealed plastic bag with moist paper towels to prevent evaporation and incubated at 27°C. It is not necessary to remove the transfection solution as Cellfectin® Reagent is not toxic to the cells. If a different lipid is used and loss of viability is observed, then remove the transfection solution after 4 hours, rinse twice with medium, and replace with 1-2 ml of fresh medium.
- [0529] The cells may be harvested, for example, at 2, 3, and 4 days post transfection and assayed for expression of the sequence of interest. Additional fresh medium need not be added to the cells if the cells are sealed in an airtight plastic bag with moist paper towels.
- [0530] Expression of a sequence of interest from the expression clone can be performed in transiently transfected cells or stable cell lines. A sample protocol to detect by Western blot a polypeptide encoded by a sequence of interest expressed as a fusion polypeptide is provided below.
- [0531] The cells from one 60 mm plate may be used for each expression experiment. A suitable cell lysis buffer may be used. One suitable buffer is 50 mM Tris, pH 7.8, 150 mM NaCl, 1% Nonidet P-40.
- [0532] The medium may be removed from the cells. If the polypeptide expressed from the sequence of interest is predicted to be secreted, save and assay both the medium and the cell pellet. Cell lysis buffer, 100 µl, may be added to the plate and the cells may be sloughed or scraped into a microcentrifuge tube. The cells may be vortexed to ensure they are completely lysed. The lysed cells may be centrifuged at maximum speed in a microfuge for 1-2 minutes to pellet nuclei and cell membranes. The

supernatant may be transferred to a new tube. If a membrane protein is expressed from the sequence of interest, it may be located in the pellet. The pellet and the lysate may be assayed. The protein concentration in the lysate may be determined, for example, by the Bradford, Lowry, or BCA assays (Pierce).

[0533] Samples may be mixed with SDS-PAGE sample buffer as follows: 30 μ l lysate with 10 μ l 4X SDS-PAGE sample buffer; the pellet may be resuspended in 100 μ l 1X SDS-PAGE sample buffer; 30 μ l medium may be mixed with 10 μ l 4X SDS-PAGE sample buffer. Because of the volume of medium, it is difficult to normalize the amount loaded on an SDS-PAGE gel. Optionally, the medium may be concentrated to facilitate normalization. Samples may be boiled for 5 minutes, centrifuged briefly, and approximately 3 to 30 μ g protein loaded per lane of an SDS-PAGE gel. The same volume of sample may be added for both the pellet sample and the lysate sample. The amount to load may be determined by one skilled in the art using routine experimentation. Samples may be separated by electrophoresis, blotted, and probed with a suitable antibody using standard techniques.

[0534] A polypeptide expressed from a sequence of interest as a fusion polypeptide may be detected by Western blot analysis, for example, with the Anti-V5 antibodies or the Anti-His(C-term) antibodies available from Invitrogen Corporation, Carlsbad, CA or an antibody that specifically recognizes the polypeptide. In addition, the Positope™ Control Protein (Invitrogen Corporation, Carlsbad, CA, Catalog no. R900-50) is available for use as a positive control for detection of fusion proteins containing a V5 epitope or a 6xHis tag.

[0535] If the pIB/V5-His-GW/lacZ plasmid is used as a positive control vector, β -galactosidase expression may be assayed by Western blot analysis or activity assay (Miller, J.H., *Experiments in Molecular Genetics*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York (1972)). Commercially available antibodies (e.g., Invitrogen Corporation, Carlsbad, CA, β -Gal Antiserum, Catalog no. R901-25), or assay kits (e.g., Invitrogen Corporation, Carlsbad, CA β -Gal Assay Kit, Catalog no. K1455-01 and β -Gal Staining Kit Catalog no. K1465-01) may be used for detection of β -galactosidase expression.

- [0536] The C-terminal peptide containing the V5 epitope and the polyhistidine tag will add approximately 5 kDa in molecular weight to a polypeptide expressed from a sequence of interest.

Selecting Stable Cell Lines

- [0537] Stable expression cell lines can be created for long-term storage and large-scale production of the desired polypeptide. Note that stable cell lines are created by multiple copy integration of the vector. Amplification as in the case with calcium phosphate transfection and hygromycin resistance in *Drosophila* is generally not observed.
- [0538] Blasticidin may be used to select for stably transformed cells. Gloves, mask, goggles, and protective clothing (e.g. a laboratory coat) should be worn when handling blasticidin. Weighing blasticidin and preparing solutions should be done in a hood. Blasticidin may be inactivated for disposal by adding sodium bicarbonate. Blasticidin is soluble in water and acetic acid. Water is generally used to prepare stock solutions of 5 to 10 mg/ml. Blasticidin may be dissolved in sterile water and filter-sterilized. Blasticidin is unstable in solutions with a pH greater than 8.0. The pH of a solution of blasticidin may be 7.0. Blasticidin solutions may be divided into aliquots in small volumes and frozen at -20°C for long-term storage or stored at +4°C for short term storage. Aqueous stock solutions are stable for 1-2 weeks at +4°C and 6-8 weeks at -20°C. Stock solutions should not be subjected to multiple freeze/thaw cycles (do not store in a frost-free freezer). Solutions should be discarded after 1-2 weeks storage at +4°C.
- [0539] Cytopathic effects should be visible within 3-5 days depending on the concentration of blasticidin in the medium. Sensitive cells will enlarge and become filled with vesicles. The outer membrane will show signs of blebbing, and cells will eventually detach from the plate. Blasticidin-resistant cells should continue to divide at regular intervals to form distinct colonies. There should not be any distinct morphological changes between blasticidin-resistant cells compared to cells not under selection with blasticidin.
- [0540] In general, concentrations of blasticidin around 10 µg/ml will kill Sf9 or Sf21 cells (in complete TNM-FH medium) and concentrations around 20

µg/ml will kill High Five™ cells (in Express Five® SFM) within one week, although a few cells may remain that exclude trypan blue. To obtain faster and more thorough killing, 50-80 µg/ml blasticidin may be used. Once blasticidin-resistant clones have been obtained, cells may be maintained in lower concentrations of blasticidin (*e.g.*, 10-20 µg/ml). An appropriate concentration of blasticidin for any specific cell type may be determined by one skilled in the art by performing a kill curve.

[0541] A suitable protocol for establishing a kill curve is provided. Assays may be conducted in 24-well tissue culture plates. Suitable medium (*e.g.*, TNM-FH medium or the serum-free medium of choice) may be prepared and supplemented with concentrations ranging from 0 to 100 µg/ml blasticidin. Generally, concentrations that effectively kill lepidopteran insect cells within a week are in the 50 to 80 µg/ml range. While 10-20 µg/ml blasticidin will kill cells within a week, higher concentrations will result in faster and more thorough killing. In addition, using higher concentrations of blasticidin may result in enrichment of clones containing multiple integrations of a sequence of interest. Test varying concentrations of blasticidin on a cell line of interest to determine the concentration that kills the cells within a week (kill curve). The concentration of drug that kills the cells of interest within a week should be used.

[0542] To isolate a stable cell line, a mock transfection and a positive control (*e.g.*, pIB/V5-His-GW/lacZ) may be used. Cells may be transfected as described above. Forty-eight hours post transfection, the transfection solution may be removed and fresh medium containing no blasticidin may be added. The cells may be split 1:5 (20% confluent) and allowed to attach overnight before adding selective medium. The medium may be removed and replaced with medium containing blasticidin at the appropriate concentration. The cells may be incubated at 27°C. The selective medium may be replaced every 3 to 4 days until foci are observed. Cloning cylinders or limiting dilution may be used to isolate clonal cell lines. Optionally, resistant cells may be allowed to continue grow out to confluence for a polyclonal cell line (2 to 3 weeks).

[0543] A polyclonal cell line may be isolated by allowing the resistant cells grow to confluence and splitting the cells 1:5. The polyclonal cell line may be tested for expression. Medium without blasticidin should be used when

splitting cells and cells should be allowed to attach before adding selective medium.

- [0544] Resistant cells may be expanded into flasks to prepare frozen stocks. Medium containing blasticidin should be used when maintaining stable lepidopteran cell lines. The concentration of blasticidin may be lowered to 10 µg/ml for maintenance.

Isolation of Clonal Cell Lines Using Cloning Cylinders

- [0545] Multiple foci may be isolated for expression testing. As in mammalian cell culture, the location of integration may affect expression of a sequence of interest. Selections may be performed in small plates or wells. Cells should not be allowed to dry out during the selection.

- [0546] The closed plate may be examined under a microscope and the location of one or more colony marked on the top of the plate. The markings may then be transferred to the bottom of the plate. Orientation marks may be included. Each colony may contain 50 to 200 cells. *Sy9* cells tend to spread more than High Five™ cells. The culture dish may be moved to a sterile cabinet and the lid removed. A thin layer of sterile silicon grease may be applied to the bottom of a cloning cylinder (Scienceware, Catalog no. 378747-00 or Belco, Catalog no. 2090-00608), using a sterile cotton-tipped wooden applicator. The layer should be thick enough to retard the flow of liquid from the cylinder, without obscuring the opening on the inside. Cloning cylinders and silicon grease can be sterilized together by placing a small amount of grease in a glass petri dish and placing the cloning cylinders upright in the grease. After autoclaving, the grease will have spread out in a thin layer to coat the bottom of the cylinders.

- [0547] The culture medium may be removed and the cylinder placed firmly and directly over the marked area. A microscope may be used to direct placement of the cylinder. 20 to 100 µl of medium (no blasticidin) may be used to dislodge the cells. The cells and medium may be removed and transferred to a microtiter plate and the cells may be allowed to attach. The medium may be removed and replaced with selective medium for culturing.

The cell line may be expanded and tested for expression of the sequence of interest.

Isolation of Clonal Cell Lines Using a Dilution Method

- [0548] Clonal cell lines may be established using a dilution method. The objective of this method is to dilute the cells so that under selective pressure only one stable viable cell per well is achieved. The higher transfection efficiency, the more the cells should be diluted. The protocol below works well with cells transfected at 5-10% efficiency.
- [0549] Forty-eight hours after transfection, cells may be diluted to 1×10^4 cells/ml in medium without blasticidin. Other dilutions of the culture may also be used as transfection efficiency will determine how many transformed cells there will be per well. 100 μ l of the cell solution may be added to 32 wells of a 96-well microtiter plate (8 rows by 4 columns). The remaining cells may be diluted 1:1 with medium without blasticidin and add 100 μ l of this solution added to the next group of 32 wells (8 x 4). The remaining cells may be diluted 1:1 with medium without blasticidin and 100 μ l of this solution added to the last group of 32 wells. Although the cells can be diluted to low numbers, cell density is critical for viability. If the density drops below a certain level, the cells will not grow.
- [0550] The cells may be allowed to attach overnight, then the medium removed and replaced with medium containing blasticidin. Removing and replacing medium may be tedious. Optionally, it is possible to dilute the cells directly into selective medium if they are handled gently.
- [0551] The plate may be wrapped and incubated at 27°C for 1 week. It is not necessary to change the medium or place in a humid environment. The plate may be checked after a week and the wells that have only one colony may be marked. The plate may be incubated until the colony fills most of the well. The cells may be harvested and transferred to a 24-well plate with 0.5 ml of fresh medium containing blasticidin. The clone may be expanded to 12- and 6-well plates, and finally to a T-25 flask.
- [0552] Each cell line may be assayed for yield of the desired polypeptide and the one with the highest yield may be scaled-up and used for purification of

recombinant polypeptide. For secreted polypeptides, the cell pellet as well as the medium may be assayed. The yield of polypeptide in the cells may be compared to the yield of polypeptide in the medium.

- [0553] Master stocks and working stocks of stable cell lines may be prepared prior to scale-up and purification.

Purification

- [0554] A polypeptide expressed from a sequence of interest may be purified using standard techniques. Stable cell lines prepared as described above may be expanded into larger flasks, spinners, shake flasks, or bioreactors to obtain the desired yield of polypeptide. If a polypeptide expressed from a sequence of interest is secreted, cells may be cultured in serum-free medium to simplify purification.
- [0555] A 6His tagged fusion polypeptide may be purified using the ProBond™ Purification System, the Ni-NTA Purification System, or a similar product. Both purification systems contain a metal-chelating resin specifically designed to purify 6xHis-tagged polypeptides.
- [0556] Cells may be maintained in a medium having a concentration of blasticidin of 10 µg/ml. Cells may be switched from complete TNM-FH medium to serum-free medium during passage.
- [0557] Adding serum-free medium directly to a metal-chelating resin such as ProBond™ to purify a secreted polypeptide from serum-free medium will strip the nickel ions from the resin. To purify 6xHis-tagged recombinant polypeptides from the culture medium, dialysis or ion exchange chromatography may be performed prior to affinity chromatography on metal-chelating resins. Dialysis allows removal of media components that strip Ni^{+2} from metal-chelating resins. Ion exchange chromatography allows removal of media components that strip Ni^{+2} from metal-chelating resins and concentration of sample for easier manipulation in subsequent purification steps.
- [0558] Conditions for successful ion exchange chromatography will vary depending on the polypeptide. For more information, refer to Coligan, J.E., *et al.*, *Current Protocols in Protein Science*, Chanda, V.B., ed., John Wiley and Sons, Inc., New York (1998), Ausubel, F.M., *et al.*, *Current Protocols in*

Molecular Biology, Unit 10 (1994), or Deutscher, M.P., "Guide to Protein Purification," in *Methods in Enzymology*, Vol. 182, Simon, M.I., ed., Academic Press, San Diego, CA (1990).

- [0559] Many insect cell proteins are naturally rich in histidines, with some containing stretches of six histidines. When using the ProBond™ Purification System or other similar products to purify 6xHis-tagged polypeptides, these histidine-rich polypeptides may co-purify with a polypeptide of interest. The contamination can be significant if the polypeptide of interest is expressed at low levels. 5 mM imidazole may be added to the binding buffer prior to addition of the polypeptide mixture to the column. Addition of imidazole may help to reduce background contamination by preventing polypeptides with low specificity from binding to the metal-chelating resin.
- [0560] If the polypeptide of interest is 6xHis-tagged and expressed intracellularly, the cells may be lysed and the lysate added directly to the ProBond™ column. 5×10^6 to 1×10^7 cells may be used for purification of a polypeptide of interest on a 2 ml ProBond™ column (see ProBond™ Purification System manual, catalog nos. R801-01, R801-15, version F, Invitrogen Corporation, Carlsbad, CA).
- [0561] A suitable protocol is to seed 2×10^6 cells in two or three 25 cm² flasks, grow the cells in selective medium until they reach confluence (4×10^6 cells); wash cells once with PBS (Phosphate Buffered Saline, pH 7.4; Invitrogen Corporation, Carlsbad, CA Catalog no. 10010-023); harvest the cells by sloughing; transfer the cells to a sterile centrifuge tube; and centrifuge the cells at 1000 x g for 5 minutes. The cells may be lysed immediately or frozen in liquid nitrogen and store at -80°C until needed.
- [0562] Many protocols are suitable for purifying polypeptides from the medium. The choice of protocol depends on the nature of the polypeptide being purified. The culture volume needed to purify sufficient quantities of polypeptide is dependent on the expression level of the polypeptide and the method of detection. One skilled in the art can develop suitable purification protocols using routine experimentation.

EXAMPLE 6

Construction of recombinant baculoviruses.

- [0563] Baculoviruses have been extremely useful tools for heterologous expression of proteins in insect cells. Improved methods for cloning genes into baculoviral genomes (*e.g.*, the 134 kb AcMNPV genome) have greatly simplified the process of recombinant baculovirus construction; however obtaining a purified viral stock still requires plaque purification and a minimum of 10-14 days. Current methods rely on recombination in insect or bacterial cells and are not well adapted for high-throughput experiments. To meet these challenges, materials and methods of the invention permit the construction of recombinant baculovirus *in vitro*. The recombinant baculovirus may be transfected directly into insect cells to generate the baculovirus stock.
- [0564] A baculovirus genome containing a recombination cassette (DEST) bounded by attR recombination sites compatible with GATEWAY™ entry vectors (Invitrogen Corporation, Carlsbad, CA) was constructed. Two transposition cassettes were constructed one with and one without the mellitin leader sequence. A schematic representation of the cassette without the mellitin sequence is provided in Fig. 19A and the sequence is provided in Table 13. A schematic representation of the cassette with the mellitin sequence is provided in Fig. 19B and the sequence is provided in Table 14. The DEST cassettes contain the HSV thymidine kinase (TK) gene driven by an immediate early promoter (IE-0 promoter) and the lacZ gene driven by a late promoter (P10 promoter). The genes permit identification of non-recombinant virus using a blue white screening protocol and selection against non-recombinant viruses using ganciclovir. The cassettes also contain the V5 epitope and a 6-Histidine sequence outside the attR2 recombination site. The sequence of the cassette contains a recognition site for the restriction enzyme *Bsu36I* (and its isoschizomer *AocI*) that is used to linearize the viral genome.
- [0565] The cassette may be inserted into a baculoviral genome such that a sequence of interest in the Entry Clone may be operably linked to a baculoviral promoter (*e.g.*, the polyhedrin promoter (ph pr in Fig. 20)) upon insertion of the sequence of interest into the viral genome. In practice, any eukaryotic cellular or viral promoter can be used to express a gene introduced

from an entry clone, e.g. promoters from any of the above named baculovirus species, whether they are early, late, or very late. Although depicted as a gene sequence in Fig. 20, any sequence of interest may be inserted; the present invention is not limited to sequences encoding polypeptides.

[0566] In one embodiment, the nucleic acid sequence of interest may be recombined directly into the baculovirus genome downstream of the polyhedrin promoter, replacing the TK and lacZ genes. With reference to Figure 20, the linearized baculoviral genome is depicted as a gapped circle. In the presence of the appropriate recombination proteins, the recombination sites (e.g., *attR1* and *attR2* sites) on the baculoviral genome will recombine with the recombination sites (e.g., *attL1* and *attL2* sites) on the nucleic acid molecule comprising the sequence of interest (Entry Clone in Fig. 20) resulting in recircularization of the baculoviral genome. The recombination reaction results in the transfer of the sequence of interest (depicted as a gene of interest (GOI) in Fig. 20) into the baculoviral genome. The transfer also results in the excision of the portions of the baculoviral genome between the *attR* recombination sites.

[0567] The resultant DNA may be directly transfected into insect cells to produce the recombinant viral stock. When the cells are grown on ganciclovir, only recombinant virus is able to replicate; replication of parental virus is prevented because of the TK gene product. The destination cassette may also be placed under the control of the CMV promoter or other promoter active in mammalian cells, for the purpose of transducing mammalian cells using baculovirus.

[0568] To demonstrate the feasibility of this system and to optimize conditions, the GFP coding sequence was first cloned into a nucleic acid molecule between two recombination sites and then transferred using recombinational cloning into a baculovirus genome comprising two compatible recombination sites. Sf21 cells were transfected with the recombination reaction mixture. After three days, the media from these cells containing budded virus produced from the first rounds of replication was used to infect a second population of cells, this time grown under ganciclovir selection. After 4 days, these cells were examined for GFP fluorescence and stained for LacZ expression. Cells infected by recombinant virus expressing

GFP were fluorescent, while cells infected with remaining parental virus stained positive for LacZ expression. Using this assay method, conditions for transfection and ganciclovir counter selection were optimized. Under ideal conditions, small scale virus stocks essentially free of parental virus were produced within 7 days post-transfection. These stocks are suitable for creation of high titer stocks or further expression studies.

[0569] The utility of this system was then demonstrated for use in a 96 well format with collections of genes cloned into an Entry vector. Multiple genes in 96 well plates were cloned and screened for expression in parallel. Within seven days, purified viral stocks were available for scale-up or further expression studies.

[0570] In some embodiments, the present invention provides a new method for baculovirus cloning based on lambda recombination that is faster, requires less hands-on time, is more reliable, and is suitable for high throughput expression in 96 well plates.

[0571] In some embodiments, the present invention provides isolated nucleic acids comprising nucleic acid sequences that function as promoters. Optionally, the nucleic acid molecules may comprise one or more sequences of interest (*e.g.*, ORFs, etc.) operably linked to one or more of the nucleic acid sequences that function as promoters. These promoters may function in any cell type, for example, mammalian, insect, etc.

[0572] In some embodiments, the promoters are tightly regulated. For example, in some embodiments, the promoters are not active unless one or more transactivators are present. In some embodiments, the nucleic acid sequences that function as promoters include, but are not limited to, the AcMNPV ORF 25 promoter sequence, the AcMNPV lef 3 promoter sequence, the AcMNPV TLP promoter sequence, the AcMNPV homologous repeat 5 sequence, other baculovirus homologous repeat sequences, and the like. The nucleic acid sequences of the AcMNPV ORF 25 promoter sequence, the AcMNPV lef 3 promoter sequence, the AcMNPV TLP promoter sequence, and the AcMNPV homologous repeat 5 sequence are provided in Table 15.

[0573] In some embodiments, the promoters discussed above are not active unless one or more transactivators are present. One suitable transactivator is the baculoviral IE-1 protein. The IE-1 promoter sequence, coding sequence,

and polypeptide sequence are provided in Table 16. The transactivator may be provided on the same nucleic acid molecule comprising the promoter sequence or on another nucleic acid molecule (*e.g.*, plasmid, virus, host cell genome, etc.). In some embodiments, the promoter sequence operably linked to a sequence of interest may be on one nucleic acid molecule (*e.g.* a plasmid) and the transactivator sequence may be on a different nucleic acid molecule (*e.g.*, a virus such as a baculovirus). The nucleic acid molecule comprising the promoter sequence operably linked to a sequence of interest may be introduced into a host cell, for example, by transfection. The sequence of interest is not expressed or is substantially not expressed in the absence of a transactivator. In some embodiments, the host cell may be a eukaryotic cell, for example, a mammalian cell or an insect cell. The host cell comprising the nucleic acid molecule comprising the promoter sequence operably linked to a sequence of interest may be further contacted with a second nucleic acid molecule comprising the a sequence encoding the transactivator. Upon expression of the transactivator, the sequence of interest is expressed. In some embodiments, the transactivator polypeptide may be directly transfected into cells comprising the nucleic acid molecule comprising the promoter sequence operably linked to a sequence of interest. Such transactivator polypeptides may be present as native polypeptides or as fusion polypeptides, for example, as fusions with the herpesvirus VP22 polypeptide.

[0574] Nucleic acid molecules comprising the promoters discussed above may be used to conditionally express any sequence of interest. In some embodiments, the sequence of interest may encode a toxic polypeptide.

[0575] In some embodiments, nucleic acid molecules comprising the promoter sequences described above may have a homologous repeat (hr) sequence in cis with the promoter. Such homologous repeat sequences may be required for hr-dependent IE-1 transactivation.

[0576] The sequences provided in Table 15 are capable of functioning as conditionally activated promoters. The present invention also comprises portions of the sequences of Table 15 that function as conditionally active promoters. Such promoters may be activated by the IE-1 polypeptide. Such portions may comprise at least 50%, 60%, 70%, 80%, 90%, 95%, or more of one or more of the sequences in Table 15.

EXAMPLE 7

[0577] In some embodiments, materials and methods of the invention may be used to create stable cell lines expressing a nucleic acid sequence of interest. One non-limiting example is the InsectSelect™ system (Invitrogen Corporation, Carlsbad, CA), which is a stable insect cell expression system that utilizes a single plasmid for expression and selection. Nucleic acid molecules of the invention (*e.g.*, InsectSelect™ vectors) may utilize different baculovirus immediate early promoters for expression of a sequence of interest and a selectable marker. Nucleic acid molecules of the invention may be constructed to be used in recombinational cloning methods. For example, pIB/V5-His (catalog no. V802001, Invitrogen Corporation, Carlsbad, CA) has been modified for using in methods involving recombinational cloning (*e.g.*, GATEWAY™ cloning). In the modified vector, a different promoter is used to drive transcription of the blasticidin resistance gene than the OpIE-1 promoter used in pIB/V5-HIS.

[0578] The OpIE-1 promoter was replaced with long or short versions of AcMNPVgp64 or pe38 promoters, using a Topoisomerase I mediated ligation strategy (Fig. 21). The AcMNPV gp64 and pe38 promoters were amplified from cosmid #58 (comprising AcMNPV bases 99803-132856 from a cosmid library of the AcMNPV genome, Harwood *et al.* Virology. 250:113-134, 1998) with promoter-specific primers that were appended at their 5' ends with antisense TOPO sites and six additional bases (Fig. 21). pIB/V5-His was amplified with primers that included an anti-sense topoisomerase site and a six base sequence that becomes an overhang following topoisomerase binding. Each promoter (gp64s is illustrated) was amplified with similarly designed primers. Following binding, the overhangs annealed and were ligated by the enzyme. The oligonucleotide sequences are given below. The antisense topoisomerase sites are underlined.

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17852 pIB Neg For  TGAGTCAAGGGCTGCCGGGCTGCAGCACTG
17853 pIB Neg Rev  CGGAACAAGGGCATGACCAAAATCCCTTAACG
17849 gp64 For    GACTCAAAGGGCTTGCTTGTGTGTTTCCTTATTG
17850 gp64s Rev   GTTCCGAAGGGTTGTGTCACGTAGGCCAGATAAC

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17851 gp64L Rev

GTTCCGAAGGGAATAATCGATTTAAGGGTGTAACTC

17857 pe38 For GACTCAAAGGGTTTGCTTATTGGCAGGCTCTCC

17858 pe38s Rev GTTCCGAAGGGTATCTGTCCCCCACTCAGGC

17859 pe38L Rev GTTCCGAAGGGTAAAGTTGATGCGGCGACGGC

[0579] The pIB/V5 His backbone was amplified using similarly designed primers. The PCR products were purified by gel electrophoresis and SNAP mini-prep columns. Following DpnI treatment to eliminate residual template vector, the PCR products were repurified by SNAP minipreps, eluted in 30 μ l water and joined using topoisomerase (Fig. 21). Topoisomerase reactions were incubated at room temperature for 10 min and contained 8 μ l of each PCR product, 50 mM Tris, pH 7.5, 0.1 μ g/ μ l enzyme in 20 μ l total volume. TOP10 *E. coli* were transformed with the joined PCR products. Following selection on ampicillin plates, resulting colonies were grown overnight, and plasmid DNA isolated by miniprep (SNAP). The presence of the promoters was confirmed by restriction digest analysis. The construct containing gp64s was ultimately chosen for GATEWAY™ adaptation (see below).

[0580] pIB/V5-His gp64 was modified to comprise recombination sites (*i.e.*, GATEWAY™ adapted) by cloning a HindIII/XbaI fragment from pDEST38 into pIB/V5-His gp64, cut with the same enzymes. The vector was fully sequenced. A plasmid map is provided (Fig. 22).

[0581] To test the modified vector in a recombinational cloning reaction, pIB/V5-His gp64Dest was used for LR reactions with attL entry vectors containing LacZ, Calmodulin, TFIIIS, and Apolipoprotein. The protocol used differed slightly from the protocol suggested in the GATEWAY™ manuals. The reaction conditions used were as follows:

2 μ L LR clonase enzyme mix (catalog no. 11791043, Invitrogen Corporation, Carlsbad, CA)
 2 μ L LR reaction buffer
 1 μ L pENTR clone (~300 ng DNA)
 1 μ L pDEST vector (~300 ng DNA)
 4 μ L 0.5 M Tris buffer (pH 7.5)

[0582] Recombination reactions were incubated for 3h at room temperature. Reactions were not proteinase K treated. 2 μ l of each recombination reaction

was used to transform 50 μ l TOP10 chemically competent bacteria. Half of the transformation mix was plated and yielded an average of 230 colonies. Thus, approximately 8000 colonies were obtained per μ g entry vector. Colonies were grown in LB/Amp overnight and DNA was isolated by SNAP miniprep.

[0583] Experiments were performed with *Sf*21 cells or HighFive cells in serum-containing or serum free media (SFM). Grace's supplemented media with 10% FBS was used for both *Sf*21 and HighFive cells. For SFM treatments, *Sf*900II or ExpressFive media were used for *Sf*21 or HighFive cells, respectively. Twenty-four well plates were seeded with 1.8×10^5 cells per well, and after 1 h attachment, washed with Grace's unsupplemented media. Transfection mixes contained 0.2 μ g DNA and 1 μ l Cellfectin® in 40 μ l Grace's unsupplemented media and incubated for 30 min at RT. The transfection mixture was then diluted to 200 μ l final volume in Grace's unsupplemented media and added to each well. Cells and transfection mix were incubated for 5 h with gentle rocking after which the mix was replaced with the appropriate media as described above. 48 h later the media was replaced with the same media containing between 10 and 25 μ g/ μ l blasticidin, depending on the experiment. Cells used from stable cultures were under selection for at least 7 days. Cells were split as needed to maintain log-phase growth. Typically, 10 μ g/ml blasticidin may be used for general purposes. However, one skilled in the art can optimize selection parameters for each construct using only routine experimentation.

[0584] Protein expression was monitored by western blot or LacZ activity assays. Cells from six well plates (approximately 10^6 per well) were washed 2x in PBS, transferred to 1.7 ml tubes, spun down, resuspended in 500 μ l lysis buffer (Tropix Galacto light kit, catalog no. T1006, Applied Biosystems, Foster City, CA), and then subjected to two freeze-thaw cycles. Lysates were microfuged at 16,000 x g for 5 min. Supernatants were stored at -20°C until used. Lysate protein concentration was measured using the BioRad protein assay against BSA as a standard. Various amounts of protein were denatured in LDS sample buffer (catalog no. NP0008, Invitrogen Corporation, Carlsbad, CA) and loaded on 4-12% NuPAGE gels (Invitrogen Corporation, Carlsbad, CA). Following electrophoresis, proteins were transferred to PVDF. The

Western Breeze kit (catalog no. WB7104, Invitrogen Corporation, Carlsbad, CA) was used to visualize protein bands using anti-V5 coupled alkaline phosphatase at a 1:5000 dilution unless noted otherwise.

[0585] Without being bound by theory, it was thought that use of a weaker promoter to drive antibiotic resistance would result in stable cultures that expressed the gene of interest at higher levels because the *bsd* gene (blasticidin resistance gene) was expressed at a lower level, integration of the plasmid containing the *bsd* gene would occur in more loci or in loci that were transcriptionally more active. Transcription of many baculovirus genes has been characterized, and suitable promoters were selected. The gp64 and pe38 promoters have both been extensively studied (Friesen, Regulation of baculovirus early gene expression, p. 141-170. *In* The Baculoviruses. L. K. Miller (ed.), Plenum Press, New York., 1997). The pe38 promoter is an immediate early promoter and thus does not require baculovirus infection for its activity. The gp64 promoter is transactivated by IE-1 but retains basal levels of activity without transactivation (Blissard, J. Virol. 65:5820-5827, 1991, Blissard, Virology. 190:783-793, 1992). The sequences responsible for IE-1 transactivation have been identified and are separable from the basal promoter (Blissard, 1992). A long (500 bp upstream of the ATG) and a short version (100 bp upstream of the ATG) for each promoter were obtained and cloned in place of the OpIE-1 promoter using TOPO-mediated ligation. LacZ was cloned into the resulting vectors. These constructs together with the OpIE1 promoter version of pIB LacZ/V5-His were transfected into Sf21 cells and polyclonal cultures were selected at two different dosages of blasticidin. The longer gp64 construct apparently did not provide sufficient levels of *bsd* expression and the cells died with the control cells. Surviving stable cultures were obtained from the other four constructs. Cells were harvested after two weeks of selection and expression levels were measured using β -galactosidase assays (Fig. 23). β -galactosidase activities for stable cell cultures established with different versions of pIB/V5-His. 20 μ g of protein was used per assay. Higher levels of expression were obtained for all three alternate promoters than obtained with the OpIE-1 promoter at both 20 and 100 μ g/ml blasticidin. There were not clear differences in LacZ activity between cultures selected at either concentration of blasticidin.

[0586] The gp64s promoter construct was used for GATEWAY™ adaptation. To examine the cloning efficiency and gene expression for the gp64s GATEWAY™ adapted version of this vector, four genes (Apolipoprotein, Calmodulin, TFIIs, and LacZ) were transferred into GATEWAY™ adapted versions of pIB/V5-His and pIB/V5-His gp64 the vector using an LR reaction. All LR reactions resulted in thousands of colonies per µg plasmid and were correct when examined by agarose gel electrophoresis. Each construct was transfected into *Sf21* cells. Transient and stable expression of Apolipoprotein was compared between the gp64 and OpIE-1 versions of pIB Apolipoprotein/V5-His GATEWAY™. Transient expression levels were equivalent between the gp64 and OpIE1 versions (Fig. 24, lanes 1 and 2), but expression was higher for the gp64 version following selection (Fig. 24, lanes 3 and 4). To be sure that the higher stable expression level observed for the gp64 promoter was a general phenomenon, expression of Calmodulin, TFIIS, and LacZ between gp64 and OpIE-1 versions of pIB/V5-His GATEWAY™ were compared (Fig. 25). Fig. 25A shows expression of calmodulin and TFIIs from *Sf21* cells stably transfected with OpIE-1 (lanes 1 and 3) and gp64s versions of pIB/V5-His. 8.6 µg total protein was loaded per lane. Fig. 25B shows expression of LacZ from *Sf21* cells stably transfected with OpIE-1 (lane 1) or gp64s (lane 2) versions of pIB/V5-His. Lane 3 is a non-transfected control. 5.7 µg of protein was loaded per lane. As for Apolipoprotein, expression of Calmodulin, TFIIS (Fig. 25A) and LacZ (Fig. 25B) was higher from the gp64 version.

[0587] The above experiments were conducted with *Sf21* cells in serum containing media. Use of a different promoter for expression of the antibiotic resistance marker could alter the dynamics of selection as a function of cell type or media used. Selection and expression from HighFive cells in serum- and serum free media was analyzed. In general, non-transfected cells were dead within a week but cells selected in SFM tended to die sooner (3-4 days) than those selected in media containing serum. As with the previous experiments, higher levels of gene expression were obtained from the gp64 construct with stably transfected HighFive cells, whether they were grown in serum or serum free media. Similar results were obtained with *Sf21* cells in SFM media. Figure 26 shows High five cells grown in serum and serum free

media transfected with Gp64 and OpIE-1 versions of pIB/V5-His. 24.5 µg total protein per assay.

[0588] A recombinational cloning adapted version of pIB/V5-His that utilizes a different baculovirus promoter for expression of the *bsd* gene has been prepared. The basal gp64 promoter presumably results in lower levels of the *bsd* gene product than the OpIE-1 promoter used in pIB/V5-His and forces integration of the plasmid into more active chromosomal loci and/ or at higher copy number.

EXAMPLE 8

[0589] In some embodiments, the present invention provides a method of making recombinant viruses using recombinational cloning. One non-limiting example is termed BaculoDirect™. Methods of this type provide a novel baculovirus cloning method that takes advantage of recombinational cloning technology (*e.g.*, GATEWAY™ cloning technology, Invitrogen Corporation, Carlsbad, CA). With BaculoDirect™, an entry clone containing a nucleic acid sequence of interest (*e.g.*, a sequence comprising a gene of interest) may be recombined into recombination-site-containing baculovirus genome in a one hour, *in vitro* reaction. The DNA product from this reaction can be transfected directly into suitable cells (*e.g.*, Sf9 or Sf21 cells) to generate recombinant viruses and screen for expression. The ability to clone the sequence of interest (*e.g.*, gene of interest (GOI)) directly into the baculovirus genome *in vitro* contrasts with existing baculovirus cloning methods in which the recombination step is performed in insect cells or bacteria. Compared with these existing baculovirus technologies, BaculoDirect™ is significantly faster, requires less hands-on time, and is more reliable. It is also easily adapted for high-throughput experiments. Thus, BaculoDirect™ offers significant advantages over current baculovirus cloning systems.

[0590] Throughout this disclosure, the term gene of interest (GOI) may be used for the sake of convenience. This should not be construed as limiting the present invention to nucleic acid sequences comprising genes. Any nucleic acid sequence of interest can be inserted into a vector of the invention using materials and methods described herein.

INTRODUCTION

- [0591] Baculoviruses are one of the most commonly used tools for eukaryotic expression of heterologous proteins. Traditionally, a GOI had to be first cloned into a transfer vector and then moved into the virus by homologous recombination into the polyhedrin locus in permissive insect cells. This occurred at low frequency. Plaque assays were tedious and required identification of polyhedrin negative plaques from among much more numerous polyhedrin-positive plaques.
- [0592] During the last 20 years, innovations have made baculovirus cloning more convenient. Use of linearized DNA and design of the recombination strategy such that recombination restored function of an essential baculovirus gene boosted the proportion of recombinant plaques obtained from 1-2% to over 90% (Kitts and Possee, 1993. *BioTechniques* 14:810-817). However, multiple rounds of plaque purification were still required and the entire process of obtaining a useful viral stock took 3-4 weeks and a substantial amount of labor. Expression kits that use this technology are marketed by BD Biosciences Pharmingen, San Diego, CA (Baculogold™), Novagen Inc., Madison, WI and Invitrogen Corporation, Carlsbad, CA (Bac-n-Blue™).
- [0593] A second method for baculovirus cloning utilizes site-specific recombination in bacteria to introduce the GOI into the baculovirus DNA (Luckow, *et al.*, 1993. *J. Virol.* 67:4566-4579). The GOI is cloned into a transfer plasmid and used to transform a specialized bacterial strain that contains the baculovirus genome propagated as an F' plasmid (bacmid). The GOI is then introduced into the bacmid by site-specific recombination between Tn7 sites on the transfer plasmid and in the baculovirus genome. Bacteria containing recombinant bacmids are then selected using antibiotic selection markers with appropriate selective media. The bacmid DNA is extracted and then transfected into insect cells. Plaque purification is, in theory, not required (except for the most rigorous applications) and the entire process from transfer plasmid to pure virus stock requires 10-12 days. Invitrogen Corporation, Carlsbad, CA markets this system under the trade name Bac to Bac™, catalog number 10359-016.
- [0594] While these advancements in baculovirus cloning have greatly simplified use of baculovirus for routine protein expression, the methods

described above still require significant “hands-on” time and are not well suited for parallel processing of multiple genes (*i.e.*, high-throughput). The present invention provides a new method that greatly simplifies and shortens the process for cloning and purification of baculovirus recombinants. One non-limiting example of the present invention is BaculoDirect™, which utilizes GATEWAY™ recombinational cloning technology (Invitrogen Corporation, Carlsbad, CA) to recombine a GOI into the baculovirus genome *in vitro* in a one hour, room temperature reaction. The resulting recombinant virus DNA is transfected directly into insect cells. In just six days, cells can be harvested for expression screening to obtain a pure viral supernatant suitable for creation of high titer stocks.

MATERIALS AND METHODS

[0595] All materials used in this study were from Invitrogen Corporation, Carlsbad, CA except restriction enzymes (Roche Applied Sciences, Indianapolis, IN or NEB, Beverly, MA) and ganciclovir sodium salt (GCV, Invivogen, San Diego, CA Catalog #sud-gcv).

Cells and Virus

[0596] Sf21 cells were cultured in Grace’s medium with supplements and 10% FBS unless stated otherwise. Infection of cells with wild type AcMNPV or other viruses was performed as described (O’Reilly *et al.*, 1992. *Baculovirus Expression Vectors: a Laboratory Manual*. W.H. Freeman Co., New York).

Plasmid and Virus Construction

[0597] Three versions of BaculoDirect™ were constructed. The first contained the melittin secretion signal, the second contained both a melittin signal and a C-term V5/His tag, and the third had a C-term V5/His tag without a secretion signal. Figs. 19A and 19B provide schematics of recombination cassettes having a C-terminal V5/His tag with (19B) and without (19A) a melittin leader.

[0598] The plasmid pVL1393 GST p10 stop (Fig. 34) was digested with *Bam*HI and *Nco*I. A 15 kb band was purified (removing the GST tag) to which was ligated, a double stranded oligonucleotide containing the melittin signal flanked by *Bam*HI and *Nco*I overhangs. The ligated products were transformed into TOP10 bacteria and the correct clones verified by restriction digestion and sequencing. This plasmid (pVL1393 Mel Stop) contained a stop codon downstream of the *att*R2 site that had to be removed by PCR directed site-specific mutagenesis. Primers *Eco*RI sense (GAATTCCAGCTGAGCGCCGGTCGCTAC) and *Bg*III antisense (AGATCTTCAATTCTCACCACCTTTGTACAAG) were used to amplify a fragment from pVL1393 Mel Stop, and the resulting 209 bp fragment was cut with *Eco*RI and *Bg*III, and then ligated to pVL1393 Mel Stop cut with the same enzymes. The correct clone was identified by restriction digestion and sequence analysis. This gave pVL1393 Mel no-Stop.

[0599] Next, a V5-His tag was added downstream of the *att*R2 site. The V5/His sequence was amplified from pIND/V5-His-TOPO (catalog no. K101001, Invitrogen Corporation, Carlsbad, CA) with primers containing *Bg*III sites at each 5' end (V5/His 5': AGATCTGGGGAAGCCTATCCCTAACCC; V5/His 3': AGATCTTCAATGGTGATGGTGATGATGACCGG). The amplicon was cloned into pCR2.1 TOPO TA and then removed by *Bg*III digestion and ligated to pVL1393 Mel no-Stop cut with *Bg*III. The correct clones were identified and verified by sequencing. This resulted in plasmid pVL1393 Mel/V5-His. The melittin signal was subsequently removed by replacing the melittin-*att*R1 sequence from pVL1393 Mel/V5-His with the *att*R1 sequence from pVL1393-Native, using *Not*I and *Bam*HI. The correct plasmid clones were verified by sequencing and dubbed pVL1393 V5/His. Fig. 27 shows a schematic of the strategy for construction of BaculoDirect™ DNA. In Fig. 27A, the GATEWAY™ counter selection cassette was cloned in the polyhedrin locus of wt AcMPPNV by homologous recombination between with pVL1393 V5-His. The resulting virus DNA contains the counter selection cassette bounded by *att*R sites, immediately downstream of the polyhedrin promoter and upstream of the V5/His tag. In Fig. 27B, LR recombination between

BaculoDirect™ DNA and an entry clone results in an expression virus in which the counter selection cassette is replaced by gene of interest.

Generation of BaculoDirect™ viruses

[0600] BaculoDirect™ viruses were created via conventional homologous recombination between wt AcMNPV and homologous recombination sequences contained in pVL1393 (Fig. 27, O'Reilly, *et al.*, 1992). Briefly, *Sf*21 cells were co-transfected with 0.5 µg wild type AcMNPV E2 virus DNA and 3-5 µg of pVL1393 V5/His. After five days, the supernatant was collected. This supernatant contained a mixture of recombinant BaculoDirect™ virus and wt virus. The recombinant virus was isolated and purified through three to four rounds of plaque purification (O'Reilly, *et al.*, 1992). Recombinant plaques could be distinguished from wt by phenotype, *i.e.*, recombinant plaques were β-Gal⁺, polyhedra(-) whereas wt plaques were β-Gal(-), polyhedra(+).

GENERATION OF RECOMBINANT EXPRESSION VIRUS

[0601] Expression viruses were generated by performing standard LR clonase reactions between BaculoDirect™ DNA and entry clones containing a GOI flanked by *att*L1 and *att*L2 (Fig. 27B, GATEWAY™ Instruction Manual Version C, 6/02, Invitrogen Corporation, Carlsbad, CA). Where indicated, BaculoDirect™ DNA was linearized using *Aoc*I (an isoschizomer of *Bsu*36I), which cuts in the 5' end of the *lacZ* gene. Reactions were performed with or without linearization. Twenty microliter LR reactions contained 300 ng viral DNA, 100 ng entry clone, 4 µl LR clonase buffer, 4 µl LR clonase, and were incubated for 1 h at room temperature. Two million *Sf*21 cells were transfected with varying amounts of completed LR reaction using 6 µl of Cellfectin® (catalog no. 10362-010, Invitrogen Corporation, Carlsbad, CA) and *Sf*900II media per the manufacturer's instructions. Five hours post-transfection, transfection buffer was replaced with the Grace's Supplemented Insect Medium containing 10% FBS and 100 µM ganciclovir. Three to five days later, the supernatant was collected and varying amounts were used to infect fresh *Sf*21 cells with or without ganciclovir selection.

High throughput (HTP) screening of expression

[0602] A method for performing LR reactions and transfection in 96 well plate format was developed. Fig. 28 provides a schematic illustration of BaculoDirect™ cloning and expression in 96 well plates. Entry vector DNAs, diluted Cellfectin®, and Sf21 cells were arrayed in 96 well plates. By arraying the components separately, the number of pipetting manipulations of the Baculovirus DNA is minimized. Following expression screening from the first generation transfection, only those wells showing expression of a protein of interest need be processed further.

[0603] Three 96 well plates were needed in this experiment. In plate A, 10 µl LR reactions were assembled in individual wells, starting with five different entry plasmids arrayed in multiple wells. The entry clones used were: pENTR APO/V5-His (Apolipoprotein), pENTR CAL/V5-His (Calmodulin), pENTR GUS, pENTR *LacZ* and pENTR CAT. Each 10 µl reaction included 50 ng entry clone, 150 ng purified linear BaculoDirect™ DNA, 2 µl LR clonase buffer, and 2 µl LR clonase. The LR reactions were incubated in the plates for 1 h at RT. During the LR incubation, Sf21 cells were seeded at 4.8×10^4 cells per well in a separate plate and allowed to attach in plate B. In plate C, 2 µl of Cellfectin® were diluted to 40 µl per well with Grace's medium. After the 1 h LR reaction, 40 µl of Grace's unsupplemented media were added to each well of plate A. Forty microliters of the Cellfectin® mixture from plate C were added to the diluted LR reactions and incubated at 27 °C for 30-45 min. After this incubation, 150 µl of Grace's un-supplemented media was added to the wells of plate A. The cells in plate B were washed twice in Grace's media and then replaced with various amounts of the transfection mixture from plate A. Plate B was incubated for 5 h at 27° C, and then the transfection mixture was removed and replaced with Grace's complete media with 100 µM ganciclovir. The cells were allowed to grow for 3-4 days. Supernatants from each well were transferred to a separate plate. The cells remaining in plate A were lysed *in situ* with 100 µl LDS lysis buffer and heated to 80 °C for 5 min. Because apolipoprotein was secreted, 15 µl of supernatant was denatured in 4x sample buffer. Protein samples were separated on SDS-PAGE gels, transferred to PVDF and visualized by western blot.

ESTIMATION OF VIRAL TITERS

[0604] Virus titers were estimated using two methods. Virus plaque assays were performed using techniques well known in the art (*e.g.*, Bac to Bac Baculovirus Expression System Manual, catalog no. 10359-016, version C, p. 27, Invitrogen Corporation, Carlsbad, CA). P1 or virus supernatants (infection from the P1 stock) using apolipoprotein-expressing versions of each virus were serially diluted ten fold from 10^{-1} to 10^{-8} and used to infect 2 million cells in six well plates. Recombinant plaques were counted and titers estimated based on the dilution factor for each plate.

[0605] TCID₅₀ (Tissue Culture Infective Dose) measurements were conducted as described (O'Reilly, *et al.*, 1992). Briefly, a 96 well plate was seeded with 4.8×10^4 Sf21 cells per well. P1 stocks or virus supernatants were as described above. 10 μ l of each dilution was added per well, twelve wells per dilution, using a multi-channel pipettor. The TCID₅₀ was calculated using the Excel (Microsoft) spreadsheet described in O'Reilly, *et al.*, 1992.

RESULTS

OPTIMIZATION OF LR CLONASE REACTIONS USING BACULODIRECT™ DNA

[0606] BaculoDirect™ DNA is the functional equivalent of a GATEWAY™ destination vector. GATEWAY™ destination vectors designed for use in bacteria, *e.g.*, *E. coli*, contain a counter-selection cassette containing the *ccdB* gene and a chloramphenicol resistance marker, bounded by *attR* sites. Recombination between an *attL* containing entry clone and the destination plasmid replaces the *ccdB* gene and *Chl(r)* marker with the gene of interest, yielding an expression clone bounded by *attB* sites. This selection scheme does not work in insect cells. To create a counter-selection cassette for use with baculovirus, wild type baculovirus DNA was engineered with a cassette containing the herpes virus TK gene (HSV *tk*) and the *lacZ* gene, both under control of baculovirus promoters, bounded by *attR* sites (Fig. 27A). The *attR* cassette was placed immediately downstream of the polyhedrin promoter. Recombination between the "destination virus" and an entry clone replaces the counter selection cassette with the GOI under polyhedrin promoter control

(Fig. 27B). Transfection of the resulting DNA creates a mixed baculovirus infection with both recombinant virus and parent virus present. Replication of the parent virus is prevented by growing the cells in the presence of ganciclovir, which is metabolized by the HSV *tk* gene into a toxic inhibitor of DNA replication (Godeau, *et al.*, 1992, *Nucl. Acids Res.* 20:6239-6246). Cells that are infected by parent virus will also express the *lacZ* gene, which can be assayed by staining infected cells, providing a method for checking the purity the virus infection.

[0607] To test if the LR reaction would work between a 3-4 kb entry clone and the 140 kb BaculoDirect™ virus DNA, an LR reaction between melittin BaculoDirect™ and a GFP entry clone was performed. GFP expression was clearly visible by fluorescence as early as 48 h post-transfection and was stronger at 72 h, demonstrating that the LR reactions were successful and that GFP was placed under control of the polyhedrin promoter. Transfection, infection and selection conditions were then optimized to minimize background resulting from residual parental virus, as evidenced by GFP fluorescence and β -galactosidase staining.

[0608] Linear and circular BaculoDirect™ DNA were compared. Thus, a standard LR reaction was performed with either linearized or circular (uncut) melittin BaculoDirect™ DNA, without ganciclovir selection. Ten, twenty or thirty microliters of LR reaction were used to transfect *Sf21* cells (only the results from the 20 μ l transfection are shown in Fig. 29). Three days later, varying amounts of supernatant (P1 stock from the “first generation”) from each transfection were used to infect new *Sf21* cells. After four days, infected cells were examined for GFP fluorescence and then stained for β -galactosidase activity. These cells are from the “second generation” and the supernatant from them is a small scale high titer stock (see titer data below). Virtually all cells in all treatments were fluorescent, demonstrating that a productive baculovirus infection had been established and that the virus was actively expressing GFP. Fig. 29 shows the results of an analysis of cells transfected with LR reaction products from the melittin version of BaculoDirect™ DNA. LR reactions between melittin BaculoDirect™ DNA were performed with *AocI* cut or circular virus DNA and a GFP entry clone. *Sf21* cells were transfected with 10 μ l, 20 μ l or 30 μ l each LR reaction, using either linear

virus DNA or circular virus DNA as indicated, without GCV selection. Cells were examined by fluorescence and β -Gal staining 72 hours following transfection. The result here shown from the 20 μ l of LR reaction was typical. Some β -Gal positive cells were found in every well examined. β -galactosidase activity (*i.e.*, background) was much higher in cells that had been infected with P1 virus derived from cells transfected with LR reactions that used circular rather than linearized BaculoDirect™ DNA (Fig. 29, upper panel). Background was much lower if cells were infected with P1 stocks derived from LR reactions with linearized BaculoDirect™ DNA, although some background was detected when more P1 stock was used for infection (Fig. 29, lower panel).

[0609] The effect of ganciclovir selection was then tested by growing cells in the presence or absence of ganciclovir. LR reactions were performed as above with circular or linearized melittin BaculoDirect™ DNA. *Sf21* cells were transfected with varying amounts of LR reaction and then grown without GCV (first generation). After 72 h, varying amounts P1 stock obtained from each transfection were used to infect new *Sf21* cells, now grown in the presence of 100 μ M GCV. After 4 days (second generation), the cells were examined for GFP fluorescence and stained for β -galactosidase. GCV did not appreciably reduce the number of cells staining positive for β -galactosidase activity when infections were derived from LR reactions using circular virus, whereas GCV reduced the number of β -gal positive cells from infections derived from LR reactions that used linearized virus.

[0610] The effectiveness of ganciclovir in eliminating background when used during the first generation, the second generation or both generations was tested. When ganciclovir was used in the first generation or the second generation, at least some blue cells were observed following the second generation. In general, more background was found when more LR reaction was transfected. However, when ganciclovir was used in both generations, no blue cells were found, suggesting that there were no cells infected by parent virus following two rounds of ganciclovir selection. Moreover, zero background was found irrespective of how much LR reaction was used during for the transfection. Representative results from these experiments are shown in Fig. 30. In the experiment shown in Fig. 30, cells were transfected and

selected during both generations as described above. Following the second generation, the cells were photographed to illustrate typical results following the selection protocol. Essentially all cells were producing GFP, but no cells stained positive for β -Gal if GCV selection was maintained during both generations. Thus, parent virus is not replicating in these cells. These results were obtained with cells grown in serum. The same result was found when serum free-adapted Sf21 cells were grown and selected with GCV in serum free media. Similar results were subsequently found using linearized V5/His virus.

HIGH-THROUGHPUT SCREENING OF EXPRESSION

[0611] The ability to clone and express genes from baculovirus without plaque purification or selection in bacteria suggests that BaculoDirect™ can be used conveniently for high-throughput screening of expression. Five pENTR clones were chosen (CAT, GUS, *LacZ*, Apolipoprotein/V5-His, and Calmodulin/V5-His) for expression. Each pENTR DNA was arrayed in multiple wells of a 96 well plate as illustrated in Fig. 28. LR clonase reaction mixes were added as described in the Materials and Methods, using linearized V5/His BaculoDirect™ DNA. All manipulations used multi-channel or repeating pipettors and thus could also be performed robotically. Following ganciclovir selection during the first generation, expression from each virus was assayed by western blot. All five genes expressed at levels sufficient to be easily detected (Fig. 31). The supernatants were stored in a separate 96 well plate and were available for second round infection and selection.

[0612] Fig. 31 shows the results of the screening of protein expression from LR reactions performed in a 96 well plate. Indicated pENTR DNAs were arrayed in a 96 well plate, and LR reactions were performed as described above. Supernatants were removed to a separate plate and then cells were lysed using 100 μ l LDS sample buffer. 15 μ l of lysate was applied per lane except for apolipoprotein, which was secreted. For apolipoprotein, 11 μ l of supernatant was used instead of cell lysate. The blot was visualized with anti V5:AP conjugate at 1:5000 and exposed to film for 15 sec.

Titer comparison

[0613] Two methods were used to compare the virus titers obtained from BaculoDirect™ with Bac to Bac™ or Bac-n-blue™. Apolipoprotein was cloned into pBlueBac 4.5/V5-His and co-transfected this with linear Bac-n-Blue DNA into Sf21 cells using well known techniques (e.g., Bac-n-blue™ manual, catalog no. K855-01, version M, Invitrogen Corporation, Carlsbad, CA). Following one round of plaque purification, a high titer stock was made. The entire apolipoprotein/V5-His reading frame was cloned into pFastBac, and bacmid DNA was generated using standard techniques (e.g., Bac to Bac™ manual, catalog nos. 11827-011, 11806-015, 11804-010 and 11807-03, version C, Invitrogen Corporation, Carlsbad, CA). The bacmid DNA was transfected into Sf21 cells, and a high titer stock was made. The apolipoprotein/V5-His reading frame was also cloned into pENTR and transferred in an LR reaction into linearized V5-His BaculoDirect™ DNA. Titers were measured following transfection and infection using plaque assay and TCID₅₀ methods. The titers obtained following infection were similar for all three baculovirus expression systems using either method and were in the range of 3×10^8 to 7×10^8 pfu/ml (Fig. 32). 10^8 pfu/ml is a typical titer for baculovirus and thus BaculoDirect™ baculoviruses replicate as well as the baculoviruses used in other systems.

[0614] Fig. 32 shows an estimation of virus titers using plaque purification and TCID₅₀ measurements. Apolipoprotein was cloned into pENTR, pFASTBAC, or pBlueBac 4.5/V5-His (catalog no. V207520, Invitrogen Corporation, Carlsbad, CA). Procedures for MaxBac and Bac to Bac were followed as described in their respective instruction manuals. Dilutions of P1 or virus stock from second generation supernatants were serially diluted and used to infect cells for agar overlay (plaque purification) or in 96 well plates (TCID₅₀). For BaculoDirect™, cells were selected on 100 μ M ganciclovir for both generations. Titers were calculated as described (O'Reilly, *et al.*, 1992).

DISCUSSION

[0615] BaculoDirect™ is functionally a GATEWAY™ adaptation of the baculovirus genome. Lambda-based recombination occurs between the attR

sites engineered in the baculovirus genome and *attL* sites surrounding the GOI in an entry clone. Following the LR clonase reaction, the counter-selection cassette containing the HSV tk gene and *lacZ* driven by baculovirus promoters and bounded on each side by *attR* sites on the baculovirus is replaced by the GOI from the entry clone. This results in re-circularization of the virus DNA. Replication of parent virus is prevented, both because it remains linearized, and because the tk gene product prevents DNA replication in the presence of ganciclovir. Linearization was highly effective at preventing replication of parental virus (Fig. 29). Virtually all cells expressed β -Gal following transfection and ganciclovir selection if the LR reaction was performed with circular virus, whereas use of linear virus boosted to greater than 95% (Fig. 29).

[0616] The presence of *lacZ* in the counter-selection cassette provides a means of judging the purity of virus stocks, since the absence of β -Gal cell staining is a good indication that a virus stock is free of contaminating parent virus.

[0617] Depending on the context, the *attB2* site can sometimes pose a problem for expression and or detection from the C-terminal V5 epitope tag. In Fig. 31, APO and CAL were cloned without an internal *attB2* site, while the remaining three genes were cloned with an internal *attB2* site between the gene and the V5-His tag. The entry clones used for APO and CAL had an encoded C-terminal V5-His. All of the genes except for CAL appeared to be expressed and detected at high levels (Fig. 31). It has been observed that CAL tends express at lower levels in most experiments. BaculoDirect™ viruses that express GUS with or without *attB2* inside the reading frame have been constructed. GUS expression was detected equally well for both versions, suggesting that the *attB2* site does not appear to interfere with expression or detection from the V5 tag in the context in which it is used in BaculoDirect™.

[0618] The addition of the GATEWAY™ cassette, presence of *attB* sites in the polyhedrin locus, or ganciclovir selection, did not appear to affect virus titers when compared to other baculovirus expression systems (Fig. 32). Titers in excess of 10^8 pfu/ml were obtained routinely following the second-generation virus infection. Since the virus stocks obtained following selection on ganciclovir were found to be essentially pure (based on the lack of infected cells that were β -Gal positive), the virus supernatants obtained at this stage

require no plaque purification and can be scaled up for production of high titer stocks. The entire process, from LR cloning to pure virus stock can be performed for multiple genes simultaneously in 96 well plates. Although the present methodology has been exemplified using just five genes, one of skill in the art will appreciate that any number of genes (*e.g.*, 20, 50, 100, 250, 500, 1,000, 2,500, 5,000, etc.) can be processed in a similar manner. The present method allows screening for expression after just three days, and continued selection and scale-up can focus on only those wells that express the desired protein product.

[0619] Fig. 33 shows a comparison of the time required for expression testing and virus purification between BaculoDirect™ and Bac to Bac. Numbers next to the arrows between steps are cumulative labor time in hours. Chronological elapsed times are indicated in days. Procedures common to both systems were given equal times, *e.g.*, 2 hours for transfection, 4 hours for expression testing.

[0620] Compared to other baculovirus expression systems, the methods described herein (*e.g.*, BaculoDirect™) require much less hands-on time and are faster chronologically. For example, Bac to Bac™ requires 10 days to obtain a purified viral stock and upwards of 17 hours of actual labor (Fig. 33). This assumes that the P1 stock obtained with Bac to Bac™ does not require plaque purification. In practice, one of skill in the art is likely to have difficulty in obtaining pure stock without plaque purification; as a result, plaque purification is now being encouraged for Bac to Bac™ users. The MaxBac baculovirus expression system relies on homologous recombination in insect cells, and, like other methods utilizing homologous recombination, requires plaque purification and even more chronological time and labor. By contrast, BaculoDirect requires only 8 hours of labor over six days to obtain a purified virus stock suitable for production of high titer stocks.

[0621] In summary, one of skill in the art with a collection of clones adapted for use in recombinational cloning methods (*e.g.*, pENTR clones adapted for GATEWAY™ methods) will be able to clone and express their genes of interest quickly in a baculoviral expression system of the present invention, using simple protocols, and in parallel reactions.

[0622] A suitable protocol for production of the recombinant baculoviruses of the invention is as follows:

- [0623] Materials: Sf9 or Sf21 cells growing in log phase; linearized BaculoDirect Virus DNA; GOI cloned into L1/L2 Entry clone (*e.g.*, pENTR CAT; LR clonase Buffer; LR clonase; and ganciclovir sodium (100 mM solution in water).
- [0624] A sequence of interest may be cloned into L1/L2 entry vector. Suitable cells (*e.g.*, Sf9 or Sf21 cells) may be plated at recommended densities (*e.g.*, Guide to Baculovirus Expression Systems and Insect Cell Culture, catalog nos. 10359016, 10360014, 10608016, 11827011, Invitrogen Corporation, Carlsbad, CA, February 27, 2002). HighFives are less preferred as they give low infectivity/titer. A suitable method may employ 6 well plates with 2 million Sf21 cells. An LR reaction may be performed between Entry vector and BaculoDirect™ linearized DNA (GATEWAY™ Manual) using 100 ng entry vector and 300 ng linearized BaculoDirect™ DNA. 1 h at room temperature. An aliquot (*e.g.*, 10 µl) of LR reaction may be transfected into the cells (*e.g.*, using Cellfectin® protocol). Transfection media may be replaced with growth media of choice, supplemented with 100 µM ganciclovir. After 72 hours, an aliquot (*e.g.*, 10 µl) of supernatant from transfected cells can be added to fresh well of cells with 100 µM ganciclovir in growth medium. Protein expression can be checked by western blot at this time. After 72 hours, supernatant can be collected (*e.g.*, in a sterile tube)x. Recommended: Stain cells with β-Gal staining kit. Viruses may be amplified as per standard protocols.

EXAMPLE 9

- [0625] In some embodiments, the present invention provides materials and methods for the construction and use of recombinant retroviruses, *e.g.*, lentiviruses. Although the present invention is exemplified using a lentivirus, any other type of retrovirus may be used in an analogous fashion to practice the present invention. A commercially available system for the construction of recombinant lentiviruses is ViraPower™ Lentiviral Expression System, available from Invitrogen Corporation, Carlsbad, CA. The ViraPower™ system provides a retroviral system for high-level expression in dividing and non-dividing eukaryotic cells, *e.g.*, mammalian cells. Examples of products

available from Invitrogen Corporation, Carlsbad, CA include the ViraPower™ Lentiviral Directional TOPO® Expression Kit catalog number K4950-00, the ViraPower™ Lentiviral GATEWAY™ Expression Kit catalog number K4960-00, and the ViraPower™ Lentiviral Support Kit catalog number K4970-00.

[0626] The present invention permits one skilled in the art to create replication-incompetent lentiviruses to deliver and express one or more sequences of interest (*e.g.*, genes). These viruses (based loosely on HIV-1) can effectively transduce dividing and non-dividing mammalian cells (in culture or *in vivo*), thus broadening the possible applications beyond those of traditional Moloney (MLV)-based retroviral systems (Clontech, Stratagene, etc.). Directional TOPO and GATEWAY™ lentiviral vectors have been created to clone one or more genes of interest with a V5 epitope, if desired. The vectors also carry the blasticidin resistance gene (*bsd*) to allow for the selection of transduced cells. Without additional modifications, these vectors can theoretically accommodate up to ~6 kb of foreign gene. Three supercoiled packaging plasmids (*gag/pol*, *rev* and *VSV-G* envelope) are provided to supply helper functions and viral proteins *in trans*. Finally, an optimized producer cell line (293FT) is provided that will facilitate production of high titer virus. A schematic representation of the production of a nucleic acid molecule comprising all or a portion of a lentiviral genome is shown in Figure 35. Plasmid maps of vectors adapted for use with GATEWAY™ and topoisomerase cloning in the production of nucleic acid molecules comprising all or a portion of a lentiviral genome are shown in Figures 36A (pLenti6/V5-DEST), 36B (pLenti6/V5-D-TOPO®), 36C (pLenti4/V5-DEST), and 36D (pLenti6/UbC/V5-DEST) respectively. The nucleotide sequences of the plasmids are provided in Tables 17-20. Plasmid maps of the three packaging plasmids pLP1, pLP2, and pLP/VSVG are shown in Figures 37A, 37B, and 37C respectively and the nucleotide sequences of these plasmids are provided as Tables 21, 22, and 23, respectively.

[0627] Retroviruses are RNA viruses that reverse transcribe their genome and integrate the DNA copy into a chromosome of the target cell. It was discovered that the retroviral packaging proteins (*gag*, *pol* and *env*) could be supplied *in trans*, thus allowing the creation of replication incompetent viral particles capable of stably delivering a gene of interest. These retroviral

vectors have been available for gene delivery for many years (Miller *et al.*, (1989) *BioTechniques* 7:980-990). One significant advantage of retroviral-based delivery is that the gene of interest is stably integrated into the genome of the host cell with very high efficiency. In addition, no viral genes are expressed in these recombinant vectors making them safe to use both *in vitro* and *in vivo*. However, one main drawback to the traditional Moloney-based retroviruses is that the target cell must undergo one round of cell division for nuclear import and stable integration to occur. Traditional retroviruses do not have an active mechanism of nuclear import and therefore must wait for the host cell nuclear membrane to breakdown during mitosis before they can access the host genomic DNA (Miller *et al.*, (1990) *Mol. Cell. Biol.* 10:4239-42).

[0628] Unlike traditional retroviruses, HIV (classified as a "lentivirus") is actively imported into the nuclei of non-dividing cells (Lewis *et al.*, (1994) *J. Virol.* 68:510-516). HIV still goes through the basic retrovirus lifecycle (RNA genome reverse transcribed in the target cell and integrated into the host genome); however, *cis*-acting elements facilitate active nuclear import, allowing HIV to stably infect non-dividing cells (for reviews see Buchschacher *et al.*, (2000) *Blood* 95:2499-2504, Naldini *et al.*, (1999) "*The Development of Human Gene Therapy*", Cold Spring Harbor Laboratory Press, pages 47-60). It is important to note that, for both lentivirus and traditional retroviruses, no gene expression occurs until *after* the viral RNA genome has been reverse transcribed and integrated into the host genome.

[0629] Similar to other retrovirus expression systems, the packaging functions of HIV can be supplied in *trans*, allowing the creation of lentiviral vectors for gene delivery. With all the viral proteins removed, the gene delivery vector becomes safe to use and allows foreign DNA to be efficiently packaged. In addition, it has been shown that lentiviral (or any retroviral) envelope proteins can be substituted for ones with broader tropism. The substitution of envelope is called pseudotyping, and allows creation of lentiviral vectors capable of infecting a wider variety of cells besides just CD4⁺ cells. Many have found that the G protein from vesicular stomatitis virus (VSV-G) is an excellent pseudotyping envelope protein that imparts a very broad host range for the virus (Yee *et al.*, (1994) *Proc. Natl. Acad. Sci. USA* 91:9564-9568). The

ability of pseudo-typed lentivirus to infect a broad range of non-dividing cells has led to its extensive use in animal gene delivery and gene therapy (Baek *et al.*, (2001) *Hum Gene Ther* 12:1551-8, Park *et al.*, (2001) *Mol Ther* 4:164-73, Peng *et al.*, (2001) *Gene Ther* 8:1456-63).

MATERIALS AND METHODS

[0630] **Vector constructions.** Lentiviral vector materials were received from Cell Genesys (Foster City, CA, see U.S. Patent Nos. 5,686,279; 5,834,256; 5,858,740; 5,994,136; 6,013,516; 6,051,427; 6,165,782 and 6,218,187, and Dull *et al.* (1998) *J. Virol.* 72(11):8463-8471) and modified to incorporate a blasticidin expression cassette and the V5 epitope tag using standard techniques to create pRRL6/V5 also referred to as pLenti6/V5. The nucleotide sequence of pRRL6/V5 is provided in Table 36. To create the GATEWAY™ Destination vector, pLenti6/V5-DEST, the Destination Vector Conversion cassette B (available from Invitrogen Corporation, Carlsbad, CA catalog #11828-019) was ligated into pRRL6/V5. This Destination vector was propagated in DB3.1 bacteria in the presence of ampicillin (100 µg/ml) and chloramphenicol (15 µg/ml) to maintain integrity. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the destination vector containing attP sites flanking the ccdB and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. It has recently been found that the use of spectinomycin selection instead of chloramphenicol selection results in an increase in the number of colonies obtained on selection plates, indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene.

[0631] To create the control Moloney retroviral vector, prKAT6/V5-DEST, prKAT (Cell Genesys) was digested with BamHI and filled-in with Klenow. This was ligated to the 2732 bp fragment, containing the DEST cassette and SV40-Bsd^R cassette, resulting from the digestion of pLenti6/V5-DEST with SpeI and Acc65I followed by Klenow fill-in and gel purification. This

Destination vector was propagated in DB3.1 bacteria in the presence of ampicillin (100 µg/ml) and chloramphenicol (15 µg/ml) to maintain integrity. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the destination vector containing attP sites flanking the ccdB and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. It has recently been found that the use of spectinomycin selection instead of chloramphenicol selection results in an increase in the number of colonies obtained on selection plates, indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene.

[0632] To create the expression control vector, pLenti6/V5-GW/lacZ, and the cognate Moloney retroviral control vector, prKAT/V5-GW/lacZ, GATEWAY™ LR reactions were performed with each of the DEST vectors and an entry vector having a copy of the lacZ gene with no stop codon according to the manufacturer's protocol.

[0633] **Directional TOPO adaptation.** The pRRL6/V5 vector was propagated in ampicillin (100 µg/ml) and blasticidin (10 µg/ml) to maintain integrity and reduce backgrounds in the TOPO adaptation. The pRRL6/V5 vector was Directionally TOPO-adapted at the EcoRI (5' end) and XhoI (3'-end) sites. EcoRI buffer (New England Biolabs, Beverly, MA) was used in the digest throughout; vectors were digested first for 3 hours with XhoI at 6 units of enzyme/µg of DNA followed by a 3 hour digestion with EcoRI at 4 units of enzyme/µg DNA. Digested DNA was purified by Phenol/Chloroform/Isoamyl alcohol (PCA) extraction, Ethanol precipitation, 80% Ethanol wash, followed by isopropanol precipitation and another 80% ethanol wash to remove the enzymes and the ~30 bp multicloning site between the EcoRI and XhoI sites. At this point, the concentration of the cut DNA was quantitated and 10 ng was transformed into chemically competent TOP10 *E. coli* to assess the amount of uncut vector (vector that had recombined to delete the multicloning site, or the original vector which "evaded" both restriction enzymes activity).

[0634] The oligonucleotides used for directional adaptation are listed below:

EcoRI (5' end): Non-regenerative site

Topo-D1 5' P-AATTGATCCCTTCACCGACATAGTACAG 3'

Topo-D2 5' P-GGTGAAGGGATC 3'

XhoI (3' end): Regenerative site

Topo-D6 5' P-TCGAGCCCTTGACATAGTACAG 3'

Topo-D7* 5' P-AAGGGC 3'

[0635] The oligonucleotides were used as pairs: Topo-D1/D2 and Topo-D6/D7 in 200 fold molar excess to vector (51 µg of Topo-D1/D2 pair and 40 µg of Topo-D6/D7 per 100 µg vector DNA).

Topo-D1 and D2 were paired in 2.3 to 1 mass ratio, respectively.

Topo-D6 and D7 were paired in 3.7 to 1 mass ratio, respectively.

[0636] 50 units of T4 DNA Ligase (New England Biolabs, Beverly, MA) per 1 µg of vector DNA was used in an overnight ligation (~16 hours) in a 14°C water bath to ligate the adapter oligonucleotides to the vectors. Subsequently, the sample was heated at 67.5°C for 15 minutes and then re-digested with EcoRI at 2 units of enzyme/µg vector DNA for 1.5 hours.

[0637] Free oligonucleotides were purified away from the oligonucleotide-adapted vector by PCA extraction and a Modified S.N.A.P. column purification protocol, as follows: The PCA extracted DNA (top aqueous phase) was added to 5 volumes of Modified Binding Buffer (MBB) [60% of S.N.A.P. Binding buffer : 40% of (100%) isopropanol], mixed and loaded onto a S.N.A.P. mini or midi (B) column; and the flow through was reloaded back onto the column once more. The column was then washed twice with SNAP Wash buffer, once with the Final Wash buffer (EtOH) and eluted in TE (60-100 µl for mini column and 750 µl for midi column) and concentration determined spectrophotometrically (OD_{260/280}) producing pLenti6/V5-D-TOPO™.

[0638] At least 50 µg of the oligo adapted vector was "Charged" with vaccinia topoisomerase in the following reactions (reagents added in the order listed):

Topo Charging with Kinase

Volume Reagent	Final Concentration
# μ l Topo adapted & purified DNA (at least 50 μ g)	
# μ l Topo D-70 annealing oligo	0.2 μ g / μ g vector DNA
50 μ l Vaccinia Topo Enzyme (1 mg/ml)	1 μ g Topo / μ g vector DNA
# μ l Water	
5.3 μ l 1 M Tris pH 7.5	15 mM
350 μ l Total	

Incubate the reaction at 37 degrees Celsius water bath for 10 minutes and then add:

Volume Reagent	Final Concentration
16.5 μ l 100 mM ATP	1.35 mM ATP (33 mM ATP/ μ g DNA)
4 μ l 1 M $MgCl_2$	10 mM $MgCl_2$
33 μ l 10 Units / μ l LTI T4 DNA Kinase	6.6 Units / 1 μ g DNA
403.5 μ l Total	

Incubate the reaction at 37 degrees Celsius water bath for 5 minutes and then load all of reaction into Q-column.

[0639] **TOPO Vector Purification.** Q-column purification was performed on the TOPO-charged sample with a 0-1M NaCl (50 mM Tris pH 7.5) gradient as reported for the TOPO-Adapted Entry vectors. DNA fluorescence characterization in the presence of Hoechst dye number 33258 (Sigma catalog #B-2883) was used to quantitate the concentration of individual or pooled fractions containing column purified TOPO-charged vector. In general, approximately 50% of the total DNA loaded onto the column is lost during the purification and the vector-TOPO complexes are eluted in ~500 mM NaCl. An equal volume of 2X TOPO-Vector Buffer (50 mM Tris 7.5, 2 mM EDTA, 2.5 mM DTT, 0.1 mg/ml BSA, 0.1% Triton X 100, 90 % glycerol) is added to the sample fractions. Therefore, the final TOPO Vector Buffer = 50 mM Tris 7.5, 1 mM EDTA, 1.25 mM DTT, 0.05 mg/ml BSA, 0.05% TritonX-100, 45% Glycerol. Samples are stored at -20 degrees Celsius until tested.

[0640] Standard Topogation reactions were set-up as follows:

- 1 μ l Topo-charged vector
- 1 μ l Directional insert PCR product*
- 1 μ l Salt Solution or 1 μ l water
- 3 μ l water

*Depending on the concentration of Topo-charged vector, PCR product insert should be adjusted to maximize yield. Ratio of 1 ng vector : 1-2 ng 750 bp insert (Or 1:10-20, vector:insert molar ratio) give good yields.

[0641] The topogation reactions were incubated at room temperature for 5 min. Two microliters of the reaction was added to TOP10 cells, incubated on ice for ~20 min, heat shocked for 40 seconds at 42°C, placed on ice, and then 250 µl of SOC was added to the transformed cells. Cells were shaken at 37°C for 1 hr and 100µl of the cell mixture was plated on LB-amp plates containing blasticidin (50 µg/ml final).

[0642] **Cell culture and growth arrest.** 293FT producer cells (available from Invitrogen Corporation, Carlsbad, CA, catalog number R7007) were cultured in DMEM/10% FBS/L-glutamine/non-essential amino acids/penicillin/streptomycin containing 500 µg/ml G418. MJ90 primary human foreskin fibroblasts, HT1080 human fibrosarcoma (ATCC #CCL-121) and HeLa cervical carcinoma cells (ATCC #CCL-2) were cultured in DMEM/10% FBS/non-essential amino acids/penicillin/streptomycin. Chinese hamster ovary cells (CHO-K1, ATCC #CCL-61) were cultured in Hams F12/10%FBS/L-glutamine/penicillin/streptomycin. For blasticidin selections, the following final concentrations were used: HT1080: 10 µg/ml, CHO: 5 µg/ml, HeLa: 2 µg/ml.

[0643] MJ90 primary cells were growth arrested by contact inhibition. Briefly, 1×10^5 cells were plated per well of a 6-well plate and media changes were performed every 3 days for 7 to 14 days, or until a quiescent monolayer was achieved. Aphidicolin (Sigma, St. Louis, MO, catalog number #A0781) was used to arrest HT1080 cells at the G1/S transition. Exponentially growing cultures were plated at 2×10^5 cells per well of a 6-well plate and the following day fresh media was supplied containing 1 µg/ml aphidicolin. Transductions of aphidicolin-arrested cells were performed in the continued presence of drug.

[0644] Primary, post-mitotic rat hippocampal and cortical neuronal tissues were received from BrainBits Inc. (Dr. Greg Brewer, University of Southern Illinois). Tissues were dissociated with a Pasteur pipette, spun down at 1100 rpm for one minute and resuspended in NeuroBasal Medium (Invitrogen Corporation, Carlsbad, CA, Gibco #21103-049) containing B27 supplement

(Invitrogen Corporation, Carlsbad, CA, Gibco #17504-010), 0.5 mM L-glutamine and 25 μ M glutamate. 5×10^4 hippocampal or 1×10^5 cortical neurons were plated per well in 24-well plates. Four days after plating, half of the medium was removed and replaced with complete NeuroBasal Medium (as above) but *without* the glutamate. The following day, cells were transduced with virus.

[0645] **Virus production.** For optimal virus production, 5×10^6 293FT cells were plated per 100 mm plate. Twenty-four hours later, the culture medium was replaced with 5 ml OptiMem/10%FBS (Opti-MEM®, catalog no. 22600050, Invitrogen Corporation, Carlsbad, CA) and cells were quadruple co-transfected, as follows. 12 μ g DNA total, at a mass ratio of 1:1:1:1 pLenti6/V5/gene:pLP-1:pLP-2:pLP/VSVG (3 μ g of each DNA) was mixed with 1.5 ml of OptiMem media. In a separate tube, 36 μ l of Lipofectamine 2000 was also mixed with 1.5 ml of OptiMem media. After a 5-minute incubation period at room temperature, the two mixtures were combined and incubated at room temperature for an additional 20 minutes. At the completion of the incubation period, the transfection mixture was added to the cells dropwise and the culture plate was gently swirled to mix. The following day the transfection complex was replaced with complete media (DMEM, 10% FBS, 1% penicillin/streptomycin, L-glutamine and non-essential amino acids). Forty-eight to seventy-two hours post transfection, the virus-containing supernatants were harvested, centrifuged at 3000 rpm for 15 minutes to remove dead cells and placed in cryovials in 1 ml aliquots. Titers were performed on fresh supernatants (see below) and the remaining viral aliquots were stored at -80 °C.

[0646] **Viral titering and transduction.** All applications of virus to cells were performed in the presence of 6 μ g/ml polybrene (Sigma, St. Louis, MO, catalog #H9268) and media changes were performed 12-24 hours post transduction. For titering virus, 6-well plates were seeded at 2×10^5 cells per well with HT1080 cells the day before transduction. One well served as an untransduced control (mock) and the remaining five wells contained 1 ml each of ten-fold serial dilutions of viral supernatant ranging from 10^{-2} to 10^{-6} (see example below). The dilutions were mixed by gentle inversion (dilutions should not be vortexed) prior to adding to cells. 6 μ g/ml of polybrene was

added to each well. The plate was gently swirled to mix. The following day, the media was replaced with complete media. Forty-eight hours post transduction, the cells were placed under 10 µg/ml blasticidin selection (Invitrogen). After 10 to 12 days of selection, the resulting colonies were stained with crystal violet: A 1% crystal violet solution was prepared in 10% ethanol. Each well was washed with 2 ml PBS followed by 1 ml of crystal violet solution for 10 minutes at room temperature. Excess stain was removed by two 2 ml PBS washes and colonies visible to the naked eye were counted to determine the viral titer of the original supernatants. In a typical example, colonies can be counted in the 10^{-5} and 10^{-6} dilutions.

[0647] **Protein analysis.** Total cell lysates were prepared using NP-40 lysis buffer (Igepal CA636, Sigma, St. Louis, MO) and the proteins (20 µg/lane) were separated on a 4-20% Novex Tris-Glycine gel. Following electrophoresis, the proteins were transferred to nitrocellulose. Western blotting was performed using the Western Breeze Chemiluminescence Kit (Invitrogen Corporation, Carlsbad, CA), using anti-large T antigen mouse monoclonal antibody (e.g., catalog no. 554149, BD Biosciences Pharmingen, San Diego, CA), anti-lacZ rabbit polyclonal antibody (1:5000 dilution, Invitrogen Corporation, Carlsbad, CA) or anti-V5 mouse monoclonal antibody (1:2000 dilution, Invitrogen Corporation, Carlsbad, CA). Beta-galactosidase activity assays were performed using the Galacto-Light Plus Kit (Tropix, Applied Biosystems, Foster City, CA) according to the manufacturer's instructions. Beta-galactosidase staining was performed using the β-Gal Staining Kit (Invitrogen Corporation, Carlsbad, CA) according to the manufacturer's instructions.

[0648] The present invention provides a production and expression kit that allows easy construction, production and use of nucleic acid molecules comprising all or a portion of a lentiviral genome (e.g., lentiviral vectors). Aspects of the invention include, but are not limited to, 1) directional TOPO® and GATEWAY™ Destination pLenti6/V5 vectors with a useful selectable marker and epitope tag, 2) optimized virus production conditions and cell lines to reproducibly achieve $>10^5$ infectious viral particles per ml, 3) stable gene delivery and expression of at least two genes into actively dividing mammalian cells, and 4) transduction of at least two non-dividing cell types.

- [0649] A four plasmid co-transfection is used to create infectious lentiviral vectors (Dull, *et al.*, (1998) *J. Virol.* 72:8463-8471). One of the vectors (pLenti6/V5-DEST, pLenti6/V5-D-TOPO[®], pLenti4/V5-DEST, or pLenti6/UbC/V5-DEST) contains the gene of interest and is packaged into the virions (for vector maps, see Figures 36A-D). The other three plasmids are co-transfected to supply the viral proteins in *trans*. None of these three vectors are packaged into the virions. Each vector and a description of its features is described in more detail below. Vector maps are provided as Figures 37A, 37B, and 37C.
- [0650] pLenti6/V5-DEST or pLenti6/V5-D-TOPO carries gene of interest and blasticidin resistance gene and is packaged into viral particles. The vector contains the RSV promoter, which enhances production of the viral genomic RNA in the producer cell and removes dependence on HIV tat protein. The vector also contains viral 5' and 3' LTRs (Long Terminal Repeats), which are required for viral packaging and reverse transcription of the viral RNA. The 3' LTR also contains polyA signal. The vector contains the Ψ (psi) packaging signal. Nuclear export of unspliced viral genomic RNA in the presence of rev occurs as a result of the RRE (Rev-Responsive Element) present in the vector. The vector also incorporates 5' and 3' splice sites that result in the removal of psi and RRE making expression of the gene of interest no longer rev-dependent in the host cell. The vector also contains Delta U3, a 400 bp deletion in the 3' LTR that gets copied to the 5' LTR after reverse transcription of the viral genome in the transduced target cell. This results in "self-inactivation" of the 5' LTR for biosafety.
- [0651] pLP1 expresses HIV-1 gag and pol genes in *trans* and is not packaged into viral particles produced with this system. The plasmid contains the RRE, which makes expression of gag/pol genes rev-dependent (for safety purposes).
- [0652] pLP2 expresses HIV-1 rev gene in *trans* and, like pLP1, is not packaged into viral particles. The plasmid encodes the rev protein, which is required for gag/pol expression and for nuclear export of the unspliced viral genome (from pLenti6/v5-DEST or D-TOPO[®]) for packaging into the virions.
- [0653] pLP/VSVG expresses the VSV-G envelope gene in *trans*. The plasmid is not packaged into viral particles, however, the VSV-G protein is

incorporated into the viral particle. VSV-G is a non-HIV envelope that broadens the host range and stabilizes the viral particles (Yee 1994).

[0654]

RESULTS AND DISCUSSION

[0655]

Vector construction. The vector pRRLsin.hCMV.GFPpre was used as the starting material from Cell Genesys. This vector contains the essential elements for lentiviral packaging (*e.g.*, 5' and 3' LTRs, psi packaging signal, rev responsive element (RRE) and necessary splice sites; see above for descriptions). In addition, it contains a deletion in the 3' LTR (called "delta U3") that results in a self-inactivation of the 5' LTR after integration of the viral genome into the genome of the target cell (Dull 1998, Zufferey *et al.*, (1998) *J. Virol.* 72:9873-80). This is an additional safety measure (see "Safety" section below) and has no effect on vector performance since the 5' LTR is only needed during viral production, not gene expression in the target cell (Zufferey 1998). Finally, all polyadenylation (polyA) functions are supplied by the 3' LTR. The 3' LTR serves as the polyA for the viral genome (driven by the RSV/5' LTR), the CMV promoter (gene of interest) and the SV40 promoter (blasticidin resistance). No heterologous polyA signals should ever be included between the LTRs or viral production will be severely compromised due to transcription termination prior to the 3' LTR. The downstream SV40 polyA in the pLenti6/V5 vectors simply enhances viral genomic RNA production in the producer cells and is not packaged into the virions.

[0656]

TOPO adaptation and purification. Fifty micrograms of TOPO-charged pLenti6/V5-D-TOPO[®] was loaded on the Q-column and fractions containing the purified vector were collected in seven 0.5 ml fractions. The peak fraction (fraction 41) contained ~20 µg of DNA by Hoechst (H 33258) dye DNA fluorescence characterization and was eluted of at ~500 mM NaCl. Only this fraction was analyzed, however fractions 39-45 also contained TOPO-charged DNA. The fractions were diluted in 2X TOPO dilution buffer, so fraction 41 contained vector at ~20 ng/µl final concentration. TOPO transformation results, using fraction 41 in two experiments (one with 750 bp insert, one with lacZ-alpha), are shown in Table 24.

Table 24. pLenti6/V5-D-TOPO® transformations.

Vector	Insert	#colonies/ μ l vector	Orientation (% correct)	% background
pLenti6/V5-D-TOPO	None	162 612	-- --	-- --
	750 bp test	1665 --	9/10 (90%) --	9.7% --
	LacZ alpha	-- 4464	-- 17/18 (94%)	-- 13.7%

Note: Transformation efficiency with pUC19: 4008 colonies/10 pg = 4.0×10^8 cfu/ μ g efficiency

[0657] **Vector instability.** While performing manipulations on the vectors, it was discovered that the presence of 182 basepairs of direct repeat present in the LTRs was triggering homologous recombination when transformed into TOP10 and plated on LB-amp. This resulted in a visible colony phenotype. In clones where LTR recombination occurred the colonies were large, while unrecombined (correct) clones resulted in small colonies. Figure 38 shows the results of an experiment in which two LR reactions were performed with either pLenti6/V5-DEST alone or pLenti/V5-DEST plus pENTR/CAT and 3 μ l of each was transformed into TOP10 cells. 100 μ l of the transformations were plated on regular LB-amp plates (No Bsd in Figure 38) or LB-amp containing 50 μ g/ml blasticidin. After overnight incubation at 37 degrees, colonies were photographed (Figure 38A) and counted (Figure 38B). Twenty-four clones (twelve each from two independent experiments) from the DEST + CAT plates (+/- Bsd) were randomly picked and screened by restriction digest to determine the percentage of correct clones.

[0658] Since blasticidin resistance (driven by the EM7 promoter) is present between the LTRs, it was found that spreading blasticidin on the bacterial plate (to a final concentration of 50 μ g/ml) resulted in all small colonies, none of which contained the LTR recombination product (not shown). This was further confirmed when GATEWAY™ LR reactions were performed using pLenti6/V5-DEST with and without including the pENTR-CAT plasmid (Figure 38B). Without blasticidin in the plate, background colonies arose from the DEST vector alone and only 50% of the DEST + CAT clones were intact. However, when blasticidin was included in the plate, the DEST vector

alone gave no background colonies and all DEST + CAT clones were correct and intact (Figure 38B). Therefore, it is recommended that one of two approaches be use when introducing a gene of interest into pLenti6/V5 vectors: 1) if high efficiency cloning (i.e. library-scale) is not required, simply pick only the small colonies for miniprep analysis; or 2) to ensure ~100% correct clones, include blasticidin (50 µg/ml) in the bacterial plate following transformation. It has been observed that once a clone is isolated and shown to be intact, it appears to remain stable over multiple rounds of large-scale propagation without blasticidin. Nevertheless, it is recommended that each DNA preparation be verified by restriction digest prior to proceeding to virus production.

[0659] **Transient transfection expression testing.** To verify protein expression and the functionality of the V5 epitope tag, the lacZ ORF (with or without a stop codon) was GATEWAY™ cloned into pLenti6/V5-DEST. The resulting attB expression clones were transiently transfected into COS cells and analyzed by anti-β-galactosidase and anti-V5 western blotting (Figure 39). COS-7 cells were transiently transfected with:

Lane 1: mock; Lane 2: pcDNA3.1/V5His/lacZ; Lane 3: pLenti6/V5-GW-lacZ (no stop); Lane 4: pLenti6/V5-GW-lacZ (with stop); and lysates were analyzed by anti-lacZ or anti-V5 western blotting as indicated.

[0660] Compared to pcDNA3.1/V5His/lacZ, pLenti6/V5-GW/lacZ expressed equally well with and without the V5 epitope tag. In addition, lacZ (no stop) resulted in an efficiently expressed V5-tagged fusion protein (lane 3). This vector can be used as an expression control vector and may be included in kits of the invention.

[0661] **Virus production optimization.** Previous reports had indicated that virus production is maximal in human 293 cells that express the SV40 large T antigen (Naldini, *et al.*, (1996) *Proc. Natl. Acad. Sci. USA* 93:11382-11388). Virus production was tested in several neomycin-resistant 293FT clones. These cell lines were created by stably transfecting 293F cells with the pCMV/Sport6-T antigen plasmid in which the SV40 origin had been deleted. 293FT clone #42 was found to produce the highest levels of infectious virus. The expression of the SV40 large T antigen was confirmed by western blot

analysis and producer cell stocks were propagated in G418 to maintain the large T antigen expression.

[0662] Since the production of virus requires a quadruple transfection, the importance of the ratio of the four plasmids was tested. Published reports suggested a variety of ratios as "optimal" (Dull 1998; Naldini 1996; Mochizuki *et al.*, (1998) *J. Virol.* 72:8873-8883), so each published ratio was evaluated and compared to the simple 1:1:1:1. Little difference was seen between the simple 1:1:1:1 and the more elaborate ratios (e.g. 4:2.6:1:1.4). The highest and most reproducible titers were generated using a simple ratio of 1:1:1:1. The most effective time course for production of virus was determined. Various genes were cloned into pLenti6/V5 and virus was produced in 293FT cells according to the following optimized protocol

Day 0 Plate 5×10^6 293FT per 100 mm plate

Day 1 Four plasmid co-transfection (ratio = 1:1:1:1)

12 μ g DNA total (3 μ g each)

36 μ l Lipofectamine 2000

Day 2 Replace media

Day 3-4 Harvest supernatant containing virus

Spin 3000 rpm x 15' and/or filter 0.45 μ m

Aliquot supernatant, use for titering and store -80°C

[0663] Independent virus productions, of either the empty vector (pLenti6/V5-DEST) or carrying lacZ, GFP, CAT or protein kinase C, were titrated on HT1080 cells by counting the number of resulting blasticidin-resistant colonies generated per ml of supernatant and the results are shown in Figure 40. The optimized protocol which included high density plating of the 293FT cells (5×10^6 cells per 100 mm plate) and the optimal lipid to DNA ratio using Lipofectamine 2000. It was found that viral supernatants can be harvested either 2 or 3 days post transfection with minimal differences in viral yield. Presumably, the short half-life of the virus in culture media at 37°C negates any advantage of viral accumulation over one extra day. For storage, aliquotting viral stocks at -80°C is recommended. Anywhere from 0 to 10% loss of viral titer for each freeze/thaw cycle of crude supernatant was observed.

[0664] The size of the inserted gene of interest can affect the viral titer. Three different genes were GATEWAY™ cloned into pLenti6/V5-DEST (lacZ, CAT and protein kinase C) and one gene was directionally TOPO cloned (GFP). Viral production was compared between these four gene-containing vectors and an empty vector, pLenti6/V5 (Figure 40). Averages from three independent experiments showed that the empty vector yielded the highest viral titer (average 1.4×10^7 cfu/ml), while the largest insert (lacZ) yielded the lowest titers (average 4.7×10^5 pfu/ml). Inserted genes of intermediate size (GFP, CAT and PKC) yielded titers somewhere in between (4×10^6 , 9×10^6 and 3×10^6 ; respectively). These data indicated that both the GATEWAY™ and TOPO versions of these vectors can produce viral supernatants that easily exceed a viral titer of 10^5 , even with the large lacZ gene. The wild type HIV-1 genome is approximately 10 kb and the elements present in pLenti6/V5 vectors add up to 3.7 kb. Therefore, the theoretical gene-packaging limit is approximately 6 kb.

[0665] **Viral gene delivery and expression.** The ability of the lentiviral vectors to deliver and express a variety of genes was further investigated. HT1080 cells were transduced with either Lenti6/V5-GW/lacZ virus (GATEWAY™) or Lenti6/V5-dT/GFP virus (D-TOPO®) and selected for 10 days with 10 µg/ml blasticidin. LacZ was visualized using the β-Gal Staining kit and GFP was visualized using the fluorescent microscope (Figure 41). Both the GATEWAY™ lacZ and the D-TOPO® GFP vectors efficiently generated heterogeneous pools of stably transduced cells in which nearly 100% of the cells expressed the heterologous gene. In addition to HT1080, HeLa and CHO cells have been stably transduced with similar efficiencies and levels of gene expression.

[0666] To confirm the above results, and to verify that a functional V5 epitope tag was efficiently added to the expressed proteins, cell lysates were prepared from HT1080 cells stably transduced with either the lacZ, CAT, GFP or protein kinase C viruses (Figure 42). HT1080 cells were transduced in duplicate with lentiviral vectors carrying genes for either lacZ, CAT, GFP or PKC and selected for 10 days with 10 µg/ml blasticidin. Cell lysates were analyzed by anti-V5 western blotting. Molecular weight markers and each V5 fusion protein are indicated. *indicates background V5 band. All four

proteins are efficiently expressed and all are properly fused to a detectable V5 epitope. In addition, the delivery and efficient expression of protein kinase C (a "relevant" gene, *i.e.*, not lacZ, GFP or CAT) indicates the robustness and broad applicability of this virus production system.

[0667] **Gene expression is correlated to MOI.** Theoretically, the multiplicity of infection (MOI = number of virus per cell) should correlate with gene delivery and expression. To investigate this, HT1080 cells were transduced in duplicate at various MOIs, ranging from 0.05 to 1 (Figure 43). HT1080 cells were transduced in duplicate with Lenti6/V5-GW/lacZ virus at multiplicities of infection (MOI) of 0.05, 0.1, 0.5 and 1. Forty-eight hours later, cells were either stained for β -Gal (Figure 43A) or harvested and analyzed for β -Gal activity (Figure 43B). As the β -Gal staining indicates, an increasing number of cells become lacZ-positive as the MOI increases. At an MOI of 1, greater than 80% of the cells express lacZ. At higher MOIs (e.g. MOI 5), 100% of the cells were transduced. When cell lysates were analyzed for lacZ activity, a near-linear dose response was observed as the MOI increased from 0.05 to 1 (Figure 43B). At higher MOIs (e.g. MOI 5), the lacZ activity continues to increase, but graph tends to flatten out.

[0668] **Lentiviral transduction of non-dividing cells.** One of the key advantages of lentiviruses over traditional retroviruses is that they are capable of stably transducing non-dividing cells. This significantly expands the potential transducible target cells to include: 1) growth- or drug-arrested cells in culture, 2) non-dividing primary cell cultures, and 3) animals/tissues. To verify that lentiviral vectors of the invention could perform under these conditions, they were using three different approaches.

[0669] **Drug-arrested cells.** Actively-growing cells in culture can be arrested at specific phases of the cell cycle using a variety of drugs. This approach is widely used in cell cycle analysis and tumor biology. One commonly used drug, aphidicolin, reversibly binds to DNA polymerase delta and is used to arrest cells at the G1/S transition (Seki *et al.*, (1980) *Biochem Biophys Acta* 610:413). To test the activity of lentiviral vectors of the invention under conditions of cell cycle arrest, aphidicolin-blocked HT1080 cells were transduced with Lenti6/V5-GW/lacZ virus (Figure 44A). HT1080 cells were either actively growing or growth arrested at G1/S by aphidicolin and

transduced at an MOI of 1, in duplicate, with either rKAT6/V6-GW/lacZ retrovirus or Lenti6/V5-GW/lacZ lentivirus. Forty-eight hours post transduction, cell lysates were analyzed for beta-galactosidase activity. The control virus, rKAT6/V5-GW/lacZ virus, is a traditional Moloney-based retrovirus carrying the same lacZ gene. Both retrovirus and lentivirus were capable of transducing actively growing cells, but only the lentiviral vector was capable of transducing the non-dividing culture.

[0670] **Quiescent primary cells.** The second approach was to apply the lentiviral vectors to non-dividing primary human cultures. A low-passage primary human foreskin fibroblast culture (MJ90, Grand Island) was plated into 6-well format and allowed to grow to confluence. Primary fibroblasts are strongly contact inhibited and can be maintained for many weeks arrested in quiescence (G_0) when maintained as a confluent culture. Contact-inhibited non-dividing quiescent primary human foreskin fibroblasts were transduced with retrovirus (rKAT6/V5-GW/lacZ) and lentivirus (Lenti6/V5-GW/lacZ) at an MOI of 1 and β -Gal stained forty-eight hours post transduction. Similar to the results in aphidicolin-arrested cells, only the lentiviral vector (and not the retroviral rKAT vector) was capable of transducing non-dividing cells. Approximately 50% of the quiescent primary cells were transduced with an MOI of 1 (Figure 44B).

[0671] **Post-mitotic primary neurons.** Neuronal research is one area where lentiviral vectors can offer significant advantages over other gene transfer methods. Neuronal cultures are typically non-dividing, "post-mitotic" cells that transfect poorly. Traditional Moloney retroviruses are not useful since the cells never go through mitosis. Lentiviral vectors are one solution to overcome these hurdles, and vectors of the invention were tested to determine if they could stably transduce these cells. Primary, post-mitotic rat neuronal tissues (cortical and hippocampal) were received from BrainBits, Inc. and then processed and plated. Four days after plating, cells were transduced at an MOI of 1 with either Lenti6/V5-GW/lacZ lentivirus or rKAT6/V5-GW/lacZ retrovirus. Three days post-transduction, cultures were stained for β -galactosidase. All wells transduced with the lentiviral vectors stained blue, with approximately 50% of the cells expressing detectable β -galactosidase. Conversely, wells transduced with the rKAT retrovirus did not show any β -

galactosidase expression. These results indicated that lentiviruses of the invention effectively transduced post-mitotic neurons of either cortical or hippocampal origin.

[0672] **Long-term gene expression from lentiviral vectors.** The stability of gene expression after delivery by lentiviral transduction was tested. HT1080 cells were transduced with either the Lenti6/V5-GW/lacZ lentivirus or the rKAT6/V5-GW/lacZ retrovirus and stably selected with 10 µg/ml blasticidin. Cultures were maintained in blasticidin and were β-Gal stained at 10 days (Figure 45A) and 6 weeks (Figure 45B) post transduction. No loss of gene expression was observed over 6 weeks in culture, indicating that lentiviral gene delivery is stable and gene expression is persistent even at 6 weeks post transduction.

[0673] The present invention describes the generation of infectious lentiviral particles based on the genome and lifecycle of HIV-1. Considerable effort has been put into developing a system that is safe to use and is as far-removed from wild type HIV as possible. Key safety features built into this “3rd generation” system are as follows:

[0674] The viral particles produced in this system are replication incompetent and only carry the gene(s) of interest. No other viral species are produced. This also means that none of the structural HIV genes (necessary for production of viral progeny) are present in the packaged viral genome. Only sequences flanked by the viral LTRs will be packaged into virions (*i.e.*, pLenti6/V5 vector). None of the three packaging plasmids contain LTRs (Figures 37A-C); so while they are expressed in the producer cell, they are never packaged into the virions. Once a cell is infected (the proper term for this event is “transduced”), the only genes that are delivered and expressed are the gene of interest and the selectable marker. Gag, pol, rev and envelope genes are not present in the viral genome and are therefore never expressed in the target cell, so no new virus can be produced.

The system described above is a four-plasmid system. The necessary HIV-1 genes (gag-pol and rev) have been separated onto individual plasmids, and the non-HIV envelope is on a third plasmid (Figures 37A-C). All four plasmids have been engineered not to contain any regions of homology with each other to prevent unwanted recombination events that could lead to the

generation of a replication competent virus (Dull 1998). In other words, multiple non-homologous recombination events would need to occur to get all necessary components into one viral genome. In addition, the expression of gag and pol (from pLP1) is rev-dependent, by virtue of the RRE in the gag/pol transcript. This prevents unwanted gag/pol expression if rev is not present (Dull 1998). In other embodiments, one or more of the genes necessary for generation of a replication-incompetent retrovirus according to the methods of the invention (*i.e.*, gag, pol, rev, and a pseudotyping envelope protein) may be expressed from the genome of a host cell. Thus, some or all of the necessary genes may be expressed from plasmids and some or all of the necessary genes may be expressed from the host cell genome. In a particular embodiment, one or more of the necessary genes may be expressed from the host cell genome and at least one gene expressed from the host cell genome may be operably linked to an inducible promoter. In another embodiment, all the genes necessary may be expressed from the genome of a host cell and one or more may be operably linked to an inducible promoter. When more than one gene is operably linked to an inducible promoter, the inducible promoters may be the same or different.

[0675] The gene transfer vector pLenti6/V5 has been modified to be "self-inactivating" (Yu *et al.*, (1986) *Proc. Natl. Acad. Sci. USA* 83:3194-3198, Yee *et al.*, (1987) *Proc. Natl. Acad. Sci. USA* 84:5197-5201, Zufferey 1998). A deletion has been made in the 3' LTR (called "delta U3") that has no effect on the generation of viral genome for packaging in the producer cell. However once the produced virus transduces a target cell, the mechanisms of reverse transcription use the 3' LTR as a template to create the 5' LTR. The end result is an integrated viral genome that is defective in both its 5' and 3' LTRs, and is no longer capable of producing packagable viral genome. This means that transduction with lentiviral vectors of the invention does not generate a productive infection, instead ending with a gene of interest integrated into the host cell genome.

[0676] Despite all of these safety features, the lentivirus produced with this system can still pose a biohazardous risk. As shown above, they are fully capable of transducing primary human cells, thus these viruses should be treated as Biosafety Level 2 organisms. Extra care should be taken when

creating viruses carrying harmful or toxic genes (such as activated oncogenes). For further information on BL-2 guidelines and lentivirus handling, please refer to: "Biosafety in Microbiological and Biomedical Laboratories", 4th Ed. Centers for Disease Control and contact the CDC.

[0677] **Conclusions.** The lentivirus production and expression system of the invention is based on the 3rd Generation lentiviral system created at Cell Genesys (Dull 1998). This system allows one skilled in the art to rapidly clone their gene of interest into a packagable lentiviral vector, via GATEWAYTM or directional TOPO, and provides materials necessary for the creation of infectious viral particles. Finally, these viruses are capable of stably delivering a variety of genes to both actively dividing and non-dividing primary and immortalized human cell lines.

EXAMPLE 10

[0678] Materials and methods of the present invention (*e.g.*, the ViraPowerTM Lentiviral Expression System) allow creation of a replication-incompetent retroviruses (*e.g.*, an HIV-1-based lentivirus), which can then be used to deliver and express a sequence of interest in either dividing or non-dividing eukaryotic (*e.g.*, mammalian) cells. In some embodiments, materials of the present invention may include, but are not limited to, expression plasmids, for example, an expression plasmid that contains the sequence of interest under the control of a suitable promoter (*e.g.*, the human cytomegalovirus (CMV) immediate-early enhancer/promoter; see Andersson, *et al.* (1989) *J. Biol. Chem.* 264, 8222-8229; Boshart, *et al.* (1985) *Cell* 41, 521-530; Nelson, *et al.* (1987) *Molec. Cell. Biol.* 7, 4125-4129) and also contains elements that allow packaging of the construct into virions. Other materials suitable for the practice of the present invention include an optimized mix of packaging plasmids (*e.g.*, pLP1, pLP2, and pLP/VSVG) which may supply the structural and replication proteins *in trans* that are required to produce a recombinant retrovirus. In some embodiments, the present invention provides a cell line (*e.g.*, 293FT), which allows production of the lentivirus following cotransfection of the expression plasmid and the plasmids in the packaging mix. In some embodiments, the present invention provides a control

expression plasmid containing the *lacZ* gene which, when packaged into virions and transduced into a mammalian cell line, expresses β -galactosidase.

[0679] Using the materials and methods of the present invention (*e.g.*, the ViraPower™ Lentiviral Expression System) to facilitate retroviral-based expression of the gene of interest provides the following advantages: 1) generates an HIV-1-based lentivirus that effectively transduces both dividing and non-dividing mammalian cells, thus broadening the potential applications beyond those of traditional Moloney Leukemia Virus (MoMLV)-based retroviral systems (Naldini, 1998); 2) efficiently delivers the gene of interest to mammalian cells in culture or *in vivo* (Dull *et al.*, 1998); 3) provides stable, long-term expression of a target gene beyond that offered by traditional adenoviral-based systems (Dull *et al.*, 1998; Naldini *et al.*, 1996); 4) produces a pseudotyped virus with a broadened host range (Yee *et al.*, 1994); and 5) includes multiple features designed to enhance the biosafety of the system.

[0680] One of skill in the art can use the teachings provided herein to: co-transfect the vectors described herein (*e.g.*, pLenti6/V5-based expression vector) and the ViraPower™ Packaging Mix into the 293FT cell line to produce a lentiviral stock; titer the lentiviral stock; use the lentiviral stock to transduce a mammalian cell line of choice; assay for “transient” expression of one or more recombinant proteins encoded by the transduced vector; and/or generate a stably transduced cell line, if desired.

[0681] Additional details and instructions to generate an expression vector using pLenti6/V5-D-TOPO® or pLenti6/V5-DEST™ are available (*e.g.*, pLenti6/V5 Directional TOPO® Cloning Kit manual, catalog no. K4955-10, version B, or pLenti6/V5-DEST™ GATEWAY™ Vector Pack manual, catalog nos. V496-10, V498-10, and V499-10, version C, Invitrogen Corporation, Carlsbad, CA). For instructions to culture and maintain the 293FT producer cell line, see Example 13 below.

[0682] Expression systems of the present invention (*e.g.*, the ViraPower™ Lentiviral Expression System) facilitate highly efficient, *in vitro* or *in vivo* delivery of a target gene to dividing and non-dividing mammalian cells using a replication-incompetent lentivirus. Based on the lentikat™ system developed by Cell Genesys (Dull *et al.*, 1998), the ViraPower™ Lentiviral Expression System possesses features which enhance its biosafety while allowing high-

level gene expression in a wider range of cell types than traditional retroviral systems.

[0683] One component of the systems of the invention is an expression vector (e.g., a pLenti6/V5-based expression vector) into which the sequence of interest (e.g., encoding a gene of interest) will be cloned. Expression of the sequence of interest is controlled by a promoter of choice, for example, the human cytomegalovirus (CMV) promoter. The vector also contains the elements required to allow packaging of the expression construct into virions (e.g. 5' and 3' LTRs, Ψ packaging signal).

[0684] Another component of a system of the invention is one or more plasmids encoding the activities necessary for packaging the RNA produced from the expression vector (e.g., the ViraPower™ Packaging Mix that contains an optimized mixture of the three packaging plasmids, pLP1, pLP2, and pLP/VSVG). These plasmids supply the helper functions as well as structural and replication proteins *in trans* required to produce the lentivirus.

[0685] An optional component of the system is an optimized cell line (e.g., the 293FT producer cell line) that may stably express the SV40 large T antigen. Expression of the SV40 large T antigen may be under the control of any promoter known in the art, for example, the human CMV promoter. Expression of the large T antigen facilitates optimal production of virus.

[0686] In an embodiment, plasmids containing the packaging activities (e.g., the ViraPower™ Packaging Mix) and an expression plasmid (e.g., the pLenti6/V5 vector containing a sequence of interest) may be co-transfected into a suitable host cell line (e.g., 293FT cells) to produce a replication-incompetent lentivirus, which can then be transduced into the mammalian cell line of interest. Once the lentivirus enters the target cell, the viral RNA is reverse-transcribed, actively imported into the nucleus (Lewis *et al.* 1994; Naldini *et al.*, 1999), and stably integrated into the host genome (Buchsacher *et al.*, 2000; Luciw, (1996) In *Fields Virology*, B. N. Fields, *et al.* eds. (Philadelphia, PA: Lippincott-Raven Publishers), pp. 1881-1975). Once the lentiviral construct has integrated into the genome, transient expression of a recombinant protein can be assayed or blasticidin selection can be used to generate a stable cell line for long-term expression.

[0687] Most retroviral vectors are limited in their usefulness as gene delivery vehicles by their restricted tropism and generally low titers. In the systems of the invention (*e.g.*, the ViraPower™ Lentiviral Expression System), this limitation has been overcome by use of the G glycoprotein gene from Vesicular Stomatitis Virus (VSV-G) as a pseudotyping envelope, thus allowing production of a high titer lentiviral vector with a significantly broadened host cell range (Burns *et al.*, (1993) *Proc. Natl. Acad. Sci. USA* 90, 8033-8037, Emi *et al.*, (1991) *J. Virol.* 65, 1202-1207, Yee *et al.*, 1994).

Cell Lines and Cell Types Tested

Cell Line or Cell Type	Description	Condition Tested
293	Human embryonic kidney (Graham <i>et al.</i> , (1977) <i>J. Gen. Virol.</i> 36, 59-74)	Actively dividing
HT1080	Human fibrosarcoma (Rasheed <i>et al.</i> , (1974) <i>Cancer</i> 33, 1027-1033)	Actively dividing Aphidicolin-arrested (at the G1/S transition)
HeLa	Human cervical adenocarcinoma	Actively dividing
CHO-K1	Chinese hamster ovary (Kao <i>et al.</i> , (1968) <i>Proc. Natl. Acad. Sci. USA</i> 60, 1275-1281)	Actively dividing
Primary foreskin fibroblasts	Human foreskin	Contact inhibited, growth-arrested (in G ₀)
Primary hippocampal neurons	Rat neuronal tissue	Non-dividing, post-mitotic
Primary cortical neurons	Rat neuronal tissue	Non-dividing, post-mitotic

[0688] The present invention is suitable for *in vivo* gene delivery applications. Many groups have successfully used lentiviral vectors to express a target gene in tissues including brain, retina, pancreas, muscle, liver, and skin (Gallichan *et al.*, (1998) *Human Gene Therapy* 9, 2717-2726; Kafri *et al.*, (1997) *Nature Genetics* 17, 314-317; Miyoshi *et al.*, (1997) *Proc. Natl. Acad. Sci. USA* 94, 10319-10323; Naldini, (1998) *Curr. Opin. Biotechnol.* 9, 457-463; Pfeifer *et al.*, (2001) *Proc. Natl. Acad. Sci. USA* 98, 11450-11455; Pfeifer *et al.*, (2001) *Mol. Ther.* 3, 319-322; Takahashi *et al.*, (1999) *J. Virol.* 73, 7812-7816). For more information about target genes that have been successfully expressed *in*

vivo using lentiviral-based vectors, refer to the references above as well as the following additional references (Baek *et al.*, 2001; Dull *et al.*, 1998; Park *et al.*, 2001; Peng *et al.*, 2001).

- [0689] The systems of the invention (*e.g.*, the ViraPower™ Lentiviral Expression System) are third-generation systems based on lentiviral vectors developed by Dull *et al.* (1998). These third-generation lentiviral systems include a significant number of safety features designed to enhance their biosafety and to minimize their relation to the wild-type, human HIV-1 virus. These safety features are discussed below.
- [0690] The expression vector (pLenti6/V5-D-TOPO® or pLenti6/V5-DEST™) contains a deletion in the 3' LTR (Δ U3) that does not affect generation of the viral genome in the producer cell line, but results in "self-inactivation" of the lentivirus after transduction of the target cell (Yee *et al.*, 1987; Yu *et al.*, 1986; Zufferey *et al.*, 1998). Once integrated into the transduced target cell, the lentiviral genome is no longer capable of producing packageable viral genome.
- [0691] The number of genes from HIV-1 that are used in the system has been reduced to three (*i.e.* *gag*, *pol*, and *rev*).
- [0692] The VSV-G gene from Vesicular Stomatitis Virus is used in place of the HIV-1 envelope (Burns *et al.*, 1993; Emi *et al.*, 1991; Yee *et al.*, 1994).
- [0693] Genes encoding the structural and other components required for packaging the viral genome are separated onto four plasmids. All four plasmids have been engineered not to contain any regions of homology with each other to prevent undesirable recombination events that could lead to the generation of a replication-competent virus (Dull *et al.*, 1998).
- [0694] Although the three packaging plasmids allow expression *in trans* of proteins required to produce viral progeny (*e.g.* *gal*, *pol*, *rev*, *env*) in the 293FT producer cell line, none of them contain LTRs or the Ψ packaging sequence. This means that none of the HIV-1 structural genes are actually present in the packaged viral genome, and thus, are never expressed in the transduced target cell. No new replication-competent virus can be produced.
- [0695] The lentiviral particles produced in this system are replication-incompetent and only carry the gene of interest. No other viral species are produced.

- [0696] Expression of the *gag* and *pol* genes from pLP1 has been rendered Rev-dependent by virtue of the HIV-1 RRE in the *gag/pol* mRNA transcript. Addition of the RRE prevents *gag* and *pol* expression in the absence of Rev (Dull *et al.*, 1998).
- [0697] A constitutive promoter (RSV promoter, Gorman *et al.* (1982). *Proc. Natl. Acad. Sci. USA* 79, 6777-6781) has been placed upstream of the 5' LTR in the pLenti6/V5 expression vector to offset the requirement for Tat in the efficient production of viral RNA (Dull *et al.*, 1998).
- [0698] Despite the inclusion of the safety features discussed above, the lentivirus produced with the systems of the invention can still pose some biohazardous risk since they can transduce primary human cells. For this reason, published guidelines for BL-2 should be followed. Furthermore, exercise extra caution when creating lentivirus carrying potential harmful or toxic genes (*e.g.* activated oncogenes).
- [0699] For more information about the BL-2 guidelines and lentivirus handling, refer to the document, "Biosafety in Microbiological and Biomedical Laboratories", 4th Edition, published by the Centers for Disease Control (CDC).
- [0700] The diagram in Figure 35 describes the general steps required to express a sequence of interest using an exemplary system of the invention.
- [0701] The present of the invention is designed to help one skilled in the art create a lentivirus to deliver and express a gene of interest in mammalian cells. For more information about retroviral biology and eukaryotic cell culture, refer to the following published reviews:
Buchsacher *et al.* (2000); Luciw (1996); Naldini (1999), Naldini (1998), and Yee (1999) Retroviral Vectors. In *The Development of Human Gene Therapy*, T. Friedmann, ed. (Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press), pp. 21-45.
- [0702] The pLP1, pLP2, pLP/VSVG plasmids are provided in an optimized mixture to facilitate viral packaging of an expression vector (*e.g.*, a pLenti6/V5-based expression vector) following cotransfection into 293FT producer cells. The amount of the packaging mix (195 µg) and Lipofectamine™ 2000 transfection reagent (0.75 ml) supplied in the kit is sufficient to perform 20 cotransfections in 10 cm plates using the

recommended protocol describe herein. To use the ViraPower™ Packaging Mix, resuspend in 195 µl of sterile water to obtain a 1 µg/µl stock.

[0703] A pLenti6/V5 expression vector containing a gene of interest in pLenti6/V5-D-TOPO® or pLenti6/V5-DEST™ can be generated using methods described herein. Once an expression construct has been created, use any method of choice to prepare purified plasmid DNA. Plasmid DNA for transfection into eukaryotic cells must be clean and free from phenol and sodium chloride as contaminants may kill the cells, and salt will interfere with lipid complexing, decreasing transfection efficiency. Suitable methods for isolating plasmid of sufficient purity include the S.N.A.P.™ MidiPrep Kit (Invitrogen Corporation, Carlsbad, CA, Catalog no. K1910-01) and CsCl gradient centrifugation.

[0704] Resuspend the purified expression plasmid (*e.g.*, a pLenti6/V5 expression plasmid) containing a gene of interest in sterile water or TE, pH 8.0 at a concentration ranging from 0.1-3.0 µg/µl. 3 µg of expression plasmid may be used for each transfection.

[0705] A suitable host cell line is the human 293FT cell line available from Invitrogen Corporation, Carlsbad, CA and supplied with the ViraPower™ Lentiviral Expression kits (Naldini *et al.*, 1996). The 293FT cell line, a derivative of the 293F cell line, stably and constitutively expresses the SV40 large T antigen from pCMVSPORT6TA_g.neo and must be maintained in medium containing Geneticin®.

[0706] Before a stably transduced cell line expressing a gene of interest can be created, a lentiviral stock (containing the packaged expression construct) must be created by cotransfecting the optimized packaging plasmid mix and an expression vector (*e.g.*, a pLenti6/V5-based expression vector) into a suitable host cell line (*e.g.*, the 293FT cell line).

[0707] One suitable protocol for generating a lentiviral stock employs the following materials: ViraPower™ Packaging Mix (supplied with the kit; resuspend in 195 µl of sterile water to a concentration of 1 µg/µl); pLenti6/V5 expression vector containing a gene of interest (0.1-3.0 µg/µl in sterile water or TE, pH 8.0); pLenti6/V5-based positive control vector (supplied with the kit; resuspend in sterile water to a concentration of 1 µg/µl); 293FT cells cultured in the appropriate medium (see Example 13); Lipofectamine™ 2000

transfection reagent (supplied with the kit; store at +4°C until use); Opti-MEM® I Reduced Serum Medium (pre-warmed; see below); Fetal bovine serum (FBS); sterile 10 cm tissue culture plates (one each for the lentiviral construct, positive control, and negative control); sterile tissue culture supplies; 15 ml sterile, capped, conical tubes; and cryovials.

[0708] Each pLenti6/V5-based expression vector kit includes a positive control vector for use as an expression control (*e.g.* pLenti6/V5-GW/*lacZ*). It is recommended that the positive control vector be included in a cotransfection experiment to generate a control lentiviral stock that may be used to help optimize expression conditions in a mammalian cell line of interest.

[0709] Any suitable transfection reagent may be used to introduce the plasmids into the producer cell line. One suitable transfection reagent is Lipofectamine™ 2000 reagent (Ciccarone *et al.*, (1999) *Focus* 21, 54-55). This reagent is a proprietary, cationic lipid-based formulation suitable for the transfection of nucleic acids into eukaryotic cells. Using Lipofectamine™ 2000 to transfect 293FT cells offers the following advantages: provides the highest transfection efficiency in 293FT cells; DNA-Lipofectamine™ 2000 complexes can be added directly to cells in culture medium in the presence of serum; and removal of complexes or medium change or addition following transfection are not required, although complexes can be removed after 4-6 hours without loss of activity.

[0710] To facilitate optimal formation of DNA-Lipofectamine™ 2000 complexes, a reduced serum medium (*e.g.*, Opti-MEM® I Reduced Serum Medium available from Invitrogen Corporation, Carlsbad, CA) may be used.

[0711] Lentiviral stocks in 293FT cells produced using the optimized transfection conditions described herein. The amount of lentivirus produced using these recommended conditions (at a titer of 1×10^5 to 1×10^7 transducing units (TU)/ml) is generally sufficient to transduce 1×10^6 to 1×10^8 cells at a multiplicity of infection (MOI) = 1.

Condition	Amount
Tissue culture plate size	10 cm (one per lentiviral construct)
Number of 293FT cells to transfect	5×10^6 cells (see below)

Condition	Amount
Amount of ViraPower™	9 µg (9 µl of 1 µg/µl stock)
Packaging Mix	
Amount of pLenti6/V5 expression plasmid	3 µg
Amount of Lipofectamine™	36 µl
2000	

- [0712] 293FT cells should be plated 24 hours prior to transfection in complete medium, and should be 90-95% confluent on the day of transfection. Make sure that cells are healthy at the time of plating.
- [0713] Follow the procedure below to cotransfect 293FT cells. Remember that the cells may be kept in culture medium during transfection. A positive control and a negative control (no DNA, no Lipofectamine™ 2000) are recommended to help evaluate results.
- [0714] The day before transfection, trypsinize and count the 293FT cells, plating them at 5×10^6 cells per 10 cm plate. Plate cells in 10 ml of normal growth medium containing serum.
- [0715] On the day of transfection, remove the culture medium from the 293FT cells and replace with 5 ml of normal growth medium containing serum (or Opti-MEM® I Medium containing serum). Do not include antibiotics.
- [0716] Prepare DNA-Lipofectamine™ 2000 complexes for each transfection sample by performing the following: Dilute 9 µg of the optimized packaging mix and 3 µg of pLenti6/V5 expression plasmid DNA (12 µg total) in 1.5 ml of Opti-MEM® I Medium without serum. Mix gently. Mix Lipofectamine™ 2000 gently before use, then dilute 36 µl in 1.5 ml of Opti-MEM® I Medium without serum. Mix gently and incubate for 5 minutes at room temperature. After the 5 minute incubation, combine the diluted DNA with the diluted Lipofectamine™ 2000. Mix gently. Incubate for 20 minutes at room temperature to allow the DNA-Lipofectamine™ 2000 complexes to form. The solution may appear cloudy, but this will not impede the transfection.
- [0717] Add the DNA-Lipofectamine™ 2000 complexes dropwise to each plate. Mix gently by rocking the plate back and forth. Incubate the cells overnight at 37°C in a CO₂ incubator.

- [0718] The next day, remove the medium containing the DNA-Lipofectamine™ 2000 complexes and replace with complete culture medium (*i.e.* D-MEM containing 10% FBS, 2 mM L-glutamine, and 1% penicillin/streptomycin). Expression of the VSV G glycoprotein causes 293FT cells to fuse, resulting in the appearance of multinucleated syncytia. This morphological change is normal and does not affect production of the lentivirus.
- [0719] Harvest virus-containing supernatants 48-72 hours posttransfection by removing medium to a 15 ml sterile, capped, conical tube. Minimal differences in viral yield are observed whether supernatants are collected 48 or 72 hours posttransfection. Remember that the supernatant contains infectious virus at this stage. Follow the recommended guidelines for working with BL-2 organisms.
- [0720] Centrifuge at 3000 rpm for 15 minutes at +4°C in a table top clinical centrifuge.
- [0721] Perform A filtration step, if desired. Pipet viral supernatants into cryovials in 1 ml aliquots. Store viral stocks at -80°C.
- [0722] If the lentiviral construct is to be used for *in vivo* applications or if the stock is to be concentrated to obtain a higher titer, filtering the viral supernatant through a sterile, 0.45 µm low protein binding filter after the low-speed centrifugation step is recommended. A suitable filter is the Millex-HV 0.45 µm PVDF filter (Millipore, Catalog no. SLHVR25LS).
- [0723] Place viral stocks at -80°C for long-term storage. Repeated freezing and thawing is not recommended as it may result in loss of viral titer. When stored properly, viral stocks of an appropriate titer should be suitable for use for up to one year. After long-term storage, it is recommended that the titer of the viral be determined before transducing a cell line of interest.
- [0724] It is possible to scale up the cotransfection experiment to produce a larger volume of lentivirus, if desired. For example, the cotransfection experiment may scaled up from a 10 cm plate to a T225 flask and up to 50 ml of viral supernatant may be harvested. To scale up, increase the number of cells plated and the amounts of DNA, Lipofectamine™ 2000, and medium used in proportion to the difference in surface area of the culture vessel.

Before proceeding to transduce the mammalian cell line of interest and express a recombinant protein, it is recommended that the titer of the lentiviral stock be determined. While this procedure is not required for some applications, it is necessary to control the number of integrated copies of the lentivirus or to generate reproducible expression results.

[0725] To determine the titer of a lentiviral stock: prepare 10-fold serial dilutions of the lentiviral stock; transduce the different dilutions of lentivirus into the mammalian cell line of choice in the presence of Polybrene®; select for stably transduced cells using blasticidin; and stain and count the number of blasticidin-resistant colonies in each dilution.

[0726] A number of factors can influence viral titers. One factor is the size of the sequence of interest inserted into the expression vector. Titers will generally decrease as the size of the insert increases. The size of the wild-type HIV-1 genome is approximately 10 kb. Since the size of the elements required for expression from pLenti6/V5 totals approximately 4 kb, the size of the gene of interest should theoretically not exceed 6 kb for efficient packaging.

[0727] Other factors that may influence viral titer are the characteristics of the cell line used for titering, the age of the lentiviral stock, the number of freeze thaw cycles that the stock has undergone, and the storage conditions of the stock. Viral titers may decrease with long-term storage at -80°C. If a lentiviral stock has been stored for 6 months to 1 year, it is recommended that the titer be determined prior to use in an expression experiment. Viral titers can decrease as much as 10% with each freeze/thaw cycle. Lentiviral stocks should be aliquotted and stored at -80°C.

[0728] The titer of a lentiviral stock may be determined using any mammalian cell line of choice. Generally, it is recommended that the same mammalian cell line be used to titer the lentiviral stock will be used to perform expression studies. However, in some instances, a different cell line may be used to titer the lentivirus (*e.g.* if performing expression studies in a non-dividing cell line or a primary cell line). In these cases, suitable cell lines with which to titer the lentivirus are those that: grow as an adherent cell line; are easy to handle; exhibit a doubling time in the range of 18-25 hours; and are non-migratory. An example of a suitable cell is the HT1080 human fibrosarcoma cell line

(ATCC, Catalog no. CCL-121) for titering purposes, but other cell lines including HeLa and NIH3T3 are also suitable.

- [0729] The titer of a lentiviral construct may vary depending on which cell line is chosen. If more than one lentiviral construct are to be used, it is recommended that the titer all of the lentiviral constructs be determined using the same cell line.

The pLenti6/V5 expression construct contains the blasticidin resistance gene (*bsd*) (Kimura *et al.*, (1994) *Biochim. Biophys. ACTA* 1219, 653-659, Izumi, *et al.* (1991) *Exp. Cell Res.* 197, 229-233.) to allow for blasticidin selection of mammalian cells that have stably transduced the lentiviral construct (Takeuchi *et al.*, (1958) *The Journal of Antibiotics, Series A* 11, 1-5; Yamaguchi *et al.*, (1965) *J. Biochem (Tokyo)* 57, 667-677.

- [0730] Since stably transduced cells are selected using blasticidin, the minimum concentration of blasticidin required to kill untransduced cells must be determined (*i.e.* perform a kill curve experiment). Typically, concentrations ranging from 2-10 µg/ml blasticidin are sufficient to kill most untransduced mammalian cell lines. For any given cell line of interest, a range of concentrations should be tested (see protocol below) to ensure that the minimum concentration necessary for the cell line is used. A suitable method to determine the appropriate concentration of blasticidin for a given cell line follows.

- [0731] Prepare a set of 6 plates. Plate cells at approximately 25% confluence. Allow cells to adhere overnight.

- [0732] The next day, substitute culture medium with medium containing varying concentrations of blasticidin (*e.g.*, 0, 2, 4, 6, 8, 10 µg/ml blasticidin).

- [0733] Replenish the selective media every 3-4 days, and observe the percentage of surviving cells.

- [0734] Determine the appropriate concentration of blasticidin that kills the cells within 10 days after addition of blasticidin.

To determine the titer of a lentiviral construct, the following materials will be needed: the lentiviral stock (store at -80°C until use); an adherent mammalian cell line of choice; complete culture medium for the cell line; hexadimethrine bromide (Polybrene®; Sigma, Catalog no. H9268; 6-well tissue culture plates; blasticidin (10 mg/ml stock solution); crystal violet (Sigma, Catalog no.

C3886; prepare a 1% crystal violet solution in 10% ethanol); and Phosphate-Buffered Saline (PBS; Invitrogen, Catalog no. 10010-023).

[0735] When adding virus to mammalian cells, Polybrene[®] is included to enhance transduction of the virus into the cell. To use Polybrene[®]: prepare a 6 mg/ml stock solution in deionized, sterile water; filter-sterilize and dispense 1 ml aliquots into sterile microcentrifuge tubes; store at -20°C for long-term storage. Stock solutions may be stored at -20°C for up to 1 year. Do not freeze/thaw the stock solution more than 3 times as this may result in loss of activity. The working stock may be stored at +4°C for up to 2 weeks.

[0736] The media contains infectious virus and appropriate safety precautions should be taken. For example, perform all manipulations within a certified biosafety cabinet. Treat media containing virus with bleach. Treat used pipets, pipette tips, and other tissue culture supplies with bleach or dispose of as biohazardous waste. Wear gloves, a laboratory coat, and safety glasses or goggles when handling viral stocks and media containing virus.

[0737] Follow the procedure below to determine the titer of a lentiviral stock using the mammalian cell line of choice. At least one 6-well plate is used for every lentiviral stock to be titered (one mock well plus five dilutions). If a lentiviral stock of the pLenti6/V5-GW/*lacZ* positive expression control has been made, it is recommended that this stock be titered as well.

[0738] The day before transduction (Day 1), trypsinize and count the cells, plating them such that they will be 30-50% confluent at the time of transduction. Incubate cells at 37°C overnight.

[0739] **Example:** When using HT1080 cells, generally plate 2×10^5 cells per well in a 6-well plate.

[0740] On the day of transduction (Day 2), thaw the lentiviral stock and prepare 10-fold serial dilutions ranging from 10^{-2} to 10^{-6} . For each dilution, dilute the lentiviral construct into complete culture medium to a final volume of 1 ml. Do not vortex. A wider range of serial dilutions (10^{-2} to 10^{-8}) may be used, if desired.

[0741] Remove the culture medium from the cells. Mix each dilution gently by inversion and add to one well of cells (total volume = 1 ml).

[0742] Add Polybrene[®] to each well to a final concentration of 6 µg/ml. Swirl the plate gently to mix. Incubate at 37°C overnight.

- [0743] The following day (Day 3), remove the media containing virus and replace with 2 ml of complete culture medium.
- [0744] The following day (Day 4), remove the medium and replace with complete culture medium containing the appropriate amount of blasticidin to select for stably transduced cells.
- [0745] Remove medium and replace with fresh medium containing blasticidin every 3-4 days.
- [0746] After 10-12 days of selection (day 14-16), no live cells in the mock well and discrete blasticidin-resistant colonies in one or more of the dilution wells should be seen. Remove the medium and wash the cells with 2 ml of PBS. Repeat the wash.
- [0747] Add 1 ml of crystal violet solution and incubate for 10 minutes at room temperature.
- [0748] Remove the crystal violet stain and wash the cells with 2 ml of PBS. Repeat wash.
- [0749] Count the blue-stained colonies and determine the titer of the lentiviral stock.
- [0750] When titering Lenti6/V5 lentiviral stocks using HT1080 cells, generally titers ranging from 5×10^5 to 2×10^7 transducing units (TU)/ml are observed. If the titer of a lentiviral stock is less than 1×10^5 TU/ml, a new lentiviral stock should be produced.
- [0751] As an example, a Lenti6/V5-GW/*lacZ* lentiviral stock was generated using the protocols described herein. HT1080 cells were transduced with 10-fold serial dilutions of the lentiviral supernatant (10^{-2} to 10^{-6} dilutions) or untransduced (mock) following the protocol described above. Forty-eight hours post-transduction, the cells were placed under blasticidin selection ($10 \mu\text{g/ml}$). After 10 days of selection, the cells were stained with crystal violet (see plate below), and colonies were counted. In the plate, the colony counts were: mock: no colonies; 10^{-2} dilution: confluent; undeterminable, 10^{-3} dilution: confluent; undeterminable, 10^{-4} dilution: confluent; undeterminable, 10^{-5} dilution: 46, and 10^{-6} dilution: 5. Thus, the titer of this lentiviral stock is 4.8×10^6 TU/ml (*i.e.* average of 46×10^5 and 5×10^6).
- [0752] Once a lentiviral stock with a suitable titer has been generated, the lentiviral construct may be transduced into the mammalian cell line of choice

and assayed for expression of a recombinant protein. An assay for expression of a gene of interest may be conducted in the following ways:

1) Pool a heterogeneous population of cells and test for expression directly after transduction (*i.e.* “transient” expression). Note that 24-48 hours must elapse after transduction before harvesting cells to allow time for the lentivirus genome to reverse transcribe and integrate into the chromosomal DNA. Integration must take place before expression of the gene of interest can occur.

2) Select for stably transduced cells using blasticidin. This requires a minimum of 10-12 days after transduction, but allows generation of clonal cell lines that stably express the gene of interest.

[0753] Stable expression of a target gene typically may be observed for at least 6 weeks following transduction and selection.

[0754] To select for stably transduced cells, the minimum concentration of blasticidin required to kill the untransduced mammalian cell line must be determined as described above. If the titer of the lentiviral construct was determined in the same cell line used to perform stable expression experiment, then the same concentration of blasticidin may be used for selection as was used for titering.

[0755] To obtain optimal expression of a gene of interest, cells must be transduced with a suitable MOI of lentivirus. MOI is defined as the number of virus particles per cell and generally correlates with the number of integration events and as a result, expression. Typically, expression levels increase linearly as the MOI increases.

[0756] A number of factors can influence determination of an optimal MOI including the nature of the cell line (*e.g.* non-dividing vs. dividing cell type), its transduction efficiency, the application of interest, and the nature of the gene of interest. If transducing a lentiviral construct into a mammalian cell line of choice for the first time, a range of MOIs should be used (*e.g.* 0, 0.05, 0.1, 0.5, 1, 2, 5) to determine the MOI required to obtain optimal expression of the recombinant protein for a particular application.

[0757] In general, 80-90% of the cells in an actively dividing cell line (*e.g.* HT1080, HeLa, CHO-K1) express a target gene when transduced at an MOI of ~1. Some non-dividing cell types transduce lentiviral constructs less

efficiently. For example, only about 50% of the cells in a culture of primary human fibroblasts express a target gene when transduced at an MOI of ~1. If transducing a lentiviral construct into a non-dividing cell type, it may be necessary to increase the MOI to achieve optimal expression levels for a recombinant protein.

[0758] If a Lenti6/V5-GW/*lacZ* control lentiviral construct has been constructed, it may be used to help determine the optimal MOI for a particular cell line and application. Once the Lenti6/V5-GW/*lacZ* lentivirus has been transduced into the mammalian cell line of choice, the gene encoding β -galactosidase will be constitutively expressed and can be easily assayed using standard techniques.

[0759] Remember that viral supernatants are generated by harvesting spent media containing virus from the 293FT producer cells. Spent media lacks nutrients and may contain some toxic waste products. If a large volume of viral supernatant is used to transduce a mammalian cell line (*e.g.* 1 ml of viral supernatant per well in a 6-well plate), note that growth characteristics or morphology of the cells may be affected during transduction. These effects are generally alleviated after transduction when the media is replaced with fresh, complete media.

[0760] It is possible to concentrate VSV-G pseudotyped retroviruses using a variety of methods without significantly affecting their transducibility. If the titer of a lentiviral stock is relatively low (less than 5×10^5 TU/ml) and an experiment requires a large volume of viral supernatant (*e.g.* a relatively high MOI), the virus may be concentrated before proceeding to transduction. For details and guidelines to concentrate the virus, refer to published reference sources (Yee, 1999).

[0761] To transduce a selected cell line, the following materials will be required: a titered stock of virus (*e.g.*, a Lenti6/V5 lentiviral stock) which should be stored at -80°C until use; a cell line of choice (*e.g.*, a mammalian cell line); complete culture medium for the cell line; hexadimethrine bromide (Polybrene[®]; 6 mg/ml stock solution); appropriately sized tissue culture plates for the intended application; and blasticidin (if selecting for stably transduced cells; 10 mg/ml stock solution).

- [0762] Follow the procedure below to transduce the mammalian cell line of choice with a lentiviral construct.
- [0763] Plate cells in complete media as appropriate for the intended application.
- [0764] On the day of transduction (Day 1), thaw the lentiviral stock and dilute (if necessary) the appropriate amount of virus (at a suitable MOI) into fresh complete medium. Do not vortex.
- [0765] Remove the culture medium from the cells. Mix the medium containing virus gently by pipetting and add to the cells.
- [0766] Add Polybrene[®] to a final concentration of 6 µg/ml. Swirl the plate gently to mix. Incubate at 37°C overnight. To reduce possible negative effects of transducing cells with undiluted viral stock, it is possible to incubate cells for as little as 6 hours prior to changing medium.
- [0767] The following day (Day 2), remove the medium containing virus and replace with fresh, complete culture medium.
- [0768] The following day (Day 3), perform one of the following: harvest the cells and assay for expression of the recombinant protein of interest if performing transient expression experiments; or remove the medium and replace with fresh, complete medium containing the appropriate amount of blasticidin to select for stably transduced cells.
- [0769] Remove medium and replace with fresh medium containing blasticidin every 3-4 days until blasticidin-resistant colonies can be identified (generally 10-12 days after selection).
- [0770] Pick at least 5 blasticidin-resistant colonies and expand each clone to assay for expression of the recombinant protein.
- [0771] Note that integration of the lentivirus into the genome is random. Depending upon the influence of the surrounding genomic sequences at the integration site, varying levels of recombinant protein expression may be seen from different blasticidin-resistant clones. Testing at least 5 blasticidin-resistant clones and selecting the clone that provides the optimal expression of the recombinant protein of interest is recommended.
- [0772] Any method of choice known to those skilled in the art may be used to detect a recombinant protein of interest including, but not limited to, functional analysis, immunofluorescence, or western blot. If the gene of

interest is cloned in frame with an epitope tag, the recombinant protein may be detected in a western blot using an antibody to the epitope tag.

[0773] Below are listed some potential problems and possible solutions that may help troubleshoot cotransfection and titering experiments.

Problem	Reason	Solution
Low viral titer	Low transfection efficiency:	<ul style="list-style-type: none"> • Use the S.N.A.P.[™] MidiPrep Kit to prepare plasmid DNA. • Use healthy 293FT cells; do not overgrow. • Cells should be 90-95% confluent at the time of transfection. • Optimize such that plasmid DNA (in μg):Lipofectamine[™] 2000 (in μl) ratio ranges from 1:2 to 1:3.
	• Poor quality of pLenti6/V5 plasmid DNA	
	• Unhealthy 293FT cells; cells exhibit low viability	
	• 293FT cells plated too sparsely	
	• Plasmid DNA:transfection reagent ratio incorrect	
	Viral supernatant too dilute	Concentrate virus using any method of choice (Yee, 1999).
	Viral supernatant frozen and thawed multiple times	DO NOT freeze/thaw viral supernatant more than 3 times.
	Poor choice of titering cell line	Use an adherent cell line with the characteristics discussed herein.
	Gene of interest is large	Viral titers generally decrease as the size of the insert increases; inserts larger than 6 kb are not recommended.
	Gene of interest is toxic to cells	Generation of constructs containing activated oncogenes or potentially harmful genes is not recommended.
	Polybrene [®] not included during transduction	Transduce the lentiviral construct into cells in the presence of Polybrene [®] .

Problem	Reason	Solution
No colonies obtained upon titering	Too much blasticidin used for selection	Determine the sensitivity of the cell line to blasticidin by performing a kill curve experiment. Use the minimum concentration of blasticidin required to kill the untransduced cell line.
	Viral stocks stored incorrectly	Aliquot and store stocks at -80°C. Do not freeze/thaw more than 3 times.
	Polybrene® not included during transduction	Transduce the lentiviral construct into cells in the presence of Polybrene®.
Titer indeterminable; cells confluent	Too little blasticidin used for selection	Increase amount of blasticidin used for selection.
	Viral supernatant not diluted sufficiently	Titer lentivirus using a wider range of 10-fold serial dilutions (e.g. 10 ⁻² to 10 ⁻⁸).

[0774] Below are listed some potential problems and possible solutions that may help troubleshoot transduction and expression experiment.

Problem	Reason	Solution
No expression	Promoter silencing	The lentiviral construct may integrate into a chromosomal region that silences the CMV promoter controlling expression of the gene of interest. Screen several blasticidin-resistant clones and select the one that demonstrates the highest expression levels of the recombinant protein.
Poor expression	Viral stocks stored incorrectly	Aliquot and store stocks at -80°C. Do not freeze/thaw more than 3 times.
	Poor transduction efficiency: <ul style="list-style-type: none"> • Polybrene® not included during transduction • Non-dividing cell type used 	<ul style="list-style-type: none"> • Transduce the lentiviral construct into cells in the presence of Polybrene®. • Transduce the lentiviral construct into cells using a higher MOI.
	MOI too low	Transduce the lentiviral construct into cells using a higher MOI.

Problem	Reason	Solution
	Too much blasticidin used for selection	Determine the sensitivity of the cell line to blasticidin by performing a kill curve experiment. Use the minimum concentration of blasticidin required to kill the untransduced cell line.
	Cells harvested too soon after transduction	Do not harvest cells until at least 24-48 hours after transduction to allow reverse transcription and integration of the lentivirus into the genome.
	Gene of interest is toxic to cells	Generation of constructs containing activated oncogenes or potentially harmful genes is not recommended.

[0775] Table 25 provides some of the characteristics of the vector pLP1. The complete sequence is provided as table 21. A plasmid map is provided as Figure 37A.

Table 25.

Feature	Benefit
Human cytomegalovirus (CMV) promoter bases 1-747, TATA box bases 648-651	Permits high-level expression of the HIV-1 <i>gag</i> and <i>pol</i> genes in mammalian cells (Andersson <i>et al.</i> , 1989; Boshart <i>et al.</i> , 1985; Nelson <i>et al.</i> , 1987).
Human β -globin intron bases 880-1320	Enhances expression of the <i>gag</i> and <i>pol</i> genes in mammalian cells.
HIV-1 <i>gag</i> coding sequence bases 1355-2857	Encodes the viral core proteins required for forming the structure of the lentivirus (Luciw, 1996).
HIV-1 <i>pol</i> coding sequence bases 2650-5661	Encodes the viral replication enzymes required for replication and integration of the lentivirus (Luciw, 1996).
HIV-1 Rev response element (RRE) bases 5686-5919	Permits Rev-dependent expression of the <i>gag</i> and <i>pol</i> genes
Human β -globin polyadenylation signal bases 6072-6837	Allows efficient transcription termination and polyadenylation of mRNA.
pUC origin of replication (<i>ori</i>) bases 6995-7668 complementary strand	Permits high-copy replication and maintenance in <i>E. coli</i> .

Feature	Benefit
Ampicillin (<i>bla</i>) resistance gene bases 7813-8673 complementary strand <i>bla</i> promoter bases 8674-8772 complementary strand	Allows selection of the plasmid in <i>E. coli</i> .

[0776] Table 26 provides some of the characteristics of the vector pLP2. The complete sequence is provided as Table 22. A plasmid map is provided as Figure 37B.

Table 26.

Feature	Benefit
RSV enhancer/promoter bases 1-271, TATA box bases 200-207, transcription initiation base 229 RSV UTR bases 230-271	Permits high-level expression of the rev gene (Gorman et al., 1982).
HIV-1 Rev ORF bases 391-741	Encodes the Rev protein which interacts with the RRE on pLP1 to induce Gag and Pol expression, and on the pLenti6/V5 expression vector to promote the nuclear export of the unspliced viral RNA for packaging into viral particles.
HIV-1 LTR polyadenylation signal bases 850-971	Allows efficient transcription termination and polyadenylation of mRNA.
Ampicillin (<i>bla</i>) resistance gene promoter bases 1916-2014 gene bases 2015-2875	Allows selection of the plasmid in <i>E. coli</i> .
pUC origin of replication (ori) bases 3020-3693	Permits high-copy replication and maintenance in <i>E. coli</i> .

[0777] Table 27 provides some of the characteristics of the vector pLP/VSVG. The complete sequence is provided as Table 23. A plasmid map is provided as Figure 37C.

[0778] Table 27.

Feature	Benefit
Human CMV promoter bases 1-747	Permits high-level expression of the VSV-G gene in mammalian cells (Andersson <i>et al.</i> , 1989; Boshart <i>et al.</i> , 1985; Nelson <i>et al.</i> , 1987).
Human β -globin intron bases 880-1320	Enhances expression of the VSV-G gene in mammalian cells.
VSV G glycoprotein (VSV-G) bases 1346-2881	Encodes the envelope G glycoprotein from Vesicular Stomatitis Virus to allow production of a pseudotyped retrovirus with a broad host range (Burns <i>et al.</i> , 1993; Emi <i>et al.</i> , 1991; Yee <i>et al.</i> , 1994).
Human β -globin polyadenylation signal bases 3004-3769	Allows efficient transcription termination and polyadenylation of mRNA.
pUC origin of replication (<i>ori</i>) bases 3927-4600 complementary strand	Permits high-copy replication and maintenance in <i>E. coli</i> .
Ampicillin (<i>bla</i>) resistance gene gene bases 4745-5606 complementary strand promoter bases 5606-5704 complementary strand	Allows selection of the plasmid in <i>E. coli</i> .

EXAMPLE 11

GATEWAY™-adapted destination vector for cloning and high-level expression
in mammalian cells using the ViraPower™ Lentiviral Expression System

ViraPower™ Lentiviral Expression Products

[0779] The pLenti6/V5-DEST™, pLenti4/V5-DEST, and pLenti6/UbC/V5-DEST vectors are designed for use with the ViraPower™ Lentiviral Expression System available from Invitrogen Corporation, Carlsbad, CA, which is discussed in some detail above. Depending on the vector chosen, the pLenti-DEST vectors are available with the human cytomegalovirus (CMV) immediate early promoter or the human ubiquitin C (UbC) promoter to control expression of the gene of interest, and the Zeocin™ resistance gene or the blasticidin resistance gene for selection in *E. coli* or mammalian cells.

- [0780] Expression of a recombinant fusion protein can be detected using an antibody to the V5 epitope. Horseradish peroxidase (HRP) or alkaline phosphatase (AP)-conjugated antibodies allow one-step detection using chemiluminescent or colorimetric detection methods. A fluorescein isothiocyanate (FITC)-conjugated antibody allows one-step detection in immunofluorescence experiments. Suitable detection reagents for fusion proteins can be obtained from Invitrogen Corporation, Carlsbad, CA, for example, Anti-V5 Antibody, catalog number R960-25, Anti-V5-HRP Antibody, catalog number R961-25, Anti-V5-AP Antibody, catalog number R962-25, Anti-V5-FITC Antibody, catalog number R963-25.
- [0781] pLenti6/V5-DEST[™] is an 8.7 kb vector adapted for use with the GATEWAY[™] Technology, and is designed to allow high-level expression of recombinant fusion proteins in dividing and non-dividing mammalian cells using Invitrogen's ViraPower[™] Lentiviral Expression System. A map of the vector is provided as Figure 36A and the sequence of the vector is provided as Table 17.
- [0782] The pLenti-DEST vectors contain the following features: Rous Sarcoma Virus (RSV) enhancer/promoter for Tat-independent production of viral mRNA in the producer cell line (Dull *et al.*, 1998); modified HIV-1 5' and 3' Long Terminal Repeats (LTR) for viral packaging and reverse transcription of the viral mRNA (Dull *et al.*, 1998; Luciw, 1996) (Note: The U3 region of the 3' LTR is deleted (Δ U3) and facilitates self-inactivation of the 5' LTR after transduction to enhance the biosafety of the vector (Dull *et al.*, 1998)); HIV-1 psi (Ψ) packaging sequence for viral packaging (Luciw, 1996); HIV Rev response element (RRE) for Rev-dependent nuclear export of unspliced viral mRNA (Kjems *et al.*, 1991, *Proc. Natl. Acad. Sci. USA* 88, 683-687; Malim *et al.*, 1989, *Nature* 338, 254-257); human CMV or UbC promoter for constitutive expression of the gene of interest from a viral or cellular promoter, respectively; two recombination sites, *attR1* and *attR2*, downstream of the CMV or UbC promoter for recombinational cloning of the gene of interest from an entry clone; chloramphenicol resistance gene (Cm^{R}) located between the two *attR* sites for counterselection; the *ccdB* gene located between the *attR* sites for negative selection; C-terminal V5 epitope for detection of the recombinant protein of interest (Southern *et al.*, 1991, *J. Gen.*

Virol. 72, 1551-1557); blasticidin (Izumi *et al.*, 1991; Kimura *et al.*, 1994; Takeuchi *et al.*, 1958; Yamaguchi *et al.*, 1965) or Zeocin[™] (Drocourt *et al.*, 1990, *Nucleic Acids Res.* 18, 4009; Mulsant *et al.*, 1988, *Somat. Cell Mol. Genet.* 14, 243-252) resistance gene for selection in *E. coli* and mammalian cells; ampicillin resistance gene for selection in *E. coli*; and the pUC origin for high-copy replication of the plasmid in *E. coli*.

[0783] A control plasmid containing the *lacZ* gene is included with each pLenti-DEST vector for use as a positive expression control in the mammalian cell line of choice.

[0784] The pLenti4/V5-DEST and pLenti6/V5-DEST vectors use the human CMV immediate early promoter to allow high-level, constitutive expression of the gene of interest in mammalian cells (Andersson *et al.*, 1989; Boshart *et al.*, 1985; Nelson *et al.*, 1987). The sequence of the pLenti4/V5-DEST plasmid is provided as Table 19. Although highly active in most mammalian cell lines, activity of the viral CMV promoter can be down-regulated in some cell lines due to methylation (Curradi *et al.*, 2002, *Mol. Cell. Biol.* 22, 3157-3173), histone deacetylation (Rietveld *et al.*, 2002, *EMBO J.* 21, 1389-1397), or both.

[0785] The pLenti6/UbC/V5-DEST vector uses the human UbC promoter to allow constitutive, but more physiological levels of expression from the gene of interest in mammalian cells (Marinovic *et al.*, 2000, *Biophys. Res. Comm.* 274, 537-541). The sequence of the pLenti6/UbC/V5-DEST plasmid is provided as Table 20. When compared to the CMV promoter, the UbC promoter is generally 2-4 fold less active. The UbC promoter is not down-regulated, making it useful for transgenic studies (Gill *et al.*, 2001, *Gene Ther.* 8, 1539-1546; Lois *et al.*, 2002, *Science* 295, 868-872; Marinovic *et al.*, 2000; Schorpp *et al.*, 1996, *Nuc. Acids Res.* 24, 1787-1788; Yew *et al.*, 2001, *Mol. Ther.* 4, 75-82). The human ubiquitin C (UbC) promoter (in pLenti6/UbC/V5-DEST) allows high-level expression of recombinant protein in most mammalian cell lines (Wulff *et al.*, 1990, *FEBS Lett.* 261, 101-105) and in virtually all tissues tested in transgenic mice (Schorpp *et al.*, 1996). The diagram below shows the features of the UbC promoter as described by Neno *et al.*, 1996 *Gene* 175, 179-185.

[0786] GATEWAY[™] is a universal cloning technology that takes advantage of the site-specific recombination properties of bacteriophage lambda (Landy,

1989) to provide a rapid and highly efficient way to move a gene of interest into multiple vector systems. To express a sequence of interest (*e.g.*, a sequence encoding a polypeptide of interest) in mammalian cells using the GATEWAY™ technology, simply: clone the sequence of interest into a GATEWAY™ entry vector of choice to create an entry clone; generate an expression clone by performing an LR recombination reaction between the entry clone and a GATEWAY™ destination vector (*e.g.* pLenti4/V5-DEST, pLenti6/V5-DEST, or pLenti6/UbC/V5-DEST); and use the expression clone in the ViraPower™ Lentiviral Expression System.

[0787] For more detailed information about GATEWAY™ System, generating an entry clone, and performing the LR recombination reaction, refer to the GATEWAY™ Technology manual available from Invitrogen Corporation, Carlsbad, CA.

[0788] The pLenti4/V5-DEST, pLenti6/V5-DEST, and pLenti6/UbC/V5-DEST vectors are supplied as supercoiled plasmids. Although the GATEWAY™ Technology Manual has previously recommended using a linearized destination vector for more efficient recombination, further testing at Invitrogen has found that linearization of pLenti6/V5-DEST™ is not required to obtain optimal results for any downstream application.

[0789] To propagate and maintain the pLenti4/V5-DEST, pLenti6/V5-DEST, or pLenti6/UbC/V5-DEST vectors, Library Efficiency® DB3.1™ Competent Cells (Catalog no. 11782-018) from Invitrogen Corporation, Carlsbad, CA are recommended for transformation. The DB3.1™ *E. coli* strain is resistant to CcdB effects and can support the propagation of plasmids containing the *ccdB* gene. To maintain integrity of the vector, select for transformants in media containing 50-100 µg/ml ampicillin and 15 µg/ml chloramphenicol. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the destination vector containing attP sites flanking the *ccdB* and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. It has recently been found that the use of spectinomycin selection instead of chloramphenicol selection results in an increase in the number of colonies obtained on selection plates,

indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene. Do not use general *E. coli* cloning strains including TOP10 or DH5 α for propagation and maintenance as these strains are sensitive to CcdB effects.

- [0790] To recombine a sequence of interest into pLenti4/V5-DEST, pLenti6/V5-DEST, or pLenti6/UbC/V5-DEST, an entry clone containing the sequence must be created. Many entry vectors including pENTR/D-TOPO[®] are available from Invitrogen Corporation, Carlsbad, CA to facilitate generation of entry clones.
- [0791] pLenti4/V5-DEST, pLenti6/V5-DEST, and pLenti6/UbC/V5-DEST are C-terminal fusion vectors. To express a fusion polypeptide of a polypeptide encoded by a sequence of interest with the V5 epitope coding sequence present in the vector, a sequence of interest must contain an ATG initiation codon in the context of a Kozak translation initiation sequence for proper initiation of translation in mammalian cells (Kozak, 1987; Kozak, 1991; Kozak, 1990). An example of a Kozak consensus sequence is (G/A)NN**ATGG**. Other sequences are possible, but the G or A at position -3 and the G at position +4 are the most critical for function (shown in bold). The ATG initiation codon is underlined. The reading frame of the polypeptide encoded by the sequence of interest must be in frame with the C-terminal tag containing the V5 epitope after recombination and the sequence of interest must not contain a stop codon in this reading frame. The C-terminal peptide containing the V5 epitope and the attB2 site will add approximately 4.5 kDa to the size of the polypeptide encoded by the sequence of interest.
- [0792] Each entry clone contains *attL* sites flanking the gene of interest. Genes in an entry clone are transferred to the destination vector backbone by mixing the DNAs with the GATEWAY[™] LR Clonase[™] Enzyme Mix available from Invitrogen Corporation, Carlsbad, CA. The resulting recombination reaction is then transformed into *E. coli* (e.g. TOP10 or DH5 α [™]-T1^R) and the expression clone selected (e.g., using ampicillin and blasticidin). Recombination between the *attR* sites on the destination vector and the *attL* sites on the entry clone replaces the chloramphenicol (Cm^R) gene and the *ccdB*

gene with the gene of interest and results in the formation of *attB* sites in the expression clone.

[0793] Any *recA*, *endA* *E. coli* strain including TOP10, DH5 α [™], or equivalent may be used for transformation. **Do not** transform the LR reaction mixture into *E. coli* strains that contain the F' episome (*e.g.* TOP10F'). These strains contain the *ccdA* gene and will prevent negative selection with the *ccdB* gene.

[0794] When transforming *E. coli* with the recombination reaction (pLenti4/V5-DEST, pLenti6/V5-DEST, or pLenti6/UbC/V5-DEST x entry clone), unwanted recombination (less than 5%) between the 5' and 3' LTRs has been observed when transformants are selected on LB agar plates containing ampicillin. These events occur less frequently when selection is performed using 100 μ g/ml ampicillin and an additional selection, for example, 50 μ g/ml blasticidin for pLenti6/V5-DEST or pLenti6/UbC/V5-DEST or 25 μ g/ml Zeocin[™] for pLenti4/V5-DEST. For Zeocin[™] to be active, the salt concentration of the bacterial medium must be <90 mM and the pH must be 7.5. Therefore, selection on LB agar plates containing 50-100 μ g/ml ampicillin and an additional selection agent is recommended. Note that transformed *E. coli* grow more slowly in LB media containing ampicillin and blasticidin, and may require slightly longer incubation times to obtain visible colonies. Transformants that contain a recombined plasmid generally give rise to larger colonies than those containing an intact plasmid.

[0795] The *ccdB* gene mutates at a very low frequency, resulting in a very low number of false positives. True expression clones will be chloramphenicol-sensitive and ampicillin- and blasticidin-resistant (for pLenti6 vectors) and ampicillin- and Zeocin[™]-resistant (for pLenti4/V5-DEST). Transformants containing a plasmid with a mutated *ccdB* gene will be ampicillin-, blasticidin- or Zeocin[™]-, and chloramphenicol-resistant, as appropriate. To check a putative expression clone, test for growth on LB plates containing 30 μ g/ml chloramphenicol. A true expression clone should not grow in the presence of chloramphenicol.

[0796] Figure 46A provides a diagram of the recombination region of pLenti6/V5-DEST[™] or pLenti4/V5-DEST after a recombination reaction with a sequence of interest. Shaded regions correspond to the sequence of interest transferred from the entry clone into the pLenti6/V5-DEST[™] vector by

recombination. Non-shaded regions are derived from the pLenti6/V5-DEST™ or pLenti4/V5-DEST vector. Bases 2448 and 4130 of the pLenti4/V5-DEST and pLenti6/V5-DEST™ sequences are marked. Restriction sites are labeled to indicate the actual cleavage site.

[0797] Figure 46B shows the recombination region of the expression clone resulting from pLenti6/UbC/V5-DEST x entry clone. Note that this diagram does not contain the complete sequence of the UbC promoter. For a diagram of the UbC promoter see Figure 46C. Shaded regions in Figure 46B correspond to those DNA sequences transferred from the entry clone into the pLenti6/UbC/V5-DEST vector by recombination. Non-shaded regions are derived from the pLenti6/UbC/V5-DEST vector. Bases 3079 and 4762 of the pLenti6/UbC/V5-DEST sequence are marked.

[0798] Once an expression clone has been generated in the pLenti6/V5-DEST backbone, maintain and propagate the plasmid in LB medium containing 50-100 µg/ml ampicillin. Addition of blasticidin is not required.

[0799] To confirm that a gene of interest is in frame with the C-terminal tag, sequence the expression construct, if desired. Refer to Figure 46 for the location of the recommended primer binding sites (CMV or UbC forward priming site and V5(C-term) reverse priming site) to use to sequence the expression construct. To sequence a pLenti4/V5-DEST or pLenti6/V5-DEST construct, the CMV forward primer 5'-CGCAAATGGGCGGTAGGCGTG-3' and V5(C-term) reverse primer 5'-ACCGAGGAGAGGGTTAGGGAT-3' can be used. To sequence a pLenti6/UbC/V5-DEST construct, the UB forward primer 5'-TCAGTGTTAGACTAGTAAATTG-3' and the V5(C-term) reverse primer 5'-ACCGAGGAGAGGGTTAGGGAT-3' can be used.

[0800] Once purified plasmid DNA of the expression construct has been obtained, a viral stock can be prepared and used to transduce a cell line of choice as described above. Host cells containing the expression clone can be propagated in LB medium with ampicillin. It is not necessary to add an additional selection agent.

[0801] High salt and acidity or basicity inactivate Zeocin™. Therefore, it is recommended that the salt in bacterial medium be reduced and the pH adjusted

to 7.5 to keep the drug active. Note that the pH and salt concentration do not need to be adjusted when preparing tissue culture medium containing Zeocin™. Store Zeocin™ at -20°C and thaw on ice before use. Zeocin™ is light sensitive. Store the drug, and plates or medium containing drug, in the dark at +4°C. Culture medium containing Zeocin™ may be stored at +4°C protected from exposure to light for up to 1 month. Wear gloves, a laboratory coat, and safety glasses or goggles when handling Zeocin™-containing solutions. Zeocin™ is toxic. Do not ingest or inhale solutions containing the drug.

[0802] The pLenti6/V5-DEST™ vector (8688 bp) contains the following features at the indicated locations. The locations of the features in the pLenti6/V5-DEST plasmid are as follows: RSV/5' LTR hybrid promoter bases 1-410; RSV promoter bases 1-229; HIV-1 5' LTR bases 230-410; 5' splice donor base 520; HIV-1 psi (ψ) packaging signal bases 521-565; HIV-1 Rev response element (RRE) bases 1075-1308; 3' splice acceptor base 1656; 3' splice acceptor base 1684; CMV promoter bases 1809-2392; *attR1* site: bases 2440-2564; Chloramphenicol resistance gene (*Cm^R*) bases 2673-3332; *ccdB* gene bases 3674-3979; *attR2* site bases 4020-4144; V5 epitope bases 4197-4238; SV40 early promoter and origin bases 4293-4602; EM7 promoter bases 4657-4723; Blasticidin resistance gene bases 4724-5122; ΔU3/3' LTR bases 5208-5442; ΔU3 bases 5208-5261; 3' LTR: bases 5262-5442; SV40 polyadenylation signal bases 5514-5645; *bla* promoter bases 6504-6602; Ampicillin (*bla*) resistance gene bases 6603-7463; and pUC origin bases 7608-8281.

[0803] The pLenti4/V5-DEST vector(8634 nucleotides) contains the following features at the indicated locations: RSV/5' LTR hybrid promoter bases 1-410; RSV promoter bases 1-229; HIV-1 5' LTR bases 230-410; 5' splice donor base 520; HIV-1 psi (ψ) packaging signal bases 521-565; HIV-1 Rev response element (RRE) bases 1075-1308; 3' splice acceptor base 1656; 3' splice acceptor base 1684; CMV promoter bases 1809-2392; *attR1* site bases 2440-2564; Chloramphenicol resistance gene (*Cm^R*) bases 2673-3332; *ccdB* gene bases 3674-3979; *attR2* site bases 4020-4144; V5 epitope bases 4197-4238; SV40 early promoter and origin bases 4293-4602; EM7 promoter bases 4621-4687; Zeocin™ resistance gene bases 4688-5062; ΔU3/3' LTR bases

5154-5388; Δ U3 bases 5154-5207; 3' LTR bases 5208-5388; SV40 polyadenylation signal bases 5460-5591; *bla* promoter bases 6450-6548; Ampicillin (*bla*) resistance gene bases 6549-7409; and the pUC origin bases 7554-8227.

[0804] The pLenti6/UbC/V5-DEST vector (9320 nucleotides) contains the following features at the indicated locations: RSV/5' LTR hybrid promoter bases 1-410; RSV promoter bases 1-229; HIV-1 5' LTR bases 230-410; 5' splice donor base 520; HIV-1 psi (ψ) packaging signal bases 521-565; HIV-1 Rev response element (RRE) bases 1075-1308; 3' splice acceptor base 1656; 3' splice acceptor base 1684; UbC promoter bases 1798-3016; *attR1* site bases 3072-3196; Chloramphenicol resistance gene (Cm^R) bases 3305-3964; *ccdB* gene bases 4306-4611; *attR2* site bases 4652-4776; V5 epitope bases 4829-4870; SV40 early promoter and origin bases 4925-5234; EM7 promoter bases 5289-5355; Blasticidin resistance gene bases 5356-5754; Δ U3/3' LTR bases 5840-6074; Δ U3 bases 5840-5893; 3' LTR bases 5894-6074; SV40 polyadenylation signal bases 6146-6277; *bla* promoter bases 7136-7234; Ampicillin (*bla*) resistance gene bases 7235-8095; and the pUC origin bases 8240-8913.

EXAMPLE 12

Five-minute, directional TOPO® Cloning of blunt-end PCR products into an expression vector for the ViraPower™ Lentiviral Expression System

[0805] The following protocol may be used to clone a nucleic acid segment using topoisomerase. Other protocols known to those skilled in the art are also suitable. An example of another suitable protocol may be found in the pENTR Directional TOPO® Cloning Kit manual available from Invitrogen Corporation, Carlsbad, CA (catalog number 25-0434).

Step	Action
Design PCR Primers	<p>Include the 4 base pair sequences (CACC) necessary for directional cloning on the 5' end of the forward primer.</p> <p>Design the primers such that a gene of interest will be optimally expressed and fused in frame with the V5 epitope tag, if desired.</p>

Step	Action										
Amplify the Gene of Interest	Use a thermostable, proofreading DNA polymerase and the PCR primers above to produce blunt-end PCR product. Use agarose gel electrophoresis to check the integrity of PCR product.										
Perform the TOPO [®] Cloning Reaction	<ol style="list-style-type: none"> Set up the following TOPO[®] Cloning reaction. <table> <tr> <td>Fresh PCR product</td><td>0.5 to 4 μl</td></tr> <tr> <td>Salt Solution</td><td>1 μl</td></tr> <tr> <td>Sterile water</td><td>add to a final volume of 5 μl</td></tr> <tr> <td>TOPO[®] vector</td><td>1 μl</td></tr> <tr> <td>Total volume</td><td>6 μl</td></tr> </table> Mix gently and incubate for 5 minutes at room temperature. Place on ice and proceed to transform One Shot[®] TOP10 chemically competent E. coli, below. 	Fresh PCR product	0.5 to 4 μ l	Salt Solution	1 μ l	Sterile water	add to a final volume of 5 μ l	TOPO [®] vector	1 μ l	Total volume	6 μ l
Fresh PCR product	0.5 to 4 μ l										
Salt Solution	1 μ l										
Sterile water	add to a final volume of 5 μ l										
TOPO [®] vector	1 μ l										
Total volume	6 μ l										
Transform One Shot [®] TOP10 Chemically Competent E. coli	<ol style="list-style-type: none"> Add 2 μl of the TOPO[®] Cloning reaction into a vial of One Shot[®] TOP10 chemically competent E. coli and mix gently. Incubate on ice for 5 to 30 minutes. Heat-shock the cells for 30 seconds at 42°C without shaking. Immediately transfer the tube to ice. Add 250 μl of room temperature SOC medium. Incubate at 37°C for 1 hour with shaking. Spread 50-200 μl of bacterial culture on a prewarmed LB agar plate containing 50-100 μg/ml ampicillin and 50 μg/ml blasticidin, and incubate overnight at 37°C. 										

[0806] Using the Control PCR Template and the Control PCR Primers included with the kit to perform a control reaction is recommended. See the protocol below for details.

[0807] The pLenti6/V5 Directional TOPO[®] Cloning Kit is shipped on dry ice and contains two boxes. Upon receipt, store the boxes as detailed below.

Box	Item	Storage
1	pLenti6/V5-D-TOPO [®] Reagents	-20°C
2	One Shot [®] TOP10 Chemically Competent E. coli	-80°C

[0808] pLenti6/V5-D-TOPO[®] reagents (Box 1) are listed below. Note that the user must supply a thermostable, proofreading polymerase and the appropriate PCR buffer.

Store Box 1 at -20°C.

Item	Concentration	Amount
pLenti6/V5-D-TOPO [®]	10 ng/μl linearized plasmid DNA in: 50% glycerol 50 mM Tris-HCl, pH 7.4 (at 25°C) 1 mM EDTA 2 mM DTT 0.1% Triton X-100 100 μg/ml BSA 30 μM bromophenol blue	20 μl
dNTP Mix	12.5 mM dATP 12.5 mM dCTP 12.5 mM dGTP 12.5 mM dTTP in water, pH 8	10 μl
Salt Solution	1.2 M NaCl 0.06 M MgCl ₂	50 μl
Sterile Water	---	1 ml
CMV Forward Sequencing Primer	0.1 μg/μl in TE Buffer, pH 8	20 μl
V5(C-term) Reverse Sequencing Primer	0.1 μg/μl in TE Buffer, pH 8	20 μl
Control PCR Primers	0.1 μg/μl each in TE Buffer, pH 8	10 μl
Control PCR Template	0.1 μg/μl in TE Buffer, pH 8	10 μl
pLenti6/V5-GW/lacZ Expression Control Plasmid	Lyophilized in TE Buffer, pH 8	10 μg

[0809] The sequences of CMV Forward and V5(C-term) Reverse sequencing primers. Two micrograms of each primer are as follows:

CMV Forward 5'-CGCAAATGGGCGGTAGGCGTG-3'

V5(C-term) Reverse 5'-ACCGAGGAGAGGGTTAGGGAT-3'

[0810] TOP10 cells have the following genotype: *F⁻ mcrA Δ(mrr-hsdRMS-mcrBC) Φ80lacZΔM15 ΔlacX74 recA1 deoR araD139 Δ(ara-leu)7697 galU galK rpsL (Str^R) endA1 nupG*. Transformation efficiency is 1×10^9 cfu/ μ g DNA and they should be stored at -80°C.

[0811] The pLenti6/V5-D-TOPO[®] vector is designed for use with the ViraPower[™] Lentiviral Expression System available from Invitrogen Corporation, Carlsbad, CA. Ordering information for the ViraPower[™] Lentiviral Expression System and other ViraPower[™] lentiviral support products and expression vectors is provided below. For more information, see the Invitrogen Corporation, Carlsbad, CA Web site.

Item	Quantity	Catalog no.
ViraPower [™] Lentiviral Directional TOPO [®] Expression Kit (includes ViraPower [™] Lentiviral Support Kit and the 293FT Cell Line)	1 kit	K4950-00
ViraPower [™] Lentiviral GATEWAY [™] Expression Kit	1 kit	K4960-00
pLenti6/V5-DEST [™] GATEWAY [™] Vector Pack	6 μ g	V496-10
ViraPower [™] Lentiviral Support Kit (includes ViraPower [™] Packaging Mix, Lipofectamine [™] 2000, and blasticidin)	20 reactions	K4970-00
293FT Cell Line	3×10^6 cells	R700-07

[0812] Some of the reagents supplied in the pLenti6/V5 Directional TOPO[®] Cloning Kit as well as other reagents suitable for use with the kit are available separately from Invitrogen Corporation, Carlsbad, CA. Ordering information for these reagents is provided below.

Item	Quantity	Catalog no.
One Shot [®] TOP10 Chemically Competent Cells	10 reactions	C4040-10
	20 reactions	C4040-03
One Shot [®] TOP10 Electrocompetent Cells	10 reactions	C4040-50
Ampicillin	200 mg	11593-019
Blasticidin	50 mg	R210-01
ThermalAce [™] DNA Polymerase	200 units	E0200
	1000 units	E1000
Platinum [®] Pfx DNA Polymerase	100 units	11708-013
Lipofectamine [™] 2000	0.75 ml	11668-027
	1.5 ml	11668-019

- [0813] The pLenti6/V5 Directional TOPO[®] Cloning Kit combines the ViraPower[™] Lentiviral Expression System with TOPO[®] Cloning technology to provide a highly efficient, rapid cloning strategy for insertion of blunt-end PCR products into a vector for expression in dividing and non-dividing mammalian cells. TOPO[®] Cloning requires no ligase, post-PCR procedures, or restriction enzymes.
- [0814] pLenti6/V5-D-TOPO[®] is a 7.0 kb expression vector designed to facilitate rapid, directional TOPO[®] Cloning and high-level expression of PCR products in mammalian cells using the ViraPower[™] Lentiviral Expression System (Catalog nos. K4950-00) available from Invitrogen Corporation, Carlsbad, CA. Features of the vector include: RSV enhancer/promoter bases 1-229; HIV-1 5' LTR bases 230-410; 5' splice donor base 520; HIV-1 psi (ψ) packaging sequence bases 521-565; HIV-1 Rev response element (RRE) bases 1075-1308; 3' splice acceptor base 1656; 3' splice acceptor base 1684; CMV promoter bases 1809-2392; CMV forward priming site bases 2274-2294; directional TOPO[®] site bases 2431-2444; V5 epitope bases 2473-2514; V5(C-term) reverse priming site bases 2482-2502; SV40 early promoter and origin bases 2569-2878; EM7 promoter bases 2933-2999; Blasticidin resistance gene bases 3000-3398; Δ U3/HIV-1 3' LTR bases 3485-3718; Δ U3: bases 3485-3537; Truncated HIV-1 3' LTR bases 3538-3718; SV40 polyadenylation signal bases 3790-3921; *bla* promoter: bases 4780-4878; ampicillin (*bla*) resistance gene bases 4879-5739; and the pUC origin: bases 5884-6557.
- [0815] The control plasmid, pLenti6/V5-GW/*lacZ*, may be used as a positive expression control in the mammalian cell line of choice.
- [0816] The ViraPower[™] Lentiviral Expression System facilitates highly efficient, *in vitro* or *in vivo* delivery of a target gene to dividing and non-dividing mammalian cells using a replication-incompetent lentivirus. Based on the lentikat[™] system developed by Cell Genesys (Dull *et al.*, 1998), the ViraPower[™] Lentiviral Expression System possesses features which enhance its biosafety while allowing high-level gene expression in a wider range of cell types than traditional retroviral systems. To express a gene of interest in mammalian cells using the ViraPower[™] Lentiviral Expression System:

1. TOPO[®] Clone a gene of interest into pLenti6/V5-D-TOPO[®] to create an expression construct.
2. Cotransfect the pLenti6/V5-D-TOPO[®] expression plasmid and the ViraPower[™] Packaging Mix into the 293FT cell line to produce lentivirus.
3. Use the lentiviral stock to transduce the mammalian cell line of choice.
4. Assay for "transient" expression of the recombinant protein or generate a stable cell line using blasticidin selection.

[0817] Detailed protocols for creating recombinant lentiviruses are known (e.g., ViraPower[™] Lentiviral Expression System manual, catalog nos. K4950-00, K4960-00, K4970-00, K4975-00, K49580-00, K49585-00, and K49590-00, version D, Invitrogen Corporation, Carlsbad, CA).

[0818] Directional joining of double-strand DNA using TOPO[®]-charged oligonucleotides occurs by adding a 3' single-stranded end (overhang) to the incoming DNA (Cheng and Shuman, 2000, *Mol. Cell. Biol.* 20, 8059-8068.). This single-stranded overhang is identical to the 5' end of the TOPO[®]-charged DNA fragment. The pLenti6/V5-D-TOPO[®] vector contains a 4 nucleotide overhang sequence.

[0819] In this system, PCR products are directionally cloned by adding four bases to the forward primer (CACC). The overhang in the cloning vector (GTGG) invades the 5' end of the PCR product, anneals to the added bases, and stabilizes the PCR product in the correct orientation. Inserts can be cloned in the correct orientation with efficiencies equal to or greater than 90%. A schematic representation of the process is shown in Figure 47.

[0820] The design of the PCR primers to amplify a gene of interest is critical for expression. Consider the following when designing PCR primers: sequences required to facilitate directional cloning; sequences required for proper translation initiation of the PCR product; and whether or not a coding sequence contained by the PCR product is to be fused in frame with the C-terminal V5 epitope tag.

- [0821] When designing a forward PCR primer, consider the points below. Refer to Figure 48 for a diagram of the TOPO[®] Cloning site for pLenti6/V5-D-TOPO[®].
- [0822] To enable directional cloning, the forward PCR primer **MUST** contain the sequence, CACC, at the 5' end of the primer. The 4 nucleotides, CACC, base pair with the overhang sequence, GTGG, in the pLenti6/V5-D-TOPO[®] vector.
- [0823] The sequence of interest should include a Kozak translation initiation sequence with an ATG initiation codon for proper initiation of translation (Kozak, 1987; Kozak, 1991; Kozak, 1990). An example of a Kozak consensus sequence is (G/A)NN**ATGG**. Other sequences are possible, but the G or A at position -3 and the G at position +4 are the most critical for function (shown in bold). The ATG initiation codon is underlined.
- [0824] Below is the DNA sequence of the N-terminus of a theoretical protein and the proposed sequence for a forward PCR primer. The ATG initiation codon is underlined.

DNA sequence: 5'-ATG GGA TCT GAT AAA

Proposed Forward PCR primer: 5'-C ACC ATG GGA TCT GAT
AAA

If the forward PCR primer is designed as above, then the primer includes the 4 nucleotides, CACC, required for directional cloning, and the ATG initiation codon falls within the context of a Kozak sequence (see boxed sequence), allowing proper translation initiation of the PCR product in mammalian cells. The first three base pairs of the PCR product following the 5' CACC overhang will constitute a functional codon.

- [0825] When designing a reverse PCR primer, consider the points below. Refer to Figure 48 for a diagram of the TOPO[®] Cloning site for pLenti6/V5-D-TOPO[®]. To ensure that the PCR product clones directionally with high efficiency, the reverse PCR primer should not be complementary to the overhang sequence GTGG at the 5' end. A one base pair mismatch can reduce the directional cloning efficiency from 90% to 50%, increasing the likelihood of the PCR product cloning in the opposite orientation (see below). Evidence of PCR products cloning in the opposite orientation from a two base pair mismatch has not been observed.

[0826] To fuse a PCR product in frame with the C-terminal tag containing the V5 epitope, the reverse PCR primer can be designed to remove the native stop codon in the gene of interest (see below). To produce a native C-terminal on an expressed polypeptide, include the native sequence containing the stop codon in the reverse primer or make sure the stop codon is upstream from the reverse PCR primer binding site.

[0827] First Example of Reverse Primer Design. Below is the sequence of the C-terminus of a theoretical protein. The stop codon is underlined.

DNA sequence: AAG TCG GAG CAC TCG ACG ACG GTG
TAG-3'

[0828] To fuse the protein in frame with the C-terminal tag in pLenti6/V5-D-TOPO[®], design the reverse PCR primer to start with the codon just up-stream of the stop codon, but the last two codons contain GTGG (underlined below), which is identical to the 4 bp overhang sequence. As a result, the reverse primer will be complementary to the 4 bp overhang sequence, increasing the probability that the PCR product will clone in the opposite orientation. This situation should be avoided.

DNA sequence: AAG TCG GAG CAC TCG ACG ACG GTG TAG-3'
Proposed Reverse PCR primer sequence: TG AGC TGC TGC CAC AAA-5'

[0829] Another solution is to design the reverse primer so that it hybridizes just down-stream of the stop codon, but still includes the C-terminus of the ORF. Note that the stop codon will need to be replaced by a codon for an innocuous amino acid such as glycine, alanine, or lysine.

[0830] Second Example of Reverse Primer Design

[0831] Below is the sequence for the C-terminus of a theoretical protein. The stop codon is underlined.

...GCG GTT AAG TCG GAG CAC TCG ACG ACT GCA TAG-3'

[0832] To fuse the ORF in frame with the C-terminal tag in pLenti6/V5-D-TOPO[®], remove the stop codon by starting with nucleotides homologous to the last codon (TGC) and continue upstream. The reverse primer will be:

5'-TGC AGT CGT CGA GTG CTC CGA CTT-3'

[0833] This will amplify the C-terminus without the stop codon and allow the ORF to be joined in frame with the C-terminal tag. To avoid joining the ORF

in frame with a C-terminal tag, design the reverse primer to include the stop codon.

5'-CTA TGC AGT CGT CGA GTG CTC CGA CTT-3'

[0834] pLenti6/V5-D-TOPO[®] accepts blunt-end PCR products. Do not add 5' phosphates to primers for PCR. This will prevent ligation into the pLenti6/V5-D-TOPO[®] vector. It is recommended that oligonucleotides be gel-purified, especially if they are long (> 30 nucleotides). Note that pLenti6/V5-D-TOPO[®] is supplied linearized with both ends adapted with topoisomerase I (see Figure 47). The sequence of pLenti6/V5-D-TOPO[™] is provided as Table 18.

[0835] Once a PCR strategy has been decided upon and primers synthesized, a blunt-end PCR product can be produced using any thermostable, proof-reading polymerase including, but not limited to, ThermalAce[™], Platinum[®] Pfx, Pfu, or Vent[®] for PCR.

[0836] Follow the manufacturer's instructions and recommendations to produce blunt-end PCR products. It is important to optimize PCR conditions to produce a single, discrete PCR product. PCR fragments may be gel purified using standard techniques.

[0837] It is recommended that a 7 to 30 minute final extension be used in the PCR reaction to ensure that all PCR products are completely extended.

[0838] After the PCR reaction, the PCR product should be checked by removing 5 to 10 µl from each PCR reaction and using agarose gel electrophoresis to verify the quality and quantity of the PCR product. Check for a single, discrete band of the correct size. If there is not a single, discrete band, follow the manufacturer's recommendations for optimizing PCR with the polymerase of choice. Alternatively, gel-purify the desired product.

[0839] Estimate the concentration of the PCR product. A 5:1 molar ratio of PCR product:TOPO[®] vector is recommended to obtain the highest TOPO[®] Cloning efficiency (e.g. use 5-10 ng of a 1 kb PCR product or 10-20 ng of a 2 kb PCR product in a TOPO[®] Cloning reaction). Adjust the concentration of the PCR product as necessary before proceeding to TOPO[®] Cloning. If ThermalAce[™] polymerase is used to produce blunt-end PCR product, note that ThermalAce[™] can generate higher yields than other proofreading polymerases. When generating PCR products in the 0.5 to 1.0 kb range, generally the PCR

reaction can be diluted 1:5 in 1X ThermalAce™ buffer before performing the TOPO® Cloning reaction. For PCR products larger than 1.0 kb, dilution may not be required.

[0840] Including salt (250 mM NaCl, 10 mM MgCl₂) in the TOPO® Cloning reaction may result in an increase in the number of transformants. Therefore, it is recommended that salt be added to the TOPO® Cloning reaction. A stock salt solution is provided in the kit for this purpose. Note that the amount of salt added to the TOPO® Cloning reaction varies depending on whether chemically competent cells (provided) or electrocompetent cells are to be transformed. For this reason two different TOPO® Cloning reactions are provided to obtain the best possible results.

[0841] Transforming Chemically Competent *E. coli*. For TOPO® Cloning and transformation into chemically competent *E. coli*, adding sodium chloride and magnesium chloride to a final concentration of 250 mM NaCl, 10 mM MgCl₂ in the TOPO® Cloning reaction increases the number of colonies over time. A Salt Solution (1.2 M NaCl, 0.06 M MgCl₂) is provided to adjust the TOPO® Cloning reaction to the recommended concentration of NaCl and MgCl₂.

[0842] Transforming Electrocompetent *E. coli*. For transformation of electrocompetent *E. coli*, the amount of salt in the TOPO® Cloning reaction should be reduced (*e.g.*, to 50 mM NaCl, 2.5 mM MgCl₂) to prevent arcing. Dilute the Salt Solution 4-fold with water to prepare a 300 mM NaCl, 15 mM MgCl₂ solution for convenient addition to the TOPO® Cloning reaction (see below).

[0843] Setting Up the TOPO® Cloning Reaction. The table below describes how to set up a TOPO® Cloning reaction (6 µl) for eventual transformation into either chemically competent One Shot® TOP10 *E. coli* (provided) or electrocompetent *E. coli*. Additional information on optimizing the TOPO® Cloning reaction can be found herein. If the PCR product was generated using ThermalAce™ polymerase, note that it may be necessary to dilute the PCR reaction before proceeding. The blue color of the TOPO® vector solution is normal and is used to visualize the solution.

Reagents*	Chemically Competent <i>E. coli</i>	Electrocompetent <i>E. coli</i>
Fresh PCR product	0.5 to 4 µl	0.5 to 4 µl
Salt Solution	1 µl	--

Reagents*	Chemically Competent <i>E. coli</i>	Electrocompetent <i>E. coli</i>
Sterile Water	add to a final volume of 5 μ l	add to a final volume of 5 μ l
TOPO [®] vector	1 μ l	1 μ l

[0844] *Store all reagents at -20°C when finished. Salt solution and water can be stored at room temperature or $+4^{\circ}\text{C}$.

[0845] Performing the TOPO[®] Cloning Reaction. Mix reaction gently and incubate for 5 minutes at room temperature ($22-23^{\circ}\text{C}$). For most applications, 5 minutes will yield plenty of colonies for analysis. Depending on needs, the length of the TOPO[®] Cloning reaction can be varied from 30 seconds to 30 minutes. For routine subcloning of PCR products, 30 seconds may be sufficient. For large PCR products ($> 1 \text{ kb}$) or TOPO[®] Cloning a pool of PCR products, increasing the reaction time may yield more colonies.

[0846] Place the reaction on ice and transform suitable host cells using standard protocols. The TOPO[®] Cloning reaction can be stored at -20°C overnight.

[0847] Transforming One Shot[®] TOP10 Competent *E. coli*. Once the TOPO[®] Cloning reaction has been performed, the pLenti6/V5-D-TOPO[®] construct is transformed into competent *E. coli*. One Shot[®] TOP10 Chemically Competent *E. coli* (Invitrogen Corporation, Carlsbad, CA) are included with the kit to facilitate transformation, however, electrocompetent cells may also be used. Protocols to transform chemically competent or electrocompetent *E. coli* are known to those skilled in the art. pLenti6/V5-D-TOPO[®] contains the ampicillin and blasticidin resistance genes for selection of transformants. Unwanted recombination (less than 5%) between the 5' and 3' LTRs has been observed when transformants are selected on LB agar plates containing ampicillin. These events occur less frequently when transformants are selected on LB agar plates containing ampicillin and blasticidin. Transformants should be selected on LB agar plates containing 50-100 $\mu\text{g/ml}$ ampicillin AND 50 $\mu\text{g/ml}$ blasticidin. Note that transformed *E. coli* grow more slowly in LB media containing ampicillin and blasticidin, and may require slightly longer incubation times to obtain visible colonies.

- [0848] Transformants that contain a recombined plasmid generally give rise to larger colonies than those containing an intact plasmid. There is no blue-white screening for the presence of inserts. Most transformants will contain recombinant plasmid with the PCR product of interest cloned in the correct orientation. Sequencing primers are included in the kit to sequence across an insert in the multiple cloning site to confirm orientation and reading frame.
- [0849] Addition of the Dilute Salt Solution to the TOPO[®] Cloning Reaction brings the final concentration of NaCl and MgCl₂ in the reaction to 50 mM and 2.5 mM, respectively. To prevent arcing of samples during electroporation, the volume of cells should be between 50 and 80 μ l (0.1 cm cuvettes) or 100 to 200 μ l (0.2 cm cuvettes). If arcing during transformation is seen, try reducing the voltage normally used to charge the electroporator by 10%, reducing the pulse length by reducing the load resistance to 100 ohms, and/or ethanol precipitating the TOPO[®] Cloning reaction and resuspending in water prior to electroporation.
- [0850] After transformation and plating, pick 5 colonies and culture them overnight in LB or SOB medium containing 50-100 μ g/ml ampicillin. Addition of blasticidin is not required. Isolate plasmid DNA using a method of choice. If ultra-pure plasmid DNA is need for automated or manual sequencing, the S.N.A.P.[™] MidiPrep Kit (Invitrogen Corporation, Carlsbad, CA Catalog no. K1910-01) may be used. Analyze the plasmids by restriction analysis to confirm the presence and correct orientation of the insert. Use a restriction enzyme or a combination of enzymes that cut once in the vector and once in the insert.
- [0851] Sequencing. The construct may be sequenced to confirm that the sequence of interest is cloned in the correct orientation and in frame with the V5 epitope. The CMV Forward and V5(C-term) Reverse primers are included in the kit and can be used to sequence the insert.
- [0852] The sequence for pLenti6/V5-D-TOPO[®] shown in Table 18 includes the overhang sequence (GTGG) hybridized to CACC.
- Analyzing Transformants by PCR. Transformants can be analyzed using PCR. For PCR primers, use a combination of the CMV Forward primer or the V5(C-term) Reverse primer and a primer that hybridizes within the insert. Appropriate amplification conditions can be determined by one

skilled in the art. Results from the PCR reaction may be verified by conducting restriction analysis in parallel. Artifacts may be obtained in the PCR reaction because of mispriming or contaminating template.

- [0853] If transformants or the correct insert are not obtained, perform the control reactions described below.
- [0854] Once the correct clone has been identified, a glycerol stock of bacteria containing the plasmid may be prepared for long term storage. Also, a stock of plasmid DNA can be prepared and stored at -20°C.
- [0855] Once a host cell containing a pLenti6/V5-D-TOPO® expression plasmid has been prepared, maintain and propagate the plasmid in LB medium containing 50-100 µg/ml ampicillin. Addition of blasticidin is not required.
- [0856] Optimizing the TOPO® Cloning Reaction. The high efficiency of TOPO® Cloning allows the cloning process to be streamlined. To speed up the process of cloning PCR products, the TOPO® Cloning reaction can be incubated for only 30 seconds instead of 5 minutes. Fewer transformants may be obtained; however, because of the high efficiency of TOPO® Cloning, most of the transformants will contain the insert. After adding 2 µl of the TOPO® Cloning reaction to chemically competent cells, incubate on ice for only 5 minutes. Increasing the incubation time to 30 minutes does not significantly improve transformation efficiency.
- [0857] When TOPO® Cloning large PCR products, toxic genes, or cloning a pool of PCR products, more transformants may be needed to obtain the desired clones. To increase the number of colonies incubate the salt-supplemented TOPO® Cloning reaction for 20 to 30 minutes instead of 5 minutes. Increasing the incubation time of the salt-supplemented TOPO® Cloning reaction allows more molecules to ligate and may increase the transformation efficiency. Addition of salt appears to prevent topoisomerase I from rebinding and nicking the DNA after it has ligated the PCR product and dissociated from the DNA.
- [0858] To clone dilute PCR products, increase the amount of the PCR product, incubate the TOPO® Cloning reaction for 20 to 30 minutes, and/or concentrate the PCR product.
- [0859] Once the sequence of interest has been TOPO® Cloned into pLenti6/V5-D-TOPO®, the ViraPower™ Lentiviral Expression System from

Invitrogen Corporation, Carlsbad, CA can be used to produce a viral stock, which may then be used to transduce a mammalian cell line of choice to express the recombinant protein (as described above).

EXAMPLE 13

GROWTH AND MAINTENANCE OF THE 293FT CELL LINE

[0860] The 293FT cell line may be transported using any technique known to those skilled in the art, for example, by freezing the cells and transporting them on dry ice. For long term storage, the cells may be stored in liquid nitrogen. The 293FT cell line is supplied as one vial containing 3×10^6 frozen cells in 1 ml of Freezing Medium.

[0861] The 293FT cell line is genetically modified and carries the pUC-derived plasmid, pCMVSPORT6TA_g.neo. A map of the vector is provided as Figure 49. The pCMVSPORT6TA_g.neo plasmid is derived from pCMVSPORT6, which has been modified to include the neomycin resistance gene for stable selection in mammalian cells (Southern and Berg, 1982, *J. Molec. Appl. Gen.* 1, 327-339). Expression of the neomycin resistance gene is controlled by the SV40 early enhancer/promoter from which the SV40 origin of replication has been removed. The plasmid also contains the gene encoding the SV40 large T antigen to facilitate optimal virus production (*e.g.* Invitrogen's ViraPower™ Lentiviral Expression System) and to permit episomal replication of plasmids containing the SV40 early promoter and origin. Expression of the SV40 large T antigen is controlled by the human cytomegalovirus (CMV) promoter.

[0862] The 293FT cell line is derived from the 293F cell line (see below) and stably expresses the SV40 large T antigen from the pCMVSPORT6TA_g.neo plasmid. Expression of the SV40 large T antigen is controlled by the human cytomegalovirus (CMV) promoter and is high-level and constitutive. For more information about pCMVSPORT6TA_g.neo, see below.

[0863] Studies have demonstrated maximal virus production in human 293 cells expressing SV40 large T antigen (Naldini *et al.*, 1996), making the 293FT cell line a particularly suitable host for generating lentiviral constructs

using the ViraPower™ Lentiviral Expression System available from Invitrogen (Catalog nos. K4950-00 and K4960-00).

[0864] The 293 cell line is a permanent line established from primary embryonal human kidney transformed with sheared human adenovirus type 5 DNA (Graham *et al.*, 1977; Harrison *et al.*, 1977, *Virology* 77, 319-329). The E1A adenovirus gene is expressed in these cells and participates in transactivation of some viral promoters, allowing these cells to produce very high levels of protein. The 293-F cell line available from Invitrogen Corporation, Carlsbad, CA (Catalog no. 11625) is a fast-growing variant of the 293 cell line, and was originally obtained from Robert Horlick at Pharmacopeia.

[0865] Antibiotic Resistance. 293FT cells stably express the neomycin resistance gene from pCMVSPORT6TA_g.neo and should be maintained in medium containing Geneticin® at the concentration listed below. Expression of the neomycin resistance gene in 293FT cells is controlled by the SV40 enhancer/promoter. Geneticin® is available separately from Invitrogen Corporation, Carlsbad, CA (catalog number 11811-023).

[0866] Media for 293FT Cells. It is recommended that 293FT cells be grown in complete medium (D-MEM (high glucose), 10% fetal bovine serum (FBS), 2 mM L-glutamine, 1% Pen-Strep (optional)). For selection 500 µg/ml Geneticin® should be included. For freezing, 90% complete medium and 10% DMSO should be used. FBS does not need to be heat-inactivated for use with the 293FT cell line. 293FT cells should be maintained in medium containing Geneticin® at the concentration listed above. If cells are split at a 1:5 to 1:10 dilution, they will generally reach 80-90% confluence in 3-4 days.

[0867] Follow the general guidelines below to grow and maintain 293FT cells. All solutions and equipment that come in contact with the cells must be sterile. Always use proper sterile technique and work in a laminar flow hood. Before starting experiments, be sure to have cells established and also have some frozen stocks on hand. Early-passage cells are recommended for experiments. Upon receipt of the cells from Invitrogen Corporation, Carlsbad, CA, grow and freeze multiple vials of the 293FT cell line to ensure that an adequate supply of early-passage cells is available.

- [0868] For general maintenance of cells, pass 293FT cells when they are 80-90% confluent (generally every 3-4 days). Avoid overgrowing cells before passaging.
- [0869] Use trypan blue exclusion to determine cell viability. Log phase cultures should be >90% viable.
- [0870] When thawing or subculturing cells, transfer cells into pre-warmed medium.
- [0871] Cells should be at the appropriate confluence and at greater than 90% viability prior to transfection.
- [0872] As with other human cell lines, when working with 293FT cells, handle as potentially biohazardous material under at least Biosafety Level 2 (BL-2) containment.
- [0873] The following protocol is designed to thaw 293FT cells to initiate cell culture. The 293FT cell line is supplied in a vial containing 3×10^6 cells in 1 ml of Freezing Medium.
- [0874] Remove the vial of cells from the liquid nitrogen and thaw quickly in a 37°C water bath. Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol, and transfer the cells to a T-75 flask containing 12 ml of complete medium without Geneticin®. Incubate the flask at 37°C for 2-4 hours to allow the cells to attach to the bottom of the flask. Aspirate off the medium and replace with 12 ml of fresh, complete medium without Geneticin®. Incubate cells overnight at 37°C. The next day, aspirate off the medium and replace with fresh, complete medium containing Geneticin® at the recommended concentration listed above. Incubate the cells and check them daily until the cells are 80-90% confluent (2-7 days).
- [0875] Passaging Cells. When cells are ~80-90% confluent, remove all medium from the flask. Wash cells once with 10 ml PBS to remove excess medium and serum. Serum contains inhibitors of trypsin. Add 5 ml of trypsin/versene (EDTA) solution to the monolayer and incubate 1 to 5 minutes at room temperature until cells detach. Check the cells under a microscope and confirm that most of the cells have detached. If cells are still attached, incubate a little longer until most of the cells have detached. Add 5 ml of complete medium to stop trypsinization. Briefly pipette the solution up and down to break up clumps of cells.

- [0876] To maintain cells in 75 cm² flasks, transfer 1 ml of the 10 ml cell suspension from above to a new 75 cm² flask and add 15 ml fresh, complete medium containing Geneticin[®]. To have the cells reach confluency sooner, split the cells at a lower dilution (i.e. 1:4).
- [0877] To expand cells into 175 cm² flasks, add 28 ml of fresh, complete medium containing Geneticin[®] to each of three 175 cm² flasks, then transfer 2 ml of the cell suspension to each flask to obtain a total volume of 30 ml.
- [0878] Incubate flasks in a humidified, 37°C, 5% CO₂ incubator.
- [0879] Passage the cells as necessary to maintain or expand cells.
- [0880] Freezing Cells. When freezing the 293FT cell line, it is recommended that the cells be frozen at a density of at least 3 x 10⁶ viable cells/ml. Use a freezing medium composed of 90% complete medium and 10% DMSO. Complete medium is medium containing serum.
- [0881] Preparing Freezing Medium. Freezing medium should be prepared immediately before use. In a sterile, conical centrifuge tube, mix together 0.9 ml of fresh complete medium and 0.1 ml of DMSO for every 1 ml needed. Place the tube on ice until use. Discard any remaining freezing medium after use.
- [0882] Freezing the Cells. Before starting, label cryovials and prepare freezing medium (see above). Keep the freezing medium on ice. To collect cells, count the cells prepared by trypsinization as described in Passaging the Cells above. Pellet cells at 250 x g for 5 minutes in a table top centrifuge at room temperature and carefully aspirate off the medium. Resuspend the cells at a density of at least 3 x 10⁶ cells/ml in chilled freezing medium. Place vials in a microcentrifuge rack and aliquot 1 ml of the cell suspension into each cryovial. Freeze cells in an automated or manual, controlled-rate freezing apparatus following standard procedures. For ideal cryopreservation, the freezing rate should be a decrease of 1°C per minute. Transfer vials to liquid nitrogen for long-term storage.
- [0883] The viability and recovery of frozen cells may be checked 24 hours after storing cryovials in liquid nitrogen by following the procedure outlined in Thawing above.
- [0884] Transfecting Cells. The 293FT cell line is generally amenable to transfection using standard methods including calcium phosphate precipitation

(Chen and Okayama, 1987, *Molec. Cell. Biol.* 7, 2745-2752; Wigler *et al.*, 1977, *Cell* 11, 223-232), lipid-mediated transfection (Felgner *et al.*, 1989, *Proc. West. Pharmacol. Soc.* 32, 115-121; Felgner and Ringold, 1989, *Nature* 337, 387-388), and electroporation (Chu *et al.*, 1987, *Nucleic Acids Res.* 15, 1311-1326; Shigekawa and Dower, 1988, *BioTechniques* 6, 742-751).

Typically cationic lipid-based transfection reagents are used to transfect 293FT cells. Lipofectamine™ 2000 (Invitrogen Corporation, Carlsbad, CA catalog number 11668-027) is recommended, but other transfection reagents are suitable.

[0885] Transient Transfection. The 293FT cell line may be transiently transfected with any plasmid. Make sure that cells are healthy at the time of plating. Overgrowth of cells prior to passaging can compromise their transfection efficiency. On the day before transfection, plate cells such that they will be approximately 60% confluent at the time of transfection. If Lipofectamine™ 2000 is to be used as a transfection reagent, plate cells such that they will be 90-95% confluent at the time of transfection. Transfect the plasmid construct into the 293FT cell line using the method of choice (see above). After transfection, add fresh medium containing 500 µg/ml Geneticin® and allow the cells to recover for 24-48 hours before proceeding to assay for expression of the gene of interest.

[0886] Generating Stable Cell Lines. 293FT cells can be used as hosts to generate a stable cell line expressing a gene of interest from most plasmids. Remember that the introduced plasmid must contain a selection marker other than neomycin resistance. Stable cell lines can then be generated by transfection and dual selection with Geneticin® and the appropriate selection agent.

[0887] Since 293FT cells stably express the SV40 large T antigen, generating stable cell lines with plasmids that contain the SV40 origin of replication is not recommended.

EXAMPLE 14

Use of Suppressor tRNAs to Transiently Label Proteins of Interest

[0888] This example describes the use of mammalian suppressor tRNAs (*e.g.*, tRNA^{ser}) that specifically recognize and decode one of the three stop codons: amber (TAG), opal (TGA) or ochre (TAA) as an amino acid (*e.g.*, serine). Expression plasmids encoding a gene of interest with one of these stop codons will express a native protein under normal conditions (see Figure 50). If the appropriate tRNA suppressor is supplied, that stop codon will be translated (*e.g.*, as serine when tRNA^{ser} is used) and translation will continue through any downstream reading frame, creating a fusion protein consisting of the protein of interest with a specific C-terminal epitope tag (see Figure 50). "Gene of interest" as used herein, refers to, for example, a nucleic acid sequence encoding a polypeptide, a protein, or an untranslated RNA, *e.g.*, tRNA, all of which are encompassed by the term.

[0889] One non-limiting example of this stop suppression technology, termed Tag-On-Demand™ available from Invitrogen Corporation, Carlsbad, CA, which allows expression of tagged or untagged proteins using a single gene expression vector. In this embodiment, recombinant adenovirus vectors carrying the amber (TAG) stop suppressor tRNA gene have been developed as well as optimized protocols for use in transiently tagging a protein of interest in mammalian cells. The specific embodiment described here is purified, titered recombinant adenovirus (Adeno-tRNA^{TAG}) and one new GATEWAY™ Destination vector (pcDNA6.2/GFP-DEST). Tag-On-Demand™ may be used with any gene of interest provided the stop codon is TAG. For example, additional Invitrogen mammalian expression vectors that are compatible with Tag-On-Demand™ are listed below.

[0890] The use of the pcDNA6.2/V5 and pcDNA6.2/GFP Destination vectors is recommended for use in Tag-On-Demand™ primarily due to the superiority of blasticidin as a selectable marker and the absence of the BGH polyA. In addition to the recommended vectors listed above, the following three Destination vectors have also been successfully used in Tag-On-Demand™ pcDNA3.2/V5-DEST, pcDNA-DEST40, and pcDNA-DEST47. The following Invitrogen vectors are all compatible with Tag-On-Demand™ and contain a

non-TAG stop codon downstream of the C-terminal epitope tag, provided the gene of interest is cloned with TAG stop in frame with the C-terminal tag: pcDNA/V5His vector family; pEF/V5His vector family; pUbC/V5His vectors; pcDNA/mycHis vector family; pEF/mycHis vector family; pcDNA3.1/CT-GFP vectors; pcDNA4/TO/mycHis vectors; pGene/V5His vectors; pIND/V5His vectors; pcDNA5/FRT/V5His; and pEF5/FRT vectors.

Materials and Methods

[0891] Vector construction. (a) pUC12-tRNA^{TAG}: Three suppressor tRNA vectors were received from Dr. Uttam RajBhandary of Massachusetts Institute of Technology. Each suppressor tRNA vector, designated pUCtS Su+ amber, opal, and ochre, is identical except for the stop anticodon (Capone *et. al.* 1985, *EMBO*, 4(1):213-221). For convenience, the pUCtS Su+ amber vector is now referred to as pUC12-tRNA^{TAG}. To create a tetracycline-regulated version, referred to herein as pUC12-TO-tRNA^{TAG}, two tetracycline operators (tetO₂) were cloned into the *SnaBI* site in pUC12-tRNA^{TAG} using the following annealed oligonucleotides:

tetO₂ Forward primer

5' GACTCGAGTCTCCCTATCAGTGATAGAGATCTCGAGGTC 3' and

tetO₂ Reverse primer

5'GACCTCGAGATCTCTATCACTGATAGGGAGACTCGAGTC3'.

In italics is a unique *BglIII* site that was introduced with the oligonucleotide. The underlined sequences are *XhoI* sites. All tRNA constructs were sequence verified.

(b) pcDNA6.2/GFP-DEST: pcDNA6.2/V5-DEST was digested with *ApaI* and *PmeI* to remove the V5 tag. pcDNA3.1/lacZ-stop^{TAG}-GFP was also digested with *ApaI* and *PmeI* to isolate the GFP fragment. The GFP fusion tag was ligated to the pcDNA6.2 DEST vector (Invitrogen Corporation, Carlsbad, CA catalog # 12489-027) and transformed into DB3.1 cells. Colonies were grown on LB-Amp plates. A clone was selected that resulted in correct band fragments when digested with *NdeI* and then sequence confirmed.

(c) pENTR CAT^{TAA,TAG,TGA} The GATEWAYTM CAT entry clones were PCR amplified followed by TOPO cloning (Invitrogen Corporation, Carlsbad, CA product manual #25-0434) into pENTR dT. Information for both vectors

may be obtained by contacting Invitrogen Corporation, Carlsbad, CA. The primer sequences used were

Forward primer: 5' CACCATGGAGAAAAAATCACTGG 3'

Reverse primer: 5' CTGCTACGCCCCGCCCTGC 3'.

The underlined sequence varied depending on which stop codon was required. Plasmid constructs were sequence verified.

(d) pcDNA3.2/V5-GW/CAT^{TAA, TAG, TGA} : pcDNA3.2/V5-DEST and pENTR CAT with each of the stops was recombined using LR clonase to generate the plasmids pcDNA3.2/V5-GW/CAT^{TAA, TAG, TGA}. Clones were identified as correct by restriction enzyme digests and sequence confirmed.

(e) pcDNA6.2/GFP-GW/CAT^{TAA, TAG, TGA} : pcDNA6.2/GFP-DEST and pENTR CAT with each of the stops was recombined using LR clonase to generate the plasmids pcDNA6.2/GFP-GW/CAT^{TAA, TAG, TGA}. Clones were identified as correct by restriction enzyme digests and sequence confirmed.

(f) pENTR p48^{TAG} : This GATEWAYTM Entry clone was obtained from the UltimateTM ORFeome Collection (Invitrogen Corporation, Carlsbad, CA) and is referred to by several names: HS8-E6 (internal Invitrogen designation), BC000141 (GenBank Accession number), or ORF 12 (used for convenience). This ORF is referred to as p48 and is a human c-myc variant (see Results section). Information for this clone may be obtained by contacting Invitrogen Corporation, Carlsbad, CA or GenBank.

(g) pcDNA6.2/GFP-GW/p48^{TAG} : pcDNA6.2/GFP-DEST and pENTR p48^{TAG} were recombined with LR clonase to generate pcDNA6.2/GFP-GW/p48^{TAG}. The recombination reaction was transformed into TOP10 cells (Invitrogen Corporation, Carlsbad, CA, catalog #C4040-10) and plated on LB Ampicillin plates. Colonies were picked and clones were identified as correct by restriction enzyme digests and functional suppression.

(h) pcDNA6.2/V5-GW/p48^{TAG} : pcDNA6.2/V5-DEST and pENTR p48^{TAG} were recombined with LR clonase to generate the plasmid pcDNA6.2/V5-GW/p48^{TAG}. The recombination reaction was transformed into TOP10 cells and plated on LB Ampicillin plates. Colonies were picked and clones were identified as correct by restriction enzyme digests and functional suppression.

(i) pENTR-TO-tRNA^{TAG} : pENTR1A (Invitrogen Corporation, Carlsbad, CA) and pUC12-TO-tRNA^{TAG} (described in (a) above) were digested with *Sall* and *EcoRI*. Following digests, the appropriate bands were gel purified and ligated. Ligations were transformed into TOP10 cells and plated on LB-Kanamycin plates. Clone 1 was selected following *Sall* and *EcoRI* diagnostic digests.

(j) pENTR-tRNA8^{TAG} : Primers were created to PCR amplify the tRNA gene from pUC12 TO tRNA^{TAG} with *EcoRI* and *XbaI* sequences at the 5' end, and *SpeI* and *HindIII* at the 3' end. The primer sequences were:

Forward primer:

5' CACCGAATTCTCTAGAGATGTCTGTGAAAAGAAACAT 3' and

Reverse primer:

5' ATATAAGCTTACTAGTCCGGATTTCTCTACCCGAGA 3'.

The tRNA PCR product was gel purified, TOPO cloned into pENTR dT, and transformed into TOP10 cells. Colonies were selected on LB Kanamycin plates. Upon confirmation of proper insertion, two separate digests were conducted. The first digest with *EcoRI* and *XbaI* opened the pENTR-tRNA^{TAG}. The second digest with *EcoRI* and *SpeI* excised the tRNA gene. Correct fragments were gel purified, the two fragments were ligated, as *XbaI* and *SpeI* have complimentary ends, thus creating a dimer of tRNA. With confirmation of proper insertion, the same two previous digests were repeated with the dimer plasmid, fragments gel purified, ligations performed creating a tetramer. A final two digests, as previously described, were repeated on the tetramer, fragments gel purified, ligations performed creating an octamer tRNA in the pENTR backbone. (Buvoli *et al.*, *Mol. Cell. Biol.* 20:3116-3124 (2000), Suppression of Nonsense Mutations in Cell Culture and Mice by Multimerized Suppressor tRNA Genes).

Adenovirus tRNA

[0892] Adenovirus carrying the suppressor tRNA^{TAG} was created using a GATEWAY™ LxR reaction. pAd/PL-DEST vector (Table 10, Figure 9) was recombined with either pENTR-tRNA^{TAG} or pENTR-tRNA8^{TAG} to create pAd-tRNA^{TAG} (Table 8) or pAd-tRNA8^{TAG} expression vectors, respectively. These vectors were subsequently cut with *PacI* and transfected into TREx 293

(Invitrogen Corporation, Carlsbad, CA, catalog #R710-07) cells to produce the initial stocks of recombinant adenovirus. Subsequent virus amplification and titering was performed in 293A cells as previously described in Example 4.

Adenovirus production and purification

[0893] Ten T-175 flasks of 293A cells were plated in 25 ml of complete medium per flask (DMEM/10%FBS/L-Glutamine/non-essential amino acids/penicillin/streptomycin). On the day of infection, the cells were 80-90% confluent. The old media was removed and replaced with 25 ml of complete media containing sufficient virus for an MOI of 5 viruses per cell. Cultures were incubated overnight at 37°C. The next day, the media was replaced with 25 ml fresh media and the cells were incubated for 2-3 days until >80% cytopathic effect (CPE) was observed. CPE is obvious: the cells swell, round-up, and begin to detach from the plate. At this point, the cells were gently dislodged using a 50 ml pipette and pooled in 250 ml sterile conical bottles. Cultures were centrifuged at 1000 rpm for 10 minutes. The supernatant was discarded, the cell pellet was dissolved in 5 ml of PBS and transferred to a 15 ml polypropylene tube. Cells were lysed to release virus by three freeze/thaws (-80°C to 37°C). Care was taken not to leave sample at 37°C any longer than necessary to melt it, as virus degradation is accelerated at 37°C. After the freeze/thaw cycles, 150 µl was removed for the wild type assay (see below). The lysates were then treated with DOC (deoxycholate, sodium salt, Sigma-Aldrich, St. Louis, MO catalogue #D 6750) to increase the virus yields. A stock of 10% DOC was prepared in water (heat was required to get it all into solution) and DOC was added to the adenovirus lysate to a final concentration of 0.2%. The lysates were incubated at room temperature for 30 minutes on a rotating platform. Insoluble materials were eliminated by centrifugation (3000 rpm for 15 minutes in table top centrifuge), and the crude high titer viral supernatant (CHT) was transferred to a fresh tube. MgCl₂ was added to 5mM final and virus was stored at -80°C or cesium chloride purified (see below).

[0894] Cesium chloride step-gradient ultracentrifugation purifies and concentrates the recombinant adenovirus by eliminating cellular contaminants in order to achieve optimum efficiency and minimal toxicity of adenovirus

gene delivery. Two cesium chloride (Molecular Biology grade CsCl, Sigma-Aldrich, St. Louis, MO, catalog #C3032) solutions were prepared with the following densities: 1.4g/ml and 1.25g/ml in 10mM Tris (pH 8.0). [1.4g/ml = 38.83g CsCl + 61ml 10mM Tris and 1.25g/ml = 26.99g CsCl + 73ml 10mM Tris] These weight/volume densities were verified by weighing 1 ml of each solution. Density was adjusted by adding more cesium chloride or 10 mM Tris as needed to achieve correct density. Each solution was filter sterilized and stored at room temperature.

[0895] To prepare the step-gradient, 2.5ml of 1.25g/ml CsCl solution was placed in one ultracentrifuge tube (SW41 Beckman centrifuge tubes, thin wall polyallomer 14x89, part#331372) and then carefully underlayered with 2.5ml of 1.4g/ml CsCl solution. A long glass Pasteur pipette was used for underlayering. Next, the step-gradient was gently overlaid with the 5ml of crude high titer viral lysate and carefully move into Beckman SW41 centrifuge rotor. Samples were spun at 35,000 rpm for one hour and ten minutes at 20°C. After ultracentrifugation, cellular lipids and cytoplasmic debris remained at the top of the tube, and the cloudy adenovirus band migrated near the interface of the two CsCl layers. The virus band was very obvious to the naked eye. The tube was clamped to a ring stand and the sides of the tube were wiped with 70% ethanol. The virus band was harvested using a 3 ml syringe fitted with a 20 or 21 gauge needle by side puncture. Virus was transferred to a 15 ml conical tube and the volume estimated. Glycerol was added to 10% final.

[0896] The cesium chloride in the recombinant adenoviral preparation was removed by four rounds of dialysis. For each ultracentrifuge tube four liters of dialysis buffer was required. The dialysis buffer consisted of 10mM Tris (pH 7.5), 1mM MgCl₂, 150mM NaCl, and 10% glycerol final and was kept at 4°C. Buffer was prepared the day before dialysis and placed in the cold room with a stir bar overnight. The recovered viral band was dialyzed at 4°C using a Spectrum Spectra/Por CE Float-A-Lyzer with a molecular weight cut-off (MWCO) of 300,000 Dalton (Fischer 3 ml size #08-700-51 or 5 ml size #08-700-64). The virus preparation was dialyzed four times. Each dialysis was conducted for one hour and in one liter, with constant gentle stirring. The final virus product was removed from Float-A-Lyzer with the plastic pipette

provided and aliquotted into eppendorf tubes. Aliquots of virus were stored at -80°C and multiple freeze/thaws were avoided.

- [0897] Typical titers from a 10-flask cesium chloride preparation range from 7×10^9 to 6.5×10^{10} pfu/ml. The volume of purified recombinant adenovirus obtained is typically around 1.0 ml, making the total virus yield from 10-flasks to be 7×10^9 to 6.5×10^{10} pfu. This stock contains enough material for one 96-well plate using an MOI of 50.

Wild-Type Assay

- [0898] The "supernatant rescue assay" is performed to detect wild-type adenovirus contamination in recombinant adenovirus stocks using standard procedures (Dion *et al.*, *J. Virol. Methods* 56:99-107 (1996)).

Reporter Cell Line

- [0899] pcDNA6/FRT/V5 was digested with *PstI* and *PmeI* to remove the nucleic acid sequence encoding the V5 tag. pcDNA3.1 lacZ stop^{TAG} GFP was digested with *PstI* and *PmeI* to isolate the fragment containing lacZ stop^{TAG} GFP. The above fragments were gel purified, ligated, and transformed into TOP10 cells. The resulting reporter plasmid, pcDNA6/FRT/lacZ-stop^{TAG}-GFP, was verified by diagnostic digests and sequencing. FlpIn CHO cells (Invitrogen Corporation, Carlsbad, CA, catalog #R758-07) were co-transfected with the vector pcDNA6/FRT/lacZ stop^{TAG} GFP and pOG44 (Invitrogen Corporation, Carlsbad, CA, catalog #V6005-20) at a ratio of 1:10. Blasticidin selection was started 4 days post transfection at a concentration of 15 $\mu\text{g/ml}$. Selection was complete after 24 days.

Co-transfections

- [0900] Six-well plates were seeded with cells one day prior to transfections. Cells tolerate transfections best if seeded at a density that allowed for greater than 90% confluency the day of co-transfection. Co-transfections were conducted for a minimum of 5 hours and up to overnight. The lipid complexes were then removed and replaced with fresh media. Co-

transfections were optimized using 1.5 µg suppressor tRNA plasmid and 0.5 µg of the corresponding reporter vector combined in 250 µl of Opti-MEM® I Reduced Serum Medium (OPTI-MEM) at room temperature for 5-10 minutes. Six microliters of Lipofectamine™ 2000 was combined with 250 µl of OPTI-MEM and allowed to sit at room temperature for 5 minutes before combining with the DNA mixture. The DNA-lipid complex was allowed to form for 20 minutes at room temperature. Subsequently, the DNA-lipid complex was added to the cells in wells containing 2 ml of media. Suppression in a GFP fused expression vector could be observed the following day and up to 72 hours post transfection. Cells were typically lysed and harvested twenty-four hours post transfection with IGE PAL CA630 lysis buffer (Sigma-Aldrich, St. Louis, MO, catalog #I-3021) or RIPA lysis buffer (10mM Tris (8.0), 150mM NaCl, 0.1% SDS, 1.0% NP-40 (or Triton X-100), 1.0% deoxycholate, 2mM EDTA) with leupeptin, pepstatin, and PMSF.

Transductions

[0901] Cells were transduced with suppressor tRNA for a minimum of five hours to a maximum of overnight in a total of 1 ml media in a six well format. Upon completion of transduction the virus was removed and 2 ml of fresh media was added. The cells were then transfected overnight or the following day. Cells were typically lysed and harvested three days post transduction with IGE PAL CA630 lysis buffer or RIPA lysis buffer with leupeptin, pepstatin, and PMSF.

Westerns

[0902] Cell lysates were centrifuged at maximum speed for 1-2 minutes. Lysates were then transferred to new tubes and pellet discarded. A Bradford protein assay was conducted to determine the protein concentration of each lysate. For western blotting, 10-30 µg of protein was loaded on a gel. Determination of percent suppression was performed using 6% Tris Glycine gels for western blotting of β-galactosidase fusion proteins to maximize resolution of high molecular weight proteins. For CAT, GFP and V5 blots, 4-20% Tris Glycine gels were used. Proteins from gels were transferred to 0.45

µm nitrocellulose using western blotting technique. Various antibodies were used in detection of proteins: anti-βgal at 1:5000, anti-CAT at 1:5000, anti-GFP at 1:5000, anti-V5 at 1:5000. Western Breeze kits and antibodies from Invitrogen Corporation, Carlsbad, CA were used throughout.

QC assay for manufactured virus

[0903] Virus produced in manufacturing should be screened for wild type virus, cesium chloride purified and plaque-assay titered. For the activity assay, COS-7 cells (ATCC #CRL-1651) were seeded at a density of 3×10^5 cells in a 6 well format with 2 ml of DMEM containing 10%FBS, 1% L-glutamine, and 1% Pen/Strep. The following day, the media was aspirated and 1 ml was added back to the culture wells to be transduced. CsCl purified Ad-tRNA8^{TAG} was added to each well at an MOI of 0, 25, 50 and 100. The transductions were allowed to proceed for 5-6 hours at 37°C. Following the transduction period, the media containing the virus was removed and 2 ml of fresh media was added back to each well. The transduced cells were allowed a day to recover before transfection of reporter plasmid. For each transfected well, two micrograms of pcDNA6.2/GFP-GW/p48^{TAG} (or pcDNA3.1/lacZ-stop^{TAG}-GFP) expression plasmid was diluted in 250 µl of OPTI-MEM and incubated at room temperature for 5 minutes. 6 µl of Lipofectamine™ 2000 was diluted in 250 µl of OPTI-MEM and incubated at room temperature for 5 minutes. The DNA and lipid dilutions may also be set-up in batch for the four wells to be transfected. The DNA and lipid were then combined and incubated at room temperature for 20 minutes to complex before adding to COS-7 cells previously transduced with Ad-tRNA8^{TAG}. The DNA-lipid complex remained on the cells between 5 to 18 hours before being removed and fresh media added to the cells. GFP fluorescence was observed on days 1-3 post transduction. The cells were lysed and harvested with ice cold 150-200 µl of RIPA lysis buffer (10mM Tris (8.0), 150mM NaCl, 0.1% SDS, 1.0% NP-40 (or Triton X-100), 1.0% deoxycholate, 2mM EDTA) containing leupeptin, pepstatin, and PMSF on day 3 post transduction. The lysates were then centrifuged for 5 minutes at maximum speed (preferably at 4°C). Lysates were transferred to new 1.5 ml eppendorf tubes and frozen at -80°C if not used immediately for western blotting. Following western blotting for anti-

myc (or anti- β -galactosidase, depending on the expression plasmid used) densitometry was performed on the Fujifilm LAS-1000 Densitometer using the software Image Reader LAS-1000 Lite v1.0 and ImageGauge v.254. Percent suppression was calculated by dividing the density of the upper band by the total (lower plus upper band). For the purpose of this example, 50% suppression was the desired level of suppression.

Results and Discussion

[0904] All three possible human tRNA suppressors (TAG, TAA and TGA) were created by mutating the anticodon of the human tRNA serine gene (Capone *et al.*, *EMBO*, 4:213-221 (1985)). This work was performed in the laboratory of Dr. RajBhandary, who also provided the pUC12-based vectors containing each of the three tRNA^{ser} suppressors. Bacterial tRNA suppressors had been identified many years previously, but the use of a mammalian tRNA suppressor allows stop suppression in mammalian cells without the need to co-express the cognate tRNA charging enzyme.

[0905] The efficiency of each tRNA suppressor was tested in several co-transfection experiments (Figures 51A-B). Three GATEWAYTM entry clones were created, with CAT as the gene of interest (GOI), followed by each of the three stop codons (pENTR-CAT^{TAA}, pENTR-CAT^{TAG} and pENTR-CAT^{TGA}). These entry clones were LR crossed into either pcDNA3.2/V5-DEST or pcDNA6.2/GFP-DEST, thus placing either V5 or GFP downstream (and in frame) of the native CAT ORF. These Destination vectors also have all three stop codons, in frame, downstream of the C-terminal tag. Having all three stop codons assures termination of translation after the tag, regardless of which tRNA suppressor is used.

V5 Epitope Tag-On-DemandTM

[0906] CHO cells were co-transfected with one of these expression vectors: pcDNA3.2/V5-GW/CAT^{TAA}, -GW/CAT^{TAG} or -GW/CAT^{TGA} in the presence or absence of its cognate tRNA suppressor: pUC12-tRNA^{TAA}, pUC12-tRNA^{TAG} or pUC12-tRNA^{TGA} (Figure 51A). Western blot analysis using antibodies against the V-5 epitope revealed easily detectable V5-epitope-

tagged protein in the presence of tRNA suppressor (left panel), which was further illustrated by the “shift” up of CAT protein on the anti-CAT western blot (right panel). The efficiency of suppression can be calculated by using densitometry to scan the intensity of the shifted and un-shifted bands. In this experiment as well as others not described herein, the TAG stop suppressor was clearly superior to the other two, demonstrating a >70% conversion of native CAT to CAT-V5 in the presence of the suppressor (Figure 51A, right panel, anti-CAT blot), as compared to only 44% and 53% for TAA and TGA, respectively.

GFP Tag-On-Demand™

[0907] 293FT cells (Invitrogen Corporation, Carlsbad, CA, catalog #R700-07) were co transfected with one of the three expression vectors (pcDNA6.2/GFP-GW/CAT^{TAA}, -GW/CAT^{TAG} or -GW/CAT^{TGA}) and one of the pUC12-tRNA vectors (Figure 51B). Anti-CAT western blotting showed a clear shift of native CAT up to CAT-GFP when the correct tRNA was supplied. Again, tRNA^{TAG} demonstrated superior stop suppression compared to the other two tRNA suppressors. It is also important to note that the stop suppression and protein tagging is very specific. In other words, when the *incorrect* tRNA suppressor is supplied, *no* stop suppression is observed and only native protein is expressed (for example, see TAA CAT reporter with tRNA^{TAG} or tRNA^{TGA}, Figure 51B). The specificity of the suppression is further demonstrated with a different reporter vector, pcDNA3.1/lacZ-stop^{TAG}-GFP (Figure 52). Only in the presence of the correct tRNA suppressor (pUC12-tRNA^{TAG}) was the β -galactosidase-GFP fusion protein expressed resulting in detectable glowing in the cells (center panels, Figure 52). When either of the other suppressors is used (tRNA^{TAA} or tRNA^{TGA}), no suppression of the TAG stop occurs, no β -galactosidase-GFP is expressed and no glowing is observed in the transfected cells (left and right panels Figure 52).

Adenovirus-tRNA delivery

[0908] Since Tag-On-Demand™ is primarily designed for the transient tagging of proteins, an ideal delivery method of the suppressor gene to

mammalian cells is using a recombinant adenovirus. Adenovirus has a very broad tropism for different mammalian cell types and transduction efficiencies can approach 100% (for review see Russell 2000, Update on adenovirus and its vectors. *J. Gen. Virol.*, 81:2573-2604). Furthermore, since the virus does not stably integrate into the host genome, expression is transient. In actively dividing cells (24 hour doubling time), gene expression from adenoviral vectors is typically detected within 24 hours and persists for 7-8 days.

[0909] The tRNA^{TAG} gene was cloned into pENTR to create pENTR-tRNA^{TAG}, and this was used in a GATEWAYTM LR reaction with pAd/PL-DEST (Table 10, Figure 9) to create pAd-tRNA^{TAG}. Several large-scale preparations of virus were performed and functional testing was done. Adenovirus proved to be a very efficient way of delivering the tRNA, however preliminary experiments required MOIs (multiplicity of infection) of several hundred to deliver biologically relevant amounts of the tRNA. The goal was to achieve at least 50% suppression using an MOI of 50 in COS cells transfected with one of the reporter genes. It is believed that the tRNAs must compete with endogenous protein "stop factors" occupying the stop codon, which may explain the more efficient suppression in the presence of multiple copies of the nucleic acid molecule encoding the suppressor tRNA sequence. In an attempt to reduce the number of viral particles required for efficient suppression, eight copies of the tRNA gene were cloned into pENTR (called pENTR-tRNA8^{TAG}) and recombined into the adenovirus promoterless Destination vector. This new adenovirus (Adeno-tRNA8^{TAG}) was compared with the original monomer virus (Adeno-tRNA^{TAG}) for stop suppression (Figure 53). As shown by both fluorescent microscopy (upper panels) and anti-β-galactosidase western blotting (lower panel), a modest increase in suppression efficiency was observed with the 8-mer tRNA, and these suppression levels are as good as those seen with the plasmid-based tRNA (lanes 2 and 4). Indeed, in all subsequent experiments, the Ad-tRNA8^{TAG} transduction performed as well or better than a pUC-tRNA^{TAG} plasmid transfection making this recombinant adenovirus configuration particularly suitable for the methods of this invention.

[0910] The initial adenovirus experiments used crude adenovirus preparations that still contained all of the debris from the lysed producer cells (roughly 10⁸

cells lysed in 5 ml PBS). This material was functional but resulted in unacceptably high toxicity to the target cells. A variety of purification methods were evaluated to attempt to remove the toxic components from the active adenovirus. Large pore dialysis (300,000 MWCO), sucrose density gradient purification, and HPLC were evaluated for use in the methods of this invention, as were traditional cesium chloride purification and two commercially available adenovirus purification columns (ViraPur, Carlsbad, CA and PureSyn, Malvern, PA). It was deduced that a modified, single-round cesium chloride step gradient purification (described above) was the least expensive option that gave the highest yields of active virus and exhibited the lowest toxicity on the target cells, making this method particularly suitable for use in the methods of this invention.

Ultimate™ ORF collection Tag-On-Demand™

[0911] The invention described herein is compatible for use with any gene or ORF of interest providing the stop codon is recognized by the provided suppressor tRNA. This stop codon may be native to the gene of interest, or it may be inserted by standard molecular biology techniques, such as described herein. Particularly suited for use in the methods of this invention are clones in the Ultimate™ ORF collection available from Invitrogen Corporation, Carlsbad, CA. This collection of genes is provided as GATEWAY™ Entry clones containing the native ORF with a TAG stop codon. Tag-On-Demand™ will allow quick and easy detection of expressed protein products (either via V5 or GFP tagging) without needing to generate antibodies against the native protein or recloning the gene to a separate expression vector.

[0912] To demonstrate the usefulness of Tag-On-Demand™ with ORFs from the Ultimate™ ORF collection, three human ORFs were chosen and recombined into either pcDNA6.2/GFP-DEST or pcDNA6.2/V5-DEST. Transduction of cells with the Ad-tRNA8^{TAG} followed by transfection of the cells with the ORF-GFP expression clone resulted in easily visible GFP positive cells (Figure 54, upper panels). Significantly, the proteins retained their normal subcellular localization that was easily detectable using fluorescence microscopy. ORF6 (BC003357) codes for a CGI-130-like protein (23.4 kD) that is primarily cytoplasmic, with nuclear exclusion

observed in some cells. ORF7 (BC000997) codes for a human mRNA splicing factor (27.4 kD) and was clearly localized to the nucleus, as expected. ORF12 (BC000141) codes for a truncated form of c-myc (48.8 kD) that is also nuclear, with specific targeting to punctate nucleolar structures. ORF12 is referred to as p48^{TAG} above, and can be the positive expression control for use in kits, as provided in the methods of the present invention. In this example, this experiment was performed by first transducing the cells with adenovirus for 6 hours, followed by transfecting with the reporter plasmid overnight. Alternatively, transduction and transfection may be performed together, or, transfection first followed by transduction. All three methods resulted in good suppression, though transduction followed by transfection to may achieve the best suppression and the least toxicity.

[0913] V5 epitope tagging of the ORFs was also successful using the methods of the present invention. Each expressed protein was easily detectable via anti-V5 western blotting, in the presence of the tRNA^{TAG}, and migrated at the correct molecular weight (Figure 54, lower panel). The addition of the V5 epitope adds approximately 4.2 kD to the protein of interest. ORF-V5 expression levels were comparable to lacZ-stop^{TAG}-V5 suppressed with tRNA^{TAG}, and surprisingly as good as a true V5 fusion protein, GFP-V5, expressed constitutively from pcDNA/GFP-V5 (last lane). This experiment was performed by co-transfection of the ORF-V5 plasmid with pUC12-tRNA^{TAG}, however similar results are obtained using Ad-tRNA8^{TAG}.

[0914] It is an important aspect of the invention that ORF expression vectors such as these may express the native protein under normal conditions, allowing the study of its native function. Application of Tag-On-Demand™ allows the use of the exact same expression construct to transiently create tagged versions of the protein. This aspect may be useful for verification of protein expression, analysis of its subcellular localization and even FACSorting of expressing cells without having to generate antibodies to the specific protein or re-cloning the ORF as a true C-terminal fusion.

Tag-On-Demand™ can be used on both transient and stable gene targets

[0915] One aspect of the present invention is the transient expression of the protein of interest with a tag to verify expression or localization, as described

herein. Another aspect of the present invention is to stably express a protein of interest, as demonstrated by the following experiment. Flp-In CHO cells stably expressing a single copy of pcDNA6/FRT/lacZ-stop^{TAG}-GFP were transduced with Adeno-tRNA^{TAG} at various MOIs (Figure 55A). Anti-lacZ western blotting revealed a dose-dependent increase in stop suppression with increasing amounts of Adeno-tRNA^{TAG}, and clearly demonstrates that Tag-On-DemandTM can be used to C-terminally tag stably expressed genes. This experiment shows that C-terminally-tagged recombinant protein is produced at all MOIs tested. As the cells are transduced with increasing MOI, the % suppression increases; however, the amount of total recombinant protein produced (untagged and GFP-tagged protein) remains nearly equivalent. The band labeled with an asterisk results from the endogenous *lacZ-Zeocin*TM fusion present in Flp-InTM-CHO cells and is derived from the construct used to create the Flp-InTM-CHO cell line. In a parallel experiment, COS cells were transduced with the same range of MOIs for 6 hours, followed by transient transfection of the lacZ-stop^{TAG}-GFP expression vector (Figure 55B). A dose-dependent increase in suppression with increasing amount of virus was shown, and an MOI as low as 19 can give suppression levels greater than 50%, demonstrating the efficiency of the invention. This experiment shows that at all MOIs tested, the % suppression achieved is >60%, resulting in production of significant levels of GFP-tagged recombinant protein. As the cells are transduced with increasing MOI, the % suppression increases; however, the amount of total recombinant protein produced (untagged and GFP-tagged protein) decreases. This may be indicative of cellular toxicity as a result of the addition of increasing amounts of virus.

Tag-On-DemandTM in common mammalian cell types

[0916] Five commonly used mammalian cells were chosen to evaluate efficiency of Tag-On-DemandTM: BHK, CHO, COS, HeLa and HT1080. Cells were transduced with Adeno-tRNA^{TAG} at an MOI of 50 for 6 hours, followed by transient transfection with the lacZ-stop^{TAG}-GFP expression vector. In all cell types tested, Tag-On-DemandTM clearly produced sufficient lacZ-GFP fusion protein to easily detect GFP fluorescence in each cell type

(Figure 56). The slight toxicity observed in these experiments most likely arose from residual cesium chloride in the purified adenovirus preparation.

Summary

[0917] One embodiment of the present invention is exemplified by the Tag-On-Demand™ system. Tag-On-Demand™ is a system that uses recombinant adenovirus to deliver a tRNA suppressor gene that results in detection of proteins from the Invitrogen Corporation, Carlsbad, CA Ultimate™ ORF collection. As described in the results section, Tag-On-Demand™ is primarily designed for: a) transient detection and localization of expressed protein products, and b) sorting or analysis of expressing cells. Any of the three stop codons normally utilized by cells can be suppressed with the correct tRNA, with TAG stop suppression being most efficient. Adenovirus has been chosen as the tRNA delivery method and was shown to be as good or better than plasmid delivery. Adenovirus is an ideal method for delivering the tRNA genes for a number of reasons:

[0918] Adenovirus has broad mammalian cell tropism. Adenovirus binds to the CAR receptor present on most mammalian cells. It is important to note, however, that not all cell types express equal levels of the required CAR receptor, so efficiency may vary from one cell type to another. Fortunately, suppression efficiency can be increased by applying more virus (see Figures 55A-B). Like any E1 deleted recombinant adenoviral vectors, use of the Tag-On-Demand™ adenovirus in 293 cells or in any mammalian cell that is expressing the E1 gene of adenovirus will lead to virus replication and possible death of the target cell.

[0919] Adenovirus does not stably integrate into the target cell's genome and its expression is transient. Stable delivery or constitutive expression of the tRNA suppressor would most likely be toxic to the cell since one third of the endogenous stop codons would be suppressed, resulting in the addition of extra amino acids to the C-termini of many cellular proteins.

[0920] Adenovirus is a very stable virus and can be produced in large quantities. Manufacturing of the virus may include a single round of cesium chloride purification (see Materials and Methods), which yields a pure viral stock with minimal toxicity to the target cell.

[0921] In summary, the Tag-On-Demand™ system allows a single expression vector to express either native protein or C-terminally tagged protein. The system is completely compatible with the GATEWAY™ cloning technology and Ultimate™ ORF collection. The present invention is particularly suited for use with the pcDNA6.2/V5 and pcDNA6.2/GFP Destination vectors, however a person of skill in the art would readily recognize that a variety of other Invitrogen vectors as well as others are also compatible with Tag-On-Demand™, including many TOPO vectors, myc and 6X-His vectors, T-REx and Flp-In as described in the Materials and Methods. Further provided in the present invention is the ability to clone any downstream “tag” for fusing to any protein of interest, provided there is a non-TAG stop codon at the end of the C-terminal tag.

EXAMPLE 15

Tag-on-Demand™ GATEWAY® Vectors

[0922] In some embodiments, the present invention provides nucleic acid molecules (*e.g.*, vectors) that may be used to express fusion polypeptides (*e.g.*, polypeptides comprising a sequence of interest and at least one additional polypeptide sequence). Non-limiting examples of vectors suitable for use in the present invention are GATEWAY®-adapted destination vectors. Such vectors may be used for high-level expression of native and C-terminally-tagged polypeptides from the same nucleic acid molecules. In some embodiments, such vectors may be used to express polypeptides (which may be fusion polypeptides) in mammalian cells. Such vectors may be used to express fusion polypeptides by introducing the vectors into a host cell and also introducing into the host cell a source of a suppressor tRNA. Any suitable source of a suppressor tRNA may be used (*e.g.*, plasmids, linear nucleic acid molecules, viruses, etc.) In some embodiments, the present invention provides an adenovirus that expresses one or more suppressor tRNA molecules and/or one or more copies of a suppressor tRNA molecule. Methods that employ an adenovirus expressing tRNAs may be referred to herein as the Tag-on-Demand™ System.

[0923] One or more of the following commercially available items may be used in connection with the methods of the invention. These items are listed with their Invitrogen Corporation, Carlsbad, CA catalog number in parenthesis: Tag-On-Demand™ Suppressor Supernatant (K400-01 or K405-01); Gateway® LR Clonase™ Enzyme Mix (11791-019 or 11791-043); Library Efficiency® DB3.1™ Competent Cells (11782-018); One Shot® TOP10 Chemically Competent *E. coli* (C4040-03); Library Efficiency DH5α™ Chemically Competent *E. coli* (18263-012); Blasticidin (R210-01); Ampicillin (Q100-16); S.N.A.P.™ MidiPrep Kit (K1910-01); Lipofectamine™ 2000 (11668-027 or 11668-019); and Phosphate-Buffered Saline (PBS), pH 7.4 (10010-023).

[0924] In some embodiments, the present invention provides methods of producing fusion polypeptides comprising a polypeptide sequence of interest fused to one or more additional polypeptide sequences. If a fusion polypeptide is produced (e.g., from pcDNA™6.2/V5-DEST or pcDNA™6.2/GFP-DEST), the fusion polypeptide can be detected using an antibody that binds to one or more of the additional polypeptide sequences (e.g., to the V5 epitope or to GFP). Commercially available antibody preparations can be used, for example, those available from Invitrogen Corporation, Carlsbad, CA such as Anti-V5 Antibody (catalog # R960-25), Anti-V5-HRP Antibody (catalog # R961-25), Anti-V5-AP Antibody (catalog # R962-25), Anti-V5-FITC Antibody (catalog # R963-25), GFP Antiserum (catalog # R970-01).

[0925] In some embodiments, methods of the invention may be used to express a fusion polypeptide comprising all or a portion of p64. To detect the p64 (human c-myc) protein expressed using methods and materials of the invention, commercially available Anti-myc Antibodies may be used (e.g., Invitrogen Corporation, Carlsbad, CA Anti-myc Antibody catalog no. R950-25, Anti-myc-HRP Antibody catalog no. R951-25, Anti-myc-AP Antibody catalog no. R952-25, and/or Anti-myc-FITC Antibody catalog no. R953-25).

[0926] Examples of nucleic acid molecules that may be used in the practice of the invention include, but are not limited to, pcDNA™6.2/V5-DEST (7.3 kb) and pcDNA™6.2/GFP-DEST (8.0 kb), which are destination vectors adapted for use with GATEWAY® Technology (Invitrogen Corporation, Carlsbad, CA)

and allow high-level, constitutive expression of recombinant polypeptides in mammalian cells. The vectors are designed for use with a suppressor tRNA producing nucleic acid molecule (*e.g.*, Invitrogen's Tag-on-Demand™ System), which allows expression of both native and C-terminally-tagged recombinant polypeptide from the same expression construct.

[0927] The pcDNA™6.2/V5-DEST and pcDNA™6.2/GFP-DEST vectors enable expression of recombinant polypeptide containing a choice of C-terminal tags. The pcDNA™6.2/V5-DEST vector encodes the V5 epitope for detection of recombinant polypeptide using the Anti-V5 antibodies. A plasmid map is provided as Figure 57 and the sequence of this vector is provided as Table 28. The pcDNA™6.2/GFP-DEST vector encodes the Cycle-3 GFP for fusion to a polypeptide sequence of interest and use as a reporter gene. A plasmid map of this vector is provided as Figure 58 and the sequence of this vector is provided as Table 29.

[0928] The pcDNA™6.2/V5-DEST and pcDNA™6.2/GFP-DEST vectors contain the following features: human cytomegalovirus (CMV) immediate early promoter for high-level constitutive expression of the gene of interest in a wide range of mammalian cells (Andersson, S., *et al.*, *J. Biol. Chem.* 264:8222-8229 (1989); Boshart, M., *et al.*, *Cell* 41:521-530 (1985); Nelson, J.A., *et al.*, *Molec. Cell. Biol.* 7:4125-4129 (1987)); two recombination sites, *attR1* and *attR2*, downstream of the CMV promoter for recombinational cloning of the DNA sequence of interest from an entry clone; the chloramphenicol resistance gene (Cm^R) located between the two *attR* sites for counterselection; the *ccdB* gene located between the *attR* sites for negative selection; the C-terminal V5 epitope for detection of the recombinant polypeptide of interest (in pcDNA™6.2/V5-DEST only) (Southern, J.A., *et al.*, *J. Gen. Virol.* 72:1551-1557 (1991)); the C-terminal cycle-3 Green Fluorescent Protein (GFP) gene for fusion of the recombinant polypeptide of interest to a reporter (in pcDNA™6.2/GFP-DEST only) (Chalfie, M., *et al.*, *Science* 263:802-805 (1994); Crameri, A., *et al.*, *Nature Biotechnol.* 14:315-319 (1996)); the Herpes Simplex Virus thymidine kinase (TK) polyadenylation sequence for efficient transcription termination and polyadenylation of mRNA (Cole, C.N., and Stacy, T.P., *Mol. Cell. Biol.* 5:2104-2113 (1985)); the Blasticidin resistance gene for selection of stable cell

lines (Kimura, M., *et al.*, *Biochim. Biophys. ACTA* 1219:653-659 (1994)); the pUC origin for high-copy replication and maintenance of the plasmid in *E. coli*; the ampicillin (*bla*) resistance gene for selection in *E. coli*. In one alternative of this aspect of the invention, the chloramphenicol resistance gene in the cassette can be replaced by a spectinomycin resistance gene (see Hollingshead *et al.*, *Plasmid* 13(1):17-30 (1985), NCBI accession no. X02340 M10241), and the pcDNA destination vector containing attP sites flanking the *ccdB* and spectinomycin resistance genes can be selected on ampicillin/spectinomycin-containing media. Use of spectinomycin selection instead of chloramphenicol selection may result in an increase in the number of colonies obtained on selection plates, indicating that use of the spectinomycin resistance gene may lead to an increased efficiency of cloning from that observed using cassettes containing the chloramphenicol resistance gene.

[0929] The location in the plasmid sequence of pcDNATM6.2/V5-DEST (7341 nucleotides) of the features discussed above are: CMV promoter bases 232-819; T7 promoter/priming site bases 863-882; *attR1* site bases 911-1035; *ccdB* gene bases 1464-1769 (c); chloramphenicol resistance gene bases 2111-2770 (c); *attR2* site bases 3051-3175; V5 epitope bases 3201-3242; V5 reverse priming site 3210-3230; TK polyadenylation signal bases 3269-3540; fl origin 3576-4004; SV40 early promoter and origin 4031-4339; EM7 promoter bases 4394-4460; Blasticidin resistance gene bases 4461-4859; SV40 early polyadenylation signal bases 5017-5147; pUC origin bases 5530-6200 (c); Ampicillin (*bla*) resistance gene bases 6345-7205 (c); *bla* promoter bases 7206-7304 (c) where (c) indicates present on the complementary strand.

[0930] The location in the plasmid sequence of pcDNATM6.2/GFP-DEST (7995 nucleotides) of the features discussed above are: CMV promoter bases 232-819; T7 promoter/priming site bases 863-882; *attR1* site bases 911-1035; *ccdB* gene bases 1464-1769 (c); Chloramphenicol resistance gene bases 2111-2770 (c); *attR2* site bases 3051-3175; Cycle-3 GFP bases 3195-3908; GFP reverse priming site 3303-3324; TK polyadenylation signal bases 3923-4194; fl origin 4230-4658; SV40 early promoter and origin 4685-4993; EM7 promoter bases 5048-5114; Blasticidin resistance gene bases 5115-5513; SV40 early polyadenylation signal bases 5671-5801; pUC origin bases 6184-

6854 (c); Ampicillin (*bla*) resistance gene bases 6999-7859 (c); *bla* promoter bases 7860-7958 (c), where (c) indicates the feature is present on the complementary strand.

[0931] In some embodiments, positive control nucleic acid molecules (*e.g.*, plasmids) may be used in conjunction with the methods of the invention. A suitable positive control nucleic acid molecule is one comprising a nucleic acid sequence encoding two polypeptide sequences in the same reading frame and having a stop codon in between the sequences. For example, the polypeptide encoded 3' to the stop codon may have a detectable activity (*i.e.*, enzymatic activity, fluorescent activity, binding activity, etc.). Examples of suitable control nucleic acid molecules include, but are not limited to, pAd/CMV/V5-GW/*lacZ*, pcDNATM6.2/V5-GW/p64^{TAG} and pcDNATM6.2/GFP-GW-p64^{TAG}, which were prepared from the corresponding vectors by conducting an LxR reaction with an entry vector containing the indicated coding sequence (*i.e.*, *lacZ* or p64 coding sequence (also known as *c-myc*). Plasmid maps of the control vectors pcDNATM6.2/V5-GW/p64^{TAG} and pcDNATM6.2/GFP-GW-p64^{TAG} are provided as Figures 59 and 60 respectively.

[0932] The GFP gene used in the pcDNATM6.2/GFP-DEST vector is described in Cramer, A., *et al.*, *Nature Biotechnol.* 14:315-319 (1996). In this paper, the codon usage was optimized for expression in *E. coli* and three cycles of DNA shuffling were used to generate a mutant form of GFP that expresses well in mammalian cells and has excitation and emission maxima that are the same as wild-type GFP (395 nm and 478 nm for primary and secondary excitation, respectively, and 507 nm for emission) and a >40-fold increase in fluorescent yield over wild-type GFP. This mutant GFP is referred to as Cycle-3 GFP to differentiate it from wild-type GFP.

[0933] Materials and methods of the invention (*e.g.*, The Tag-on-DemandTM System, Invitrogen Corporation, Carlsbad, CA) facilitate transient expression of C-terminally-tagged and untagged recombinant polypeptides from a single expression construct such as one prepared using GATEWAYTM. The System is based on stop suppression technology originally developed by RajBhandary and colleagues (Capone, J.P., *et al.*, *EMBO J.* 4:213-221 (1985)), and consists of two major components: an expression vector into which the gene of interest will be cloned and a nucleic acid molecule (or composition comprising such a

nucleic acid molecule) expressing one or more suppressor tRNAs (*e.g.*, the Tag-on-Demand™ Suppressor Supernatant). The vector (*e.g.*, pcDNA™6.2/V5-DEST or pcDNA™6.2/GFP-DEST) must be in a configuration that is compatible with expression of C-terminally-tagged recombinant polypeptide by introducing a suppressor tRNA to suppress a stop codon (*e.g.*, by using the Tag-on-Demand™ System). In one non-limiting embodiment, (*i.e.*, the Tag-on-Demand™ Suppressor Supernatant) a suppressor tRNA molecule may be introduced into a host cell by transducing the host cell with a replication-incompetent adenovirus containing the human tRNA^{ser} suppressor. This tRNA suppressor has been mutated to recognize the TAG (amber stop) codon and decode it as a serine. When added to mammalian cells, the Tag-on-Demand™ Suppressor Supernatant is transduced and provides a transient source of the tRNA^{ser} suppressor.

[0934] When an expression vector encoding a gene of interest with the TAG stop codon is transfected into mammalian cells, the stop codon will be translated as serine, allowing translation to continue through any downstream reading frame (*e.g.*, a C-terminal tag), and resulting in production of a fusion polypeptide containing the polypeptide encoded by the gene of interest fused to the amino acids encoded 3' to the stop codon (*e.g.*, a marker or tag sequence). One skilled in the art will appreciate that, in similar fashion, a nucleic acid molecule (*e.g.*, a replication-incompetent adenovirus) expressing a suppressor tRNA that suppresses TAA (ochre) or TGA (opal) stop codons can be prepared and used in the practice of the present invention.

[0935] To recombine a DNA sequence of interest into a nucleic acid molecule of the invention (*e.g.*, pcDNA™6.2/V5-DEST or pcDNA™6.2/GFP-DEST), an entry clone containing the DNA comprising a sequence of interest may be prepared. In an entry clone, a sequence of interest may be flanked by recombination sites (*e.g.*, sites compatible with those in one or more destination vector). Many entry vectors are available from Invitrogen to facilitate generation of entry clones. Examples include, but are not limited to, pENTR/D-TOPO® (catalog number K2400-20), pENTR/SD/D-TOPO® (catalog number K2420-20), pENTR™1A (catalog number 11813-011), pENTR™2B (catalog number 11816-014), pENTR™3C (catalog number

11817-012), pENTRTM4 (catalog number 11818-010), and pENTRTM11 (catalog number 11819-018).

[0936] In some embodiments, the present invention encompasses the expression of fusion polypeptides comprising all or a portion of a human polypeptide. One suitable source of nucleic acid molecules encoding human polypeptides is the UltimateTM Human ORF (hORF) Clone collection available from Invitrogen Corporation, Carlsbad, CA. To express a human gene of interest from pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST, an UltimateTM Human ORF (hORF) Clone available from Invitrogen Corporation, Carlsbad, CA can be used. Each UltimateTM hORF Clone is a fully-sequenced clone provided in a GATEWAY® entry vector that is ready-to-use in an LR recombination reaction with pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST. In addition, each UltimateTM hORF Clone contains a TAG stop codon, making it fully compatible for use in the Tag-on-DemandTM System. For more information about the UltimateTM hORF Clones available, see the Invitrogen Corporation, Carlsbad, CA Web site or contact Invitrogen Corporation, Carlsbad, CA.

[0937] When generating an entry clone, a nucleic acid sequence encoding a polypeptide of interest in the entry clone must contain an ATG initiation codon in the context of a Kozak consensus sequence for proper initiation of translation in mammalian cells as discussed above.

[0938] To enable expression of both a native and C-terminally-tagged recombinant polypeptide of interest from pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST using the Tag-on-DemandTM System, the gene of interest in the entry clone may contain a stop codon. This stop codon may be encoded by the nucleotides, TAG. In addition, the gene should be in frame with the C-terminal tag after recombination. Those skilled in the art will appreciate that other stop codons can be similarly used by constructing a vector expressing a suppressor tRNA that recognizes the other stop codons.

[0939] The recombination region of pcDNATM6.2/V5-DEST and pcDNATM6.2/GFP-DEST are provided as Figures 61A and 61B respectively. In Figure 61A, shaded regions correspond to those DNA sequences transferred from the entry clone into the pcDNATM6.2/V5-DEST vector by recombination. Non-shaded regions are derived from the pcDNATM6.2/V5-DEST vector. The

sequences encoded by the gene of interest are boxed. To facilitate use with the Tag-on-Demand™ System, a gene of interest must contain a TAG stop codon and be in-frame with the C-terminal tag. Bases 918 and 3161 of the pcDNA™6.2/V5-DEST sequence are marked. Note that TAA and TGA stop codons are included downstream of the V5 epitope to allow translation termination in the Tag-on-Demand™ System. In Figure 61B, the recombination region of the expression clone resulting from pcDNA™6.2/GFP-DEST x entry clone is shown. The shaded regions correspond to those DNA sequences transferred from the entry clone into the pcDNA™6.2/GFP-DEST vector by recombination. Non-shaded regions are derived from the pcDNA™6.2/GFP-DEST vector. The sequences encoded by the gene of interest are boxed. To facilitate use with the Tag-on-Demand™ System, the gene of interest should contain a TAG stop codon. Bases 918 and 3161 of the pcDNA™6.2/GFP-DEST sequence are marked. TAA and TGA stop codons are included downstream of the GFP gene to allow translation termination in the Tag-on-Demand™ System (not shown).

[0940] To generate an expression clone: an LR recombination reaction using the *attL*-containing entry clone and the *attR*-containing pcDNA™6.2/V5-DEST or pcDNA™6.2/GFP-DEST vector may be performed. Both the entry clone and the destination vector may be supercoiled or linear. After the LR reaction has been performed, all or a portion of the reaction mixture may be used to transform a suitable *E. coli* host. The expression clones can be selected for using ampicillin and/or blasticidin.

[0941] The pcDNA™6.2/V5-DEST and pcDNA™6.2/GFP-DEST vectors are supplied as supercoiled plasmids. Although the GATEWAY® Technology manual has previously recommended using a linearized destination vector for more efficient recombination, it has been found that linearization of pcDNA™6.2/V5-DEST and pcDNA™6.2/GFP-DEST is not required to obtain optimal results for any downstream application.

[0942] Nucleic acid molecules of the invention, (*e.g.*, destination vectors) may be lyophilized for long term storage. Lyophilized plasmids may be resuspended in a suitable buffer (*e.g.*, TE, pH 8.0). In some embodiments, the vectors may be lyophilized in a buffer (*e.g.*, TE, pH 8.0) and may be resuspended by the addition of sterile water. A suitable concentration for

solutions of nucleic acid molecules to be used in the practice of the invention is about 150 ng/ μ l although other concentrations may be used.

[0943] In some embodiments, nucleic acid molecules of the invention may be propagated in suitable host cells. To propagate and maintain the pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST vectors, Library Efficiency[®] DB3.1TM Competent Cells (Invitrogen Corporation, Carlsbad, CA Catalog no. 11782-018) can be used. The DB3.1TM *E. coli* strain is resistant to CcdB effects and can support the propagation of plasmids containing the *ccdB* gene. To maintain integrity of the vector, select for transformants in media containing 50-100 μ g/ml ampicillin and 15-30 μ g/ml chloramphenicol. General *E. coli* cloning strains including TOP10 or DH5 α should not be used for propagation and maintenance as these strains are sensitive to CcdB effects.

[0944] Once an entry clone containing a gene of interest has been prepared, perform an LR recombination reaction between the entry clone and pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST, and transform the reaction mixture into a suitable *E. coli* host. A negative control (no entry vector) is recommended to help evaluate results. Any *recA*, *endA* *E. coli* strain including TOP10, DH5 α TM, or equivalent for transformation can be used. Do not transform the LR reaction mixture into *E. coli* strains that contain the F' episome (e.g., TOP10F'). These strains contain the *ccdA* gene and will prevent negative selection with the *ccdB* gene.

[0945] The pcDNATM6.2/V5-DEST and pcDNATM6.2/GFP-DEST vectors contain the ampicillin and Blasticidin resistance genes to allow selection of *E. coli* transformants using ampicillin or Blasticidin, respectively. To select for transformants using Blasticidin, use Low Salt LB agar plates containing 100 μ g/ml Blasticidin. For Blasticidin to be active, the salt concentration of the medium must remain low (<90 mM) and the pH must be 7.0. Low salt plates may be prepared by mixing 10 g Tryptone, 5 g NaCl, 5 g Yeast Extract and adding deionized, distilled water to 950 ml. Adjust pH to 7.0 with 1 N NaOH. Bring the volume up to 1 liter. For plates, add 15 g/L agar before autoclaving. Autoclave on liquid cycle at 15 psi and 121°C for 20 minutes. Allow the medium to cool to at least 55°C before adding the blasticidin to 100 μ g/ml final concentration. Store plates at +4°C in the dark. Plates containing

blasticidin are stable for up to 2 weeks. Blasticidin is available from Invitrogen Corporation, Carlsbad, CA.

[0946] An LR recombination reaction may be performed with purified plasmid DNA of an entry clone (50-150 ng/ μ l in TE, pH 8.0); pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST vector (150 ng/ μ l in TE, pH 8.0); LR ClonaseTM enzyme mix (Invitrogen, Catalog no. 11791-019; keep at -80°C until immediately before use); 5X LR ClonaseTM Reaction Buffer (supplied with the LR ClonaseTM enzyme mix); TE Buffer, pH 8.0 (10 mM Tris-HCl, pH 8.0, 1 mM EDTA); 2 μ g/ μ l Proteinase K solution (supplied with the LR ClonaseTM enzyme mix; thaw and keep on ice until use); an appropriate competent *E. coli* host and growth media for expression; SOC Medium; and selective plates (e.g., LB agar plates containing 100 μ g/ml ampicillin or Low Salt LB plates containing 100 μ g/ml Blasticidin).

[0947] Add the following components to 1.5 ml microcentrifuge tubes at room temperature and mix.

Component	Sample	Negative Control
Entry clone (100-300 ng/reaction)	1-10 μ l	--
Destination vector (300 ng/reaction)	2 μ l	2 μ l
5X LR Clonase TM Reaction Buffer	4 μ l	4 μ l
TE Buffer, pH 8.0	to 16 μ l	10 μ l

[0948] Remove the LR ClonaseTM enzyme mix from -80°C and thaw on ice (~ 2 minutes). Vortex the LR ClonaseTM enzyme mix briefly twice (2 seconds each time). To each sample above, add 4 μ l of LR ClonaseTM enzyme mix. Mix well by pipetting up and down. Return LR ClonaseTM enzyme mix to -80°C immediately after use. Incubate reactions at 25°C for 1 hour. Extending the incubation time to 18 hours typically yields more colonies. Add 2 μ l of the Proteinase K solution to each reaction. Incubate for 10 minutes at 37°C. Transform 1 μ l of the LR recombination reaction into a suitable *E. coli* host (follow the manufacturer's instructions) and select for expression clones. The LR reaction may be stored at -20°C for up to 1 week before transformation, if desired.

[0949] If *E. coli* cells with a transformation efficiency of 1×10^8 cfu/mg are used, the LR reaction should give approximately >5,000 colonies if the entire transformation is plated.

- [0950] The *ccdB* gene mutates at a very low frequency, resulting in a very low number of false positives. True expression clones will be ampicillin-resistant and chloramphenicol-sensitive. Transformants containing a plasmid with a mutated *ccdB* gene will be ampicillin- and chloramphenicol-resistant. To check a putative expression clone, test for growth on LB plates containing 30 µg/ml chloramphenicol. A true expression clone should not grow in the presence of chloramphenicol.
- [0951] To confirm that a gene of interest is in the correct orientation and in frame with the C-terminal fusion tag, the expression construct can be sequenced. The following primers can be used to sequence an expression construct. Figures 61A and 61B provide the location of the primer binding sites in each vector. For sequencing the pcDNATM6.2/V5-DEST vector, an oligonucleotide that binds to the T7 promoter/priming site (*e.g.*, 5'-TAATACGACTCACTATAGGG-3') and an oligonucleotide that binds to the V5(C-term) reverse priming site (*e.g.*, 5'-ACCGAGGAGAGGGTTAGGGAT-3') can be used. To sequence the pcDNATM6.2/GFP-DEST vector, an oligonucleotide that binds to the T7 promoter/priming site (*e.g.*, 5'-TAATACGACTCACTATAGGG-3') and an oligonucleotide that binds to the GFP reverse priming site (*e.g.*, 5'-GGGTAAGCTTTCCGTATGTAGC-3') can be used.
- [0952] Once an expression clone has been prepared, plasmid DNA for transfection may be prepared. Plasmid DNA for transfection into eukaryotic cells must be very clean and free from phenol and sodium chloride. Contaminants will kill the cells, and salt will interfere with lipid complexing, decreasing transfection efficiency. Plasmid DNA can be isolated using the S.N.A.P.TM MidiPrep Kit (Invitrogen Corporation, Carlsbad, CA Catalog no. K1910-01) or CsCl gradient centrifugation.
- [0953] For established cell lines (*e.g.*, COS, HeLa), consult original references or the supplier of the cell line for the optimal method of transfection. It is recommended that the protocol developed for individual cell lines be followed. Factors that may influence transfection efficiencies include medium requirements, when to pass the cells, and at what dilution to split the cells. Further information is provided in *Current Protocols in Molecular Biology*

(Ausubel, F.M., *et al.*, *Current Protocols in Molecular Biology*, Greene Publishing Associates and Wiley-Interscience, New York (1994)).

[0954] Methods for transfection include calcium phosphate (Chen, C., and Okayama, H., *Mol. Cell. Biol.* 7:2745-2752 (1987); Wigler, M., *et al.*, *Cell* 11:223-232 (1977)), lipid-mediated (Felgner, P.L., *et al.*, *Proc. West. Pharmacol. Soc.* 32:115-121 (1989); Felgner, P.L., and Ringold, G.M., *Nature* 337:387-388 (1989)) and electroporation (Chu, G., *et al.*, *Nucleic Acids Res.* 15:1311-1326 (1987); Shigekawa, K., and Dower, W.J., *BioTechniques* 6:742-751 (1988)). If a cationic lipid-based reagent for transfection is used, one suitable reagent is Lipofectamine™ 2000 Reagent available from Invitrogen Corporation, Carlsbad, CA (Catalog no. 11668-027). Other suitable transfection reagents may also be used.

[0955] pcDNA™6.2/V5-GW/p64^{TAG} or pcDNA™6.2/GFP-GW/p64^{TAG} is provided as a positive control vector for mammalian cell transfection and expression and may be used to optimize recombinant protein expression levels in a particular cell line. These vectors allow expression of native or C-terminally-tagged recombinant human c-myc (p64) protein that may be detected by Western blot. If using these vectors as expression controls, be aware that the p64 protein is naturally associated with nucleolar structures and requires ionic detergents (RIPA or SDS gel loading buffer) to adequately solubilize in total cell lysates prior to western blot analysis.

[0956] To propagate and maintain each of the control plasmids resuspend the vector in 10 µl sterile water to prepare a 1 µg/µl stock solution and use the stock solution to transform a *recA*, *endA* *E. coli* strain like TOP10, DH5α™, or equivalent. Transformants can be selected on LB agar plates containing 100 µg/ml ampicillin or Low Salt LB agar plates containing 100 µg/ml Blasticidin. A glycerol stock of a transformant containing plasmid can be prepared for long-term storage.

[0957] The methods described herein (*e.g.*, the Tag-on-Demand™ System) can be used to express both native and C-terminally-tagged recombinant polypeptide in mammalian cells from the same pcDNA™6.2/V5-DEST or pcDNA™6.2/GFP-DEST expression construct. To use the Tag-on-Demand™ System, add the Tag-on-Demand™ Suppressor Supernatant to mammalian cells at a specified time

- [0958] In some embodiments, particularly those in which an adenovirus is used to transduce a host cell in order to express a suppressor tRNA, the host cell may be transduced with the adenovirus followed immediately by transfection with the expression construct containing a sequence of interest encoding a polypeptide of interest. Embodiments of this type may be used to quickly screen for expression (or localization, if possible) of a recombinant polypeptide or to screen for expression of a large number of polypeptides. Embodiments of this type will be discussed in greater detail in the following example.
- [0959] In some embodiments, it may be desirable to generate a stable cell line comprising a nucleic acid molecule encoding a polypeptide of interest. In embodiments of this type, a nucleic acid molecule encoding a suppressor tRNA may be introduced into the stable cell line to produce fusion polypeptides comprising the polypeptide of interest fused to an additional polypeptide sequence (*e.g.*, a tag sequence, etc.). For example, a stable cell line may be transduced with an adenoviral vector expressing one or more suppressor tRNAs (*e.g.*, the Tag-On-Demand™ Suppressor Supernatant) to produce a C-terminally-tagged recombinant polypeptide.
- [0960] In some embodiments (*e.g.*, the Tag-on-Demand™ Suppressor Supernatant), nucleic acid molecules of the invention may be purified, titered, replication-incompetent, recombinant adenoviruses containing a human tRNA^{TAG} suppressor. Transduction of the adenovirus into mammalian cells facilitates transient stop suppression at the TAG codon in a gene of interest, enabling production of C-terminally-tagged recombinant polypeptide.
- [0961] In some embodiments (*e.g.*, the Tag-on-Demand™ Suppressor Supernatant), a nucleic acid molecules of the invention may be recombinant adenovirus that is deleted in the E1 region. Such an adenovirus is replication-incompetent in any mammalian cells that do not express the E1 proteins. Using such adenoviruses in 293 cells or in any cell line that expresses the adenovirus E1 gene (Graham, F.L., *et al.*, *J. Gen. Virol.* 36:59-74 (1977); Kozarsky, K.F., and Wilson, J.M., *Curr. Opin. Genet. Dev.* 3:499-503 (1993); Krougliak, V., and Graham, F.L., *Hum. Gene Ther.* 6:1575-1586 (1995)) results in viral replication and will lead to rapid death of the target cell within 1-2 days after infection.

[0962] Using methods of the invention, fusion polypeptides may be expressed transiently or stably. To express a recombinant fusion polypeptide transiently, nucleic acid molecules encoding the fusion polypeptide of interest and encoding and nucleic acid molecules encoding a suppressor tRNA may be introduced into a host cell. One skilled in the art will appreciate that the sequences encoding the fusion polypeptide of interest and the suppressor tRNA may be on the same or different nucleic acid molecules. In embodiments where an adenovirus is used to express a suppressor tRNA, cells may be transduced with the adenovirus and then transfected with the expression construct (*i.e.* nucleic acid molecule encoding the fusion polypeptide).

[0963] To express a recombinant fusion polypeptide from a stable cell line, a stable cell line comprising a nucleic acid molecule encoding the fusion polypeptide of interest may be created using any standard technique or one or more of the techniques described herein (*e.g.*, using lentiviral vectors or transfecting the mammalian cell line with the pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST expression construct, etc.). A nucleic acid molecule encoding a suppressor tRNA (*e.g.*, an adenovirus expressing a suppressor tRNA) may be introduced into the stable cell line to produce a fusion polypeptide.

[0964] In some embodiments, nucleic acid molecules of the invention (*e.g.*, pcDNATM6.2/V5-DEST and pcDNATM6.2/GFP-DEST vectors) may contain one or more selectable markers that may be used to select for stable cell lines. In one embodiment, nucleic acid molecules of the invention may contain the Blasticidin resistance gene to allow selection of stable cell lines. Some methods of the invention may entail creating stable cell lines by transfecting a construct into a mammalian cell line of choice and selecting for foci using Blasticidin. Methods of creating stable cell lines may also comprise linearizing a nucleic acid molecule of the invention (*e.g.*, pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST expression constructs) before transfecting them into a host cell. While linearizing the vector may not improve the efficiency of transfection, it increases the chances that the vector does not integrate in a way that disrupts elements necessary for expression in mammalian cells. Linearizing may comprise digesting the construct with a

restriction enzyme that cuts at a unique site that is not located within a critical element or within the gene of interest.

[0965] In some embodiments, methods of generating a stable cell line expressing a polypeptide of interest may comprise determining the minimum concentration of Blasticidin required to kill the untransfected host cell line by performing a kill curve experiment using any one of the protocols described herein. Typically, concentrations ranging from 2.5 to 10 $\mu\text{g/ml}$ Blasticidin are sufficient to kill most untransfected mammalian cell lines.

[0966] Once the appropriate Blasticidin concentration to use for selection has been determined, a stable cell line expressing a fusion polypeptide of interest (e.g., a pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST construct) can be generated. Methods of creating a stable cell line may comprise transfecting a mammalian cell line of interest with a nucleic acid molecule of the invention (e.g., a pcDNATM6.2/V5-DEST or pcDNATM6.2/GFP-DEST construct) using any transfection method of choice and selecting a stable cell line. Selecting may comprise 24 hours after transfection, washing the cells and adding fresh growth medium. 48 hours after transfection, splitting the cells into fresh growth medium such that they are no more than 25% confluent. If the cells are too dense, the antibiotic will not kill the cells. Antibiotics work best on actively dividing cells. Selecting may further comprise incubating the cells at 37°C for 2-3 hours until they have attached to the culture dish, removing the growth medium and replacing with fresh growth medium containing Blasticidin at the predetermined concentration required for the cell line. Methods of creating a stable cell line may also comprise feeding the cells with selective media every 3-4 days until Blasticidin-resistant colonies can be identified. Pick at least 5 Blasticidin-resistant colonies and expand them to assay for recombinant polypeptide expression.

[0967] Methods of the invention may comprise detecting a fusion polypeptide of the invention. For example, V5 fusion polypeptides expressed from pcDNATM6.2/V5-DEST can be detected using Western blot, immunofluorescence, or a functional assay specific the polypeptide of interest. A time course of expression may be prepared to optimize expression of the recombinant polypeptide (e.g. 24, 48, 72 hours, etc.). Anti-V5 Antibodies are available from Invitrogen Corporation, Carlsbad, CA and can be used to detect

V5-tagged recombinant fusion polypeptides: For Western blot analysis, the Anti-V5-Horseradish Peroxidase (HRP) Antibody or the Anti-V5-Alkaline Phosphatase (AP) Antibody may be used for detection. For immunofluorescence, the Anti-V5-Fluorescein Isothiocyanate (FITC) Antibody can be used for detection.

[0968] Methods of detecting a fusion polypeptide may comprise performing a Western blot. Such a method may comprise preparing a cell lysate from transfected cells. Any suitable protocol for preparing a cell lysate known to those skilled in the art may be used. Preparing a cell lysate may comprise washing cell monolayers (*e.g.*, $\sim 5 \times 10^5$ to 1×10^6 cells may be washed once with Phosphate-Buffered Saline, PBS, Invitrogen Corporation, Carlsbad, CA, Catalog no. 10010-023). Preparing a cell lysate may further comprise scraping cells into a buffer and centrifuging the cells. For example, cells may be scraped into 1 ml PBS and cells may be centrifuged at $1500 \times g$ for 5 minutes to form a cell pellet. Methods of preparing a cell lysate may comprise re-suspending a cell pellet in a lysis buffer. For example, cells may be re-suspended in 50 μ l Cell Lysis Buffer (*e.g.*, 50 mM Tris, pH 7.8, 150 mM NaCl, 1% Nonidet P-40). Other cell lysis buffers known to those skilled in the art are also suitable. Re-suspending may comprise mixing (*e.g.*, vortexing) the cell pellet in the lysis buffer to form a cell suspension and incubating the cell suspension (for example, at 37°C for 10 minutes) under conditions suitable to lyse the cells. Cells may be lysed at room temperature or on ice if degradation of polypeptide is a potential problem. Methods of preparing a cell lysate may further comprise centrifuging the cell lysate, for example, at $10,000 \times g$ for 10 minutes at $+4^\circ\text{C}$ to pellet nuclei and transferring the supernatant to a fresh tube.

[0969] Lysates prepared according to the invention may be further analyzed using techniques well known in the art, for example, lysates may be assayed for protein concentration. Those skilled in the art will appreciate that protein assays utilizing Coomassie Blue or other dyes should not be used if the lysis buffer comprises NP-40 since NP-40 interferes with the binding of the dye with the protein.

[0970] Methods of performing a Western blot may comprise mixing an aliquot of a cell lysate with an SDS-PAGE. For example, SDS-PAGE sample buffer

can be added to cell lysate to form a mixture and the mixture may be boiled, for example, for 5 minutes. An amount of the mixture comprising about 20 µg of protein may be loaded onto an SDS-PAGE gel and electrophoresed. One skilled in the art can select the appropriate concentration of acrylamide to be used to prepare the gel based upon the expected size of the fusion polypeptide.

[0971] One skilled in the art will recognize that a C-terminal tag containing the *attB2* site and the V5 epitope will add approximately 4 kDa to the polypeptide of interest. Fusion polypeptides of the invention may also comprise additional amino acids located between the polypeptide of interest and the additional polypeptide sequence (*e.g.*, a tag sequence such as the V5 epitope).

[0972] In some embodiments, methods of the invention may comprise detecting the presence of a fusion polypeptide comprising all or a portion of the p64 polypeptide. Methods of this type (*e.g.*, fusion polypeptides expressed from the pcDNATM6.2/V5-GW/p64TAG control), may utilize any suitable detection means, for example, any of the Anti-V5 Antibodies and/or anti-myc antibodies discussed above. Methods of preparing a cell lysate from a cell expressing a fusion polypeptide comprising all or a portion of p64 may comprise the use of harsher extraction conditions since procedures using NP-40 lysis are not effective in releasing p64 protein. Since p64 is localized in the nucleoli, harsher lysis procedures using RIPA or SDS-PAGE sample buffer are required to adequately solubilize p64 in total cell lysates. Methods of this type may comprise washing cell monolayers, for example, once with Phosphate-Buffered Saline (PBS, Invitrogen Corporation, Carlsbad, CA Catalog no. 10010-023). Methods may further comprise add 1X SDS-PAGE Sample Buffer to each well containing cells. 1 X SDS-PAGE buffer can be prepared by mixing 2.5 ml 0.5 M Tris-HCl, pH 6.8, 2 ml of glycerol (100%), 0.4 ml of β-mercaptoethanol, 0.02 g Bromophenol Blue, 0.4 g SDS and enough sterile water to bring the volume to 20 ml. For a 24-well plate, use 100 µl of 1X SDS-PAGE Sample Buffer per well. Methods may further comprise removing the cells from the plate, for example, a pipette tip may be used to loosen lysed cells from plate. Lysed cells may be transferred to a 1.5 ml microcentrifuge tube. Lysates prepared according to this method are typically viscous. Methods may further comprise heating samples, for

example, at 70°C for 10 minutes and mixing samples, for example, by vortexing every few minutes and briefly centrifuging the sample.

[0973] Methods may further comprise loading an aliquot of the cell lysate, for example, 5 µl of cell lysate, onto an SDS-PAGE gel and electrophoresing. One skilled in the art will appreciate that the V5-tagged p64^{TAG} protein has a molecular weight of approximately 53 kDa.

[0974] To detect the polypeptides expressed from as cycle-3 GFP fusion polypeptides from pcDNATM6.2/GFP-DEST, fluorescence, Western blot analysis, or a functional assay specific for the polypeptide of interest may be used. A time course may be prepared to optimize expression of the recombinant polypeptide (e.g. 24, 48, 72 hours, etc.). Any suitable technique, including those discussed herein, may be used to evaluate expression.

[0975] Cycle-3 GFP fusion polypeptides may be detected *in vivo* using fluorescence microscopy. The CMV promoter used to control expression of the cycle-3 GFP fusion polypeptide from pcDNATM6.2/GFP-DEST is a strong promoter and typically cycle-3 GFP fluorescence may be detected about 24 hours after transfection or transduction.

[0976] Methods of the invention may comprise methods of detecting fluorescent cells. In the practice of such methods, it is important to pick the best filter set to optimize detection. The primary excitation peak of cycle-3 GFP is at 395 nm. There is a secondary excitation peak at 478 nm. Excitation at either of these wavelengths yields a fluorescent emission peak with a maximum at 507 nm. Note that the quantum yield can vary as much as 5- to 10-fold depending on the wavelength of light that is used to excite the GFP fluorophore.

[0977] Use of the best filter set will insure that the optimal regions of the cycle-3 GFP spectra are excited and passed. Suitable filter sets include those designed to detect fluorescence from wild-type GFP (e.g., Omega Optical XF76 filter; see www.omegafilters.com). FITC filter sets may be used to detect cycle-3 GFP fluorescence, but note that these are not optimal and fluorescent signal may be weaker. For example, a FITC filter set may excite cycle-3 GFP with light from 460 to 490 nm, covering the secondary excitation peak and pass light from 515 to 550 nm. A set of this type may allow detection of most but not all of the cycle-3 GFP fluorescence.

- [0978] Most tissue culture media fluoresce because of the presence of riboflavin (Zylka, M.J., and Schnapp, B.J., *BioTechniques* 21:220-226 (1996)) and may interfere with detection of cycle-3 GFP fluorescence. Medium can be removed and replaced with Phosphate-Buffered Saline (PBS, Invitrogen Corporation, Carlsbad, CA, Catalog no. 10010-023) during the assay to alleviate this problem. If cells will be cultured further after assaying, remove the PBS and replace with fresh growth medium prior to re-incubation.
- [0979] To detect expression of a cycle-3 GFP fusion polypeptide by Western blot, an antibody to the polypeptide of interest or an antibody to cycle-3 GFP may be used. GFP Antiserum is available separately from Invitrogen Corporation, Carlsbad, CA (Catalog no. R970-01) for detection. The GFP Antiserum is a purified, polyclonal rabbit antiserum raised against recombinant cycle-3 GFP, and can detect both cycle-3 GFP and wild-type GFP protein.
- [0980] The C-terminal tag containing the *attB2* site and cycle-3 GFP will add approximately 28.3 kDa to the size of the fusion polypeptide. Fusion polypeptides of the invention may further comprise additional amino acids located between the polypeptide of interest and cycle-3 GFP.

EXAMPLE 16

- [0981] In some embodiments, the present invention provides materials and methods for the expression of fusion polypeptides. In one aspect, the same nucleic acid molecule is used to express a polypeptide of interest and a fusion polypeptide comprising the polypeptide of interest. In some aspects, this is accomplished by introducing into a host cell a nucleic acid molecule encoding a fusion polypeptide comprising a polypeptide of interest in the same reading frame as an additional polypeptide sequence. Typically, the nucleic acid molecule encoding the fusion polypeptide may comprise one or more stop codons, one of which may be located between the portion of the nucleic acid sequence encoding the polypeptide of interest and the portion of the nucleic acid sequence encoding the additional polypeptide sequence. In the presence of a nucleic acid molecule expressing one or more nucleic acid sequences encoding suppressor tRNA molecules, the stop codon between the two polypeptide sequences is suppressed and a fusion polypeptide is expressed.

[0982] Thus, in one aspect, the present invention comprises nucleic acid molecules (and/or compositions comprising such molecules) from which tRNA molecules (*e.g.*, suppressor tRNA molecules) can be expressed. Nucleic acid molecules from which tRNA molecules can be expressed may be any type nucleic acid molecule known to those skilled in the art, for example, plasmids, linear nucleic acid molecules, viruses and the like. In a particular embodiment, the present invention provides a virus (*e.g.*, an adenovirus, a lentivirus, a baculovirus etc.) from which a tRNA molecule may be expressed. In a specific embodiment, the present invention provides an adenovirus from which one or more tRNA molecule may be expressed.

[0983] In one embodiment, the present invention provides an adenovirus that expresses one or more suppressor tRNA molecules. One non-limiting example of such an adenovirus can be found in the Tag-On-Demand™ System commercially available from Invitrogen Corporation, Carlsbad, CA catalog number K400-01. Methods of the invention may employ an adenoviral-based stop suppression technology to allow expression of an untagged (*i.e.* native) or C-terminally-tagged recombinant polypeptide of interest in host cells from a single expression vector. In some embodiments, nucleic acid molecules of the invention may include Tag-On-Demand™ GATEWAY® vectors and/or other vectors, which may be used to generate an expression construct.

[0984] In one aspect, materials and methods of the present invention may be used to facilitate transient expression of a C-terminally-tagged recombinant polypeptide of interest in host cells (*e.g.*, mammalian cells). Materials and methods of the invention may be used to provide a means to easily detect the expression or localization of a recombinant polypeptide(s) for which there is no specific antibody available. This may be useful, for example, in that once tagged recombinant polypeptide expression is verified, native polypeptide expression experiments may be performed with the same construct.

[0985] In some aspects, the present invention uses adenovirus as a delivery vehicle, enabling efficient delivery of suppressor tRNAs to a large variety of host cell types (*e.g.*, mammalian cell types). Typically, in methods of the invention, suppressor tRNAs may be delivered transiently to cells to minimize toxicity.

- [0986] In some aspects of the invention, methods of the invention may be used for high-throughput applications including rapid screening of a large number of genes for expression in a particular cell type.
- [0987] In some embodiments, materials and methods of the invention may be used to transiently express C-terminally-tagged and native recombinant polypeptides in mammalian cells from a single expression construct. Suppressor tRNAs that function in mammalian cells have been described (see Capone, *et al.* (1985) *EMBO J.* 4, 213-221).
- [0988] In one aspect, the present invention provides nucleic acid molecules (*e.g.*, mammalian expression vectors) into which the a nucleic acid sequence encoding a polypeptide of interest will be cloned. Preferably, a nucleic acid sequence encoding a polypeptide of interest may be cloned into a nucleic acid molecule of the invention (*e.g.* pcDNA™ 6.2/V5-DEST or pcDNA™ 6.2/GFP-DEST) in a configuration that is compatible with expression of C-terminally-tagged recombinant polypeptide by suppression of one or more stop codons.
- [0989] In another aspect, the present invention provides nucleic acid molecules (*e.g.*, replication-incompetent adenoviruses) comprising a nucleic acid sequence from which a suppressor tRNA can be expressed (*e.g.*, the human tRNA^{ser} suppressor gene). In some embodiments, a suppressor tRNA may be a tRNA mutated to recognize one or more stop codons, for example, the TAG (amber stop) codon, and decode it as a serine. Nucleic acid molecules according to this aspect of the invention may be introduced into host cells to provide a transient source of the tRNA^{ser} suppressor. If the expression construct encoding a gene of interest with a TAG stop codon is present in the host cells, the stop codon will be translated as serine, allowing translation to continue through any downstream reading frame (*i.e.* C-terminal tag). This results in production of a C-terminally-tagged fusion polypeptide.
- [0990] In one aspect, a nucleic acid molecule from which a suppressor tRNA molecule may be expressed may be a recombinant adenovirus and may be constructed as follows. A vector containing the gene encoding the tRNA^{ser} gene with its native promoter and terminator may be obtained, for example, from Dr. Uttam RajBhandary at the Massachusetts Institute of Technology. This tRNA^{ser} gene has been mutated such that the anticodon recognizes the TAG (amber) stop codon, and is referred to as the tRNA^{ser} suppressor gene

(see Capone, *et al.* (1985)). The tRNA^{ser} suppressor gene may be PCR amplified and TOPO[®] Cloned into the pENTR/D-TOPO[®] vector available from Invitrogen Corporation, Carlsbad, CA (Catalog no. K2400-20) to generate a GATEWAY[®] entry clone. Using the entry clone above and a multimerization procedure described in Buvoli *et al.*, 2000 (Buvoli, *et al.* (2000) *Mol. Cell. Biol.* 20, 3116-3124), a GATEWAY[®] entry clone containing 8 tandem copies of the tRNA^{ser} suppressor gene can be generated. One such entry clone has been constructed and is named pENTR[™]-tRNA8^{TAG}. The pENTR[™]-tRNA8^{TAG} entry clone was recombined with Invitrogen's pAd/PL-DEST[™] destination vector (Catalog no. V494-20) using the GATEWAY[®] LR recombination reaction to generate the adenoviral expression clone, pAd/GW-tRNA8^{TAG}. The pAd/GW-tRNA8^{TAG} expression construct was used in Invitrogen's ViraPower[™] Adenoviral Expression System (Catalog no. K4940-00) to produce recombinant adenovirus, which was CsCl-purified and titered to generate the Tag-On-Demand[™] Suppressor Supernatant.

[0991] In some embodiments, a nucleic acid molecule expressing a suppressor tRNA molecule may be a recombinant adenovirus, which may be used, for example, to deliver the tRNA^{ser} suppressor to host cells (*e.g.*, mammalian cells). Although adenovirus has a very broad tropism and can be used to deliver the tRNA^{ser} suppressor to a large variety of host cell lines and cell types, materials and methods of the invention are not limited to those cells that can be transduced with adenovirus. Thus, materials and methods of the invention may be used with any host cell line or type known to those skilled in the art.

[0992] In the practice of the invention, nucleic acid molecules expressing one or more suppressor tRNA molecules may be introduced into host cells. When such nucleic acid molecules are introduced into host cells, they may be introduced into from about 25% to about 100% of the cell population, or from about 25% to about 90%, from about 25% to about 80%, from about 25% to about 70%, from about 25% to about 60%, from about 25% to about 50%, from about 50% to about 100%, from about 60% to about 100%, from about 70% to about 100%, from about 80% to about 100%, from about 90% to about 100%, from about 50% to about 90%, from about 50%, to about 80%, from about 50% to about 75%, from about 50% to about 70%, or from about 50% to

about 60%. In some embodiments, nucleic acid molecules expressing one or more suppressor tRNAs may be adenoviruses and may transduce mammalian cells with extremely high efficiency, resulting in delivery of the tRNA^{scr} suppressor to nearly 100% of mammalian cells.

[0993] In some embodiments, nucleic acid molecules expressing suppressor tRNAs of the invention may not integrate into the host genome and expression of the suppressor tRNA may be transient and only persist for as long as the nucleic acid molecule (*e.g.*, viral genome) is present (typically 7-8 days after transduction). Typically, once a nucleic acid molecule expressing a suppressor tRNA is introduced into host cells, the suppressor tRNA is expressed within 24 hours.

[0994] In some embodiments, a nucleic acid molecule expressing a suppressor tRNA molecule may be a virus (*e.g.*, adenovirus). As is known in the art, viruses may possess the ability to bind to one or more receptors that may be present on a cell surface. For example, adenovirus enters target cells by binding to the Coxsackie/Adenovirus Receptor (CAR) (see, Bergelson, *et al.* (1997) *Science* 275, 1320-1323) and internalizing via integrin-mediated endocytosis (see, Russell, W. C. (2000) *J. Gen. Virol.* 81, 2573-2604). Once internalized, the recombinant adenovirus is actively transported to the nucleus, and begins to express the suppressor genes. Thus, when the nucleic acid molecule expressing a suppressor tRNA is an adenovirus, the host cell line should contain CAR. Most mammalian cell types express CAR, but levels vary. Depending on the amount of the CAR expressed in a specific target cell line, transduction efficiencies may vary when an adenovirus is used to express a suppressor tRNA. One skilled in the art will appreciate that other viruses may be used to express suppressor tRNAs in the practice of the invention, for example, vaccinia virus, herpes virus, adeno-associated virus, baculovirus, retroviruses (*e.g.*, lentivirus), plant viruses (*e.g.*, tobacco mosaic virus, cauliflower mosaic virus, etc.), negative stranded RNA viruses (*e.g.*, Sendai virus, etc.), positive stranded RNA viruses (*e.g.*, alphaviruses, etc.). One skilled in the art can readily select an appropriate virus to infect any desired type of target cell based on the known tropisms of specific viruses for specific cell types.

- [0995] In some embodiments, nucleic acid molecules expressing a suppressor tRNA of the invention may be adenoviruses. Adenoviruses for use in this aspect of the invention may have one or more deletions in the adenoviral genome compared to a wild-type adenoviral genome (*e.g.*, Ad2, Ad5, etc.). For example, an adenovirus for use in the invention may be deleted in the E1 and/or E3 regions. In some embodiments, the entire E1 and E3 regions may be deleted. Such viruses may be replication-incompetent when transduced into mammalian cells that do not express the E1a or E1b proteins (see, Graham, *et al.* (1977) *J. Gen. Virol.* 36, 59-74; Kozarsky and Wilson (1993) *Curr. Opin. Genet. Dev.* 3, 499-503; and Krougliak and Graham (1995) *Hum. Gene Ther.* 6, 1575-1586).
- [0996] As is known in the art, adenovirus does not integrate into the host genome upon transduction. Because the virus is replication-incompetent, the presence of the viral genome is transient and will eventually be diluted out as cell division occurs. Because the adenovirus is present transiently in mammalian cells, the production of C-terminally-tagged polypeptide resulting from stop suppression is also transient. As levels of adenovirus decrease, levels of C-terminally-tagged polypeptide produced decrease.
- [0997] In some embodiments, viruses used to express suppressor tRNAs in accordance with the invention may be replication-incompetent in the cell type in which they express the suppressor tRNA. Viruses for use in this aspect of the invention may be screened for the presence of wild-type replication-competent viruses using techniques known in the art. For example, a population of adenovirus for use in the present invention may be screened for the presence of replication-competent adenovirus (RCA) contamination using a supernatant rescue assay (see, Dion, *et al.* (1996) *J. Virol. Methods* 56, 99-107) with a detection sensitivity of one wild-type RCA per 10^9 recombinant adenovirus. In some embodiments, a viral preparation to be used to express one or more suppressor tRNA molecules in accordance with the methods of the invention may contain no detectable wild-type RCA.
- [0998] In some embodiments, the present invention provides methods to express a C-terminally-tagged fusion polypeptide, comprising transducing a host cell with a virus expressing one or more suppressor tRNA molecules, transfecting the transduced cells with one or more nucleic acid molecules

encoding all or a portion of a fusion polypeptide, and incubating the host cell under conditions sufficient to express a C-terminally tagged fusion polypeptide. A schematic representation of an embodiment of this type is provided in Figure 62. In another embodiment, the present invention provides methods to express a C-terminally tagged fusion polypeptide, comprising transducing a stable cell line comprising a nucleic acid molecule encoding all or a portion of a fusion polypeptide with a virus expressing one or more suppressor tRNA molecules and incubating the transduced cell under conditions sufficient to express a C-terminally tagged fusion polypeptide. A schematic representation of an embodiment of this type is shown in Figure 63.

[1000] Methods of the invention may entail the use of stocks of viruses, for example, viruses expressing one or more suppressor tRNA molecules. As will be appreciated by those skilled in the art, stocks of viruses may be stored at -80°C. In general, stocks stored under these conditions are stable for at least 6 months. If a viral stock has been stored at -80°C for longer than 6 months, the titer of the stock may be determined using standard techniques as viral titers may decrease with long-term storage. Viral stocks should not be repeatedly thawed and re-frozen as viral titers can decrease with more than 3 freeze/thaw cycles.

[1001] One skilled in the art is aware that the handling of materials containing viruses should be performed following the applicable Federal and institutional guidelines for working with potentially hazardous organisms. For example, all manipulations should be performed within a certified biosafety cabinet, all media containing virus should be treated with bleach, all material that comes into contact with virus (*e.g.*, pipettes, pipette tips, and other tissue culture supplies) should be treated with bleach or disposed of as biohazardous waste, and persons handling material containing virus should wear appropriate safety clothing (*e.g.*, gloves, a laboratory coat, and safety glasses or goggles).

[1002] In some embodiments, methods of the invention may be used to create a nucleic acid molecule encoding a fusion polypeptide. According to one aspect of the invention, a nucleic acid molecule encoding a fusion polypeptide may be constructed by combining a first nucleic acid molecule having a first nucleic acid sequence encoding a polypeptide sequence (*e.g.*, a polypeptide of interest) with a second nucleic acid molecule having a second nucleic acid

sequence encoding an additional polypeptide sequence (*e.g.*, a polypeptide tag sequence). A nucleic acid molecule encoding a polypeptide of interest should contain an ATG initiation codon in the context of a Kozak consensus sequence for proper initiation of translation in mammalian cells (Kozak, 1987; Kozak, 1991; Kozak, 1990). An example of a Kozak consensus sequence is (G/A)NN**ATGG**, where the ATG initiation codon is underlined. Other sequences are possible, but the G or A at position -3 and G at position +4 are the most critical for function (shown in bold).

- [1003] Typically, a nucleic acid molecule encoding a polypeptide of interest will contain a stop codon, for example, encoded by the nucleotides, TAG, TAA or TGA. One skilled in the art will appreciate that an appropriate suppressor tRNA (*i.e.*, with an anti-codon that corresponds to the stop codon) must be used.
- [1004] One skilled in the art will appreciate that, after joining of the first and second nucleic acid molecules to produce a nucleic acid molecule encoding a fusion polypeptide, the sequence encoding the polypeptide of interest must be in the same reading frame as the additional polypeptide sequence.
- [1005] Second nucleic acid molecules encoding an additional polypeptide sequence will typically comprise one or more stop codons after the sequence encoding the additional polypeptide sequence. In general, the stop codon on the second nucleic acid molecule will be different from the stop codon on the first nucleic acid molecule.
- [1006] A wide variety of nucleic acid molecules are suitable for use as second nucleic acid molecules in accordance with the present invention. Non-limiting examples of such nucleic acid molecules include vectors commercially available from Invitrogen Corporation, Carlsbad, CA. Examples of such vectors are provide with their Invitrogen Corporation, Carlsbad, CA catalog number in parenthesis. Such vectors include, but are not limited to, pLenti4/V5-DEST[™] (K4980-00), pLenti6/V5-DEST[™] (K4950-00), pLenti6/Ubc/V5-DEST[™] (K4990-00), pLenti6/V5-D-TOPO[®] (K4960-00), and pAd/CMV/V5-DEST (K4930-00), which may be used for viral expression; pcDNA5/FRT/V5-His-TOPO[®] (K6020-01), pSecTag/FRT/V5-His-TOPO[®] (K6025-01), pEF5/FRT/V5-DEST[™] (V6020-20), and pEF5/FRT/V5-D-TOPO[®] (K6035-01), which may be used for expression from

a specific genomic locus using the Flp-In™ System; pcDNA™ 4/TO/myc-His (K1030-01), pGene/V5-His (K1060-01), which may be used for inducible expression; pcDNA™ 6.2/V5-DEST (K420-01), pcDNA™ 3.2/V5-DEST (12489-019), pcDNA™ -DEST40 (12274-015), pcDNA6.2/V5-GW/D-TOPO® (K2460-20), pcDNA3.2/V5-GW/D-TOPO® (K2440-20), pcDNA3.1D/V5-His-TOPO® (K4900-01), pcDNA™ 3.1/V5-His-TOPO® (K4800-01), pcDNA™ 3.1/V5-His (V810-20), pcDNA™ 3.1/myc-His (V800-20), pcDNA™ 3.1(-)/myc-His (V855-20), pcDNA™ 4/V5-His (V861-20), pcDNA™ 4/myc-His (V863-20), pcDNA™ 6/V5-His (V220-20), and pcDNA™ 6/myc-His (V221-20), which may be used for constitutive expression from the CMV promoter; pEF6/V5-His-TOPO® (K9610-20), pEF1/V5-His (V920-20), pEF1/myc-His (V921-20), pEF4/V5-His (V941-20), pEF4/myc-His (V942-20), pEF6/V5-His (V961-20), and pEF6/myc-His (V962-20), which may be used for constitutive expression from the EF-1 α promoter; pUB6/V5-His (V250-20), which may be used for constitutive expression from the UbC promoter; pSecTag2 (V900-20), and pSecTag2/Hygro (V910-20), which may be used for constitutive secreted expression; and pcDNA™ 6.2/GFP-DEST (K410-01), pcDNA™ -DEST47 (12281-010), and pcDNA3.1/CT-GFP-TOPO® (K4820-01), which may be used for fusion to the GFP reporter gene.

[1007] A variety of factors may be optimized to produce fusion polypeptides according to the methods of the invention. Factors include, but are not limited to, characteristics of the host cell line; the health of the cells and experimental cell culture conditions; the transfection method used to introduce nucleic acid molecules into the host cell line; the transduction procedure used; and the amount of nucleic acid encoding a suppressor tRNA introduced into the host cells (*e.g.*, multiplicity of infection when the nucleic acid molecule encoding a suppressor tRNA is a virus such as an adenovirus).

[1008] In some embodiments, a fusion protein of the invention may be expressed in any host cell type known to those skilled in the art. In some embodiments, a host cell line may be a mammalian host cell line. When an adenovirus is used in the practice of the invention to express one or more suppressor tRNA molecules, a host cell line preferably expresses one or more receptors allowing efficient transduction of the cell line by the adenovirus. An example of a suitable receptor is the Coxsackie/Adenovirus Receptor (CAR)

(see, Bergelson, *et al.* (1997) *Science* 275, 1320-1323). Most mammalian cell types express CAR, but levels vary. One skilled in the art will appreciate that transduction efficiencies of cell lines will vary depending on the amount of the CAR expressed in a given cell line and can adjust either the multiplicity of infection and/or the cell line used as necessary for any particular application using routine experimentation.

[1009] In some embodiments, cells lines used in the practice of the present invention may not express viral proteins necessary for replication of a virus used to introduce suppressor tRNAs into the host cells. For example, when an adenovirus is used to introduce suppressor tRNAs into host cells, the host cells may not express the adenovirus E1 proteins.

[1010] Typically, host cells used in the practice of the invention may be amenable to efficient transfection. For example, it may be possible to introduce a nucleic acid molecule encoding a fusion polypeptide invention into a high percentage of cells using standard techniques. For example, using lipid-mediated transfection (for example, with Lipofectamine 2000), it may be possible to introduce a nucleic acid molecule encoding a fusion polypeptide of the invention into from about 25% to about 100%, from about 25% to about 99%, from about 25% to about 95%, from about 25% to about 80%, from about 25% to about 70%, from about 25% to about 60%, from about 25% to about 50%, from about 25% to about 40%, from about 40% to about 95%, from about 50% to about 95%, from about 60% to about 90%, from about 75% to about 95%, or from about 80% to about 95% of the cells of a given sample of cells (*e.g.*, the cells in a well of a tissue culture plate). Examples of suitable cell lines include, but are not limited to, COS-7, CHO-S, HeLa, HT1080, and BHK-21, primary rat hippocampal and cortical neurons.

[1011] In some embodiments, nucleic acid molecules encoding suppressor tRNAs for use in the present invention may be adenoviruses. Such adenoviruses may be deleted in the E1 region, rendering them replication-incompetent in any cells that do not express the E1 proteins. Typically methods of the invention are not performed in cells that express the adenovirus E1 protein (*e.g.*, 293 cells or derivatives) as viral replication may occur in these cells, leading to rapid death of the target cell within 1-2 days after

infection. In some instances it may be desirable to practice methods of the invention in cells expressing the adenovirus E1 protein.

[1012] One skilled in the art will appreciate that the health of the cells to be used in methods of the invention may affect the expression of fusion polypeptide of the invention expressed in these cells. In general, in methods of the invention, cells should be healthy (*i.e.* exhibit > 95% viability) at the time of plating. Poor quality cell stock (*e.g.* cells consistently allowed to become overgrown or confluent before passaging, growth media allowed to become yellow before re-feeding) can negatively impact suppression efficiency and the amount of fusion polypeptide expressed. Generally, freshly prepared media may be used in the practice of methods of the invention.

[1013] Methods of the invention may entail introducing one or more nucleic acid molecules into one or more host cells. Any method of choice may be used to transfect nucleic acid molecules into cells. Suitable methods include, but are not limited to, calcium phosphate (see Chen and Okayama (1987); Wigler *et al.* (1977)), lipid-mediated (see, Felgner *et al.* (1989); Felgner and Ringold (1989)), and electroporation (see Chu, *et al.* (1987); Shigekawa and Dower (1988)). Suitable conditions (*e.g.*, reagents, incubation conditions, etc.) for introducing nucleic acid molecules into any specific cell line may be determined by consulting published literature, consulting the provider of the cell line in question, and/or by routine experimentation. In some embodiments, methods of the invention may entail introducing one or more nucleic acid molecules into one or more cells using lipid-mediated transfection with a suitable lipid reagent (*e.g.*, a lipid reagent from Invitrogen Corporation, Carlsbad, CA such as a cationic lipid-based reagent, Lipofectamine™ 2000 Reagent).

[1014] In some embodiments, methods of the invention may comprise introducing one or more nucleic acid molecules into one or more cells using Lipofectamine™ 2000 Reagent (see, Ciccarone, *et al.* (1999) *Focus* 21, 54-55) a cationic lipid-based formulation designed for transfection of nucleic acids into eukaryotic cells. Methods of this type may comprise forming a complex comprising nucleic acid molecules and Lipofectamine™ 2000 Reagent and contacting cells with the complexes in culture medium in the presence of serum. Such methods may not comprise removal of complexes or medium

change or addition following transfection. Alternatively, methods may comprise removal of complexes or medium change or addition following transfection, for example, at 4-6 hours after contacting cells with the complexes.

[1015] In some embodiments, the present invention may include a method of screening for expression of a polypeptide comprising introducing into a host cell a nucleic acid molecule expressing a suppressor tRNA and a nucleic acid molecule encoding the polypeptide; and detecting the presence of the polypeptide. In some embodiments such methods may be used to screen for expression or localization of the polypeptide or to screen for expression of a large number of genes. Such methods may involve the use of an adenovirus expressing a suppressor tRNA and may involve transducing a host cell with the adenovirus and transfecting a nucleic acid molecule encoding the polypeptide. Typically, transfection of the nucleic acid molecule is done as soon as practical after transduction with the adenoviruses. In some embodiments, the cells may be contacted with a solution comprising the adenovirus and then nucleic acid molecules (*e.g.*, in complex with a transfection reagent) may be added to the solution comprising the adenovirus. Optionally, a nucleic acid molecule encoding a polypeptide of the invention may be introduced into a host cell prior to transduction of the host cell with an adenovirus expressing one or more suppressor tRNAs. One skilled in the art will appreciate that transducing a host cell with an adenovirus and simultaneously (*i.e.* as soon as practically possible) transfecting with plasmid encoding a polypeptide of the invention can increase plasmid-derived gene expression as well as reduce toxicity to the cell (see, Cotten, *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89, 6094-6098; Curiel, *et al.* (1991) *Proc. Natl. Acad. Sci. USA* 88, 8850-8854; Guy, *et al.* (1995) *Mol. Biotechnol.* 3, 237-248; Honda, *et al.* (1996) *J. Virol. Methods* 58, 41-51; and Merwin, *et al.* (1995) *J. Immunol. Methods* 186, 257-266).

[1016] In some embodiments, it may be desirable to create a stable cell line comprising a nucleic acid molecule encoding a polypeptide of the invention using standard techniques and to transduce the stable cell line with an adenovirus expressing one or more suppressor tRNAs.

[1017] In methods of the invention that comprise transducing a host cell with a virus expressing one or more suppressor trans (*e.g.*, an adenovirus), cells may be transduced with any desired amount of virus. For example, cells may be transduced with virus at a multiplicity of infection (MOI) of from about 0.1 to about 500, from about 0.25 to about 500, from about 0.5 to about 500, from about 0.75 to about 500, from about 1 to about 500, from about 2 to about 500, from about 3 to about 500, from about 4 to about 500, from about 5 to about 5000, from about 10 to about 500, from about 25 to about 500, from about 50 to about 500, from about 75 to about 500, from about 100 to about 500, from about 200 to about 500, from about 300 to about 500, from about 400 to about 500, from about 1 to about 250, from about 1 to about 200, from about 1 to about 150, from about 1 to about 100, from about 1 to about 75, from about 1 to about 50, from about 1 to about 25, from about 1 to about 20, from about 1 to about 15, from about 1 to about 10, from about 1 to about 5, from about 10 to about 400, from about 10 to about 300, from about 10 to about 200, from about 10 to about 100, from about 10 to about 75, from about 10 to about 70, from about 10 to about 65, from about 10 to about 60, from about 10 to about 55, from about 10 to about 50, from about 10 to about 45, from about 10 to about 40, from about 10 to about 35, from about 10 to about 30, from about 10 to about 25, from about 10 to about 20, or from about 10 to about 15. Thus, cells may be transduced at an MOI of about 1, about 5, about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, or about 100. MOI is defined as the number of virus particles per cell and generally correlates with expression. Depending on the cell line used and the nature of the gene of interest, MOI may be varied to optimize expression of a fusion polypeptide of the invention using routine experimentation.

[1018] As an example, for the cell lines tested (*i.e.* COS-7, CHO-S, HeLa, HT1080, BHK-21, and primary rat hippocampal and cortical neurons), transduction at an MOI of 50 followed by immediate transfection of the nucleic acid molecule encoding a fusion polypeptide of the invention generally results in 50-80% suppression. This means that 50-80% of the polypeptide expressed from the nucleic acid molecule encoding the fusion polypeptide is

expressed as the fusion polypeptide. Note that 100% suppression cannot be achieved at any MOI. Some untagged polypeptide will always be expressed.

[1019] One of skill in the art will appreciate that the % suppression (*i.e.* suppression efficiency) achieved when cells are transduced at a particular MOI (*e.g.* MOI = 50) can vary and is dependent on a number of factors including: the amount of CAR expressed in the mammalian cell; the nature of the gene being expressed; the health of the cells at the time of transduction; phenotypic changes to the cells resulting from stop codon suppression.

[1020] Depending on the suppression efficiency and consequently, the amount of fusion polypeptide expressed, the % suppression achieved can be optimized by varying the MOI using routine experimentation. It is important to note that while the % suppression achieved can be increased by increasing the MOI, doing so may increase the likelihood of phenotypic changes to the cells.

[1021] Expression of a suppressor tRNA in a host cell may result in phenotypic changes in the cell (*e.g.* toxicity) since 1/3 of the endogenous stop codons (*i.e.* all genes containing the stop codon recognized by the suppressor tRNA) can be suppressed. This leads to potential addition of extra amino acids to the C-termini of cellular proteins other than the fusion polypeptide of interest. In some embodiments, it may be desirable to optimize methods of the invention in order to minimize phenotypic effects in a particular cell line of interest.

[1022] In embodiments of the invention where one or more suppressor tRNAs are expressed from an adenovirus, the adenovirus may deliver a transient source of the suppressor tRNA to the target cell. When the adenovirus is replication-incompetent, it does not stably integrate into the genome of the target cell, and will be diluted out gradually as cell division occurs. This results in an overall decrease in suppressor tRNA expression over time. In some embodiments, it may be desirable to detect fusion polypeptide of the invention from about 1 hour to about 5 days, from about 1 hour to about 4 days, from about 1 hour to about 3 day, from about 1 hour to about 2 day, from about 1 hour to about 1 day, from about 1 hour to about 20 hours, from about 1 hour to about 16 hours, from about 1 hour to about 12 hours, from about 1 hour to about 8 hours, from about 1 hour to about 4 hours, from about

1 hour to about 3 hours, from about 1 hour to about 2 hours, from about 8 hours to about 72 hours, from about 8 hours to about 60 hours, from about 8 hours to about 48 hours, from about 8 hours to about 36 hours, from about 8 hours to about 24 hours from about 8 hours to about 20 hours, from about 8 hours to about 16 hours, or from about 8 hours to about 12 hours after transduction of the cells of interest. Thus fusion polypeptide may be detected at about 4 hours, about 8 hours, about 12 hours, about 16 hours, about 20 hours, about 24 hours, about 28 hours, about 32 hours, about 36 hours, about 40 hours, about 44 hours, about 48 hours, about 52 hours, about 56 hours, about 60 hours, about 64 hours, about 68 hours, about 72 hours, about 76 hours, about 80 hours, about 84 hours, about 88 hours, or at about 92 hours after transduction of the cells.

[1023] Methods of the invention may be practiced using any number of cells grown in any apparatus for that purpose known in the art (*e.g.*, tissue culture plates, tissue culture flasks, roller bottles, bioreactors, etc.) In some embodiments, tissue culture plates may be used (*e.g.*, 6-well, 24-well, or 96-well plates). For high-throughput applications, 96-well plates may be used. When cells are grown in tissue culture plates for use in the present invention, cells may be plated such that they are be 90% confluent at the time of transduction. As an example, the cells may be transduced with an adenovirus expressing a suppressor tRNA at an MOI of 50 and transfected with a plasmid encoding a fusion polypeptide of the invention. As discussed above, any suitable transfection protocol and/or reagents may be used. The amounts of nucleic acid molecule encoding a fusion polypeptide of the invention and transfection reagent may be adjusted to comport with the number of cells to be transfected using techniques well known in the art.

[1024] In a non-limiting example, COS-7 cells may be plated at, for example, 8×10^4 COS-7 cells/per well of a 24-well plate and cultured overnight at 37°C. On the following day, the cells may be transduced (*e.g.*, with an adenovirus expressing a suppressor tRNA), for example, using an MOI = 50. It may be assumed that the number of cells has doubled during overnight culture so that the number of cells is $2 \times 8 \times 10^4 = 1.6 \times 10^5$ cells. If the titer of the viral stock is, for example, 1×10^9 pfu/ml, the amount of viral stock to add to the cells may be calculated as follows:

$50 \text{ pfu/cell} \times 1.6 \times 10^5 \text{ cells} = 8 \times 10^6 \text{ pfu} / 1 \times 10^9 \text{ (pfu/ml)} = .008 \text{ ml} = 8 \text{ } \mu\text{l}$ to add to each well.

[1025] In some embodiments, it may be desirable to include one or more controls in the methods of the invention. For examples, methods of the invention may comprise transducing a host cell with a virus expressing a suppressor tRNA, transfecting the cells with a control nucleic acid molecule (*e.g.*, a nucleic acid molecule encoding a fusion polypeptide with a reporter activity), and detecting a reporter activity. For example, pcDNA™ 6.2/GFP-GW/p64^{TAG} may be used as a positive control for transduction, transfection, and expression. In this plasmid, the p64 protein (human c-myc) containing a TAG stop codon is cloned in frame with the cycle-3 GFP reporter gene (see (Chalfie, M., *et al.*, *Science* 263:802-805 (1994); Cramer, A., *et al.*, *Nature Biotechnol.* 14:315-319 (1996)). Including pcDNA™ 6.2/GFP-GW/p64^{TAG} plasmid when conducting transduction and transfection methods of the invention allows detecting a reporter gene activity (*e.g.*, assaying for cycle-3 GFP expression using fluorescence microscopy or c-myc expression using Western blot analysis), and evaluating transfection and/or transduction conditions.

[1026] In one non-limiting example, methods of the invention may be used to express a fusion polypeptide of the invention. Methods of the invention may comprise seeding cells into a suitable tissue culture vessel at a suitable density (*e.g.*, at a density such that the cells will be approximately 90% confluent at the time of transduction). Optionally, cells may be incubated (*e.g.*, at 37°C overnight) after seeding. When a 24-well tissue culture plate is used, cells may be seeded in 500 μl of complete medium. Methods of the invention may comprise, on the day of transduction, removing the growth medium from each well of cells and replacing with fresh growth medium (for a 24-well plate, 250 μl of medium may be used). Methods may further comprise contacting the cells with a nucleic acid molecule encoding a suppressor tRNA (*e.g.*, transducing the cells with an adenovirus expressing a suppressor tRNA). When the nucleic acid molecule expressing a tRNA is a virus, any suitable MOI may be used (*e.g.*, 50). Methods may further comprise returning the transduced cells to an incubator.

[1027] In embodiments where the host cell line is a stable cell line comprising a nucleic acid molecule encoding a fusion polypeptide of the invention, methods of the invention may comprise incubating cells (*e.g.*, for 5-6 hours at 37°C) after introduction of a nucleic acid molecule encoding a suppressor tRNA (*e.g.*, after transduction with an adenovirus expressing a suppressor tRNA). Typically, cells are incubated for at least 5 hours as transduction efficiency will be decreased at shorter times. Longer incubation time is possible (*e.g.* overnight), but will not increase the transduction efficiency and may increase cell toxicity. Methods may further comprise removing the medium containing virus from the cells (*e.g.*, after 5-6 hours), washing the cells (*e.g.*, with 500 µl of fresh, complete growth medium in a 24-well plate), adding complete growth medium (*e.g.*, 500 µl of fresh, complete growth medium in a 24-well plate) and incubating the cells under conditions sufficient to express a fusion polypeptide of the invention (*e.g.* at 37°C in an incubator for a suitable period of time). Methods of the invention may further comprise detecting the fusion polypeptide.

[1028] In some embodiments, a nucleic acid molecule encoding a fusion polypeptide of the invention may be introduced into a host cell (*e.g.*, after the cell has been transduced as described above). For example, after transduction, a suitable amount of a nucleic acid molecule encoding a fusion polypeptide of the invention may be mixed in a suitable medium. For example, for a well of a 24 well plate 500 ng of plasmid DNA may be dissolved in 50 µl of Opti-MEM® I Reduced Serum Medium without serum and a suitable amount of a transfection reagent (*e.g.*, a cationic lipid transfection reagent) may be mixed with a suitable amount of a medium (*e.g.*, for a well of a 24 well plate, 1.5 µl of Lipofectamine™ 2000 may be mixed in 50 µl of Opti-MEM® I Reduced Serum Medium). Both mixtures (*i.e.*, DNA:medium and reagent:medium) may be incubated, for example, for 5 minutes at room temperature. The two mixtures may be combined and incubated, for example, for 20 minutes at room temperature to allow the formation of nucleic acid:transfection reagent complexes (*e.g.*, DNA-Lipofectamine™ 2000 Reagent complexes). Methods of the invention may comprise adding the complexes (*e.g.*, DNA-Lipofectamine™ 2000 Reagent complexes) directly to the growth medium containing viruses used to transduce the host cells. Methods may comprise

incubating the cells (for example, for 5-6 hours at 37°C). Typically, cells are incubated for at least 5 hours as transduction efficiency will be decreased at shorter times. Longer incubation time is possible (*e.g.* overnight), but will not increase the transduction efficiency and may increase cell toxicity. Methods may further comprise removing the medium containing virus from the cells (*e.g.*, after 5-6 hours), washing the cells (*e.g.*, with 500 µl of fresh, complete growth medium in a 24-well plate), adding complete growth medium (*e.g.*, 500 µl of fresh, complete growth medium in a 24-well plate) and incubating the cells under conditions sufficient to express a fusion polypeptide of the invention (*e.g.* at 37°C in an incubator for a suitable period of time). Methods of the invention may further comprise detecting the fusion polypeptide.

[1029] One skilled in the art can readily adjust the volumes of the various reagents described above to transduce cells in different tissue culture formats (*e.g.*, vary the amounts of cells and medium used) in proportion to the difference in surface area of the tissue culture plates used. For example, a 96-well plate may be used having a surface area per well of 0.3 cm², cells may be seeded in a volume of 100 µl and transduced in a volume of 50 µl; a 6-well plate may be used having a surface area per well of 10 cm², cells may be seeded in a volume of 2 ml and transduced in a volume of 1 ml.

[1030] As a non-limiting example, for methods of the invention using COS-7 cells, the following seeding densities and reagent quantities for transduction and transfection may be used in different tissue culture formats. Note that the suggested DNA quantities are for transfection using Lipofectamine™ 2000 Reagent.

Condition	6-well	24-well	96-well
Seeding density	3 x 10 ⁵ cells	8 x 10 ⁴ cells	1 x 10 ⁴ cells
MOI = 50	3 x 10 ⁷ virus	8 x 10 ⁶ virus	1 x 10 ⁶ virus
Amount of plasmid DNA per well	2 µg	500 ng	320 ng
Amount of Lipofectamine™ 2000 Reagent per well	6 µl	1.5 µl	1 µl

[1031] In methods of the invention in which an adenovirus is used to express a suppressor tRNA in a host cell, one skilled in the art will recognize that such expression is transient. Accordingly, expression of a fusion polypeptide of the invention from a transiently transfected plasmid generally peaks within 24-48

hours following transfection. To obtain maximal levels of fusion polypeptide of the invention, cells may be harvested and assayed for fusion polypeptide of the invention expression between 24 and 48 hours post-transfection. Since expression conditions will vary depending on the nature of a particular fusion polypeptide of the invention and its half-life, conditions described above may be optimized using routine experimentation to obtain maximal levels of fusion polypeptide expression.

[1032] Methods of the invention may comprise detecting a fusion polypeptide of the invention. In some embodiments, detection may be by Western blot and/or immunofluorescence. In methods of this type, an antibody that specifically binds to the polypeptide of interest portion of the fusion protein may be used. One skilled in the art will appreciate that this allows detection of fused and un-fused forms of the polypeptide of interest. Alternatively, an antibody that specifically bind to the additional polypeptide sequences of the fusion polypeptide of the invention allows detection of only the fusion polypeptide and not the polypeptide of interest lacking the additional polypeptide sequences.

[1033] In some embodiments, additional polypeptide sequences in a fusion polypeptide of the invention may be a fluorescent polypeptide (*e.g.*, the green fluorescent protein (GFP)). In embodiments of this type, it may be desirable to detect the fluorescence of the fluorescent polypeptide. In a specific embodiment, methods of the invention may comprise detecting GFP. GFP may be detected, for example, *in vivo* using fluorescence microscopy. An example of a fusion polypeptide of the invention is the GFP-tagged p64^{TAG} fusion protein expressed from the plasmid pcDNATM 6.2/GFP-GW/p64^{TAG}. Since the GFP-tagged p64^{TAG} protein is expressed from the strong CMV promoter, fusion protein is generally detectable within 24 hours after transfection.

[1034] To detect fluorescent cells, suitable filter sets to optimize detection may be employed. The primary excitation peak of cycle-3 GFP is at 395 nm. There is a secondary excitation peak at 478 nm. Excitation at either of these wavelengths yields a fluorescent emission peak with a maximum at 507 nm. Note that the quantum yield can vary as much as 5- to 10-fold depending on the wavelength of light that is used to excite the GFP fluorophore.

- [1035] Use of the best filter set will insure that the optimal regions of the cycle-3 GFP spectra are excited and passed (emitted). A filter set designed to detect fluorescence from wild-type GFP (*e.g.* Omega Optical XF76 Filter) may be used. Alternatively, FITC filter sets may be used to detect cycle-3 GFP fluorescence. One skilled in the art will appreciate that these filter sets are not optimal and fluorescent signal may be weaker. For example, a typical FITC filter set excites cycle-3 GFP with light from 460 to 490 nm, covering the secondary excitation peak. The filter set passes light from 515 to 550 nm, allowing detection of most but not all of the cycle-3 GFP fluorescence.
- [1036] Most tissue culture media fluoresce because of the presence of riboflavin (see, Zylka, M. J., and Schnapp, B. J. (1996) *BioTechniques* 21, 220-226) and may interfere with detection of cycle-3 GFP fluorescence. To alleviate this problem, methods of the invention may comprise removing the growth medium and replacing the growth medium with Phosphate-Buffered Saline (PBS; Invitrogen Corporation, Carlsbad, CA, Catalog no. 10010-023) before assaying for GFP fluorescence. If cells are being cultured further after assaying, methods of the invention may comprise removing the PBS and replacing with fresh growth medium prior to re-incubation.
- [1037] In some embodiments, methods of the invention may comprise detecting a fusion polypeptide of the invention comprising a polypeptide sequence of the GFP by Western blotting. For example, GFP-tagged p64^{TAG} fusion polypeptide can be detected by Western analysis using the following antibodies available from Invitrogen Corporation, Carlsbad, CA: to detect both untagged and GFP-tagged p64^{TAG} protein, an antibody that specifically binds to the p64 portion of the fusion polypeptide (*i.e.*, one of the Anti-*myc* Antibodies) can be used; to detect GFP-tagged p64^{TAG} protein only, an antibody that specifically binds to the GFP portion of the fusion polypeptide (*e.g.*, the GFP Antiserum) may be used.
- [1038] In methods of the invention that comprise detecting a fusion polypeptide of the invention by Western blotting, a lysate of host cells expressing the fusion polypeptide may be prepared. For example, a cell lysate to assay for native or GFP-tagged p64 protein may be prepared. One skilled in the art will appreciate that procedures using NP-40 lysis are not effective in releasing p64 protein. Since p64 is localized in the nucleoli, harsher lysis

procedures using RIPA or SDS-PAGE sample buffer may be used to adequately solubilize p64 in total cell lysates. Methods of preparing a cell lysate to assay for p64 protein, may comprise washing cell monolayers (*e.g.*, washing once with Phosphate-Buffered Saline Invitrogen Corporation, Carlsbad, CA, PBS, Catalog no. 10010-023); adding a suitable lysis buffer (*e.g.*, 1X SDS-PAGE Sample Buffer) to each well containing cells (*e.g.*, for a 24-well plate, 100 μ l of 1X SDS-PAGE Sample Buffer per well may be used); loosening lysed cells from the plate (*e.g.*, a pipette tip can be used to loosen lysed cells from plate); and transferring the cells to a centrifuge tube (*e.g.*, for cells from one well of a 24-well plate a 1.5 ml microcentrifuge tube). Lysates will be viscous.

[1039] Methods of preparing a lysate may further include heating samples at 70°C for 10 minutes. Optionally, methods may include mixing (*e.g.* using a vortex mixer) one or more times and briefly centrifuging the sample. A lysate prepared by methods of the invention may be further processed or analyzed using techniques well known in the art. For example, an aliquot of the lysate (*e.g.*, 5 μ l of cell lysate) may be loaded onto an SDS-PAGE gel and electrophoresed. The GFP-tagged p64^{TAG} protein has a molecular weight of approximately 77.2 kDa.

[1040] One example of a suitable lysis buffer is 1 X SDS-PAGE Sample Buffer, which may be prepared by combining the following reagents in the amounts indicated: 0.5 M Tris-HCl, pH 6.8 (2.5 ml); Glycerol (100%) (2 ml); β -mercaptoethanol (0.4 ml); Bromophenol Blue (0.02 g); SDS (0.4 g); and sterile water to a final volume of 20 ml. Aliquots of the buffer may be frozen at -20°C until needed.

[1041] In one specific embodiment, the following ORFs were amplified to contain a TAG stop codon, and cloned into the pENTR/D-TOPO[®] Gateway[®] vector to generate entry clones. The entry clones were then transferred into the pcDNA[™] 6.2/GFP-DEST vector using the Gateway[®] LR recombination reaction to create expression clones: 1) human CGI-130 (GenBank Accession # BC003357), which localizes to the cytoplasm; 2) human nuclear splicing factor (GenBank Accession # BC000997), which localizes in the nucleus; and 3) human c-myc (GenBank Accession # BC000141), which localizes with the nucleoli. COS-7 cells were transduced with the an adenovirus expressing

suppressor tRNA molecules (*i.e.*, Tag-On-Demand™ Suppressor Supernatant, Invitrogen Corporation, Carlsbad, CA) at an MOI of 50 followed by transfection with the pcDNA™ 6.2/GFP-DEST expression constructs using the procedure described above. Twenty-four hours post-transfection, GFP fluorescence was assayed using fluorescence microscopy. Fluorescent micrographs for each expression construct are shown in Figure 64. For all three proteins above, methods of the invention result in expression of detectable levels of GFP-tagged recombinant protein as measured by GFP fluorescence by 24 hours post-transfection. Also, the GFP-tagged recombinant protein is correctly localized to the appropriate cellular organelle. The expression construct containing ORF3 (BC000141) is the same construct as the control pcDNA™ 6.2/GFP-GW/p64^{TAG} plasmid described above.

[1042] In some instances, cell toxicity may be observed when transduction and transfection are performed sequentially with a 5-6 hour incubation period after transduction. Although suppression may be clearly observed under these circumstances, the cells may appear unhealthy and may detach from the plate. This phenomenon is not due to either virus alone or transfection alone.

[1043] It has been demonstrated that adenovirus transduction performed *simultaneously* with plasmid transfection results in reduced toxicity and increased plasmid-derived gene expression (see Cotten *et al.*, *Proc Natl Acad Sci U S A* 89(13):6094-8, (1992); Curiel *et al.*, *Proc Natl Acad Sci U S A* 88(19):8850-4, (1991); Guy *et al.*, *Mol Biotechnol.* 3(3):237-48, (1995); Honda *et al.*, *J Virol Methods* 58(1-2):41-51, (1996); Merwin *et al.*, *J Immunol Methods* 186(2):257-66, (1995); Zatloukal *et al.*, *Verh Dtsch Ges Pathol.* 78:171-6, (1994)).

[1044] A series of experiments were performed to directly compare the method of sequential transduction-transfection with a simultaneous transduction/transfection method. In addition to being easier to perform, the simultaneous method resulted in cells that were clearly healthier (normal morphologies and proper adherence) with no evidence of toxicity (Figure 66, right panels) as compared to the sequential method (left panels). As an added benefit, transfection efficiencies were higher making detection of fluorescent cells easier. In Fig. 66, 8 x 10⁴ COS-7 cells were plated in 24-well format and transfected/transduced as follows: Sequential Method (left panels): Cells

were transduced with an adenovirus expressing a suppressor tRNA molecule (Ad-tRNA^{TAG}) at an MOI of 50 for 5 hours, media was replaced and cells were grown overnight. The following morning, cells were transfected with 0.5 µg pcDNA6.2/GFP-GW/p64^{TAG} using 1.5 µl Lipofectamine 2000 for 6 hours, media was replaced and cells grown overnight. GFP fluorescence and brightfield microscope photos were taken the following day. Simultaneous Method (right panels): Cells were transfected/transduced simultaneously. Adenovirus expressing a suppressor tRNA (Ad-tRNA^{TAG}) at an MOI of 50 was applied to cells and pre-formed DNA:Lipid complexes (0.5 µg DNA + 1.5 µl Lipofectamine 2000) were added directly to the virus and cells for 5 hours. Media was replaced and GFP fluorescence and brightfield microscope photos were taken the following day.

[1045] A variety of lipid/DNA ratios were also evaluated using the simultaneous transduction/transfection method (Figure 67). All lipid/DNA ratios tested resulted in healthy, normal looking cells. Western blotting revealed that all ratios tested gave stop suppression greater than 50%, even at MOI 25, with suppression levels ranging from 63% to 87% when simultaneous transduction/transfection was used (Figure 67, upper panels). In Fig. 67, 8 x 10⁴ COS-7 cells were plated in 24-well format and transfected/transduced as described for Sequential and Simultaneous methods above. Various Lipid/DNA ratios were tested, as indicated. 24 hours post transduction/transfection, 5 µl of each total cell lysate was analyzed on 4-12% NuPage gel, MOPS running buffer, transferred to PVDF membrane and Western blot probed with anti-myc antibody. Percent suppression was determined by densitometry

[1046] Gene expression levels were noticeably higher with the simultaneous method and there was no MOI-dependent shut-down of gene expression (*i.e.* no MOI-dependent toxicity) which was visible with the sequential method (Figure 67, compare upper western blot panels with lower panels).

[1047] Thus, methods of producing a fusion polypeptide according to the invention may comprise transducing a host cell with an adenovirus expressing a suppressor tRNA and introducing a nucleic acid molecule encoding a fusion polypeptide into the host cell wherein the host cell is contacted with the adenovirus and the nucleic acid molecule at the same time. Such a method

may comprise seeding host cells, transducing host cells with an adenovirus and contacting host cells with one or more complexes comprising one or more nucleic acid molecules and one or more transfection reagents. Adenovirus may be used at any suitable MOI as discussed above (for example, about 50). Methods may comprise incubating cells in the presence of adenovirus and complexes for from about 10 minutes to about 48 hours, from about 10 minutes to about 36 hours, from about 10 minutes to about 24 hours, from about 10 minutes to about 20 hours, from about 10 minutes to about 16 hours, from about 10 minutes to about 12 hours, from about 10 minutes to about 8 hours from about 10 minutes to about 7 hours, from about 10 minutes to about 6 hours, from about 10 minutes to about 5 hours, from about 10 minutes to about 4 hours, from about 10 minutes to about 3 hours, from about 10 minutes to about 2 hours, from about 10 minutes to about 1 hour, from about 10 minutes to about 45 minutes, from about 10 minutes to about 30 minutes, from about 1 hour to about 48 hours, from about 1 hour to about 36 hours, from about 1 hour to about 24 hours, from about 1 hour to about 20 hours, from about 1 hour to about 16 hours, from about 1 hour to about 12 hours, from about 1 hour to about 8 hours, from about 1 hour to about 7 hours, from about 1 hour to about 6 hours, from about 1 hour to about 5 hours, from about 1 hour to about 4 hours, from about 1 hour to about 3 hours, from about 1 hour to about 2 hours, from about 2 hours to about 48 hours, from about 3 hours to about 48 hours, from about 4 hours to about 48 hours, from about 5 to about 48 hours, from about 6 hours to about 48 hours, from about 7 hours to about 48 hours, from about 8 hours to about 48 hours, from about 9 hours to about 48 hours, or from about 10 hours to about 48 hours. Thus, cells may be incubated in the presence of virus and nucleic acid molecule about 1 hour, about 2 hours, about 3 hours about 4 hours, about 5 hours, about 6 hours, about 7 hours, about 8 hours, about 9 hours, about 10 hours about 12 hours, about 16 hours, about 20 hours, about 24 hours, about 36 hours or about 48 hours.

[1048] Methods may further comprise removing the solution comprising virus and complexes; contacting the cells with a suitable medium (*e.g.*, a complete medium); and incubating the cells under conditions sufficient to produce a fusion polypeptide of the invention. Methods may further comprise detecting

the fusion polypeptide using any technique known to those skilled in the art (e.g., fluorescence microscopy, western blotting, etc). In some instances, toxicity may be observed at later time points and may be cell type specific. Cells should be closely monitored for evidence of toxicity if incubations are carried out for extended periods of time (e.g., longer than about 24-48 hours).

- [1049] Suitable amounts of cells, media, virus, DNA, and transfection reagent (Lipofectamine 2000 = LF2K) for various size tissue culture trays are as follows

	6-well	24-well	96-well
COS cells seeded per well	3×10^5 cells	8×10^4 cells	1×10^4
media/well for culturing	2 ml	500 μ l	100 μ l
MOI 50	3.0×10^7 virus	8×10^6 virus	1×10^6 virus
MOI 25	1.5×10^7 virus	4×10^6 virus	5×10^5 virus
media/well during tdx/tx	1 ml	250 μ l	50 μ l
Transfect DNA/well	2 μ g	500 ng	320 ng
LF2K/well	6 μ l	1.5 μ l	1 μ l

- [1050] Methods of the invention may comprise one or more incubation steps that may be performed in complete medium as described herein, unless otherwise indicated. Cell numbers are for COS-7 cells. Other cell types may require different cell numbers. Cells should be ~90% confluent on the day of virus transduction. Assume that the number of cells double from the time of seeding to the time of transduction for the purpose of calculating MOI.

- [1051] An example of a complete media is DMEM (high glucose) supplemented with FBS to a final concentration of 10%, L-glutamine to a final concentration of 4 mM, and MEM non-essential amino acids to a final concentration of 0.1 mM. These reagents are commercially available from, for example, Invitrogen Corporation, Carlsbad, CA (DMEM (high glucose) catalog no. 11960-044, FBS catalog no. 16000-044, L-glutamine (200 mM) catalog no. 25030-081, MEM non-essential amino acids (10 mM = 100X) catalog no. 11140-050). Media should not be warmed in a water bath. Media should be allowed to come to room temperature in the dark.

- [1052] As discussed above, in some embodiments, methods of the invention may comprise making a cell lysate. One suitable method for making a lysate entails removing media from the wells, adding a suitable lysis buffer (e.g., for a 24-well plate, 100 μ l of 2X NuPAGE® LDS Sample Buffer (4X NuPAGE® LDS sample buffer is available from Invitrogen Corporation, Carlsbad, CA,

catalog no. NP0007 and can be diluted with water)) with 1/50th volume of β -mercaptoethanol to each well; loosening the cells from the plate (*e.g.*, using a pipette tip in a swirling motion to loosen lysed cells from plate); and transferring to a centrifuge tube (*e.g.*, 1.5 ml eppendorf tube). Typically, lysates may be viscous. If it is too viscous, more 2X NuPage LDS Sample Buffer with β -mercaptoethanol can be added up to a total of 200 μ l. Samples should be stored at -80 °C until conclusive western blotting has been completed.

[1053] Methods may further entail heating samples (*e.g.*, at 70 °C for 10 minutes); mixing samples one or more times (*e.g.*, vortexing and centrifugation throughout); loading an aliquot of the sample on an SDS-PAGE gel (*e.g.*, loading 5 μ l (per 100 μ l harvested) on a 4-12% NuPage Bis-Tris gel, Invitrogen Corporation, Carlsbad, CA, catalog no. NP0322BOX). Each gel may contain one or more controls molecular weight markers (*e.g.*, 5 μ l of Magic Mark, Invitrogen Corporation, Carlsbad, CA, catalog no. LC5600, 10 μ l of See Blue Plus 2, Invitrogen Corporation, Carlsbad, CA LC5925), and Western blot controls (*e.g.*, 10 μ l of Positope, Invitrogen Corporation, Carlsbad, CA R90050) heated at 70 °C. Electrophoresis may be performed using, for example, 1X NuPage MOPS SDS Sample Running Buffer (Invitrogen Corporation, Carlsbad, CA, catalog no. NP0001). Add 500 μ l of NuPage Antioxidant (Invitrogen Corporation, Carlsbad, CA, catalog no. NP0005) to the sample running buffer in the "inner core." Electrophoresis may be performed for a suitable period of time under suitable conditions (*e.g.*, for approximately 50 minutes at 200 volts).

[1054] For Western blotting of gel, make up a suitable transfer buffer (*e.g.*, 1X NuPage Transfer Buffer with 20% methanol, Invitrogen Corporation, Carlsbad, CA catalog no. NP0006). Add 1 ml of Antioxidant to 1 liter of 1X NuPage Transfer Buffer. Wet PVDF membranes (Invitrogen Corporation, Carlsbad, CA catalog no. LC2002) in methanol, rinse with H₂O, and then equilibrate in Transfer Buffer. Transfer to PVDF membrane for 90 minutes at 30 volts. Follow all procedures and recommendations in NuPage Bis-Tris gel package insert (Invitrogen Corporation, Carlsbad, CA).

[0999] Following transfer, wash membrane 2X with 20 ml of H₂O. Block membrane using a suitable blocking solution (*e.g.*, that provided in the anti-

mouse Western Breeze Chemiluminescent Kit, Invitrogen Corporation, Carlsbad, CA, catalog no. WB7104). Blocking can be done for 30 minutes up to overnight. Dilute suitable antibody in an appropriate buffer (*e.g.*, for detecting myc protein, anti-myc antibody (Invitrogen Corporation, Carlsbad, CA, catalog nos. R95025, R95225, or 95325) can be diluted 1:5000 in PVDF primary antibody diluent. Incubate antibody solution with membrane, wash, and detect bound antibody using standard techniques suitable for the antibodies used (*e.g.*, chemiluminescent detection, fluorogenic detection, radiolabel detection, etc.). Such techniques are well known to those skilled in the art.

- [1055] When using myc protein that becomes tagged with GFP upon suppression of the stop codon between the coding region of the two proteins (*e.g.*, as expressed from pcDNA6.2/GFP-GW/p64^{TAG}), un-tagged myc (no virus control) should band around Magic Marks 55 kDa and myc tagged with GFP should band around Magic Marks 80 kDa. Densitometry can be done to determine % shift from untagged myc to GFP tagged myc (*i.e.*, percent suppression).
- [1056] Other suitable lysis techniques may be used. For example, harvest cells from 24 well plate with 100 μ l of 1X Tris-Glycine Sample Buffer (Invitrogen Corporation, Carlsbad, CA, catalog no. LC2676) containing 1/50th volume of β -mercaptoethanol to each well. Use a pipette tip in a swirling motion to loosen lysed cells from plate and transfer to a 1.5 ml eppendorf tube. Lysates will be viscous, this is normal. If it is too viscous, more 1X Tris Glycine Sample Buffer with β -mercaptoethanol can be added up to a total of 200 μ l. Samples can be stored at 4 °C. Heat samples at 100 °C for 10 minutes (with vortexing and centrifugation throughout) prior to loading 5 μ l (per 100 μ l harvested) on a 4-20% Tris Glycine gel (Invitrogen Corporation, Carlsbad, CA, EC60252BOX). Western blot analysis may be performed as above or using other suitable techniques know to those skilled in the art.
- [1057] Another suitable lysis technique is as follows. Harvest cells from 24 well plate with 100 μ l of 1X RIPA lysis buffer containing Complete Protease Inhibitor Cocktail (Roche, catalog no. 1 697 498) 50X in H₂O) & Pepstatin (Roche, catalog no. 253 286) 1000X in EtOH). Use pipette tip in a swirling motion to loosen lysed cells from plate and transfer to a 1.5 ml eppendorf tube.

Lysates will be viscous. If it is too viscous, more RIPA lysis buffer can be added up to 150 μ l total. These lysates can be analyzed as above or using other techniques known to those skilled in the art. Bradford Protein assay can be conducted with this lysis buffer to quantitate total amount of protein loaded. Store samples at -80°C until ready to use. Thaw at room temperature, and then keep on ice.

[1058] One suitable protocol for conducting the Bradford protein assay is as follows:

In a 96 well U-bottom flexible polyvinyl chloride plate (Falcon Cat. No. 35-3911)

Perform a 1:10 dilution of cell lysates (e.g., prepared as described above) directly in the wells (9 μ L of H_2O and 1 μ L of lysate).

Load 10 μ L of BSA standard curve to the 96 well plate (1000 $\mu\text{g/ml}$ serial diluted 1:2 down to 15.625 $\mu\text{g/ml}$)

Add 190 μ L Bradford reagent to 10 μ L of diluted lysates and standard curve (1:5 dilution of BioRad Protein Assay Solution, Bio-Rad Corporation, Hercules, CA, catalog no 500-002, 1 ml Solution and 4 ml H_2O)

Read at endpoint wavelength 595 on plate reader and display Reduced numbers.

Use 4 parameter fit for Graph.

[1059] The methods described above may be scaled up or down as appropriate for the number of cells to be used. In some embodiments, particularly those involving high-throughput applications, it may be desirable to analyze a large number of samples in a 96-well format. The protocol for 96-well late applications is the same as the 24 well format described previously with the following modifications.

Seed COS-7 cells at 1×10^4 cells/well in 100 μ l/well in a 96-well plate.

Assume doubling of cells in 24 hour period to 2×10^4 cells/well.

Transduction & Transfection are conducted in 50 μ l/well volumes (100 μ l total- 50 culture media, 50 complexes) for 5 hours. Complexes may be formed using 25 μ l medium (e.g., 1X OPTIMEM) and 1 μ l transfection reagent (e.g., Lipofectamine 2000) and 25 μ l medium and 320 ng DNA incubated separately for 5 minutes at room temperature and then combined and incubated for 20 minutes at room temperature.

Cells may be harvested with 30-60 μ l of Sample Buffer or 30-50 μ l Lysis Buffer, depending upon viscosity.

Load 10 μ l (per 30 μ l harvested) of samples harvested with Sample Buffer on gel.

- [1060] The most likely sources of low suppression efficiency include poor quality of cell stock at time of plating experiment (*i.e.*, cells very confluent, media not pink) and the use of old media. Media should be freshly prepared for use in transduction/transfection.
- [1061] In another specific example of methods of the invention, COS-7 cells were transduced with an adenovirus expressing suppressor tRNA molecules (*i.e.*, the Tag-On-Demand™ Suppressor Supernatant) at various MOIs following the procedures described above and simultaneously transfected with the pcDNA™ 6.2/GFP-GW/p64^{TAG} plasmid using Lipofectamine™ 2000 Reagent and the procedure described above. Twenty-four hours post-transfection, cell lysates were prepared and analyzed by Western blot using the Anti-myc Antibody and the WesternBreeze® Chemiluminescent Anti-Mouse Kit (Catalog no. WB7104) to detect native and GFP-tagged p64^{TAG} (c-myc) protein. The results are shown in Figure 68. In Figure 68, Lane 1 contains MagicMark™ MW Standard, lane 2 contains untransfected COS-7 cells, lane 3 contains cells transduced at an MOI = 0, lane 4 contains cells transduced at an MOI = 50, lane 5 contains cells transduced at an MOI = 100, lane 6 contains cells transduced at an MOI = 200. GFP-tagged c-myc protein is produced and detectable by Western blot within 24 hours post-transfection. The % suppression achieved is > 80% when transducing cells at an MOI \geq 50. In this experiment, increasing the MOI has little effect on the suppression efficiency. Maximal levels of GFP-tagged c-myc protein are produced using an MOI = 50.
- [1062] In another working example of methods of the invention, 96 of Invitrogen's Ultimate™ Human ORF Clones encoding 96 different kinases were transferred into the pcDNA™ 6.2/V5-DEST vector using the Gateway® LR recombination reaction to generate expression clones. The expression constructs were purified, and the plasmid DNA (ranging from 20 ng to 300 ng) was transfected using Lipofectamine™ 2000 Reagent into COS-7 cells (plated in 96-well format) that had been transduced with the Tag-On-Demand™

Suppressor Supernatant at an MOI of 50 following the procedure described above. Forty-eight hours post-transfection, cell lysates were prepared and analyzed by Western blot using the Anti-V5 Antibody (Invitrogen, Catalog no. R961-25) and the WesternBreeze[®] Chemiluminescent Anti-Mouse Kit (Catalog no. WB7104) to detect V5-tagged fusion polypeptide. Using this antibody, native polypeptide is not detected. V5-tagged fusion polypeptide is produced and detectable by Western blot within 48 hours post-transfection. The levels of V5-tagged fusion polypeptide produced vary widely from gene to gene. This is expected since transfection and expression conditions are not optimized for each gene and can vary depending on the nature of the gene of interest. In this working example, the amount of plasmid DNA transfected and the amount of cell lysate loaded on the polyacrylamide gel were not quantitated for each sample (*i.e.* transfection and expression conditions were not optimized). In addition, antibodies to each of the 96 different kinase proteins do not exist. This example demonstrates the utility of methods of the invention to quickly screen and analyze the expression of large numbers of recombinant proteins for which antibodies do not currently exist.

[1063] As discussed above, methods of the invention may be optimized using routine experimentation in order to produce a desired quantity of fusion polypeptide of the invention. A variety of factors may be considered when optimizing experimental conditions. For example, in some initial experiments, low expression of the desired fusion polypeptide may be observed. This may be due to any one or more or a number of reason such as 1) low suppression efficiency; 2) phenotypic effects observed; 3) poor transfection efficiency; and 4) improper timing of the assay (*i.e.*, assayed too early or too late).

[1064] Low suppression efficiency may result in a reduced production of a desired fusion polypeptide and may be observed when the host cells used were not healthy and/or were not plated at the correct density. One skilled in the art can optimize this factor by ensuring that cells are healthy and > 95% viable before plating and are plated at the proper density. Low suppression efficiency may be observed when the media used was not fresh. This factor can be optimized by preparing fresh media for use in the practice of the present invention. Low suppression efficiency may be observed if the host cells are transduced with too little virus (*i.e.* low MOI). One skilled in the at

can optimize transduction by testing varying MOIs starting at about 50. Low suppression efficiency may be observed when host cells express low levels of CAR. One skilled in the art can optimize this factor by using a cell line that expresses suitable levels of CAR (*e.g.* COS-7, CHO, HeLa). Low suppression efficiency may be observed if host cells are not transduced for an optimum length of time. One skilled in the art can optimize this factor by transducing for various periods of time, for example, about 5-6 hours.

[1065] Phenotypic effects on host cells caused by methods of the invention may result in reduced production of a desired fusion polypeptide of the invention. Factors that may be optimized to mitigate phenotypic effects include the length of incubation after transduction and transfection. One skilled in the art can optimize this factor by assaying for fusion polypeptide at various times after transduction and transfection (*e.g.*, 24-48 hours). Phenotypic effects may be observed if host cells used are sensitive to transduction and transfection procedure. One skilled in the art can optimize this factor by performing methods of the invention in a different host cell line and/or by making a stable cell line containing the nucleic acid molecule encoding the fusion polypeptide and subsequently introducing a nucleic acid molecule encoding a suppressor tRNA (*e.g.*, transducing with a virus expressing a suppressor tRNA).

[1066] Poor transfection efficiency may result in a reduced production of a desired fusion polypeptide of the invention. One skilled in the art can readily optimize this factor by testing various transfection reagent to identify one that provides a high transfection efficiency for the cell line being used.

[1067] Reduced production of a fusion polypeptide of the invention may be observed when fusion polypeptide expression is assayed at a sub-optimal time (*i.e.*, too early or too late). One skilled in the art can optimize this factor by assaying at various times to determine when optimum expression is observed (*e.g.*, by conducting a time course of expression).

EXAMPLE 17

[1068] In some embodiments, the invention provides nucleic acid molecules comprising all or a portion of a viral genome that comprise transcriptional

regulatory sequences (*e.g.*, promoters, repressors, etc.). In one specific embodiment, the invention provides nucleic acid molecules comprising all or a portion of a viral genome (*e.g.*, a retroviral genome) that comprise a repressor sequence. A repressor sequence may inhibit or prevent transcription of a nucleotide sequence to which it is operably linked.

[1069] A repressor sequence may bind or may be bound by one or more molecules (*e.g.*, peptides, small molecules, etc.). In one embodiment, a repressor sequence may bind a protein (*e.g.*, a repressor protein). One example of a repressor to which binds a repressor protein is the tetracycline operator to which binds the tetracycline repressor protein. In the absence of tetracycline, the repressor protein binds to the tetracycline operator and prevents or inhibits transcription of a nucleotide sequence to which it is operably linked. In the presence of tetracycline, the repressor protein binds tetracycline and no longer binds to the repressor sequence.

[1070] In some embodiments, a repressor sequence and a promoter sequence may be operably linked to a sequence of interest. In embodiments of this type, the repressor sequence may prevent transcription of the sequence of interest from the promoter under some conditions (*e.g.*, when a repressor protein is bound to the repressor sequence) and not under other conditions (*e.g.*, in the absence of repressor protein or under conditions in which the repressor protein is not bound to the repressor sequence).

[1071] In one embodiment of the invention, a nucleic acid molecule comprising all or a portion of a lentiviral genome may also comprise a repressor sequence (*e.g.*, the tetracycline operator) and/or may comprise a nucleic acid sequence encoding a polypeptide that binds to a repressor (*e.g.*, the tetracycline repressor protein). Embodiments of this type may be used to construct host cells and/or host cell lines comprising a nucleic acid sequence of interest operably linked to a repressor sequence. Optionally, such host cells and/or host cell lines may comprise a nucleic acid sequence encoding a polypeptide that binds to the repressor sequence. In a particular embodiment, the present invention encompasses host cells and/or host cell lines in which a sequence of interest is operably linked to a tetracycline repressor sequence and a promoter sequence and further comprise a nucleic acid sequence encoding the tetracycline repressor protein. Such host cell lines provide the ability to

regulate the transcription of the sequence of interest, *i.e.*, in the absence of tetracycline, the sequence of interest is not transcribed or is transcribed at an insignificant level while in the presence of tetracycline the sequence of interest is transcribed at a much higher level (*i.e.*, transcription is induced by tetracycline).

[1072] Host cells and/or host cell lines according to the invention may be any type of cell (*e.g.*, dividing or non-dividing cells) and may be isolated cells or may be within a larger organism. Methods of the invention allow controlled gene expression in tissue culture cells and whole organisms.

[1073] In some embodiments, the present invention provides a method of making a cell expressing a repressor protein and cells made by such methods. Methods may comprise introducing into a cell a nucleic acid molecule comprising all or a portion of a viral genome and encoding a repressor protein. Such methods may also comprise selecting for a cell stably expressing the repressor.

[1074] In some embodiments, the present invention comprises methods of expressing a sequence of interest comprising introducing into a host cell expressing a repressor protein, one or more nucleic acid molecules comprising a sequence of interest operably linked to a repressor and a promoter. In some embodiments, a nucleic acid molecule comprising a sequence of interest operably linked to a repressor and a promoter may comprise all or a portion of a viral genome (*e.g.*, a lentiviral genome). Methods may further comprise incubating the cell under conditions in which the repressor protein does not bind to the repressor sequence. Such conditions may include incubation in the presence of a molecule that prevents the repressor protein from binding to the repressor sequence. For example, when the repressor sequence is the tetracycline operator and the repressor protein is TetR, such conditions may comprise incubating the cell in the presence of tetracycline.

[1075] In one particular embodiment, the present invention provides two nucleic acid molecules (*e.g.*, plasmids, viral vectors etc.) that may be used in the practice of the invention. A first nucleic acid molecule comprises a repressor sequence and a promoter and may comprise a sequence of interest operably linked to the repressor and promoter. A first nucleic acid molecule may also comprise one or more recognition sequences (*e.g.*, recombination

sites, topoisomerase sites, restriction enzyme sites, etc.). One non-limiting example of a first nucleic acid molecule is the plasmid, pLenti4/TO/V5-DEST, which contains two copies of the tetracycline operator sequence (TO) within the CMV promoter (CMVTetO₂). A map of this vector is provided as Figure 70A and the nucleotide sequence is provided in Table 31. This plasmid also contains two recombination sites that do not recombine with each other. A sequence of interest may be operably linked to the promoter and repressor using any technique known in the art. In one embodiment, a sequence of interest may be operably linked to the promoter and repressor by conducting a recombination reaction between a sequence of interest flanked by recombination sites and the nucleic acid molecule of the invention. For example, pLenti4/TO/V5-DEST (Figure 70A) can be reacted with a sequence of interest flanked by *attR1* and *attR2* sites to operably link the sequence of interest to the CMV promoter and tetracycline operator in a LR-recombination reaction. The reaction places the sequence of interest downstream of CMVTetO₂ for regulated expression in the presence of the tetracycline repressor protein.

[1076] A second nucleic acid molecule of the invention may express one or more proteins that interact with repressor sequences. One non-limiting example of a repressor protein is the tetracycline repressor protein (TetR). One example of a suitable second nucleic acid molecule is the repressor plasmid pLenti6/TR, which expresses TetR. A map of this vector is provided as Figure 69 and the nucleotide sequence is provided as Table 32. TetR binds the tetracycline operator sites in CMVTetO₂ promoter on the expression vector and blocks transcription from the promoter in the absence of inducer. When tetracycline inducer binds TetR, however, the latter dissociates from the promoter and transcription proceeds.

[1077] Methods of the of the invention may be use to regulate the expression of a sequence of interest in transformed dividing cells and in difficult-to-transfect growth-arrested primary cells. Methods of the invention may be used for transient or stable gene regulation. Induction of expression may be from about 2-fold to about 100-fold, from about 5-fold to about 100-fold, from about 10-fold to about 100-fold, from about 25-fold to about 100-fold, from about 50-fold to about 100, from about 75-fold to about 100-fold, from about

5-fold to about 5-fold to about 70-fold, from about 10-fold to about 70-fold, from about 25-fold to about 70-fold, from about 50-fold to about 70-fold, or from about 60-fold to 70-fold. Thus, gene expression may be induced about 5-fold, about 10-fold, about 20-fold, about 30-fold, about 40-fold, about 50-fold, about 60-fold, about 70-fold, about 80-fold, about 90-fold, or about 100-fold.

[1078] In some embodiments of the invention, the present invention may comprise a viral stock that may be used to transduce host cells. Stocks may be at any suitable concentration of virus. For example, pLenti6/TR may be used to create a viral stock at about 1×10^5 cfu/ml or greater, which may be used to stably transduce TetR into target cells in blasticidin-containing media. Cells transduced in this fashion will typically express TetR protein at a level detectable by Western blot.

[1079] In another aspect of the invention, the present invention provides nucleic acid molecules comprising a promoter sequence and a repressor sequence to which a sequence of interest may be operably linked. Such nucleic acid molecules may be used to create a viral stock. For example, recombinational cloning of the lacZ gene into pLenti4/TO/V5-DEST and packaging the resulting pLenti4/TO/V5-GW/lacZ vector may be used to produce a viral stock at 1×10^5 cfu/ml or greater. Such a viral stock may be used to transduce a host cell and express a sequence of interest (*e.g.*, the lacZ sequence).

[1080] In some embodiments, nucleic acid molecules comprising a promoter and repressor sequence operably linked to a sequence of interest and nucleic acid molecules comprising a sequence encoding a polypeptide that binds to the repressor sequence may be introduced into a host cell. In methods of this type, nucleic acid molecules may be introduced simultaneously or sequentially. Typically, once both types of nucleic acid molecule have been introduced into a host cell, expression of the sequence of interest will be inducible. For example, transient co-transduction of Lenti6/TR and Lenti4/TO/V5-GW/LacZ may show at least 20-fold induction in HT1080 cells. In some embodiments, after both types of nucleic acid molecule are introduced into a host cell, a stable cell line may be produced, for example, by selecting for cells expressing both the sequence of interest and the repressor protein. In one example, cell

lines may be made that contain Lenti6/TR and Lenti4/TO/V5-GW/lacZ and such cells may show at least 20-fold induction.

[1081] One aspect of the present invention is the capability to regulate the expression of a sequence of interest in a non-dividing cell. In a specific embodiment, the present invention provides non-dividing host cells containing a sequence of interest, the expression of which is regulatable, for example, is inducible by the addition of tetracycline to the growth medium of the cell. The present invention contemplates compositions comprising such cells and further comprising one or more component selected from a group consisting of an inducer (*e.g.*, tetracycline), a growth medium, and a buffer.

[1082] A nucleic acid molecule expressing the tetracycline repressor protein may be constructed using any technique known in the art. For example, a nucleic acid fragment containing the tetracycline repressor coding sequence can be cloned using any technique known in the art. The nucleotide sequence of a nucleic acid fragment containing the coding sequence for the tetracycline repressor is provided as Table 35. The 1.4 kb fragment also contains the β -globin intron. The 1.4 kb TetR-containing fragment was cloned into pLenti6/V5 (Invitrogen Corporation, Carlsbad, CA). A map of pLenti6/V5 is provided as Figure 71 and the nucleotide sequence is provided as Table 33. The resulting plasmid, pLenti6/TR, was verified by restriction digest and sequence analyses. A map of pLenti6/TR is shown in Figure 69. pLenti6/TR can be used to generate blasticidin resistant mammalian cells that stably express the tetracycline repressor, TetR.

[1083] Nucleic acid molecules comprising a promoter sequence and a repressor sequence can be constructed using any techniques known in the art. For example, pLenti4/TO/V5-DEST was created from pLenti3/V5-TREx (Invitrogen Corporation, Carlsbad, CA), by replacing the neomycin resistance gene of the latter with the zeocin resistance gene. pLenti3/V5-TREx contains the CMV promoter and Tet operators of pT-REx-DEST30 (Invitrogen Corporation, Carlsbad, CA catalog no. 12301016). A map of pLenti3/V5-TREx is provided as Figure 72 and the nucleotide sequence is provided in Table 34.

[1084] pLenti3/V5/TREx was digested with Sall, filled in using Klenow and then digest with KpnI and the 5917 bp vector backbone was gel isolated.

Next, pLenti4/V5-DEST (Invitrogen Corporation, Carlsbad, CA catalog nos. K498000 and V49810) was digested with SpeI, Klenow filled-in, then digested with KpnI. A 2682 bp fragment of pLenti4/V5-DEST containing a GATEWAY™ Destination cassette, SV40 promoter and Zeocin resistance cassette, was gel isolated and ligated to the SalI-Klenow-KpnI processed pLenti3/V5-TREx. The ligation mixture was transformed into DB3.1 and selected on LB media containing Amp (100 µg/ml) and chloramphenicol (15 µg/ml). Colonies of the transformants were analyzed by restriction analysis. A map of pLenti4/TO/V5-DEST is shown in Figure 70A. The GATEWAY™ Destination vector pLenti4/TO/V5-DEST contains the tet-regulated CMVTetO₂ T-REx promoter (consisting of CMV promoter and two tet operator sites). TetR protein binds the tetO sites to inhibit gene transcription; tetracycline relieves the transcription inhibition. pLenti4/TO/V5-DEST confers zeocin resistance and allows in-frame fusion of genes-of-interest to the V5 epitope tag.

[1085] pLentiTO/V5-GW/lacZ was generated by standard Gateway LxR reaction between pLenti4/TO/V5-DEST and pENTR/dT-lacZ no stop (Invitrogen Corporation, Carlsbad, CA). Clones of pLenti4/TO/V5-GW/lacZ were confirmed by restriction and sequence analyses. A map of pLenti4/TO/V5-GW/lacZ is shown in Figure 70B.

[1086] 293FT (Invitrogen Corporation, Carlsbad, CA catalog no. R70007) and GripTite 293 (Invitrogen Corporation, Carlsbad, CA catalog no. R79507) cells were cultured in DMEM/10% FBS/L-glutamine/non-essential amino acids/penicillin/streptomycin containing 500 µg/ml G418. MJ90 primary human foreskin fibroblasts (Grand Island) and HT1080 human fibrosarcoma (ATCC #CCL-121) were cultured in DMEM/10% FBS/non-essential amino acids/penicillin/streptomycin. 10 µg/ml blasticidin was used to select for stable pLenti6/TR-transduced cells. MJ90 primary cells were growth arrested by contact inhibition. Briefly, 1×10^5 cells were plated per well of a 6-well plate and media changes were performed every 3 days for 7 to 14 days, or until a quiescent monolayer was achieved.

[1087] For virus production, 5×10^6 293FT cells were plated per 100 mm plate. Twenty-four hours later, the culture medium was replaced with 5 ml OptiMem/10%FBS and cells were quadruple co-transfected, as follows. 12 µg

DNA total, at a mass ratio of 1:1:1:1 pLenti6/TR or pLentiTO/V5-GW/lacZ :pLP1:LP2:pLP/VSVG (3 µg of each DNA) was mixed with 1.5 ml of OptiMem media. In a separate tube, 36µl of Lipofectamine 2000 was also mixed with 1.5 ml of OptiMem media. After a 5-minute incubation period at room temperature, the two mixtures were combined and incubated at room temperature for an additional 20 minutes. At the completion of the incubation period, the transfection mixture was added to the cells dropwise and the culture plate was gently swirled to mix. The following day the transfection complex was replaced with complete media (DMEM, 10% FBS, 1% penicillin/streptomycin, L-glutamine and non-essential amino acids). Forty-eight to seventy-two hours post transfection, the virus-containing supernatants were harvested, centrifuged at 3000 rpm for 5 minutes to remove dead cells and placed in cryovials in 1 ml aliquots. Titers were performed on fresh supernatants (see below) and the remaining viral aliquots were stored at -70 °C.

[1088] All applications of virus to cells were performed in the presence of 6 µg/ml polybrene (Sigma #H9268) and media changes were performed 12-24 hours post transduction. For titering virus, 6-well plates were seeded at 2×10^5 cells per well with HT1080 cells the day before transduction. One well served as an untransduced control (mock) and the remaining five wells contained 1 ml each of ten-fold serial dilutions of viral supernatant ranging from 10^{-2} to 10^{-6} . The dilutions were mixed by gentle inversion prior to adding to cells. 6 µg/ml of polybrene was added to each well. The plate was gently swirled to mix. The following day, the media was replaced with complete media. Forty-eight hours post-transduction, the cells were placed under 10 µg/ml blasticidin or 100 µg/ml zeocin selection, as appropriate. In particular, Zeocin selection was done as follows: 24-hour post-transduction cells were trypsinized from 6-well plates and expanded into 100 mm plates. 24 hrs after expansion into 100 mm plates, 100 µg/ml Zeocin was added to the transduced cell culture medium for selection. After 7 to 10 days of blasticidin selection, or two-to-three weeks of zeocin selection, the resulting colonies were stained with crystal violet : A 1% crystal violet solution was prepared in 10% ethanol. Each well was washed with 2 ml PBS followed by 1 ml of crystal violet solution for 10 minutes at room temperature. Excess stain was

removed by two 2 ml PBS washes and colonies visible to the naked eye were counted to determine the viral titer of the original supernatants.

- [1089] Cell lysates for western blot and Tropix Assays were prepared as follows: Culture media were aspirated and cells were washed 1x with PBS and followed by incubation in Versene (Invitrogen Corporation, Carlsbad, CA catalog no. 15040066) for 2 minutes at room temperature. Detached cells were pelleted in Eppendorf Tubes and lysed in ice-cold 100 μ l NP-40 lysis buffer (50 mM Tris, 150 mM NaCl, 1% NP-40, pH 8.0) containing protease inhibitors. Lysates were centrifuged at 14000 rpm for 5 min to pellet cellular debris; the supernatant was collected and frozen at -70° C until needed for assays. Protein concentrations were determined using BioRad Protein Assay protocol according to the manufacturer's (Biorad) recommendations.
- [1090] Western blots were performed using 20 μ g of normalized protein in a 4X loading dye. Samples were run on a Novex^R Tris-Glycine 4-20% Gel (Invitrogen Corporation, Carlsbad, CA catalog no. EC60252BOX), at 200 volts for 45 minutes. Protein was transferred to a nitrocellulose membrane and were detected with a WesternBreeze^R Chemiluminescent Kit (Invitrogen Corporation, Carlsbad, CA catalog no. WB7104) using polyclonal anti-TetR and monoclonal anti-V5 (Invitrogen Corporation, Carlsbad, CA catalog no. R96025) primary antibodies, as appropriate.
- [1091] pLenti6/TR and pLenti4/TO/V5-GW/lacZ were transfected into 293FT cells, in the presence of Virapower Packaging Mix, to produce the respective viruses. pLenti6/TR and pLenti4/TO/V5-GW/lacZ produced viral titers of 6×10^5 and 1×10^5 cfu/ml respectively. Thus introduction of β -globin intron and TetR into pLenti6, and introduction of Tet Operators into pLenti4-DEST, do not compromise virus packaging and transduction efficiency.
- [1092] Materials and methods of the invention may be used for a wide variety of purposes. For example, a nucleic acid molecule expressing a repressor protein (e.g., Lenti6/TR virus) may be used to generate repressor expressing cell lines. Such cell lines may be transduced with a nucleic acid molecule comprising promoter and repressor sequences operably linked to a sequence of interest (e.g., Lenti4/TO/V5-GW/sequence of interest) and then expression of the sequence of interest may be regulated (e.g., using tetracycline). Another use of the materials and methods of the invention is to simultaneously

cotransduce a nucleic acid molecule encoding a repressor and a nucleic acid molecule comprising promoter and repressor sequences operably linked to sequence of interest (*e.g.*, Lenti6/TR and Lenti4/TO/V5-GW/sequence of interest) into primary non-dividing cells, then regulate expression of the sequence of interest (*e.g.*, using tetracycline).

[1093] HT1080 cells were transduced with Lenti6/TR virus at MOI of 1, 10 or 32 and were selected in blasticidin medium until mock transduced cells had died-off. The blasticidin-resistant cells were next transduced with Lenti4/TO/V5-GW/lacZ virus at MOI of 5. Twenty-four hours after transducing with Lenti4/TO/V5-GW/lacZ, 1 μ g/ml tetracycline was added to the culture medium. Cells were incubated in the inducer-supplemented medium for 48 hrs. Thereafter, cell lysates were prepared and analyzed for gene expression by (i) assaying for lacZ activity; (ii) performing western blot for lacZ-V5 fusion using anti-V5 antibody; (iii) western blot for TetR.

[1094] Increasing the amount of transduced TetR virus reduced lacZ expression in the absence of tetracycline. Tetracycline at 1 μ g/ml induces lacZ expression to levels approaching full-strength CMV promoter activity. To determine fold induction, the ratios of β -galactosidase activities in the presence and absence of tetracycline (for a given MOI) were calculated. Induction of lacZ expression was 4-, 17- and 27-fold at TetR MOI of 1, 10 and 32, respectively, indicating that induction was dependent on the amount of TetR. Western blot analyses using anti-V5 antibody was consistent with the β -galactosidase enzymatic activity data. Expression of TetR protein was confirmed by western blot using polyclonal anti-TetR antibody.

[1095] These results confirm CMVTetO₂ and TetR in the lentiviral vectors to be functional and responsive to tetracycline. The relatively high level basal transcription from CMVTetO₂ at lower Lenti6/TR MOIs could result from the fact that not all blasticidin resistant cells generated at the low TetR MOIs actually express TetR. Those cells that do not express TetR would express lacZ from CMVTetO₂ promoter without inhibition and produce a high background. By contrast, at high Lenti/TR MOIs, close to 100% of blasticidin-resistant cells generated would express TetR, inhibit transcription from CMVTetO₂ promoter and produce lower background lacZ expression.

- [1096] The data in HT1080 cells showed that lower basal transcription in a cell population is achieved at higher TetR levels. Therefore when testing induction in GripTite 293 cells, Lenti6/TR was transduced at MOI = 10 and MOI = 32 to generate blasticidin-resistant GripTite-10 and GripTite-32 populations, respectively. These populations were transduced with Lenti4/TO/V5-GW/lacZ virus at MOI = 1 or MOI = 5 and tested for lacZ induction. Tetracycline was used at 1 µg/ml or at 5 µg/ml to determine if inducer was limiting at higher TetR concentrations.
- [1097] TetR effectively inhibited lacZ expression in GripTite-10 cells in the absence of inducer and this repression was relieved by tetracycline. 1 µg/ml tetracycline was nearly as effective as 5 µg/ml tetracycline in inducing gene expression. Fold induction was calculated as induced:uninduced ratios at a given Lenti4/TO/V5-GW/lacZ MOI and tetracycline concentration. LacZ expression was induced over 27-fold at Lenti4/TO/V5-GW/lacZ MOI = 5 compared to just above 7-fold at Lenti4/TO/V5-GW/lacZ MOI = 1. Western blot analyses using anti-V5 antibody reflected β-gal enzymatic Tropix data. Expression of TetR protein was confirmed by western blot using polyclonal anti-TetR antibody.
- [1098] The results in GripTite 293-10 cells were recapitulated in GripTite-32 cells. As in GripTite 293-10 cells, 1 µg/ml was nearly as effective as 5 µg/ml tetracycline in inducing gene expression in GripTite-32 cells. The fold lacZ induction was significantly higher in GripTite 293-32 cells however and ranged from 57 to 72 fold at Lenti4/TO/V5-GW/lacZ MOI = 1 and Lenti4/TO/V5-GW/lacZ MOI = 5, respectively.
- [1099] The data indicate that 1 µg/ml tetracycline is not limiting in inducing lacZ expression. LacZ induction was higher at Lenti4/TO/V5-GW/lacZ MOI = 5 than at MOI = 1. Thus the amount of expression may be adjusted by altering the MOI of the virus containing a sequence of interest operably linked to a promoter and repressor sequence (*e.g.*, higher MOI for higher expression level when de-repressed, lower MOI for lower expression level when de-repressed). The increased MOI has little effect on background uninduced levels when TetR is not limiting (*e.g.*, MOI of 10 and 32).
- [1100] In one particular embodiment, materials and methods of the invention may be used to regulate gene expression in non-dividing primary cells. MJ90

cells are contact-inhibited primary fibroblasts that undergo growth arrest at confluence and are refractory to both lipid transfection and transduction by Moloney retroviral vectors. MJ90 cells were transduced with 2×10^6 cfu/well Lenti6/TR virus for 24 hrs followed by transduction with 2×10^6 cfu/well of Lenti4/TO/V5-GW/lacZ virus (estimated MOI = 7.5 each). Twenty-four hours after transducing with Lenti4/TO/V5-GW/lacZ, lacZ expression was induced with 1 μ g/ml tetracycline for 48 hrs. Lysates from transduced cells were analyzed for protein induction. TetR repressed expression of lacZ over 90%, resulting in a 10-fold induction. It is worth noting that the preceding experiment was performed with equal MOI of Lenti6/TR and Lenti4/TO/V5-GW/lacZ. Higher Lenti6/TR MOI, or different Lenti6/TR : Lenti4/TO/V5-GW/lacZ ratios may be used to give higher inducibility. The demonstration that the present invention can regulate gene expression in quiescent primary cells is significant especially since the cells are hard to transfect and resist transduction by Moloney retroviral vectors.

- [1101] Nucleic acid molecules of the invention comprising promoter and repressor sequences operably linked to a sequence of interest may be used in conjunction with any nucleic acid molecule expressing a repressor protein. For example, Lenti4/TO/V5-GW/lacZ virus was transduced into the Flp-In T-REx 293 product cell line (Invitrogen Corporation, Carlsbad, CA catalog no. R78007) at MOIs of 1 and 2.5. Gene expression was induced with 1 μ g/ml tetracycline for 48 hrs. Tetracycline induced lacZ expression from Lenti4/TO/V5-GW/lacZ in Flp-In T-REx 293 cells. Increasing the amount of transduced Lenti4/TO/V5-GW/lacZ from MOI=1 to MOI=2.5 increased induction from 16-fold to 24-fold, respectively similar to the results in GripTite-10 and GripTite-32 populations.

EXAMPLE 18

- [1102] In some embodiments, the present invention provides a method of covalently attaching an enzyme (*e.g.*, a topoisomerase) to a nucleic acid molecule. In one aspect, a nucleic acid molecule for use in methods of this type may comprise a restriction enzyme recognition sequence (*e.g.*, a TypeIIIs restriction enzyme recognition) and a topoisomerase recognition sequence. In

some embodiments, a TypeII_s recognition sequence may be located adjacent to a topoisomerase recognition sequence. In this regard, adjacent means that the cleavage sites of the two enzymes may be within from about 1 to about 50, from about 1 to about 45, from about 1 to about 40, from about 1 to about 35, from about 1 to about 30, from about 1 to about 25, from about 1 to about 20, from about 1 to about 15, from about 1 to about 10, from about 1 to about 9, from about 1 to about 8, from about 1 to about 7, from about 1 to about 6, from about 1 to about 5, from about 1 to about 4, from about 1, to about 3, or from about 1 to about 2 base pairs from each other. Any TypeII_s enzyme may be used. In some embodiments, a suitable TypeII_s enzyme may leave a 3'-overhanging sequence. Suitable TypeII_s enzymes include *BaeI*.

[1103] With reference to Figure 73, a nucleic acid molecule of the invention may comprise two topoisomerase recognition sites and two TypeII_s recognition sites, for example with the two restriction enzyme sites between the two topoisomerase sites. Optionally a nucleic acid molecule of this type may comprise nucleic acid sequence between the restriction enzyme sites. The nucleic acid sequence between the restriction enzyme sites may encode a polypeptide, for example, a selectable marker such as the *ccdB* gene.

[1104] The restriction enzyme sites may be located such that a 3'-overhang of a desired length is produced on the strand containing the topoisomerase cleavage site (after the 3'-T in Fig. 73). The location of the topoisomerase cleavage site may be varied with respect to 3'-most nucleotide of the strand containing the cleavage site. This may be useful in generating a 5'-overhang on the opposite strand after topoisomerase cleavage in order to generate a sequence that can invade a double-stranded insert (see Figure 47).

[1105] After restriction enzyme cleavage, the cleaved vector may be contacted with an oligonucleotide that anneals to the 3'-overhanging sequence and/or may be contacted with a topoisomerase.

[1106] In some embodiments, methods of the invention may comprise digesting a nucleic acid molecule of the invention (*e.g.*, 20 µg) with a TypeII_s restriction enzyme (*e.g.*, 100 Units of *BaeI*, New England Biolabs, catalog no. R0613S), for example, in a final volume of 250 µl. Any other restriction enzyme known in the art may be also be used. The reaction may be carried out in a suitable buffer (*e.g.*, 1X NEBuffer 2 with 100 µg/ml of BSA and 20

μ M S-adenosylmethionine, New England Biolabs) under suitable conditions (e.g., at 37 °C for 6 hours). The digestion may be terminated, for example, with the addition of 250 μ l of Phenol/Chloroform (Invitrogen Corporation, Carlsbad, CA, Cat. #15593-031) and mixing. The organic and aqueous phases may be separated by centrifugation at 14,000 X g at 4 °C for 5 minutes. The aqueous (top) layer may be transferred to a new tube and 25 μ l of 3M sodium acetate (pH 5.2) may be added and mixed. This may be followed by 625 μ l of 100% ethanol and incubation in ice for 5 minutes. Precipitated DNA may be harvested by centrifugation at 14,000 X g for 5 minutes at 4 °C. The DNA pellet may be washed with 500 μ l of 70% ethanol, harvested by centrifugation at 14,000 X g for 5 minutes at 22 °C. The pellet may be allowed to dry and then resuspended in 100 μ l of TE. The DNA concentration may be determined by its optical density at 260 nm.

[1107] The digested vector may be contacted with an oligonucleotide that anneals to all or a portion of the 3'-overhang produced by the restriction enzyme and/or with a suitable topoisomerase enzyme (e.g., Vaccinia DNA Topoisomerase) in a suitable buffer (e.g., 1X NEBuffer #1, New England Biolabs), for example, in a final volume of 50 μ l. The reaction may be incubated under suitable conditions (e.g., 25 °C for 15 minutes). Then reaction may be terminated with the addition of 5 μ l of 10X Stop Buffer. The topoisomerase-linked vector may be purified by gel electrophoresis (see, Heyman, *et al. Genome Research* 9:383-392 (1999)).

[1108] Having now fully described the present invention in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious to one of ordinary skill in the art that the same can be performed by modifying or changing the invention within a wide and equivalent range of conditions, formulations and other parameters without affecting the scope of the invention or any specific embodiment thereof, and that such modifications or changes are intended to be encompassed within the scope of the appended claims.

[1109] All publications, patents and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains, and are herein incorporated by reference to the

same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference.

Table 6: Nucleotide sequence of pAd/CMV/V5-DEST.

catcatcaataatataacctatttttgattgaagccaatatgataatgagggggtggagtttgtgacgtggcgcgggcggtgggaacgg
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tgccattttcgcggaactgaataagagggaagtgaatctgaataattttgtttactcatagcgcgtaattttgtctaggccgcg
gggactttgacctttacgtggagactcggccaggtgttttctcaggtgtttccgcgttcgggtcaaaagtggcgttttattattatagtc
agtcgaagcttgatccgggtacctctagaattctcgagcgccgctagcgacatcgatcccgatcccctatggtcgactctcagta
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cgagacccattggggccaata

Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

cgccccgctttcttcttccccaccccccccccaagttcgggtgaaggcccagggtcgcagccaacgtcggggcggcaggcc
ctgccatagcagatccgattcgacagatcactgaaatgtgtggcgctggttaagggtgggaaagaatataagttgggggtcttatg
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Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

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Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

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Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

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Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

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 gtgtccacttaactcaggcacaacctccgcggcagctcgggaagtttctactccacaggtcgcgcaccatcaccacacgcgttag
 caggtcggggccgcatatctgaagtcgagttggggcctccgcctgcgcgcgagttgcgatacacagggttcagcagcactgga
 acatatcagcgcc

Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

ggggtggtgcacgctggccagcacgctctgtcggagatcagatccgcgtccaggtcctccgcgttgcagggcgaacggagtgaa
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 gtctggcggttaggatacagcgctgcataaaagccttgatctgtttaaaggccacctgagccttgcgccttcagagaagaacatgcc
 gcaagacttgcggaaaactgattggccggacaggccgcgtcgtgcacgcagcaccttgcgtcgggtgttgagatctgcaccacatt
 cggccccaccgggttcttcacgatcttggccttgcctagactgctccttcagcgcgcgtgcccgttttcgtcgtcacatccattcaatcac
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 tttagagttcgcctttagatcgttatccacgtggtactgtccatcagcgcgcgcagcctccatgcccttctccacgcagacacgatc
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 cctcggactcgatacggcctcatccgctttttggggcgcccggggagggcgggcgacggggacggggacgacacgtcctc
 catggttgggggacgtcgcgccgaccgcgtccgcgtcgggggtggttgcgtcctccttcccgactggccatttcttctcct
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 cgtatagtgggacaccactggaaccaggggcggttaagtccaagcagccgccggttagcccaaggaacaacagcgccaaggc
 taccgtcatggc

Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

gcgggcacaaagccatagttgcttgctgcaagactgtgggggcaacatctcttcgcccgcgctttctctaccatcacggc
 gtggccttccccgtaacatcctgcattactaccgtcatctctacagccatactgcaccggcgagcggcagcggcgaacagc
 agcggccacacagaagcaaggcgaccggatagcaagactctgacaaagcccaagaaatccacagcggcgagcagcaggag
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 agatgctaactcactttggtcttaacaaaatgtggcagtcataactgtgtacagtttgcgtgttaaggcagtttggtccaat
 atctggaacagttcaaaag

Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

tgctcatcttattataagatttgacgaaaatggagtgctactaaacaattcctcctggacccagaatattggaactttagaatggagatct
 tactgaaggcagcagcctatacaaacgctgttgatttatgcctaacctatcagcttatccaaaatctcacggtaaaactgccaaaagtaac
 attgtcagtgcaagtttacttaaacggagacaaaactaaacctgtaacactaacattacactaaacgggtacacaggaaacaggagacac
 aactccaagtcatactctatgtcattttcatgggactggctggccacaactacattaatgaaatattgccacatcctctacacttttcat
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 cccaccaccacatagcttatacagatcaccgtacctaatacaactcacagaaccctagtattcaacctgccacctccctcccaacaca
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 cagcgatgattcgcaccgccgcagcataaggcgcctgtcctcgggcacagcagcgcaccctgatctcacttaaatcagcacagt
 aactgcagcacagcaccacaatattgttcaaatccacagtgcaaggcgctgtatccaaagctcatggcggggaccacagaacca
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 aacacctgaaaaacctcctgcctaggcaaaatgacacctcccgctccagaacaacatacagcgttccacagcggcagccataac
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 actataaagataccaggcgtttccccctggaaagctccctcgtgcgtctcctgttccgacctgccgcttaccggataacctgtccgcttt
 ctcccttcgggaagc

Table 6 (continued) Nucleotide sequence of pAd/CMV/V5-DEST.

gtggcgcttttcaatgctcacgctgtaggtatctcagttcgggtgtaggtcgttcgctccaagctgggctgtgtgcacgaacccccgttc
agcccgaccgctgcgccttatccggttaactatcgtcttgagtcgaacccggtaagacacgacttatccactggcagcagccactgg
taacaggattagcagagcgaggatgtagggcgtgctacagagttctgaagtgggtggcctaactacggctacactagaaggacagta
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Table 7 Nucleotide sequence of pAd-GW-TO/tRNA.

catcatcaataataacatttttggattgaagccaatatgataatgagggggtggagtttgacgtggcgcgggcggtgggaacgg
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 aagtgggagg

Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

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 tcgcaaggtagg

[illegible]

Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

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Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

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Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

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gtccccggtctggcggttaggatacagcgctgcataaaagccttgatctgcttaaaagccacctgagcctttgcgcctcagagaaga
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tcaatcacgtgctccttattatcataatgcttccgtgtagacacttaagctcgccttcgatctcagcgcagcgggtgcagccacaacgcgc
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gtgcgccattatctgcgacgcgttgaagagcgcagcgtgtgcccctcgcctatagcggatgtcagccttgcctacgaacgccaccta
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gcgcgcctagccgtactaaaacgcagcatcaggtcaccacttgcctaccggcacttaacctacccccaaaggtcatgagcaca
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cagttggcgacgagcagctagcgcgtggttcaaacgcgcgagcctgccacttggaggagcgacgcaaaactaatgatggccgc
agtgtcgttaccgtggagcttgagtgatgcagcgggttcttctgacccggagatgcagcgaagctagaggaaacattgcactac
accttctgacagggtacg

Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

tacgccaggcctgcaagatctcaacgtggagctctgcaacctggctcctaccttggaaatttgacgaaaaccgccttgggcaaaac
 gtgcttattccacgctcaagggcgaggcgccgcgactacgtccgcactgcgttactatttctatgctacacctggcagacggc
 catgggctgttggcagcagtgcttggaggagtgcaacctcaaggagctgcagaaactgctaaagcaaaactgaaggacctatggac
 ggcttcaacgagcgtccgtggccgcgacctggcggacatcatttcccccgaacgcctgcttaaacctgcaacagggctgcc
 agacttcaccagtc aaagcatgttgcaaaccttaggaacttatacctagagcgctcaggaatcttggccgacacgtctgtgacttct
 agcgactttgtgccattaagtagccgaatgccctccgcccgttggggccactgctaccttctgcagctagccaactaccttgctac
 cactctgacataatggaagcgtgagcggtgacggtctactggagtgctactgtcgtcgaacctatgcaccccgaccgctccctgg
 ttgcaattcgagctgcttaacgaaagtcaaattatcggtacctttagctgaggggtccctcgctgacgaaaagtccgggctccgg
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 acggagcaggagggaactactgggacagtcaggcagaggaggtttggacgaggaggaggagacatgatggaagactgggagag
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 agcagcgccacacagaagcaaaaggcgaccggatgcaagactcagcaaaagcccaagaaatccacagcgccggcgagcagcag
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 caacagagcaggggccaagaacaagagctgaaaataaaaaacaggtctctgcgatccctcaccgcagctgcctgtatcacaaga
 cgaagatcagcttggcgacgctggaagacgggaggtctcttcagtaaaactgcgcgtgactcttaaggactagtttcgccc
 ctttctcaaatgaagcgcaaaactacgtcatctccagcgccacacccggcgccagcacctgtcgtcagcgccattatgagcaagg
 aaattccacgcccctacatgtggagttaccagccacaatgggacttgcggctggagctgcccagactactcaacccgaataaacta
 catgagcgccgggacccacatgatccgggtcaacgggaatccgcccaccgaaaccgaattcttgaacaggcggtatttac
 caccacacctcgaataacctaataccccgtagttggccgctgcctgggtgtagcaggaaagtcgctccaccactgtggtacttcc
 cagagacgcccaggccgaagttcagatgactaactcagggggcgagcttgcggggcggttctgcacagggtgcggtcggcggg
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 ggacattcagatcgccggcgccggcgtctcattcacgcctcgcaggaatcctaactctgcagacctcgtcctctgagccgcgc
 tctggaggcattggaactctgcaatttattgaggagttgtgccatcggtctactttaacccttctcgggacctcccgccactatccgga
 tcaatttattcctaactttgacgcggtaaaggactcggcggagcgctacgactgaatgttaagtggagaggcagagcaactgcgcctg
 aaacacctgggtcactgtcggccgcaagtgcttggccgactccggtagtgggttctactttgaattgccgaggatcatactgag
 ggccggcgacggcggtccggtaccgcccaggagagcttggcgtgagctgattcgggagttaccagcgccccctgctagt
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 gtataataatacagaaattaaataatactggggctcctatcgcaactgttaaacgccaccgcttcaccgcccgaagcaaacagg
 cgaaccttacctgttacttttaacatctctccctctgtgatttacaacagtttaacccagacggagtgagctacgagagaacctctccga
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 attattacagagcagcgctgtagaaagacgcagggcagggcgagcaacagcgcatgaatcaagagctcaagacatggttaa
 ctgacacagtcaaaagggtatctttgtctgtaaaagcaggccaaagtcacctacgacagtaataaccacggacaccccttagct
 acaagttgccaaccaagcgtcagaattggtggtcatggtgggagaaaagccattaccataactcagcactcggtagaaaccgaag
 gctgcattcactcactgtcaaggacctgaggatctctgcaccccttataagacctgtgcggctcacaagatcttattccctttaaactaat
 aaaaaaaataataaagcatcacttacttaaaatcagttagcaaatctgtccagtttattcagcagcacctccttgcctcctccagctc
 tggattgca

Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

gcttctctctggctgcaaacctttctccacaatctaataaggatgtcagtttctctgttctgtccatccgacccactatcttcatgttggtg
 cagatgaagcgcgcaagaccgtctgaagataccttcaaccccggtatccatgatgacacggaaacgggtcctccaactgtgccttttctt
 actctccctttgtatcccccaatgggttcaagagagtgccctggggtactcttgcgcctatccgaacctctagtacctccaatgg
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 ctcaaaaaaaccaagtcacaataaacctggaatatctgcacccctcacagttacctcagaagccctaactgtggctgcccgcac
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 gacccctcacagtgctcagaaggaaagctagccctgcaaacatcaggccccctaccaccaccgatagcagtagccttactatcactgc
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 cttaacttgatgttagttatccgtttgatgctcaaaaccaactaaatctaagactaggacaggccctcttttataaactcagcccacaac
 ttgatattaactacaacaaggccttacttctgttacagcttcaacaattccaaaagcttgaggtaacctaaagcactgccaaaggggtt
 gatgttgacgtacagccatagccattaatgcaggagatgggcttgaatttggttcacctaattgcaccaaacacaaatccccctcaaac
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 acagtaggaacaaaaataatgataagctaactttgtgaccacaccagctccatctcctaactgtagactaaatgcagagaaagatgc
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 aatggagatcttactgaaggcacagcctatacaaacgctgttggatttatgcctaacctatcagcttatccaaaatctcacggtaaaactg
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 acaggagacacaactccaagtgcatactctatgtcattttcatggactgtgtgcccacaactacattaatgaaatatttgcacatctc
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 cagtggtctcctcagcgatgattgcaccgcccgcagcataaggcgcttgcctcgggcacagcagcgccacctgatctcacttaa
 atcagcacagtaactgcagcacagcaccacaataattgtcaaatccacagtgcaaggcgctgtatccaaagctcatggcggggac
 cacagaaccacgtggccatcataccacaagcgaggttagtaagtggcgaccctcataaacacgctggacataaacattactct
 ttggcatgttgaattcaccacctcccgggtaccatataaaccttgattaaacatggcgccatccaccaccatcctaaaccagctggcca
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 tccaaaacctcaaatgaagatctattaagtgaacgcgtccccctccggtggcgtggtcaactctacagccaaagaacagataatgg
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 aggtctgcacggaccagcgcggccacttccccgccaggaaacttgacaaaagaaccacactgattatgacacgcatactcggagct
 atgctaaccag

Table 7 (continued) Nucleotide sequence of pAd-GW-TO/tRNA.

cgtagccccgatgaagctttgttgcattggcgccgcatataaaatgcaagggtgctgctcaaaaaatcaggcaaaagcctcgcgcaaaaa
 agaaagcacatcgtagtcatgctcatgcagataaaggcaggttaagctccggaaccaccacagaaaaagacaccattttctcctcaaac
 atgtctcggggtttctgcataaacacaaaaataaaataacaaaaaacatttaaacattagaagcctgtcttacacaggaaaaacaaccc
 ttataagcataagacggactacggccatgccggcgtgaccgtaaaaaaactggtcaccgtgattaaaaagcaccaccgacagctcct
 cggatcatgctcggagtcataatgaagactcggtaaacacatcaggttgattcacatcggtcagtgtctaaaaagcaccaccgaaatagccc
 gggggaatacataccgcgagcgtagagacaacattacgccccataggaggtataacaaaattaataggagagaaaaacacata
 aacacctgaaaaaccctcctgcctaggcaaaatagcacctcccgctccagaacaacatacagcgctccacagcggcagccataac
 agtcagccttaccagtaaaaaagaaaacctattaaaaaacaccactcgacacggcaccagctcaatcagtcacagtgtaaaaaagg
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 cctacgccagaaacgaagccaaaaaacccacaacttcctcaaatcgtcactccgttttcccacgttacgtcacttccattttaagaa
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 ggccatgctgtccagcaggtgatgacgaccatcagggacagcttcacggccagcaaaaggccaggaaccgtaaaaaggccgc
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 gcagcagccactggtaacaggattagcagagcgaggatgttagggcgtgctacagagttcttgaagtgggtgacctaacggctac
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 tctaaagtatatatgagtaaaacttggtctgacagttaccaatgcttaacagtgaggcacctatctcagcgaatctgtctatttcgttcacat
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 cgctcaccggctccagattatcagcaataaaccagccagccgggaaggccgagcgcagaaagtggtcctgcaactttatccgcctcc
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 ggtgtacgctcgtcgtttggtatggcttcattcagctccggttcccaacgatcaaggcgagttacatgatccccatgttggtgcaaaaaa
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 tcgtctcaaggatccgaattccgggagagctcgatcgcgtggtttaaattaattaa

Table 8 Nucleotide sequence of pAdenoTAG tRNA.

1 catcatcaat aatatacctt attttggatt gaagccaata tgataatgag ggggtggagt
 61 ttgtgacgtg gcgcggggcg tgggaacggg gcgggtgacg tagtagtgg gcggaagtgt
 121 gatgttgcaa gtgtggcgga acacatgtaa gcgacggatg tggcaaaagt gacgttttg
 181 gtgtgcgccg gtgtacacag gaagtgacaa tttcgcgcg gtttaggcg gatgtttag
 241 taaatttggg cgtaaccgag taagatttgg ccatttcgc gggaaaactg aataagagga
 301 agtgaaatct gaataatttt gtgttactca tagcgcgtaa tatttgtcta gggccgcggg
 361 gactttgacc gtttacgtgg agactcgccc aggtgtttt ctcagggtgt tccgcgttc
 421 cgggtcaaag ttggcgtttt attattatag tcagtcgaag ctggatccg gtaccttag
 481 aatttcgag cggccgctag cgacatcgat cacaagttg tacaaaaaag caggcttaa
 541 aggaaccaat tcagtcgact ctaggagatc gaaaccatcc tctgctatat ggccgcatat
 601 attttacttg aagactagga ccctacagaa aagggtttt aaagtaggcg tgctaaacgt
 661 cagcggacct gaccggtga agaaccaca aggtatcctg ttggaaatgc gcatttgtag
 721 gcttcaatat ctgtaacct actaattagg ttggagagc tticagccag ttcttaggt
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 841 tcagacaagt ctgcaattt gcaatatcic tttagccaa tatctaaatc tttaaattt
 901 tgattttgtt ttaccacag gatgagagac attccagagt tgtaccttg tcaaaataaa
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 1021 ttgaaggagg tacgacctcg agatctctat cactgatagg gagactcgag ttagtctgtg
 1081 gccgagtggg taaggcgatg gactctaaat ccattggggg tccccgcgc aggttcgaat
 1141 cctgccgact acggcgtgct tttttactc tcgggtagag gaaatccggt gcactacctg
 1201 tgcaatcaca cagaataaca tggagtagta ctttttatt tctgttatt atctttctc
 1261 ataaaagtgg aaccagataa ttttagttct ttgtgtaac aagactagag atttttgaa
 1321 gtgttacatt ggaaagcact tgaacacaca agtaattct gacactgcta taaaaatgat
 1381 ggaaaaacgc tcaagttgtt ttgccttca gtcttctga aatgctgtct ccctatctga
 1441 aatccagctc acgtctgact tcaaaaaccg tgcttcctt taactatgg aataaatatc
 1501 tcaaacagat ccccgggcga gctcgaattc gcggccgcac tcgagatc tagaccagc
 1561 tttctgtac aaagtgtga tcgattcgac agatcactga aatgtgtggg cgtggcttaa
 1621 ggggtggaaa gaatatataa ggtgggggtc ttatgtagt ttgtatctgt ttgcagcag
 1681 ccgcgcgcgc catgagcacc aactcgttg atggaagcat tgtgagctca tatttgacaa
 1741 cgcgcgatgc cccatgggcc ggggtgcgc agaattgtat gggctccagc attgatggc
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 2041 tttctcagca gctgttggat ctgcgcagc aggtttctg cctgaaggct tctccctc
 2101 ccaatgcggg ttaaaacata aataaaaaac cagactctgt ttgatttgg atcaagcaag
 2161 tgtcttctg tctttattt ggggttttgc gcgcgcggtg ggccgggac cagcggctc
 2221 ggtcgttgag ggtcctgtgt atttttcca ggacgtggtg aaggtgactc tggatgttca
 2281 gatacatggg cataagcccg tctctggggg ggaggttagc cactgcaga gcttcagct
 2341 gcgggggtgt gttgtagatg atccagtcgt agcaggagcg ctgggcgtgg tgcctaaaaa
 2401 tgtcttcag tagcaagctg attgccaggg gcaggccctt ggtgtaagtg ttacaaagc
 2461 ggtaagctg ggtggtgtc atacgtgggg atatgagatg catcttgac tgtatttta
 2521 ggttggtat gttcccagcc atatccctc ggggattcat gttgtcaga accaccagca
 2581 cagtgtatcc ggtgcacttg ggaatttgt catgtagctt agaaggaaat gcgtggaaga
 2641 acttgagac gccctgtga cctccaagat ttccatgca ttctccata atgatggcaa
 2701 tgggcccacg ggcggcgccc tggcgaaga tatttctggg atcactaacg tcatagttgt
 2761 gttccaggat gagatcgtca taggccattt ttacaaagcg cgggcggagg gtgccagact
 2821 gcggtataat ggttccatcc ggcccagggg cgtagttacc ctcacagatt tgcatttccc

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

2881 acgctttgag ttcagatggg gggatcatgt ctacctgagg ggcgatgaag aaaacgggtt
 2941 ccggggtagg ggagatcagc tgggaagaaa gcaggttctt gacgagctgc gacttaccgc
 3001 agccggtagg cccgtaaatc acacctatta ccgggtgcaa ctgtagtta agagagctgc
 3061 agctgccgtc atccctgagc aggggggcca ctctgtaag catgtccctg actgcatgt
 3121 ttccctgac caaatccgcc agaaggcgct cgccgccag cgatagcagt tcttgaagg
 3181 aagcaaagt tttaacggg ttgagaccgt ccgcctagg catgctttg agcgtttgac
 3241 caagcagttc caggcggtcc cacagctcgg tcacctgctc tacggcatct cgatccagca
 3301 tatctctcg ttctcgggg tggggcggt ttctgttac ggcagtagtc ggtgctcgtc
 3361 cagacgggcc agggctcatgt cttccacgg gcgcagggtc ctctcagcg tagtctgggt
 3421 cacgggtgaag ggggtgcgtc cgggctgcgc gctggccagg gtgcgttga ggctggtcct
 3481 gctggtgtg aagcgtgcc ggtcttcgcc ctgcgctcg gccaggtagc atttgacct
 3541 ggtgtcatag tccagcccct ccgcggcgtg gcccttggcg cgcagcttc ccttgaggga
 3601 ggcgcgcac gaggggcagt gcagactttt gagggcgtag agcttggcg cgagaaatac
 3661 cgattccggg gtaggcat ccgcgccgca ggcccgcag acggtctcg attccagag
 3721 ccaggtgagc tctggcgtt cggggtcaaa aaccagggtt ccccatgct tttgatcg
 3781 ttcttacct ctggttcca tgagccgtg tccacgtcg gtacgaaaa ggctgtcgt
 3841 gtccccgtat acagactga gaggcctgtc ctgagcggg gtccgcggg cctctctga
 3901 tagaaactcg gaccactctg agacaaaggc tcgctccag gccagcacga aggaggctaa
 3961 gtgggagggg tagcggctgt tctccactag ggggtccact cgtccaggg tgtgaagaca
 4021 catgtgccc tcttcggcat caaggaagg gattggttg taggttagg ccacgtgacc
 4081 ggggttctt gaaggggggc tataaaagg ggtggggggc gttctgtct cactcttc
 4141 cgcacgtcg tctgcgagg ccagctgtt gggtagtac tccctctga aagcgggcat
 4201 gacttctcg ctaagattgt cagtttcaa aaacaggag gatttgat tccactggc
 4261 cgcggtgat ccttgaggg tggccgcat catctgttca gaaaagaca tctttgtt
 4321 gtcaagcttg gtggcaaac acccgtagag ggcgttggac agcaacttg cgatggagcg
 4381 cagggttgg ttttctgc gatcggcgc ctcttggcc gcgatgtta gctgcacga
 4441 ttgcgcgca acgcaccgc attcgggaaa gacggtgtg cgtctcgg gcaccagtg
 4501 cagcgccaa ccgcggtgt gcagggtgac aaggtcaac ctggtggta cctctccgc
 4561 taggcgtcg ttgtccagc agaggcgcc gccctgcgc gagcagaat gcgtagggg
 4621 gtctagctgc gtctctcg ggggtctgc gtccacgga aagaccccg gcagcaggc
 4681 cgcgtcgaag tagtctatc tgcactctg caagtctag gcctgtgcc atgcgcggc
 4741 ggcaagcgc cgtctgtat ggttagtg gggacccat ggcatgggt ggtgagcgc
 4801 ggaggcgta atgcgcgaaa tctctgaaac gtagagggg tctctgata ttcaagata
 4861 ttagggtag catctccac cgcggtgct ggcgcgacg taatctata gtctgtcga
 4921 gggagcgagg aggtcgggag cgaggttct acggcgggc tctctgctc ggaagactat
 4981 ctgcctgaag atggcatgt agttggatga tatggttga cgtggaaga cgtgaagct
 5041 ggcgtctgt agacctacc cgtcacgcac gaaggaggcg taggagtcgc gcagcttgt
 5101 gaccagctc gcggtgacct gcacgtctag ggcgcagtag tccagggtt ccttgatgat
 5161 gtcatactt tctgtccct tttttcca cagctcgcg ttgaggaaa actctcgcg
 5221 gtcttccag tactcttga tcggaaccc gtcggcctcc gaacggtta agcctagcat
 5281 gtagaactgg ttgacggcct gtaggcgca gcatccctt tctacggga gcgcgtatgc
 5341 ctgcgcggcc ttccggagcg aggtgtgggt gagcgcaaa gtgtccctga ccatgactt
 5401 gaggtactgg tatttgaagt cagtgtctc gcatccgcc tctccaga gcaaaaagtc
 5461 cgtgcgctt ttggaacgc gatttggcag ggcgaagggt acatcgttga agagtatct
 5521 tcccgcgca ggcataaagt tgcgtgtgat gcggaagggt cccggcacct cggaaagggt
 5581 gtaattacc tggcgggcga gcacgatct gtcaaagccg ttgatgtt ggccacaat
 5641 gtaaagtcc aagaagcgc gtagtccct gatggaaggc aatttttaa gtctctga
 5701 ggtgagctct tcaggggagc tgagccgtg ctctgaaagg gccagctc caagatgagg

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

5761 gttggaagcg acgaatgagc tccacaggct acgggccatt agcatttgca ggtggcgcg
 5821 aaaggtccta aactggcgac ctatggccat ttttctggg gtgatgcagt agaaggttaag
 5881 cgggtcttgt tcccagcggc cccatccaag gtgcgcggct aggtctcgcg cggcagtcac
 5941 tagaggctca tctccgccga acttcatgac cagcatgaag ggcacgagct gcttcccaaa
 6001 ggcccccatc caagtatagg tctctacatc gtaggtgaca aagagacgct cggcgcgagg
 6061 atgcgagccg atcggggaaga actggatctc ccgccaccaa ttggaggagt ggctattgat
 6121 gtggtgaaag tagaagtccc tgcgacgggc cgaacactcg tgctggcttt tgtaaaaacg
 6181 tgcgcagtac tggcagcggg gcacgggctg tacatcctgc acgaggttga cctgacgacc
 6241 gcgcacaagg aagcagagtg ggaatttgag cccctcgcct ggcgggttg gctggtggtc
 6301 ttctacttcg gctgcttgc ctgaccgc tggctgctc aggggagta cggcggatcg
 6361 gaccaccacg ccgcgcgagc ccaaagtcca gatgccgcg cgcggcggc ggagcttgat
 6421 gacaacatcg cgcagatggg agctgtccat ggtctggagc tcccgcggcg tcaggtcagg
 6481 cgggagctcc tgcaggttta cctcgcatag acgggtcagg gcgcgggcta gatccagggtg
 6541 atacctaatt tccaggggct ggttgggtgc ggcgtcgatg gcttgcaaga ggccgcatcc
 6601 ccgcggcgcg actacggtag cgcgcggcgg gcgggtggcc gcgggggtgt ccttggatga
 6661 tgcatctaaa agcggtagc cgggcgagcc cccggaggta gggggggctc cggaccgcgc
 6721 gggagagggg gcaggggcac gtcggcgccg cgcgcgggca ggagctggtg ctgcgcgct
 6781 aggttgctgg cgaacgcgac gacgcggcgg ttgatctct gaatctggcg cctctgcgtg
 6841 aagacgacgg gcccggtgag cttagcctg aaagagagti cgacagaatc aatttcggtg
 6901 tcgttgacgg cggcctggcg caaaatctcc tgcacgtctc ctgagttgct ttgataggcg
 6961 atctcgcca tgaactgctc gatctctcc tcttgagat ctccgcgtcc ggctcgtcc
 7021 acgggtggcg cgaggtcgtt ggaaatgcgg gccatgagct gcgagaaggc gttgagcct
 7081 cctcgttcc agacgcggct gtagaccacg ccccttcgg catcggggc gcgcagacc
 7141 acctgcgca gattgagctc cagtgccgg gcgaagacgg cgtagtttcg caggcgctga
 7201 aagaggtagt tgagggtggt ggcggtgtgt tctgccacga agaagtacat aaccagcgt
 7261 cgcaacgtgg attcgttgat atcccccaag gcctcaaggc gctccatggc ctctagaag
 7321 tccacggcga agttgaaaaa ctgggagttg cgcgccgaca cggttaactc ctctccaga
 7381 agacggatga gtcggcgac agtgtcgcgc acctcgcgt caaaggctac aggggcctct
 7441 tcttctctt caatctctc ttccataagg gcctccctt ctctcttc tggcggcggt
 7501 gggggagggg ggacacggcg gcgacgacgg cgcaccggga ggcggctcgc aaagcgctcg
 7561 atcatctccc cgcggcgacg gcgcagtgct tcggtgacgg cgcggcgtt ctgcggggg
 7621 cgcagttgga agacggccc cgtcatgtcc cgttatggg ttggcgggg gctgccatgc
 7681 ggcagggata cggcgctaac gatgcatctc aacaattgt gttaggtac tccgccggc
 7741 agggacctga gcgagtcgc atcgaccgga tcggaaaacc tctcgagaaa ggcgtctaac
 7801 cagtcacagt cgcaaggtag gctgagcacc gtggcggcg gcagcggcg gcggtcggg
 7861 ttgtttcgg cggaggtgct gctgatgat taattaaagt aggcggtct gagacggcg
 7921 atggtcgaca gaagcaccat gtcttgggt ccggcctgct gaatgcgcag gcggtcgcc
 7981 atgccccagg ctctgtttg acatcgcgcg aggtctttg agtagtctg catgagcct
 8041 tctaccgca ctctcttc tcttctct tctctgcat ctctgcatc tatcgtgcg
 8101 gcggcgcgcg agtttggcg taggtggcg cctctctc ccatcggtgt gacccgaag
 8161 cccctcatcg gctgaagcag ggctaggtcg gcgacaacgc gctcggctaa tatggcctgc
 8221 tgcacctcgc tgagggtaga ctggaagtca tccatgtcca caaagcgggtg gtatgcgcc
 8281 gtgttgatgg tgaagtga gttggccata acggaccagt taacggtctg tgacccggc
 8341 tgcgagagct cgggtacct gagacgcgag taagccctcg agtcaatac gtagctgtg
 8401 caagtcgca ccaggtactg gtatccacc aaaagtgcg gcggcggtg gcggtagagg
 8461 ggccagcgtg ggtggccgg ggtccgggg gcgagatctt ccaacataag gcgatgat
 8521 ccgtagatgt acctggacat ccaggtgat cggcgggcg tggtagggc gcgcggaaag
 8581 tcgcgagcgc ggttcagat gttgcgagc ggcaaaaagt gctccatgt cgggacgctc

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

8641 tggccgggtca ggcgcgcgca atcgttgacg ctctagaccg tgcaaaagga gagcctgtaa
 8701 gcgggcactc tccgtggtc tggaggataa attcgcaagg gtatcatggc ggacgaccgg
 8761 ggttcgagcc ccgtatccgg ccgtccgcgc tgatccatgc ggttaccgcc cgcgtgtcga
 8821 acccaggtgt gcgacgtcag acaacggggg agtgctcctt ttggcttctt tccaggcgcg
 8881 gcggctgtg cgttagctt ttggccact ggccgcgcgc agcgtaacgc gttaggctgg
 8941 aaagcgaaag cattaagtgg ctgctccct gtagccggag ggttatttc caagggttga
 9001 gtcgcgggac ccccggttcg agtctcggac cggccggact gcggcgaacg ggggtttgcc
 9061 tcccgtcat gcaagacccc gcttgcaaat tctccggaa acaggagcga gcccctttt
 9121 tgctttccc agatgcatcc ggtgctcggg cagatgcgcc cccctctca gcagcggcaa
 9181 gagcaagagc agcggcagac atcgaggga cctccctc ctcctaccgc gtcaggaggg
 9241 gcgacatccg cgttgacgc ggcagcagat ggtgattacg aacccccgc gcgccggcc
 9301 cggcactacc tggacttga ggaggcgag ggcctggcg gcctaggagc gccctctct
 9361 gagcgtacc caagggtga gctgaagcgt gatacgcgtg aggcgtacgt gccgcggcag
 9421 aacctgttc gcgaccgca gggagaggag cccgaggaga tgcgggatcg aaagtccac
 9481 cgaggcgcg agtgcggca tggcctgaat cgcgagcgtg tctgcgcga ggaggacttt
 9541 gagccgacg cgcgaaccgg gattagtccc gcgcgcgcac acgtggcggc cgcggacctg
 9601 gtaaccgcat acgagcagac ggtgaaccag gagattaact ttcaaaaag cttaacaac
 9661 cacgtgcgta cgcttggtgc gcgcgaggag gtggctatag gactgatgca tctgtgggac
 9721 ttgttaagcg cgctggagca aaaccaaata agcaagccgc tcatggcgca gctgttcct
 9781 atagtgcagc acagcaggga caacgaggca ttacgggatg cgctgctaaa catagtagag
 9841 cccgaggggc gctggctgct cgatttgata aacatcctgc agagcatagt ggtgcaggag
 9901 cgcagcttga gcttggtga caagtggtgc gccatcaact attcatgct tagcctgggc
 9961 aagttttac cccgaagat ataccatacc cttacgttc ccatagacaa ggaggtaaag
 10021 atcgagggtg tctacatgc catggcgctg aagtgctta ccttgagcga cgacctgggc
 10081 gttatcgca acgagcgcat ccacaaggcc gtgagcgtga gccggcggcg cgagctcagc
 10141 gaccgcgagc tgatgcacag cctgcaaagg gccctggctg gcacgggcag cggcgataga
 10201 gaggccgagt cctacttga cgcggcgct gacctgcgt gggcccaag ccgacgcgcc
 10261 ctggaggcag ctggggccgg acctgggctg gcggtggcac ccgcgcgcgc tggcaacgtc
 10321 ggcggcgtgg aggaatatga cgaggacgat gactacgagc cagaggacgg cgagtactaa
 10381 gcggtgatgt ttctgatcag atgatgcaag acgcaacgga cccggcggtg cggcgggcgc
 10441 tgcagagcca gccgtccggc cttaactcca cggacgactg gcgccaggtc atggaccgca
 10501 tcatgtcgt gactgcgcgc aatcctgacg cgttccggca gcagccgcag gccaaccggc
 10561 tctccgcaat tctggaagcg gtggtccgg gcgcgcgaaa cccacgcac gagaaggtgc
 10621 tggcgatcgt aaacgcgctg gccgaaaaca gggccatccg gcccgacgag gccggcctgg
 10681 tctacgacgc gctgttcag cgcgtggctc gttacaacag cggcaacgtg cagaccaacc
 10741 tggaccggct ggtgggggat gtgcgcgagg ccgtggcgca gcgtgagcgc gcgcagcagc
 10801 agggcaacct gggctccatg gttgactaa acgcttctt gactacacag cccgccaacg
 10861 tgccgcgggg acaggaggac tacaccaact ttgtgagcgc actgcggcta atggtgactg
 10921 agacaccgca aagtgagggtg taccagtctg ggccagacta tttttccag accagtagac
 10981 aaggcctgca gaccgtaaac ctgagccagg ctttcaaaa cttgcagggg ctgtgggggg
 11041 tgcgggtcc cacaggcgac cgcgcgaccg tcttagctt gctgacgcc aactgcgcgc
 11101 tgttctgct gtaatatcg ccttcacgg acagtggcag cgtgtccgg gacacatacc
 11161 taggtcactt gctgacactg taccgcgagg ccataggctc ggcgcagtgt gacgagcata
 11221 cttccagga gattacaagt gtcagccgcg cgctggggca ggaggacacg ggcagcctgg
 11281 aggcaacctt aaactacctg ctgaccaacc ggcggcagaa gatccccctg ttgcacagt
 11341 taaacagcga ggaggagcgc atttgcgt acgtgcagca gagcgtgagc cttaacctga
 11401 tgcgcgacgg gtaacgccc agcgtggcgc tggacatgac cgcgcgcaac atggaaccgg
 11461 gcatgtatgc ctcaaacgg ccgttatca accgcctaata ggactacttg catcgcgcgg

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

11521 ccgccgtgaa ccccgagtat ttaccaatg ccatcttgaa cccgcactgg ctaccgcccc
 11581 ctggtttcta caccggggga ttgaggtgc ccgagggtaa cgtgggattc ctctgggacg
 11641 acatagacga cagcgtgttt tccccgaac cgcagaccct gctagagttg caacagcgcg
 11701 agcaggcaga ggcggcgctg cgaaaggaaa gcttccgcag gccaaagcagc ttgtccgac
 11761 taggcgctgc ggccccgcgg tcagatgcta gtagccatt tccaagcttg atagggtctc
 11821 ttaccagcac tcgaccacc cgcccgcgcc tgcgtggcga ggaggagtac ctaacaact
 11881 cgtgctgca gccgcagcg gaaaaaacc tgcctccgc atttccaac aacgggatag
 11941 agagcctagt ggacaagatg agtagatga agacgtacgc gcaggagcac agggacgtgc
 12001 caggccccgcg cccgccacc cgtcgtcaaa ggcacgaccg tcagcggggt ctggtgtggg
 12061 aggacgatga ctggcagac gacagcagcg tcttgattt gggaggagt ggcaaccgt
 12121 ttgcacacct tcgccagg ctggggagaa tgtttaaaa aaaaaaagc atgatcaaa
 12181 ataaaaaact caccaaggcc atggcaccga gcgttggtt tctgtattc cccttagtat
 12241 gcggcgcgcg gcgatgatg aggaaggctc tctccctcc tacgagagtg tggtagcgc
 12301 ggcgccagtgc gcggcgcgcg tgggtctcc ctctgatgct cccctggacc cgccgttgt
 12361 gctccgcgg tacctgcgc ctaccggggg gagaacagc atccgttact ctgagtggc
 12421 accctattc gacaccacc gtgtgtacct ggtggacaac aagtcaacgg atgtggcatc
 12481 cctgaactac cagaacgacc acagcaact tctgaccac gtcattcaa acaatgacta
 12541 cagcccgggg gaggcaagca cacagaccat caatctgac gaccggcgc actggggcgg
 12601 cgacctgaaa accatcctgc ataccaacat gccaatgtg aacgagtca tgtttacaa
 12661 taagtttaag gcgcgggtga tgggtgcgc ctgtcctact aaggacaac aggtggagct
 12721 gaaatacgag tgggtggagt tcacgtgcc cgagggaac tactccgaga ccatgaccat
 12781 agacctatg aacaacgcga tcgtggagca ctactgaaa gtgggcagac agaacgggt
 12841 tctgaaagc gacatcgggg taaagttga ccccgcaac ttcagactgg ggttgacc
 12901 cgtcactgt ctgtcatgc ctgggtata tacaacgaa gccttccatc cagacatcat
 12961 tttgtgcca ggatgcggg tggactcac ccacagccgc ctgagcaact tgttggcat
 13021 ccgaagcgg caaccttcc aggagggtt taggatcacc tacgatgatc tggagggtg
 13081 taacattccc gactgttg atgtggacgc ctaccaggcg agcttgaag atgacaccga
 13141 acaggcgggg ggtggcgag gcggcagcaa cagcagtggc agcggcgcg aagagaactc
 13201 caacgcggca gccgcggcaa tgcagccgt ggaggacatg aacgatcatg ccattcggg
 13261 cgacacctt gccacacgg ctgaggagaa gcgcgtgag gccgaagcag cggccgaagc
 13321 tgcgcccc gctgcgcaac ccgaggtcga gaagcctcag aagaaccgg tgatcaaac
 13381 cctgacagag gacagcaaga aacgcagta caacctata agcaatgaca gcacctcac
 13441 ccagtaccgc agctgtgacc ttgcataca ctacggcgac ctcagaccg gaatccgctc
 13501 atggaccctg cttgcatc ctgacgtaac ctgcggctcg gagcaggtct actggtggt
 13561 gccagacatg atgcaagacc ccgtgacctt ccgtccacg gccagatca gcaacttcc
 13621 ggtggtggc gccgagctgt tggcgtgca ctcaagagc tctacaacg accaggccgt
 13681 ctactccaa ctatccgcc agttacctc tctgaccac gtgttaac ctttccga
 13741 gaaccagatt ttggcgccc cgccagccc caccatcac accgtcagtg aaacgttcc
 13801 tctctcaca gatcacggga cgtaccgt gcgcaacagc atcgaggag tccagcgagt
 13861 gaccattact gacgccagac gccgcacctg cccctacgt tacaaggccc tgggcatagt
 13921 ctgccgcgc gtcctatcga gccgcactt ttgagcaagc atgtccatcc ttatatgcc
 13981 cagcaataac acaggctggg gcctgcgtt cccaagcaag atgtttggcg gggccaagaa
 14041 gcgtccgac caacaccag tgcgctgctg cgggcactac cgcgcgccct ggggcgcga
 14101 caaacgcggc gcactgggc gcaccaccgt cgtacgccc atcgacgcgg tggtaggagga
 14161 ggcgcgcaac tacacgccc cgccgccacc agtgtccaca gtggacgcgg ccattcagac
 14221 cgtgtgctgc ggagccggc gctatgctaa aatgaagaga cggcgaggc gcgtagcacg
 14281 tcgccaccgc cgccgaccgc gactgccc ccaacgcgc gggcgggccc tgcctaaccg
 14341 cgcacgtgc accggccgac gggcgggcat gggggccgt cgaaggctgg ccgcgggtat

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

14401 tgtcactgtg cccccaggt ccaggcgacg agcgggccgc gcagcagccg cggccattag
 14461 tgctatgact cagggtcgca ggggcaacgt gtattgggtg cgcgactcgg ttagcggcct
 14521 gcgcgtgccc gtgcgcaccc gccccccgcg caactagatt gcaagaaaaa actacttaga
 14581 ctctactgtg tgtatgtatc cagcggcggc ggcgcgcaac gaagctatgt ccaagcgcaa
 14641 aatcaaagaa gagatgctcc aggtcatcgc gccggagatc tatggcccc cgaagaagga
 14701 agagcaggat tacaagcccc gaaagctaaa gcgggtcaaa aagaaaaaga aagatgatga
 14761 tgatgaactt gacgacgagg tggaactgct gcacgctacc gcgcccaggc gacgggtaca
 14821 gtggaaaggt cgacgcgtaa aacgtgtttt gcgaccggc accaccgtag tctttacgcc
 14881 cggtgagcgc tccaccgcga cctacaagcg cgtgtatgat gaggtgtacg gcgacgagga
 14941 cctgcttgag caggccaacg agcgctcgg ggagtttgc tacggaaagc ggcataagga
 15001 catgctggcg ttcccgctgg acgagggcaa cccaacacct agcctaaagc ccgtaacct
 15061 gcagcagggt ctgcccgcgc ttgcaccgtc cgaagaaaag cgcggcctaa agcgcgagtc
 15121 ttgtgacttg gcaaccaccg tgcagctgat ggtaccaag cgccagcgac tggaagatgt
 15181 ctggaaaaa atgaccgtgg aacctgggtt ggagcccgag gtccgcgtgc ggccaatcaa
 15241 gcaggtggcg ccgggactgg gcgtgcagac cgtggacgtt cagataccca ctaccagtag
 15301 caccagtatt gccaccgcca cagagggcat ggagacacaa acgtccccgg ttgcctcagc
 15361 ggtggcggat gccgcgtgc aggcggcgc tgcggccgcg tccaagacct ctacggaggt
 15421 gcaaacggac ccgtggatgt ttgcgtttc agcccccg cgcccgcgcg gtccgaggaa
 15481 gtacggcgcc gccagcgcg tactgcccga atatgcccta catccttcca ttgcgctac
 15541 ccccggtat cgtggctaca cctaccgcc cagaagacga gcaactacc gacgccgaac
 15601 caccactgga acccgccgc gccgtcgccg tcgccagccc gtgctggccc cgatttccgt
 15661 gcgcagggtg gtcgcgaag gaggcaggac cctggtgctg ccaacagcg gctaccaccc
 15721 cagcatcgtt taaaagccgg tctttgtgt tctgcagat atggccctca cctgccgcct
 15781 ccgtttccc gtgcgggat tccgaggaag aatgcaccgt aggaggggca tggccggcca
 15841 cggcctgacg ggcggcatgc gtcgtgcgca ccaccggcg cggcgcgct cgcaccgtcg
 15901 catgcgcggc ggtatcctgc cctccttat tccactgac gccgcggcga ttggcgccgt
 15961 gcccgaatt gcatccgtgg ccttcagcgc gcagagacac tgataaaaa caagttgcat
 16021 gtggaaaaa caaaataaaa agtctggact ctacgctcg cttggtcctg taactatgtt
 16081 gtagaatgga agacatcaac ttgcgtctc tggccccgc acacggctcg cggccgttca
 16141 tgggaaactg gcaagatc ggcaccagca atatgagcgg tggcgcttc agctggggct
 16201 cgctgtggag cggcattaaa aatttcggtt ccaccgttaa gaactatggc agcaaggcct
 16261 ggaacagcag cacaggccag atgctgaggg ataagttgaa agagcaaaat ttccaacaaa
 16321 aggtggtaga tggcctggcc tctggcatta gcggggtggt ggacctggcc aaccaggcag
 16381 tgcaaaataa gattaacagt aagcttgat cccgccctcc cgtagaggag cctccaccgg
 16441 ccgtggagac agtgtctcca gaggggcgtg gcgaaaagcg tccgcgcccc gacaggggag
 16501 aaactctggt gacgcaata gacgagcctc cctgtacga ggaggcacta aagcaaggcc
 16561 tgcccaccac ccgtccatc gcgcccattg ctaccggagt gctgggcccag cacacaccgg
 16621 taacgtgga cctgcctccc ccgcccaga cccagcagaa acctgtgctg ccaggcccga
 16681 ccgcccgtgt tgaaccctg cctagccgcg cgtccctgcg ccgcccgcg agcggtcgc
 16741 gatcgttgcg gcccgtagcc agtggaact ggcaaaagc actgaacagc atcgtgggtc
 16801 tgggggtgca atccctgaag cgccgacgat gcttctgaat agctaactg tcgtatgtgt
 16861 gtcattgatg cgtccatgt gccgcccag gagctgtga gccgcccgc gcccgcttc
 16921 caagatggct accccttca tgatccgca gtggtcttac atgcacatc cgggcccagga
 16981 cgctcggag tacctgagcc ccgggctggt gcagtttgc cgccaccg agacgtactt
 17041 cagcctgaat aacaagtta gaaacccac ggtggcgctt acgcagcagc tgaccacaga
 17101 ccggtcccag cgttgacgc tgcggttcat cctgtggac cgtgaggata ctgcgtactc
 17161 gtacaaggcg cgttcaccc tagctgtggg tgataaccgt gtgtgggaca tggcttccac
 17221 gtactttgac atccgcgcg tgctggacag gggccctact ttaagccct actctggcac

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

17281 tgcctacaac gccctggctc ccaagggtgc cccaaatcct tgcgaatggg atgaagctgc
 17341 tactgtctctt gaaataaacc tagaagaaga ggacgatgac aacgaagacg aagtagacga
 17401 gcaagctgag cagcaaaaaa ctacgtatt tgggcaggcg ccttattctg gtataaatat
 17461 taaaaaggag ggtattcaaa taggtgtcga aggtcaaca cctaaatatg ccgataaaac
 17521 atttcaacct gaacctcaaa taggagaatc tcagtggtag gaaactgaaa ttaatcatgc
 17581 agctgggaga gtccctaaaa agactacccc aatgaaacca tgttacggtt catatgcaaa
 17641 acccacaat gaaaatggag ggcaaggcat tctgttaaag caacaaaatg gaaagctaga
 17701 aagtcaatg gaaatgcaat ttttcaac tactgaggcg accgcaggca atggtgataa
 17761 cttgactcct aaagtggat tgtacagtga agatgtatg atagaaacc cagacactca
 17821 tatttcttac atgccacta ttaaggagg taactcacga gaactaatg gccacaatc
 17881 tatgcccaac aggcctaatt acattgctt tagggacaat ttattggtc taatgtatta
 17941 caacagcacg ggtaatatg gtgtctggc ggccaagca tcgcagtga atgctgtgt
 18001 agatttcaa gacagaaaca cagagcttc ataccagctt ttgctgatt ccattggtga
 18061 tagaaccagg tacttttcta tgtggaatca ggctgtgac agctatgac cagatgttag
 18121 aattattgaa aatcatggaa ctgaagatga acttccaaat tactgttct cactgggagg
 18181 tgtgattaat acagagactc ttaccaaggt aaacctaata acaggtcagg aaaatggatg
 18241 ggaaaaagat gctacagaat ttccagataa aaatgaaata agagtggaa ataatttgc
 18301 catggaaatc aatctaaatg ccaacctgtg gagaaattc ctgtactcca acatagcgt
 18361 gtatttggc gacaagctaa agtacagtcc ttccaacgta aaaatttctg ataaccctaa
 18421 cacctacgac tacatgaaca agcagtggtt ggctccggg ttagtgact gctacattaa
 18481 ccttgagaca cgctggctcc ttgactatat ggacaacgtc aaccattta accaccacg
 18541 caatgtggc ctgcgtacc gctcaatgtt gctgggcaat ggctgctatg tgccttcca
 18601 catccaggtg cctcagaagt tcttgccat taaaacctc ctctcctgc cgggctcata
 18661 cacctacgag tggaaactca ggaaggatgt taacatggtt ctgcagagct ccttaggaaa
 18721 tgacctagg gttgacggag ccagcattaa gttgatagc attgacctt acgccacct
 18781 ctccccatg gccacaaca ccgctccac gcttgaggcc atgcttagaa acgacacca
 18841 cgaccagtc tttaacgact atctctccg cgccaacatg ctctacccta taccgccaa
 18901 cgctaccaac gtgcccata cctccccct ccgcaactgg cgggcttcc cgggctggg
 18961 ctccagcgc cttaagacta aggaaacccc atcactggc tcgggctacg accctatta
 19021 cacctactt ggctctata cctacctaga tggaaactt tacctcaacc acaccttaa
 19081 gaaggtggc attaccttg actctctgt cagctggcct ggcaatgacc gcctgttac
 19141 ccccaacgag ttgaaatta agcgtcagt tgacggggag ggttacaac ttgccagt
 19201 taacatgacc aaagactgt tctgtgata aatgtagct aactacaaca ttggtacca
 19261 gggtcttat atccagaga gctacaagga ccgcatgtac tcttctta gaaactcca
 19321 gccatgagc cgtcaggtg tggatgata taaatacaag gactaccaac aggtgggcat
 19381 cctacacca cacaacaact ctggaattgt tggctacct gccccacca tgcgcgaag
 19441 acaggcctac cctgctaact tccctatcc gcttataggc aagaccgag ttgacagcat
 19501 taccagaaa aagttctt gcgacgcac ctttggcg atccattct ccagtaact
 19561 tatgtccat ggcgactca cagacctgg ccaaacctt ctctacgcca actccgcca
 19621 cgcgctagc atgactttg aggtggatcc catggacgag cccaccttc ttatgttt
 19681 gttgaagtc ttgacgtg tccgtgtgca ccggccgac cgcggcgta tcgaaaccg
 19741 gtacctgac acgcccctt cgccgggcaa cgccacaaca taaagaagca agcaacatca
 19801 acaacagct ccgcatggg ctccagtga caggaactga aagccattgt caaagatct
 19861 ggtgtgggc catattttt gggcacctat gacaagcgt ttccaggctt tgtttcca
 19921 cacaagctc cctgcgcat agtcaatac gccggtcgc agactgggg cgtacactg
 19981 atggccttg cctggaacc gactcaaaa acatgtacc tcttgagcc ctttgctt
 20041 tctgaccag cactcaagca ggttaccag ttgagtac agtactcct gcgcgtagc
 20101 gccattgct ctccccca ccgctgtata acgtggaa agtccacca aagctacag

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

20161 gggcccaact cggccgctg tggactattc tgctgcatgt ttctccacgc ctttgccaac
 20221 tggccccaaa ctccatgga tcacaacccc accatgaacc ttattaccgg ggtacccaac
 20281 tccatgctca acagtcccca ggtacagccc accctgcgtc gcaaccagga acagctctac
 20341 agcttcttgg agcgccactc gccctacttc cgcagccaca gtgcgcagat taggagcgcc
 20401 acttcttttt gtcactttaa aaacatgtaa aaataatgta ctagagacac tttaataaa
 20461 ggcaaatgct ttattttgta cactctcggg tgattattta cccccacct tgcgctcgc
 20521 gccgtttaa aatcaaagg gttctgccgc gcatcgctat gcgccactgg cagggacacg
 20581 ttgcgatact ggtgtttagt gctccactta aactcaggca caaccatccg cggcagctcg
 20641 gtgaagtttt cactccacag gctgcgcacc atcaccaacg cgtttagcag gtcgggcgcc
 20701 gatatcttga agtcgcagtt ggggcctccg cctgcgcgc gcgagttgcg atacacaggg
 20761 ttgcagcact ggaacactat cagcgccggg tggcgcacgc tggccagcac gctcttgctg
 20821 gagatcagat ccgctccag gtctccgcg ttgctcagg gcaacggagt caactttgtg
 20881 agctgccttc caaaaagg gcgctgccc ggttttagt tgcactcgca ccgtagtggc
 20941 atcaaaagg gaccgtgcc ggtctggcg ttaggataca gcgcctgcat aaaagccttg
 21001 atctgcttaa aagccactg agcctttgcg ccttcagaga agaacatgcc gcaagacttg
 21061 ccggaanaact gattggccgg acagcccgcg tcgtgcacgc agcaccttgc gtcggtgttg
 21121 gagatctgca ccacattcg gcccaccgg ttcttcacga tcttggcctt gctagactgc
 21181 tcttcagcg cgcgtgccc gtttgcgtc gtcacatcca ttcaatcac gtgctcctta
 21241 tttatcataa tgcttcgctg tagacactta agctcgcctt cgatcagc gcagcggcgc
 21301 agccacaacg cgcagcccgt gggctcgtga tgcctttagg tcaccttgc aaacgactgc
 21361 aggtacgcct gcaggaatcg ccccatcatc gtcacaaagg tcttgttgc ggtgaaggtc
 21421 agctgcaacc cgcggtgctc ctcttcagc caggtcttgc atacggccgc cagagcttcc
 21481 acttggctcag gcagtagttt gaagtcgcc tttagatcgt tatccacgtg gtacttgc
 21541 atcagcgcg gcgcagcctc catgcccttc tcccacgcag acacgatcgg cacactcagc
 21601 ggggtcatca ccgtaattc actttccgt tcgctgggt cttctcttc ctcttgcgc
 21661 cgcataccac gcgccactgg gtcgtcttca ttcagccgc gactgtgag ctacctctct
 21721 ttgccatgct tgattagcac cgggtgggtg ctgaaacca ccattttag cgccacatct
 21781 tctcttctt cctcgtctc cagcattacc tctggtgatg gcgggcgctc gggcttggga
 21841 gaaggcgct tcttttctt ctggggcgca atggccaaat ccgccgccga ggtcgtatggc
 21901 cgcgggcttg gtgtgcgcgg caccagcgcg tcttgtgatg agtcttctc gtcctcggac
 21961 tcgatacgcc gcctatccg ctttttggg ggcgccggg gaggcggcg gcagggggac
 22021 ggggacgaca cgtctccat ggttggggga cgtcgcgcg caccgcgtcc gcgctcgggg
 22081 gtggttgcg gctgctctc ttccgactg gccatttct tctctatag gcagaaaaag
 22141 atcatggagt cagtcagaaa gaaggacagc ctaccgcc cctctgagtt cgccaccacc
 22201 gcctccaccg atgcgccaa cgcgcctacc acctccccg tcgaggcacc cccgcttag
 22261 gaggaggaag tgattatga gcaggaccca ggtttgtaa gcgaagacga cgaggaccgc
 22321 tcagtacaa cagaggataa aaagcaagac caggacaacg cagaggcaaa cgaggaacaa
 22381 gtcgggcggg gggacgaaag gcatggcgac tacctagatg tgggagacga cgtgctgttg
 22441 aagcatctgc agcgccagt gcgcattatc tgcgacgcgt tgcaagagcg cagcgatgtg
 22501 cccctcgcca tagcggatgt cagccttgc tacgaacgcc acctattct accgcgcgta
 22561 ccccccaaac gccaagaaa cggcacatgc gagcccaacc cgcgcctcaa cttctacccc
 22621 gtatttgccg tgccagaggt gcttgccacc tatcacatct tttccaaaa ctgcaagata
 22681 cccctatcct gccgtgcaa ccgcagccga gcggacaagc agctggcctt gcggcagggc
 22741 gctgtcatc ctgatatgc ctgcctaac gaagtgcga aaatcttga ggtcttggga
 22801 cgcgacgaga agcgcgcggc aaacgctctg caacaggaaa acagcgaaaa tgaaagtcac
 22861 tctggagtgt tgggtgaact cgagggtgac aacgcgcgcc tagccgtact aaaacgcagc
 22921 atcagaggtc cccacttgc ctaccgggca cttacctac ccccaaggt catgagcaca
 22981 gtcatgagt agctgatcgt gcgccgtgc cagcccttg agagggatgc aaatttcaa

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

23041 gaacaacag aggagggcct acccgaggtt ggcgacgagc agctagcgcg ctggcttcaa
 23101 acgcgcgagc ctgccgactt ggaggagcga cgaaactaa tgatggccgc agtgctcgtt
 23161 accgtggagc ttgagtgcat gcagcgggtc ttgtctgacc cggagatgca gcgcaagcta
 23221 gaggaacat tgcactacac cttcgacag ggctacgtac gccaggcctg caagatctcc
 23281 aacgtggagc tctgcaacct ggtctctac cttggaattt tgcacgaaa cgccttggg
 23341 caaacgtgc ttattccac gctcaagggc gaggcgcgcc gcgactacgt ccgcgactgc
 23401 gtttacttat ttctatgcta cacctggcag acggccatgg gcgttggca gcagtgttg
 23461 gaggagtga acctcaagga gctgcagaaa ctgctaaagc aaaacttgaa ggacatatgg
 23521 acggccttca acgagcgtc cgtggccgcg cacctggcgg acatcattt cccgaacgc
 23581 ctgttaaaa ccctgcaaca gggctgcca gacttacca gtcaaagcat gttgcagaac
 23641 tttaggaact ttatctaga gcgctcagga atcttgccc ccacctgtg tgcacttct
 23701 agcgactttg tgccattaa gtaccgcga tgccctccgc cgcttgggg ccaactgtac
 23761 cttctgcagc tagccaacta cctgcctac cactctgaca taatggaaga cgtgagcgtt
 23821 gacggtctac tggagtgtca ctgtcgtgc aacctatga cccgcaccg ctccctggtt
 23881 tgaattcgc agctgttaa cgaagtcaa attatcggtta cttttagct gcagggtccc
 23941 tcgctgacg aaaagtccgc ggctccgggg ttgaaacta ctccggggct gtggacgtcg
 24001 gcttaccttc gcaatttgt acctgaggac taccagccc acgagattag gttctacgaa
 24061 gaccaatccc gcccgccaaa tgcggagctt accgctcgc tcattacca gggccacatt
 24121 cttggccaat tgcaagccat caacaagcc cgcaagagt ttctgtacg aaaggacgg
 24181 ggggtttact tggacccca gtccggcgag gagctcaacc caatcccccc gccgccgag
 24241 ccctatcagc agcagccgcg ggcccttgt tccaggatg gcacccaaaa agaagctga
 24301 gctgccgccg ccaccacgg acgaggagga atactgggac agtcaggcag aggaggttt
 24361 ggacgaggag gaggaggaca tgatggaaga ctgggagagc ctgacgagg aagcttcca
 24421 ggtcgaagag gtgtcagacg aaacaccgtc accctcgtc gattccctt cgcggcgcc
 24481 ccagaatcg gcaaccggtt ccagcatggc tacaacctc gctcctcagg cgcggccgc
 24541 actgcccgtt cgcggacca accgtatgat ggacaccact ggaaccaggg ccggttaagt
 24601 caagcagccg ccgctgttag ccaagagca acaacagcg caaggctacc gtcctggcg
 24661 cgggcacaag aacgccatag ttgctgtt gcaagactgt gggggcaaca tctcttcgc
 24721 ccgcgcttt cttcttacc atcagggcgt ggcctcccc cgtaacatcc tgcattacta
 24781 ccgtatctc tacagcccat actgcaccgg cggcagcggc agcggcagca acagcagcg
 24841 ccacacagaa gcaaggcga ccgcatagca agactctgac aaagcccaag aaatccacg
 24901 cggcggcagc agcaggagga ggagcgtgc gtcggcgcc caacgaacc gtatcgacc
 24961 gcgagcttag aaacaggatt ttccactc tctatgctat attcaacag agcaggggcc
 25021 aagaacaaga gctgaaaata aaaaacaggt ctctcgatc cctacccgc agctgcctgt
 25081 atcacaaaag cgaagatcag cttcggcgca cgctggaaga cgcggaggct ctcttcagta
 25141 aatctcgcg gctgactctt aaggactagt ttcgcgcct ttctcaaatt taagcgcga
 25201 aactacgtca tctcagcgg ccacaccgg cgcagcacc tctcgtcagc gccattatga
 25261 gcaaggaaat tcccagccc tacatgtgga gttaccagcc acaaatggga cttgcggctg
 25321 gagctgcca agactactca acccgaataa actacatgag cgcgggacc ccatgatata
 25381 cccgggtcaa cggatccgc gccaccgaa accgaattct cttggaacag gcggctatta
 25441 ccaccacacc tcgtaataac cttaatccc gtagtggcc cgctgccctg gtgtaccag
 25501 aaagtccgc tcccaccact gtgtacttc ccagagacgc ccaggccgaa gttcagatga
 25561 ctaactcagg ggcgcagctt cggggcggtt ttgtcagag ggtgcggtc cccgggcagg
 25621 gtataactca cctgacaatc agaggcgag gtaactcag caacgacgag tcggtgagct
 25681 cctcgttgg tctcgtccg gacgggacat ttcatcgg cggcgccggc gtccttcat
 25741 tcacgcctcg tcaggcaatc ctaactctgc agacctcgtc ctctgagccg cgctctggag
 25801 gcattggaac tctgcaattt attgaggagt ttgtccatc ggtctactt aacccttct
 25861 cgggacctcc cggccactat ccgatcaat ttattcctaa cttgacgcg gtaaaggact

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

25921 cggcggacgg ctacgactga atgttaagt gagaggcaga gcaactgcgc ctgaaacacc
 25981 tggctccactg tcgccgccac aagtgccttg cccgcgactc cggtgagttt tgctactttg
 26041 aattgcccgga ggatcatatc gagggcccgg cgcacggcgt cgggcttacc gccagggag
 26101 agcttgcccg tagcctgatt cgggagttta cccagcgcgc cctgctagtt gagcgggaca
 26161 ggggaccctg tgttctact gtgatttga actgtcctaa ccttggtta catcaagatc
 26221 tttgttgcca tctctgtgt gagtataata aatacagaaa ttaaaatata ctggggctcc
 26281 tatcgccatc ctgtaaagc caccgtctc acccgcccaa gcaaaccaag gcgaacctta
 26341 cctgttactt ttaacatctc tccctctgtg attacaaca gttcaaccc agacggagt
 26401 agtctacgag agaacctctc cgagctcagc tactccatca gaaaaaacac caccctcct
 26461 acctgccggg aacgtacgag tgcgtcaccg gccgtgcac cacacctacc gcctgaccgt
 26521 aaaccagact tttccggac agacctcaat aactctgtt accagaacag gagtgagct
 26581 tagaaaaccc ttagggtatt aggccaaagg cgcagctact gtggggttta tgaacaattc
 26641 aagcaactct acgggctatt ctaattcagg tttctctaga aatggacgga attattacag
 26701 agcagcgctt gctagaaaga cgcagggcag cggccgagca acagcgcgtg aatcaagagc
 26761 tccaagacat ggtaacttg caccagtga aaagggtat ctttgtctg gtaaagcagg
 26821 ccaaaagcac ctacgacagt aataccaccg gacaccgcct tagctacaag ttgccaacca
 26881 agcgtcagaa attggtggtc atggtgggag aaaagcccat taccataact cagcactcgg
 26941 tagaaaccga aggctgcatt cactcacctt gtaaggacc tgaggatctc tgcacctta
 27001 ttaagacctt gtgcggctc aaagatctta tccctttaa ctaataaaaa aaaataataa
 27061 agcatcactt acttaaaatc agttagcaaa tttctgtcca gttattcag cagcacctcc
 27121 ttgccctcct cccagctctg gtattgcagc ttctcctgg ctgcaactt tctccacaat
 27181 ctaaatggaa tgcagttc ctctgttcc tgtccatccg caccactat ctctatgtg
 27241 ttgcagaiga agcgcgcaag accgtctgaa gatacctca acccgtgta tccatatgac
 27301 acggaaaccg gtctccaac tgtgcctttt ctactcctc ctttgtatc ccccaatggg
 27361 tttcaagaga gtccccctgg ggtactctct ttgcgctat ccgaacctct agttacctc
 27421 aatggcatgc ttgcgtcaa aatgggcaac ggcctctctc tggacgaggc cggcaacctt
 27481 acctcccaaa atgtaaccac tgtgagccca cctctcaaaa aaaccaagtc aaacataaac
 27541 ctggaatat ctgcaccctc cacagttacc tcagaagccc taactgtggc tgcggccgca
 27601 cctctaatgg tcgcgggcaa cacactcacc atgcaatcac agggcccgct aaccgtgcac
 27661 gactccaaac ttgcatgac cacccaagga cccctcacag tgcagaagg aaagctagcc
 27721 ctgcaaacat caggccccct caccaccacc gatagcagta cccttactat cactgcctca
 27781 cccctctaa ctactgccac tggtagcttg ggcattgact tgaagagcc catttataca
 27841 caaaatggaa aactaggact aaagtacggg gtcctttgc atgtaacaga cgacctaaac
 27901 actttgaccg tagcaactgg tccaggtgtg actattaata atacttctt gcaactaaa
 27961 gttactggag ccttgggttt tgattcaca ggcaatatgc aacttaagt agcaggagga
 28021 ctaaggattg attctcaaaa cagacgcctt atactgatg ttagttatcc gtttgatgct
 28081 caaaaccaac taaatctaag actaggacag ggccctctt ttaaaactc agcccacaac
 28141 ttggatatta actacaacaa aggcctttac ttgttacag cticaacaa ttcaaaaag
 28201 cttgaggta acctaagcac tgccaagggg ttgatgttg acgtacagc catagccatt
 28261 aatgcaggag atgggctga atttggtca cctaagcac caaacacaaa tccctcaaa
 28321 acaaaaattg gccatggcct agaattgat tcaaacagg ctatggttcc taaactagga
 28381 actggcctta gtttgacag cacaggtgcc attacagtag gaaacaaaa taatgataag
 28441 ctaacttgtt ggaccacacc agtccatct cctaactgta gactaaatgc agagaaagat
 28501 gctaaactca ctttggtctt aacaaaatgt ggcagtcaaa tacttgctac agtttcagt
 28561 ttggctgta aaggcagttt ggctccaata tctggaacag ttcaaagtgc tcatcttatt
 28621 ataagatttg acgaaaatgg agtgctacta aacaattcct tcttgaccc agaattattg
 28681 aactttagaa atggagatct tactgaaggc acagcctata caaacgctgt tggatttatg

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

28741 cctaacctat cagcttatcc aaaatctcac ggtaaaactg ccaaaagtaa cattgtcagt
 28801 caagtttact taaacggaga caaaactaaa cctgtaacac taaccattac actaaacggt
 28861 acacaggaaa caggagacac aactccaagt gcatactcta tgtcatttc atgggactgg
 28921 tctggccaca actacattaa tgaatatatt gccacatcct ctacacttt ttacataatt
 28981 gcccaagaat aaagaatcgt ttgtgtatg ttcaacgtg ttatttttc aattgcagaa
 29041 aatttcgaat cttttttcat tcagtagtat agccccacca ccacatagct tatacagatc
 29101 accgtacctt aatcaaaactc acagaaccct agtattcaac ctgccacctc cctcccaaca
 29161 cacagagtac acagtctttt ctccccggct ggccttaaaa agcatcatat catgggtaac
 29221 agacatattc ttaggtgtta tattccacac ggtttctgt cgagccaaac gctcatcagt
 29281 gatattaata aactccccgg gcagctcact taagttcatg tcgctgtcca gctgctgagc
 29341 cacaggctgc tgtccaactt gcggttgctt aacgggcggc gaaggagaag tccacgccta
 29401 catgggggta ggtcataat cgtgcatcag gataggcggg tgggtgtgca gcagcgcgag
 29461 aataaaactgc tgcgcggcc gctcgtcct gcaggaatac aacatggcag tggctcctc
 29521 agcgatgatt cgcaccgccc gcagcataag gcgcctgtc ctccgggcac agcagcgcac
 29581 cctgatctca cttaaatcag cacagtaact gcagcacagc accacaatat tgttcaaat
 29641 cccacagtgc aaggcgtgt atccaaagct catggcgggg accacagaac ccacgtggcc
 29701 atcataccac aagcgcaggt agattaagtg gcgacccctc ataaacacgc tggacataaa
 29761 cattacctct ttggcatgt tgtaattcac cacctccggg taccatataa acctctgatt
 29821 aaacatggcg ccatccacca ccatcctaaa ccagctggcc aaaacctgcc cgcgggctat
 29881 aactgcagg gaaccgggac tggaacaatg acagtggaga gccaggact cgtaacatg
 29941 gatcatcatg ctgctcatga tatcaatgtt ggcaaacac aggcacacgt gcataactt
 30001 cctcaggatt acaagctcct ccgcggttag aaccataacc cagggaacaa cccattcctg
 30061 aatcagcgtg aatccacac tgcagggaag acctgcacg taactcacgt tgtcattgt
 30121 caaagtgtta cattcgggca gcagcggatg atcctccagt atggtagcgc gggtttctgt
 30181 ctcaaaagga ggtagacgat cctactgta cggagtgcgc cgagacaacc gagatcgtgt
 30241 tggctgtagt gtcatccaa atggaacgcc ggacgtatg atatttctg aagcaaaacc
 30301 aggtgcgggc gtgacaaaca gatctgcgtc tccggtctcg ccgcttagat cgctctgtgt
 30361 agtagttgta gtatatccac tctctcaaag catccaggcg cccctggct tcgggttcta
 30421 tgtaaaactc ttcatgcgcc gctgccctga taacatccac caccgcagaa taagccacac
 30481 ccagccaacc tacacattcg ttctgcgagt cacacacggg aggagcggga agagctggaa
 30541 gaaccatgtt tttttttt ttccaaaaga ttatccaaaa cctcaaatg aagatctatt
 30601 aagtgaacgc gctccccctc ggtggcgtgg tcaaaactcta cagccaaaga acagataatg
 30661 gcatttgtta gatgttcac aatggcttc aaaaggcaaa cgccctcac gtccaagtgg
 30721 acgtaaaggc taaaccttc aggtgaatc tccttataa acattccagc acctcaacc
 30781 atgcccaaat aattctcatc tcgccacct ctcaatatat cttaagcaa atcccgaata
 30841 ttaagtcgg ccattgtaaa aatctgctc agagcgcct ccacctcag cctcaagcag
 30901 cgaatcatga ttgaaaaa ttaggttct cacagacctg tataagattc aaaagcggaa
 30961 cattaacaaa aataccgga tccgtaggt ccttcgcag ggccagctga acataatcgt
 31021 gcaggtctgc acggaccagc gcggccactt ccccgccagg aaccttgaca aaagaacca
 31081 cactgattat gacacgcata ctggagcta tgtaaccag cgtagccccg atgtaagctt
 31141 tgttgcatgg gcggcgatat aaaatgcaag gtgctgtca aaaaatcagg caaagcctcg
 31201 cgcaaaaaag aaagcacatc gtatgcatg tcatgcagat aaaggcaggt aagctccgga
 31261 accaccacag aaaaagacac ctttttctc tcaaacatgt ctgcgggtt ctgcataaac
 31321 acaaaataaa ataacaaaa aacatttaaa cattagaagc ctgtcttaca acaggaaaaa
 31381 caaccttat aagcataaga cggactacgg ccatgccggc gtgaccgtaa aaaaactggt
 31441 caccgtgatt aaaaagcacc accgacagct cctcgtcat gtccggagtc ataagtgaag
 31501 actcggtaaa cacatcaggt tgattcacat cgtcagtcg taaaaagcga ccgaatagc
 31561 ccgggggaat acataccgc agcgtagag acaacattac agccccata ggaggtataa

Table 8 (continued) Nucleotide sequence of pAdenoTAG tRNA.

31621 caaaattaat aggagagaaa aacacataaa cacctgaaaa accctcctgc ctaggcaaaa
 31681 tagcacccct ccgctccaga acaacataca gcgctccac agcggcagcc ataacagtca
 31741 gccttaccag taaaaaagaa aacctattaa aaaaacacca ctgcacacgg caccagctca
 31801 atcagtcaca gtgtaaaaaa gggccaagtg cagagcgagt atatatagga ctaaaaaatg
 31861 acgtaacggg taaagtccac aaaaaacacc cagaaaaccg cacgcgaacc tacgccaga
 31921 aacgaaagcc aaaaaaccca caacttcctc aaatcgtcac ttccgtttc ccacgttacg
 31981 tcacttccca ttttaagaaa actacaattc ccaacacata caagttactc cgccctaaaa
 32041 cctacgtcac ccgccccgtt cccacgcccc gcgccacgtc acaaactcca cccctcatt
 32101 atcatattgg ctcaatcca aaataaggta tattattgat gatgttaatt aatttaaatc
 32161 cgcattgcgat atcgagctct cccgggaatt cggatctgcg acgcgaggct ggatggcctt
 32221 cccattatg attctctcgc ctccggcgg catcgggatg cccgcgttc aggccatgct
 32281 gtccaggcag gtagatgacg accatcaggg acagcttcac ggccagcaaa aggccaggaa
 32341 ccgtaaaaag gccgcgttcg tggcgtttt ccataggctc cgccccctg acgagcatca
 32401 caaaaatcga cgctcaagtc agagggtggc aaaccgcaca ggactataaa gataccaggc
 32461 gtttccctt ggaagctccc tcgtgcgtc tcctgttccg accctgccgc ttaccggata
 32521 cctgtccgcc ttctccctt cgggaagcgt ggcgtttct caatgctcac gctgtaggta
 32581 tctcagttcg gtgtaggctg ttcgtccaa gctgggctgt gtgcacgaac ccccggtca
 32641 gcccgaccgc tgcgccttat ccggtacta tcgtctgag tccaaccgg taagacacga
 32701 ctatcgcca ctggcagcag ccactggtaa caggattagc agagcgagg atgtaggcgg
 32761 tgctacagag ttcttgaagt ggtggcctaa ctacggctac actagaagga cagtattgg
 32821 tatctgcgt ctgtgaagc cagtaccct cggaaaaaga gttgtagct ctgatccgg
 32881 caaacaacc accgctggta gcggtggtt tttgtttc aagcagcaga ttacgcgcag
 32941 aaaaaaagga tctcaagaag atcctttagt ctttctacg gggctgacg ctacgtggaa
 33001 cgaaaactca cgtaaggga tttggtcat gagattatca aaaaggatct tcacctagat
 33061 cttttaaat caatctaaag tatatatgag taaacttgt ctgacagta ccaatgcta
 33121 atcagttagg cacctatctc agcgatctgt ctattcgtt catccatagt tgcctgactc
 33181 cccgtcgtgt agataactac gatacgggag ggcttaccat ctggccccag tgcgtcaatg
 33241 ataccgcgag acccacgtc accggctcca gattatcag caataaacca gccagccgga
 33301 agggccgagc gcagaagtgg tctgcaact ttatccgct ccatccagtc tattaattgt
 33361 tgcggggaag ctagagtaag tagttcgcca gtaaatagt tgcgcaacgt tgttgccatt
 33421 gntgcaggca tcgtgtgtc acgtcgtcg ttggtatgg ctctcattcag ctccggttc
 33481 caacgatcaa ggcgagttac atgatcccc atgtgtgca aaaaagcgg tagctcttc
 33541 ggtcctccga tcgtgtcag aagtaagtg gccgcagtgt tatcactcat gggtatggca
 33601 gcactgcata attctcttac tgcattgcca tccgtaagat gctttctgt gactggtgag
 33661 tactcaacca agtcattctg agaatagtgt atgcggcgac cgagttgctc ttgcccgcg
 33721 tcaacacggg ataataccgc gccacatagc agaacttaa aagtgtcat cattggaaaa
 33781 cgttcttcgg ggcgaaaact ctcaaggatc ttaccgtgt tgagatccag ttcatgtaa
 33841 ccaactcgtg caccactg atctcagca tctttactc tcaccagcgt ttctgggtga
 33901 gcaaaaacag gaaggcaaaa tgcgcgcaaaa aagggaataa gggcgacacg gaaatgtga
 33961 atactcatal tcttctttt tcaatattat tgaagcatt atcagggtta ttgtctatg
 34021 agcgataca tatttgaatg tatttagaaa aataaataa taggggttc gcgcacatt
 34081 ccccgaaaag tgccacctga cgtctaagaa accattatta tcatgacatt aacctataa
 34141 aataggcgta tcacgagcc ctttctctt caaggatccg aattccggg agagctcgt
 34201 atcgcattcg gatftaatt aattaa

Table 9 Nucleotide sequence of a Sau3A fragment used to construct vectors comprising suppressor tRNA sequences.

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1  ctagaggatc gaaaccatcc tctgctatat ggccgcatat attttacttg aagactagga
61  ccctacagaa aaggggtttt aaagtaggcg tgctaaacgt cagcggacct gacccgtgta
121 agaatccaca aggtatcctg gtggaaatgc gcatttgtag gcttcaatat ctgtaatcct
181 actaattagg tgtggagagc tttcagccag tttcgtagggt ttggagacca tttagggggt
241 ggcgtgtggc cccctcgtaa agtccttcgt acttcctaca tcagacaagt cttgcaatct
301 gcaatatctc ttttagccaa tatctaaatc tttaaaatct tgattttgtt ttttaaccag
361 gatgagagac attccagagt tgttaccttg tcaaaataaa caaatttaaa gatgtctgtg
421 aaaagaaaca tatattcctc atgggaatat atccagggtg ttgaaggagg tacactcgag
481 tctccctatc agtgatagag atctcgagggt cgtagtcgtg gccgagtggg taaggcgatg
541 gactctaaat ccattggggg ctccccgcgc aggttcgaat cctgccgact acggcgtgct
601 ttttttactc tcgggtagag gaaatccggt gcactacctg tgcaatcaca cagaataaca
661 tggagtagta ctttttattt tcctgttatt atctttctcc ataaaagtgg aaccagataa
721 ttttagttct tttgtgtaac aagactagag attttttgaa gtgttacatt ggaaagcact
781 tgaaaacaca agtaatttct gacactgcta taaaaatgat ggaaaaacgc tcaagttggt
841 ttgcctttca gtcttcttga aatgctgtct ccctatctga aatccagctc acgtctgact
901 tccaaaaccg tgcttgcctt taacttatgg aataaatatc tcaaacagat cccc

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Table 10 Nucleotide sequence of pAd/PL-DEST™.

CATCATCAATAATATACCTTATTTTGGATTGAAGCCAATATGATAATGAGGGGGTGGAGTTTGTGACGTG
 GCGCGGGGCGTGGGAACGGGGCGGGTGACGTAGTAGTGTGGCGGAAGTGTGATGTTGCAAGTGTGGCGGA
 ACACATGTAAGCGACGGATGTGGCAAAAGTGACGTTTTTGGTGTGCGCCGGTGTACACAGGAAGTGACAA
 TTTTCGCGCGGTTTTAGGCGGATGTTGTAGTAAATTTGGGCGTAACCGAGTAAGATTGGCCATTTTCGC
 GGGAAAACCTGAATAAGAGGAAGTGAATCTGAATAATTTGTGTTACTCATAGCGCGTAATATTTGTCTA
 GGGCCGCGGGGACTTTGACCGTTTACGTGGAGACTCGCCAGGTGTTTTCTCAGGTGTTTTCCGCGTTC
 CGGGTCAAAGTTGGCGTTTTATTATTATAGTCAGTCGAAGCTTGATCCGGTACCTCTAGAATTCTCGAG
 CGGCCGCTAGCGACATCGATCACAAAGTTTGTACAAAAAGCTGAACGAGAAACGTAAAAATGATATAAATA
 TCAATATATATAAATTAGATTTTGCATAAAAAACAGACTACATAATACTGTAAAACACAACATATCCAGTC
 ACTATGGCGGCGGCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTT
 TGAGTTAGGATCCGGCGAGATTTTCAGGAGCTAAGGAAGCTAAAAATGGAGAAAAAATCACTGGATATAC
 CACCGTTGATATATCCCAATGGCATCGTAAAGAACATTTTGAGGCATTTTCAGTCAGTTGCTCAATGTACC
 TATAACCAGACCGTTTCAGCTGGATATTACGGCCTTTTTAAAGACCGTAAAGAAAAATAAGCACAAAGTTT
 ATCCGGCCTTTATTACATTTCTTGCCCGCTGTATGAATGCTCATCCGAATTCCGTATGGCAATGAAAGA
 TCGTGAAGCTGGTATATGGGATAGTGTTCACCTTTGTTACACCGTTTTCCATGAGCAAACTGAAACGTTT
 TCATCGCTCTGGAGTGAATACCACGACGATTTCCGGCAGTTTCTACACATATATTCGCAAGATGTGGCGT
 GTTACGGTGAAAACTGGCCTATTTCCCTAAAGGGTTTATTGAGAAATATGTTTTCTGCTCAGCCAATCC
 CTGGGTGAGTTTACCAGTTTTGATTAAACGTGGCAATATGGACAACTTCTTCGCCCCCGTTTTTACC
 ATGGGCAAATATATACGCAAGGCGACAAGGTGCTGTATGCCGCTGGCGATTTCAGGTTTCATCATGCCGTCT
 GTGATGGCTTCCATGTGGCGAATGCTTAATGAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGG
 GTAAACGCGTGGATCCGGCTTACTAAAAGCCAGATAACAGTATGCGTATTTGCGCGCTGATTTTGGCGG
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 AGTGACAGTTTGACAGCGACAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAA
 GCACAACCATGCAGAATGAAGCCCGTCTGCTGCGTGCCGAACGCTGGAAGCGGAAATCAGGAAGGGAT
 GGCTGAGGTGCGCCGGTTTTATTGAAATGAACGGCTCTTTGCTGACGAGAACAGGGACTGGTGAAATGCA
 GTTTAAGGTTTTACCTATAAAAGAGAGAGCCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATT
 GACACGCCCCGGGCGACGGATGGTGATCCCCCTGGCCAGTGACGCTCTGCTGTGATATAAGTCTCCCGTG
 AACTTTACCCGGTGGTGATATCGGGGATGAAAGCTGGCCAGTGATGACACCGATATGGCCAGTGTGCC
 GGTCTCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCAGCGGAAATGACATCAAAAACGCCATTAAAC
 CTGATGTTCTGGGGAATATAAATGTCAAGGCTCCGTTATACACAGCCAGTCTGCAGGTGACCATAGTGAC
 TGGATATGTTGTGTTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTAATTTAATATATGTATAT
 TTATATCATTTTTACGTTTCTCGTTTCAGCTTTCTTGTACAAAGTGGTGATCGATTTCGACAGATCACTGAAA
 TGTGTGGGCGTGGCTTAAGGGTGGGAAAGAATATATAAGGTGGGGGTCTTATGTAGTTTTGTATCTGTTT
 TGCAGCAGCCGCCCGCCCATGAGCACCACCTCGTTTTGATGGAAGCATTGTGAGCTCATATTTGACAACG
 CGCATGCCCCCATGGGCCGGGTGCGTCAGAATGTATGGGCTCCAGCATGATGGTGCAGCCCGTCTGCG
 CCGCAAACCTCTACTACCTTGACCTACGAGACCGTGTCTGGAACGCCGTTGGAGACTGCAGCCTCCGCCGC
 CGCTTTCAGCCGCTGCAGCCACCGCCCGCGGATTGTGACTGACTTTGCTTTCCTGAGCCCGCTTGCAAGC
 AGTGACGCTTCCCGTTTCATCCGCCCGCATGACAAGTTGACGGCTCTTTTGGCACAATTGGATTCTTTGA
 CCCGGGAACCTTAATGTCGTTTTCTCAGCAGCTGTTGGATCTGCGCCAGCAGGTTTCTGCCCTGAAGGCTTC
 CTCCCTCCCAATGCGGTTTTAAACATATAAATAAAACCAGACTCTGTTTGGATTGTGATCAAGCAAGTG
 TCTTGTGTCTTTATTTAGGGGTTTTGCGCGCGCGGTAGGCCCGGGACAGCGGTCTCGGTCTGTGAGG
 TCTGTGTATTTTTTCCAGGACGTGGTAAAGGTGACTCTGGATGTTTCAGATACATGGGCATAAGCCCGTCT
 TCTGGGGTGGAGGTAGCACCCTGACAGCTTTCATGCTGCGGGGTGGTGTGTAGATGATCCAGTCTGTAG
 CAGGAGCGCTGGGCGTGGTGCTTAAATGTCTTTTCAGTAGCAAGCTGATTGCCAGGGGCAGGCCCTTGG
 TGTAAGTGTTTACAAAGCGGTTAAGCTGGGATGGGTGCATACGTGGGGATATGAGATGCATCTTGGACTG
 TATTTTTAGGTTGGCTATGTTCCAGCCATATCCCTCCGGGGATTTCATGTTGTGCAAGCACCAGCACA
 GTGTATCCGGTGCATTTGGGAAATTTGTATGTAGCTTAGAAGGAAATGCGTGGAAGAACTTGGAGACGC
 CCTTGTGACCTCCAAGATTTTCATGCATTCGTCATAATGATGGCAATGGGCCACGGGCGGCGGCTG
 GCGGAAGATATTTCTGGGATCACTAACGTCATAGTTGTGTTCCAGGATGAGATCGTCATAGGCCATTTT
 ACAAGCGCGGGCGGAGGGTGCCAGACTGCGGTATAATGTTCCATCCGGCCAGGGGCGTAGTTACCTT
 CACAGATTTGCATTTCCACGCTTTGAGTTCAGATGGGGGATCATGTCTACCTGCGGGGCGATGAAGAA
 AACGGTTTTCCGGGTAGGGGAGATCAGCTGGGAAGAAAGCAGGTTCTTGAGCAGCTGCGACTTACCGCAG
 CCGGTGGGCCCCGTAAATCACACCTATTACCGGGTGCAACTGGTAGTTAAGAGAGCTGCAGCTGCCGTCAT
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 GCGCATCTCGATCCAGCATATCTCCTCGTTTCGCGGGTTGGGGCGGCTTTCGCTGTACGGCAGTAGTCCGG
 TGCTCGTCCAGACGGGCCAGGGTCATGTCTTCCACGGGCGCAGGGTCTCGTCAGCGTAGTCTGGGTCA
 CGGTGAAGGGGTGCGCTCCGGGCTGCGCGCTGGCCAGGGTGCCTTGAGGCTGGTCTGCTGGTGTGAA

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

GCGCTGCCGGTCTTCGCCCTGCGCGTCGGCCAGGTAGCATTTGACCATGGTGTTCATAGTCCAGCCCCCTCC
 GCGCGTGGCCCTTTGGCGCGCAGCTTGCCCTTGGAGGAGCGCGCCGACGAGGGGAGTGCAGACTTTTGA
 GGGCGTAGAGCTTGGGCGCGAGAAATACCGATTCCGGGGAGTAGGCATCCGCGCGCAGGCCCCGAGAC
 GGTCTCGCATTCACGAGCCAGGTGAGCTCTGGCCGTTCCGGGGTCAAAAACAGGTTTCCCCCATGCTTT
 TTGATGCGTTTCTTACCTCTGGTTTCCATGAGCCGGTGTCCACGCTCGGTGACGAAAAGGCTGTCCGTGT
 CCCCCTATACAGACTTGAGAGGCCTGTCTCGAGCGGTGTCCGCGGTCTCTCTCGTATAGAACTCGGA
 CCACTCTGAGACAAAGGCTCGCGTCCAGGCCAGCACGAAGGAGGCTAAGTGGGAGGGGTAGCGGTCTGTG
 TCCACTAGGGGGTCCACTCGCTCCAGGGTGTGAAGACACATGTCCGCCCTCTTCGGCATCAAGGAAGGTGA
 TTGGTTTGTAGGTGTAGGCCACGTGACCGGGTGTCTCTGAAGGGGGGTATATAAAGGGGGTGGGGCGCG
 TTCGTCTCACTCTCTTCCGCATCGCTGTCTGCGAGGGCCAGCTGTTGGGGTGAAGTACTCCCTCTGAAAA
 GCGGGCATGACTTCTCGCCTAAGATTGTCAAGTTTCAAAAACGAGGAGGATTGATATTACCTGGCCCCG
 CGGTGATGCCCTTTGAGGGTGGCCGCATCCATCTGGTTCAGAAAAGACAATCTTTTGTGTCAAGCTTGGT
 GGCAAAACGACCCGTAGAGGGCGTGGACAGCAACTTGGCGATGGAGCGCAGGGTTGGTTTGTGCGCA
 TCGGCGCGCTCTTGGCCGCGATGTTTAGCTGCACGTATTCGCGCGCAACGCACCGCCATTTCGGGAAAGA
 CGGTGTGCGCTCGTCCGGCACCAGGTGCACGCGCAACCGCGGTGTGTCAGGGTGACAAGGTCAACGCT
 GGTGGCTACCTCTCCGCGTAGGCGCTCGTTGGTCCAGCAGAGGCGGCGCCCTTCGCGAGCAGAAATGGC
 GGTAGGGGGTCTAGCTCGCTCTCGTCCGGGGGTCTGCGTCCACGGTAAAGACCCCGGCAGCAGCGCG
 CGTCGAAGTAGTCTATCTTGCATCTTGAAGTCTAGCGCTGCTGCCATGCGCGGGCGGCAAGCGCGCG
 CTCGTATGGGTTGAGTGGGGGACCCCATGGCATGGGGTGGGTGAGCGCGGAGGCGTACATGCCGCAATG
 TCGTAAACGTAGAGGGGCTCTCTGAGTATTCCAAGATATGTAGGGTAGCATCTTCCACCGCGGATGCTGG
 CGCGCACGTAATCGTATAGTTCTGTGCGAGGGAGCGAGGAGGTCCGGACCGAGGTTGCTACGGGCGGGCT
 CTCTGCTCGGAAGACTATCTGCCTGAAGATGGCATGTGAGTTGGATGATATGGTTGGACGCTGGAAGACG
 TTGAAGCTGGCGTCTGTGAGACCTACCGCGTCACGCACGAAGGAGCGGTAGGAGTCCGCGAGCTTGTGA
 CCAGCTCGGCGGTGACCTGCAGCTCTAGGGCGCAGTAGTCCAGGGTTTCTTGATGATGTCACTATTATC
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 GGAAACCCGTCGGCCTCCGAACGGTAAGAGCCTAGCATGTAGAACTGGTTGACGGCTGGTAGGCGCAGC
 ATCCCTTTTCTACGGGTAGCGGTATGCCGTGCGCGCCCTTCGCGAGCGAGGTGTGGGTGAGCGCAAAGGT
 GTCCCTGACCATGACTTTGAGGTACTGGTATTGGAAGTCAGTGTCTGCGCATCCGCCCTGCTCCCAGAGC
 AAAAAGTCCGTGCGCTTTTGGAAACGCGGATTGTCAGGGCGAAGGTGACATCGTTGAAGAGTATCTTTT
 CCGCGCGAGGCATAAAGTTGCGTGTGATGCGGAAGGGTCCCGGCACCTCGGAACGGTTGTTAATTACCTG
 GGCGGCGAGCACGATCTCGTCAAGCCGTTGATGTTGTGGCCACAAATGTAAAGTTCCAAGAAGCGCGGG
 ATGCCCTTGATGGAAGGCAATTTTGAAGTCTCTCGTAGGTGAGCTCTTCAGGGGAGCTGAGCCCGTGCT
 CTGAAAGGGCCAGTCTGCAAGATGAGGGTTGGAAGCGACGAATGAGCTCCACAGGTACAGGGCCATTAG
 CATTTGACAGTGGTTCGCGAAAGTCTTAAACTGGCGACCTATGGCCATTTTCTGGGGTGTATGAGTAG
 AAGGTAAGCGGGTCTTGTTCACAGCGGTCCCATCAAGGTTTCGCGGTAGGTCTCGCGCGGCAGTCACTA
 GAGGCTCATCTCCGCCGAACCTCATGACCAGCATGAAGGGCACGAGCTGCTTCCAAAGGCCCCATCCA
 AGTATAGGTCTCTACATCGTAGGTGACAAAGAGACGCTCGGTGCGAGGATGCGAGCCGATCGGGAAGAAC
 TGGATCTCCCGCCACCAATTGAGGAGTGGCTATTGATGTGGTGAAGTAGAAGTCCCTGCGACGGGCGG
 AACACTCGTGTGCTGCTTTTGTAAACAGTGGCGAGTACTGGCAGCGGTGCACGGGCTGTACATCCTGCAC
 GAGGTTGACCTGACGACCGCGCACAAAGGAAGCAGAGTGGGAATTTGAGCCCTCGCTGCGCGGTTTGGC
 TGGTGGTCTTCTACTTCCGCTGCTTGTCTTGACCGTCTGGCTGCTCGAGGGGAGTTACGGTGGATCGGA
 CCACCACGCGCGCGAGCCCAAAGTCCAGATGTCCGCGCGCGGGTCCGAGCTTGATGACAACATCGCG
 CAGATGGGAGCTGTCCATGGTCTGAGCTCCCGCGCGGTACAGTCCAGGCGGGAGCTCCTGCAGGTTTACC
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 CGTCGATGGCTTGCAAGAGGCGCATCCCCGCGCGGCGACTACGGTACCGCGCGGGCGGGTGGGCGCG
 GGGGGTGTCTTGGATGATGATCTAAAGCGGTGACGCGGGCGAGCCCCGAGGTAGGGGGGGCTCCG
 GACCCGCGGGGAGAGGGGGCAGGGGCAGTCCGCGCGCGCGCGGGCAGGAGCTGGTGTGCGCGCGTAG
 GTTGTGCGGAACGCGACGACGCGCGGTTGATCTCTGAATCTGGCGCTCTGCGTGAAGACGACGGGC
 CCGGTGAGCTTGAGCTGAAAGAGAGTTGACAGAAATCAATTTCCGTGTCTTGACGGCGGCTGGCGCA
 AAATCTCTGACGCTCTCTGAGTTGTCTTGATAGGCGATCTCGGCCATGAAGTGTCTCGATCTCTTCTC
 CTGGAGATCTCCGCGTCCGGCTCGCTCCAGGTGGCGGAGGTCTGTGAAATGCGGGCATGAGCTGC
 GAGAAGGCGTTGAGGCTCCCTCGTTCCAGACGCGGTGTAGACCACGCCCCCTTCGGCATCGCGGGCGC
 GCATGACCACTGCGCGAGATTGAGCTCCACGTGCCGGCGAAGACGGCGTAGTTTCGAGGCGGTGAAA
 GAGGTAGTTGAGGGTGGTGGCGGTGTGTCTGCCACGAAGAAGTACATAACCCAGCGTCGCAACGTGGAT
 TCGTTGATATCCCCAAGGCCTCAAGGCGCTCCATGGCTCGTAGAAGTCCACGGCGAAGTTGAAAACT
 GGGAGTTGCGCGCGGACACGGTTAACTCTCTCTCCAGAAGACGGATGAGCTCGGCGACAGTGTGCGGCAC
 CTCGCGCTCAAAGGCTACAGGGGCTCTCTTCTCTTCAATCTCTCTTCCATAAGGGCTCTCTCTCT
 TCTTCTTCTGGCGCGGTGGGGGAGGGGGACGCGCGGCGACGAGCGCGACCGGGAGGGGTGACAA
 AGCGCTCGATCATCTCCCGCGCGACGCGCGCATGGTCTCGGTGACGGCGCGCGCTTCTCGCGGGGCG

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

CAGTTGGAAGACGCCGCCGTCATGTCCCGTTATGGGTTGGCGGGGGGCTGCCATGCGGCAGGGATACG
 GCGCTAACGATGCATCTCAACAATTGTTGTGTAGGTACTCCGCCGCCGAGGGACCTGAGCGAGTCCGCGAT
 CGACCGGATCGGAAAACCTCTCGAGAAAGGCGTCTAACCAGTCACAGTCGCAAGGTAGGCTGAGCACCGT
 GCGGGCGGCAGCGGGCGGCGGTCCGGGTTGTTTCTGGCGGAGGTGCTGCTGATGATGTAATTAAAGTAG
 GCGGTCTTGAGACGGCGGATGGTCGACAGAAGCACCATGTCCTTGGGTCCGGCCTGCTGAATGCGCAGGC
 GGTCCGCCATGCCCCAGGCTTCGTTTGGACATCGGCGCAGGTCTTTGTAGTAGTCTTGCATGAGCCTTTC
 TACCGGCACTTCTTCTCTCTCTCTCTTGTCTGTCATCTCTTGCATCTATCGCTGCGGCGCGCGGAG
 TTTGGCCGTAGGTGGCGCCCTCTTCTCTCCATGCGGTGTGACCCCGAAGCCCTCATCGGCTGAAGCAGGG
 CTAGGTCCGGCAGACAAAGCGCTCGGCTAATATGGCCTGCTGCACCTGCGTGAGGGTAGACTGGAAGTCATC
 CATGTCCACAAAGCGGTGGTATGCGCCCGTGTGATGGTGAAGTGCAGTTGGCCATAACGGACCAAGTTA
 ACGGTCTGGTGACCCGGCTGCGAGAGCTCGGTGTACCTGAGACGCGAGTAAGCCCTCGAGTCAAAATACGT
 AGTCGTTGCAAGTCCGACCAAGTACTGGTATCCCAACAAAGTGGCGCGCGGCTGGCGGTAGAGGGG
 CCAGCGTAGGGTGGCGCGGCTCCGGGGCGAGATCTTCCAACATAAGGCGATGATATCCGTAGATGTAC
 CTGGACATCCAGGTGATGCCGCGCGGTGGTGGAGGCGCGGAAAGTCGCGGACGCGGTTCCAGATGT
 TGCGCAGCGGCAAAAGTGCTCCATGGTCGGGACGCTCTGGCCGGTCAAGCGCGCAATCGTTGACGCT
 CTAGACCGTGCAAAAGGAGAGCCTGTAAGCGGGCACTCTTCCGTGGTCTGGTGGATAAATTCGCAAGGGT
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 CGTGTCCGAACCCAGGTGTGCGACGTGACACAACGGGGGAGTGCTCCTTTTGGCTTCCTTCCAGGCGCGGC
 GGCTGTGCGCTAGCTTTTGGCCACTGGCCGCGCGCAGCGTAAGCGGTTAGGCTGGAAAGCGAAAGCA
 TTAAGTGGCTCGCTCCCTGTAGCCGGAGGGTTATTTTCAAGGGTTAGTTCGCGGACCCCGGTTCCGAG
 TCTCGGACCGCCGACTGCGGCGAAGCGGGGTTTGCTTCCCGTTCATGCAAGACCCCGCTTGCAAAATC
 CTCCGGAACAGGGACGAGCCCTTTTGGCTTTCACAGATGATCCCGTGTGCGGAGATGCGCCCC
 CCTCTCAGCAGCGGCAAGAGCAAGAGCAGCGGAGACATGCAGGGCACCTCCCTCTCTTACCGCGT
 CAGGAGGGGCGACATCCGCGGTTGACGCGGCGAGCAGATGGTGATTACGAACCCCGCGGCGCGGGCCCG
 GCACTACCTGGACTTGGAGGAGGGCGAGGGCTGGCGCGGCTAGGAGCGCCCTCTCTGAGCGGTACCCA
 AGGGTGCAGCTGAAGCGTGATACGCGTGAGGCGTACGTGCCGCGGCGAGAACCTGTTTCGCGACCGCGAGG
 GAGAGGAGCCCGAGGAGATGCGGGATCGAAAGTTCCACGAGGGCGCGAGCTGCGGCATGGCCTGAATCG
 CGAGCGGTTGCTGCGCGAGGAGGACTTTGAGCCCGACGCGCGAACCAGGATTAGTCCCGCGCGCGCACAC
 GTGGCGGCGCGGACCTGGTAACCGCATACGAGCAGCGGTGAACAGGAGATTAACTTTCAAAAAGCT
 TGTAAGCGCGCTGGAGCAAAACCAATAGCAAGCCGCTCATGGCGCAGCTGTTCTTATAGTGCAGCAC
 AGCAGGGACAACGAGGCATTGAGGATGCGCTGCTAAACATAGTAGAGCCCGAGGGCCGCTGGCTGCTCG
 ATTTGATAAACATCCTGCAGAGCATAGTGGTGCAGGAGCGCAGCTTGAGCCTGGCTGACAAGGTGGCCGC
 CATCAACTATTCCATGCTTAGCCTGGGCAAGTTTACGCCCGCAAGATATACCATAACCCCTTACGTTCCC
 ATAGACAAGGAGGTAAAGATCGAGGGGTTCTACATGCGCATGGCGCTGAAGGTGCTTACCTTGAGCGAGC
 ACCTGGGCGTTTATCGCAACGAGCGCATCCACAAGCCGCTGAGCGTGAGCCGCGCGGAGCTCAGCGA
 CCGCGAGCTGATGCACAGCCTGCAAAAGGCGCTGGCTGGCACGGGCGAGCGGCGATAGAGAGGCGGAGTCC
 TACTTTGACGCGGGCGCTGACCTGCGCTGGGCCCCAAGCCGACGCGCCCTGGAGGCGAGTGGGGCCGAC
 CTGGGCTGGCGGTGGCACCCGCGCGCTGGCAACGTCGCGCGCTGGAGGAATATGACGAGGACGATGA
 GTACGAGCCAGAGGACGGCGAGTACTAAGCGGTGATGTTCTGATCAGATGATGAAGACGCAACGGACC
 CGGCGGTGCGGGCGCGCTGAGAGCCAGCCGTCGCGCTTAACCTCACGGACGACTGGCGCCAGGTGAT
 GGACCGCATCATGTGCTGACTGCGCGCAATCCTGACGCGTTCCGGCAGCAGCCGAGGCCAACCCTC
 TCCGCAATTCTGGAAGCGGTGGTCCCGCGCGCGCAACCCACGACGAGAGAAGGTGCTGGCGATCGTAA
 ACGCGCTGGCCGAAAACAGGGCCATCCGGCCCGACAGGCGCGCCTGGTCTACGACGCGCTGCTTCAGCG
 CGTGGCTCGTTACAACAGCGGCAACGTGACAGCAACCTGGACCGGCTGGTGGGGATGTGCGCGAGGCC
 GTGGCGCAGCGTGAGCGCGCGCAGCAGAGGGCAACCTGGGCTCCATGGTTGCACTAAACGCCCTTCTGA
 GTACACAGCCCGCAACGTGCCGCGGGACAGGAGGACTACACCAACTTTGTGAGCGCACTGCGGCTAAT
 GGTGACTGAGACACCGCAAAGTGAGGTGTACAGTCTGGGCCAGACTATTTTCCAGACCAGTAGACAA
 GGCTGCGAGACCGTAAACCTGAGCCAGGCTTTCAAAAACCTTGAGGGGCTGTGGGGGTGCGGGCTCCCA
 CAGGCGACCGCGGACCGTGTCTAGCTTGCTGACGCCCAACTCGCGCTGTTGCTGCTGCTAATAGCGCC
 CTTACAGGACAGTGGCAGCGTGTCCCGGACACATACCTAGGTCACTTGTGACACTGTACCGCGAGGCC
 ATAGGTACGGCGCATGTGAGAGCATATTTCCAGGAGATTACAAGTGTGAGCCGCGCGCTGGGGCAGG
 AGGACACGGGCGAGCTGGAGGCAACCTAAACTACCTGCTGACCAACCGGCGCGAGAAGATCCCTCGTT
 GCACAGTTTAAACAGCGAGGAGGAGCGCATTTTGGCGTACGTGACGAGAGCGTGAGCCTTAACCTGATG
 CGCGACGGGGTAACGCCAGCGTGGCGCTGGACATGACCGCGCGCAACATGGAACCGGGCATGTATGCCT
 CAAACCGGCGGTTTATCAACCGCTAATGGACTACTGTCATCGCGCGGCGCGGTGAACCCGAGTATTT
 CACCAATGCCATCTTGAACCGCACTGGCTACCGCCCTGGTTTCTACACCGGGGATTGAGGTGCCC
 GAGGGTAACGATGGATTCTCTGGGACGACATAGACGACAGCGTGTTCCTCCCGCAACCGCAGACCTGC
 TAGAGTTGCAACAGCGCGAGCAGGCGCGCTGCGAAAGGAAAGCTTCCGCGAGGCCAAGCAGCTT

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

GTCCGATCTAGGCGCTGCGGCCCGCGGTGAGATGCTAGTAGCCATTTCCTCAAGCTTGATAGGGTCTCTT
 ACCAGCACTCGCACCACCCGCGCGCTGCTGGGCGAGGAGGTACCTAAACAACCTGCTGCTGCAGC
 CGCAGCGCGAAAAAACCCTGCCTCCGGCATTTCCCAACAACGGGATAGAGAGCCTAGTGGACAAGATGAG
 TAGATGGAAGACGTACGCGCAGGAGCACAGGGACGTGCCAGGCCCGCGCCCGCCACCCGTCGTCAAAGG
 CACGACCGCTCAGCGGGGTCTGGTGTGGGAGGACGATGACTCGGCAGACGACAGCAGCGTCTCGGATTGG
 GAGGGAGTGGCAACCCGTTTGGCGACCTTCGCCCCAGGCTGGGGAGAATGTTTTAAAAAAGCAT
 GATGCAAAATAAAAACTCACCAGGCCATGGCACCAGCGTGGTGGTTTCTTGTATTCCCTTAGTATGC
 GGCGCGCGCGATGTATGAGGAAGGTCTCTCCCTCCCTACGAGAGTGTGGTGAGCGCGCGCCAGTGGC
 GGCGCGCTGGGTCTCCCTTCGATGCTCCCTGGACCCGCGTGTGTGCTCCGCGGTACCTGCGGCCT
 ACCGGGGGAGAAACAGCATCCGTTACTCTGAGTTGGCACCCTATTTCGACACCACCCGTGTGTACCTGG
 TGGACAACAAGTCAACGGATGTGGCATCCCTGAACCTACGAGAACGACCACAGCAACTTTCTGACCACGGT
 CATTCAAAACAATGACTACAGCCCGGGGGAGGCAAGCACACAGACCATCAATCTTGACGACCGGTGCGAC
 TGGGGCGGCGACCTGAAAACCATCTGCATACCAACATGCCAAATGTGAACGAGTTCATGTTTACCAATA
 AGTTTAAGGCGCGGTGATGGTGTGCGCTTGCCTACTAAGGACAATCAGGTGGAGCTGAAATACGAGTG
 GGCTGGGTTTCACTGACCGGAGGGCACTACTCCGAGACCATGACCATAGACCTTATGAACACCGCATC
 GTGGAGCACTACTTGAAGTGGGCAGACAGAACGGGGTCTGGAAGCGACATCGGGGTAAAGTTTGACA
 CCCGCAACTTCAGACTGGGGTTTGACCCCGTCACTGGTCTTGTGTCATGCCCTGGGGTATATACAAACGAAGC
 CTTCCATCCAGACATCATTTTGGCTGCCAGGATGCGGGTGGACTTCACCCACAGCCGCTGAGCAACTTG
 TTGGGCATCCGCAAGCGGCAACCTTCCAGGAGGGCTTTAGGATCACCTACGATGATCTGGAGGGTGGTA
 ACATTCCCGCACTGTTGGATGTGGACGCTTACAGGCGAGCTTGAAAGATGACACCGAACAGGGCGGGGG
 TGGCGCAGGCGGCGAGCAACAGCAGTGGCAGCGGCGGAGAGAACTCCAACGCGGCGAGCCGCGCAATG
 CAGCCGGTGGAGGACATGAACGATCATGCCATTGCGGCGACACCTTTGCCACACGGGCTGAGGAGAAGC
 GCGCTGAGGCGGAGCAGCGGCGGAGCTGCGGCCCGCTGCGCAACCCGAGGTGAGAGCCTCAGAA
 GAAACCGGTGATCAAAACCTGACAGAGGACAGCAAGAAACGAGTTACAACCTAATAAGCAATGACAGC
 ACCTTCACCCAGTACCGCAGCTGGTACCTTGCATACAACTACGGCGACCTCAGACCGGAATCCGCTCAT
 GGACCTGCTTTGCACTCCTGACGTAACCTGCGGCTCGGAGCAGGTCTACTGGTGGTGGCAGACATGAT
 GCAAGACCCGCTGACCTTCCGCTCCACGCGCCAGATCAGCAACTTTCGGTGGTGGGCGCGGAGCTGTTG
 CCCGTGCACTCCAAGAGCTTCTACAACGACAGGCGCTTACTCCCACTCATCCGCGAGTTTACCTCTC
 TGACCGGCTGATTCAAGCTGCTTTCCCGAGAACCAGATTTTGGCGCGCCCGCCAGCCCCACCATCACAC
 CGTCAGTGAAAACGTTCTGCTCTCACAGATCACGGGACGCTACCGCTGCGCAACAGCATCGGAGGAGTC
 CAGCGAGTGACCACTACTGACGCCAGACGCGCACCTGCCCTACGTTTACAAGGCCCTGGGCATAGTCT
 CGCGCGCGCTCCTATCGAGCCGCACTTTTGGAGCAAGCATGTCCATCCTTATATCGCCAGCAATAACAC
 AGGCTGGGGCGCTGCGCTTCCCAAGCAAGATGTTTGGCGGGGCAAGAAGCGCTCCGACCAACACCCAGTG
 CGCGTGCGCGGCACTACCGCGCGCTTGGGGCGCGCAAAACGCGGCCGCACTGGGCGCACCAACCGTGC
 ATGACGCCATCGACCGGTGGTGGAGGAGGCGCGCACTACACGCCACGCGCCAGCCAGTGTCCAGT
 GGACCGGCTTCAAGACCGTGGTGGCGGAGCCGCGCTATGCTAAATGAAGAGACGGCGGAGGCGC
 GTAGCAGCTCGCCACCGCGCGGACCCGCACTGCCGCCAACGCGCGGCGGCGGCGCTGCTTAACCGCG
 CACGTGCGACCGCGCGACGGGCGGCCATGCGGGCGCTCGAAGGCTGGCCGCGGTATTGTCACTGTGCC
 CCCAGGTCCAGGCGAGCGGCGCGCGCAGCAGCCGCGCCATTAGTGTATGACTCAGGGTTCGCGAGG
 GGCAACGTGTATTGGGTGCGGACTCGGTTAGCGGCTGCGCGTGCCTGCGCACCCGCGCCCCCGCGCA
 ACTAGATTGCAAGAAAAAATACTTAGACTCGTACTGTTGTATGTATCCAGCGGCGGCGCGCAACGA
 AGCTATGTCCAAGCGCAAAATCAAAGAAGAGATGCTCCAGGTATCGCGCGGAGATCTATGGCCCCCG
 AAGAAGGAAGAGCAGGATTACAAGCCCCGAAAGCTAAAGCGGGTCAAAAAGAAAAAGAAAGATGATGATG
 ATGAACCTTGACGACGAGGTGGAACGTGTCACGCTACCGCGCCAGGCGACGGGTACAGTGGAAAGGTG
 ACGCGTAAACGTGTTTTGCGACCCGCGCACACCGTAGTCTTTACGCGCGGTGAGCGCTCCACCCGCACC
 TACAAGCGCGTGTATGATGAGGTGTACGCGACGAGGACCTGCTTGAGCAGGCCAACGAGCGCTCGGGG
 AGTTTGCCTACGGAAAGCGGCATAAGGACATGCTGGCGTTGCCGCTGGACGAGGGCAACCAACACCTAG
 CCTAAAGCCCGTAACACTGCAGCAGGTGCTGCCCGCGCTTGACCGTCCGAAGAAAGCGCGGCTAAAG
 CGCGAGTCTGGTACTTGGCACCACCGTGCAGCTGATGGTACCAAGCGCCAGCGACTGGAAGATGTCT
 TGGAAAAAATGAGCTGGAACCTGGGCTGGAGCCGAGGTCCGCGTGGCGCAATCAAGCAGGTGGCGCC
 GGGACTGGGCGTGACAGCCGTGGACGTTTCAATACCCACTACAGTAGCACCAGTATTGCCACCGCCACA
 GAGGGCATGGAGACACAAACGTCCCCGTTGCTCAGCGGTGGCGGATGCCGCGGTGACGGCGGTGCTG
 CGGCGCGCTCAAGACCTCTACGGAGGTGCAACCGACCCGTGGATGTTTCGCGTTTTCAGCCCCCGCG
 CCCGCGCGGTTCGAGGAAGTACGGCGCGCCAGCGCGTACTGCCCGAATATGCCCTACATCCTTCCATT
 GCGCTACCCCCGCTATCGTGGCTACACCTACCGCCCCAGAAGACGAGCAACTACCCGACGCGCAACCA
 CCACTGGAACCCGCGCGCGTCCGCTCGCCAGCCGCTGCTGGCCCGGATTTCGCTGCGCAGGGTGGC
 TCGCGAAGGAGGACGACCTGGTGTGCGCAACAGCGCGTACCACCCAGCATCGTTTTAAAGCCGGTGC
 TTTGTGGTCTTTCAGATATGGCCCTCACCTGCGCGCTCCGTTTCCCGGTGCGGGGATTCGAGGAAGAA
 TGCACCGTAGGAGGGCATGGCCGGCCACGGCTGACGGGCGCATGCGTGGTGGCACCACCGGCGCG

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

GCGCGCGTCGCACCGTCGCATGCGCGGCGGTATCCTGCCCTCCTTATTCCACTGATCGCCGCGGCGATT
 GGCGCCGTGCCCCGAATTGCATCCGTGGCCTTGCCAGGCGCAGAGACACTGATTAACAAACAAGTTGCATGT
 GGAAAAATCAAAAATAAAAAGTCTGGACTCTCAGCTCGCTTGGTCTGTAACTATTTGTAGAATGGAAG
 ACATCAACTTTGCGTCTCTGGCCCCGCGACACGGCTCGCGCCCGTTCATGGGAACTGGCAAGATATCGG
 CACCAGCAATATGAGCGGTGGCGCCTTCAGCTGGGGCTCGCTGTGGAGCGGCATTAAAAATTTTCGGTTCC
 ACCGTTAAGAACTATGGCAGCAAGGCTTGAACAGCAGCACAGGCCAGATGCTGAGGGATAAGTTGAAAG
 AGCAAAATTTCCAAACAAAGGTGGTAGATGGCCTGGCCTCTGGCATTAGCGGGGTGGTGGACCTGGCCAA
 CCAGGCAGTGCAAAATAAGATTAACAGTAAGCTTGATCCCCGCCCTCCCGTAGAGGAGCCTCCACCGGCC
 GTGGAGACAGTGTCTCCAGAGGGGCGTGGCGAAAAGCGTCCGCGCCCCGACAGGGAAGAACTCTGGTGA
 CGCAATAGACGAGCCTCCCTCGTACGAGGAGGCACTAAAGCAAGGCTGCCACCACCCGTCCTATCGC
 GCCCATGGCTACCGGAGTGTGGGCGAGCACACCCGTAACGCTGGACCTGCCTCCCCCGCCGACACC
 CAGCAGAAACCTGTGTCTGCCAGGCCGACCGCGCTTGTGTAAACCGTCTAGCCGCGCGTCCCTGCGCC
 CGCGCGCCAGCGGTCCGCGATCGTTGCGGCCCCGTAGCCAGTGGCAACTGGCAAAGCACACTGAACAGCAT
 CGTGGGTCTGGGGGTGCAATCCCTGAAGCGCCGACGATGCTTCTGAATAGCTAACGTGTGATGTGTGT
 CATGTATGCGTCCATGTGCGCGCCAGAGGAGCTGCTGAGCCGCGCGCGCCCGCTTTTCAAGATGGCTAC
 CCCTTCGATGATGCCGCGAGTGGTCTTACATGCACATCTCGGGCCAGGACGCCTCGGAGTACCTGAGCCCC
 GGGCTGGTGCAGTTTGCCCGCGCCACCGAGACGTACTTCAGCCTGAATAACAAGTTTAGAAACCCACCG
 TGGCGCCTACGCACGACGTGACCACAGACCGGTCCAGCGTTTGACGCTGCGGTTTATCCCTGTGGACCG
 TGAGGATACTGCGTACTCGTACAAGGCGCGGTTTACCCTAGCTGTGGGTGATAACCGTGTGTGACATG
 GCTTCCACGTCTTTGACATCCGCGCGGTGCTGGACAGGGGCCCCACTTTTAAGCCCTACTCTGGCACTG
 CCTACAACGCTCTGGCTCCCAAGGGTGCCCCAAATCCTTGCGAATGGGATGAAGCTGCTACTGCTCTTGA
 AATAAACCTAGAAGAAGAGGACGATGACAACGAGCAAGTAGACGAGCAAGCTGAGCAGCAAAAACT
 CACGTATTTGGGCAGGCGCCTTATTCTGGTATAAATATTACAAAGGAGGGTATTCAAATAGGTGTGGAAG
 GTCAAACACCTAAATATGCCGATAAAACATTTCAACCTGAACCTCAAATAGGAGAATCTCAGTGGTACGA
 AACTGAAATTAATCATGCAGCTGGGAGAGTCCCTTAAAAAGACTACCCCAATGAAACCATGTTACGGTTCA
 TATGCAAAACCCACAAATGAAATGGAGGGCAAGGCATTCTTGTAAAGCAACAAATGGAAAGCTAGAAA
 GTCAAGTGGAAATGCAATTTTCTCACTACTGAGGCGACCCGAGGCAATGGTGATAACTTGACTCTTAA
 AGTGGTATTGTACAGTGAAGATGTAGATATAGAAACCCAGACACTCATATTTCTTACATGCCCACTATT
 AAGGAAGGTAACCTACGAGAACTAATGGGGCCCAACATCTATGCCCAACAGGCCTAATTACATTGCTTTTA
 GGGACAATTTTATTGGTCTAATGTATTACAACAGCAGCGGTAAATATGGGTGTTCTGGCGGGCCAAGCATC
 GCAGTTGAATGCTGTTGTAGATTGTGAAGACAGAAACACAGAGCTTTCATACCAGCTTTTGCTTGATTCC
 ATTGGTGTAGAACACAGGTACTTTTCTATGTGAATCAGGCTGTGACAGCTATGATCCAGATGTTAGAA
 TTATTGAAATCATGGAACCTGAAGATGAACCTCCAAATTAAGTCTTTCCACTGGGAGGTGTGATTATAC
 AGAGACTCTTACCAAGGTAACCTAAACAGGTGAGGAAATGGATGGGAAAAAGATGCTACAGAATTT
 TCAGATAAAAAATGAATAAGAGTTGGAATAATTTGGCCATGGAATCAATCTAAATGCCAACCTGTGGA
 GAAATTTCTGTACTCCAACATAGCGCTGTATTTGCCGCAAGCTAAAGTACAGTCTTCCAACTGAAA
 AATTTCTGATAACCCAAACACCTACGACTACATGAACAAGCGAGTGGTGGCTCCCGGGTTAGTGGACTGC
 TACATTAACCTTGGAGCACGCTGGTCCCTTGACTATATGGACAACGTCAACCCATTTAACCACCACCGCA
 ATGCTGCGCTGCGCTACCGCTCAATGTTGCTGGGCAATGGTTCGCTATGTGCCCTTCCACATCCAGGTGCC
 TCAGAAGTTCTTTGCCATTAAAAACCTCCTTCTCCTGCCGGGCTCATACACCTACGAGTGGAACTTCAGG
 AAGGATGTTAATACATGGTTCTGACAGCTCCCTAGGAAATGACCTAAGGGTTGACGGAGCCAGCATTAAGT
 TTGATAGCATTGGCTTTACGCCACCTTCTCCCATGGGCCACAACACCGCCTCCACGCTTGAGGCCAT
 GCTTAGAAACGACACCAACGACAGTCTTTAACGACTATCTCTCCGCCGCAACATGCTCTACCTATA
 CCCGCCAACGCTACCAACGTGCCCATATCCATCCCTCCCGCAACTGGGCGGCTTTCCGCGGTGGGCCCT
 TCACGCGCCTTAAGACTAAGGAAACCCCATCACTGGGCTCGGGCTACGACCCTTATTACACCTACTCTGG
 CTCTATACCCTACCTAGATGGAACCTTTTACCTCAACCACACCTTTAAGAAGGTGGCCATTACCTTTGAC
 TCTTCTGTGCTGCGCTGGCCTGGCAATGACCGCTGCTTACCCCAACGAGTTTGAATTAAGCGCTCAGTTG
 ACGGGGAGGGTTACAACGTTGCCAGTGTAACATGACCAAGACTGGTTCTTGGTACAAATGCTAGCTAA
 CTACAACATTGGCTACCGAGGCTTCTATATCCAGAGAGCTACAAGGACCGCATGTACTCCTTCTTTAGA
 AACTTCCAGCCCATGAGCGCTCAGGTGGTGGATGATACTAAATAACAAGGACTACCAACAGGTGGGCATCC
 TACACCAACACAACAACTCTGGATTTGTTGGCTACCTTGCCCCCACCATGCGCGAAGGACAGGCCATACC
 TGCTAACTTCCCCTATCCGCTTATAGGCAAGACCGCAGTTGACAGCATTACCCAGAAAAAGTTTCTTTGC
 GATCGCACCTTTTGGCGCATCCCATTTCCAGTAACCTTATGTCCATGGGCGCACTCACAGACCTGGGCC
 AAAACCTTCTCTACGCCAACTCCGCCACGCGCTAGACATGACTTTTGAGGTGGATCCCATGGACGAGCC
 CACCCTTCTTTATGTTTGTGTTGAAGTCTTTGACGTGGTCCGTGTGCACCGGCCGACCGCGGCGTCACT
 GAAACCTGTGTGCGCAGCGCTTCTCGGCCGCAACGCCACAACATAAAGAAGCAAGCAACATCAAC
 AACAGCTGCCGCCATGGGCTCCAGTGAGCAGGAACCTGAAAGCCATTGTCAAAGATCTTGGTTGTGGGCCA
 TATTTTTTGGGCACCTATGACAAGCGCTTTCCAGGCTTTGTTTCTCCACACAAGCTCGCCTGCGCCATAG
 TCAATACGGCCGCTCGCGAGACTGGGGCGGTACACTGGATGGCCTTTGCCTGGAACCCGCACTCAAAAAC

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

ATGCTACCTCTTTGAGCCCTTTGGCTTTTCTGACCAGCGACTCAAGCAGGTTTACCAGTTTGAGTACGAG
TCACTCCTGCGCGGTAGCGCCATTGCTTCTTCCCCGACCGCTGTATAACGCTGGAAAAGTCCACCCAAA
GCGTACAGGGGCCCCAACTCGGCCGCTGTGGACTATTCTGCTGCATGTTTCTCCACGCTTTGCCAACTG
GCCCCAACTCCCATGGATCACAACCCCAACCATGAACCTTATTACCGGGGTACCCAACTCCATGCTCAAC
AGTCCCCAGGTACAGCCACCCCTGCGTCGCAACCAGGAACAGCTCTACAGCTTCTCTGGAGCGCCACTCGC
CCTACTTCCGACGCCACAGTGGCAGATTAGGAGCGCCACTTCTTTTGTCACTTGAAAAACATGTAAAA
ATAATGTACTAGAGACACTTTCAATAAAGGCAAATGCTTTTATTGTACACTCTCGGGTGATTATTTACC
CCACCCCTTGCCGCTTGTGCGCGTTTAAAAATCAAAGGGGTTCTGCGCGCATCGCTATGCGCGCACTGGCA
GGGACACGTTGCGATACTGGTGTTTAGTGCTCCACTTAAACTCAGGCACAACCATCCGCGGCAGCTCGGT
GAAGTTTTCACTCCACAGGCTGCGCACCATCACCACGCGTTTAGCAGGTGCGGCGCGATATCTTGAAG
TCGCAAGTTGGGGCTCCGCCCTGCGCGCGAGTTGCGATACACAGGGTTGCAGCACTGGAACACTATCA
GCGCCGGGTGTTGACGCTGGCCAGCAGCTCTTGTGCGAGATCAGATCCGCGTCCAGGTCTCCGCGTT
GCTCAGGGCGAACGGAGTCAACTTTGGTAGCTGCCTTCCCAAAAAGGGCGCGTGGCCAGGCTTTGAGTTG
CACTCGCACCGTAGTGCCATCAAAGGTGACCGTGCCCGGTCTGGGCGTTAGGATACAGCGCCTGCATAA
AAGCCTTGATCTGCTTGAAGCCACTGAGCCTTTGCGCCTTCAAGAGAAGAACATGCCGCAAGACTGGCC
GGAAAACTGATTGGCCGACAGGCCGCGTCTGTCACGACGACCTTGCCTCGGTGTTGGAGATCTGCACC
ACATTTGGGCCCCACCGGTTCTTACGATCTTGGCCTTGCTAGACTGCTCCTTACGCGCGCTGCCCGT
TTTCGCTCGTCACATCCATTTCAATCAGTGCTCCTTATTTATCATAATGCTTCCGTGTAGACACTTAAG
CTCGCCTTCGATCTCAGCGCAGCGGTGCGGCCACAACGCGCAGCCCGTGGGCTCGTGATGCTTGTAGGTC
ACCTCTGCAAACGACTGCAGGTACGCTGCGAGGAATCGCCCCATCATCGTCACAAAGGTCTTGTGCTGG
TGAAGGTCACTGCAACCCGCGGTGCTCCTCGTTGACGAGGTCTTGCTACAGGCGCCAGAGCTTCCAC
TTGGTCAGGCAGTAGTTTGAAGTTCGCTTTAGATCGCTTATCCACGTGGTACTTGTCCATCAGCGCGCG
GCAGCCTCCATGCCCTTCTCCACGACAGACGATCGGCACACTCAGCGGGTTTATCACCCTAATTTTAC
TTTCCGCTTCGCTGGGCTCTTCTCTTCTTCTGCTCCGCATACCACGCGCCACTGGGTGCTTCTATT
CAGCCGCGCACTGTGCGCTTACCTCCTTTGCCATGCTTGATTAGCACCGGTGGGTGCTGAAACCCACC
ATTTGTAGCGCCACATCTTCTCTTCTTCTTCTGCTGTCACGATTACCTCTGGTGATGGCGGGCGCTCGG
GCTTGGGAGAAAGGGCGCTTCTTTTCTTCTTGGGCGCAATGGCCAAATCCGCGCGCGAGGTGCGATGGCCG
CGGGCTGGGTGTGCGCGGCACAGCGCGTCTTGTGATGAGTCTTCTCGTCTCGACTCGATACGCGCG
CTCATTCTGCTGCGAGCGTTTGGGGGCGCCCGGGAGGCGCGGCGACGCGGACGACAGCTCTCCATGG
TTGGGGGACGTCGCGCGCACCGCGTCCGCGCTCGGGGTGGTTTCGCGCTGCTCCTCTTCCGACTGGC
CATTTCTTCTCTATAGGCAGAAAAAGATCATGGAGTCAGTCGAGAAGAAGGACAGCCTAACCGCCCC
TCTGAGTTTCCGACCCACCGCCTCCACCGATGCCGCCAACGCGCTTACCACCTTCCCGTGCAGGACCCCC
CGCTTGAGGAGGAGGAAGTGATTATCGAGCAGGACCCAGGTTTGTAGCGAAGACGACGAGGACCGCTC
AGTACCAACAGAGGATAAAAAGCAAGACCAGGACAACGACAGGCAACGAGGAACAAGTCGGGCGGGGG
GACGAAGGCATGGCGACTACCTAGATGTGGGAGACGAGTGTGTTGAAGCATCTGCAGCGCCAGTGCG
CCATTATCTGCGACGCGTTTGCAAGAGCGCAGCGATGTGCCCTCGCCATAGCGGATGTGACGCTTGCCTA
CGAACGCCACCTATTCTACCGCGCTACCCCCCAAACGCCAAGAAAACGGCACATGCGAGCCCCAACCCG
CGCCTCAACTTCTACCCCGTATTGCGCGTCCAGAGGTGCTTGCCACCTATCACATCTTTTCCAAACT
GCAAGATACCCCTATCCTGCGGTGCCAACCGCAGCCGAGCGGACAAGCAGCTGGCCTTGGCGCAGGGCGC
TGTATACCTGATATCGCCTCGCTCAACGAAGTGCCAAAATCTTTGAGGGTCTTGGACGCGACGAGAAG
CGCGCGGCAACGCTCTGCAACAGGAAAACAGCGAAAATGAAAGTCACTCTGGAGTGTGGTGGAACCTCG
AGGGTGACAACGCGCGCTTAGCCGTACTAAAACGACGATCGAGGTACCCACTTTGCCTACCCGGCACT
TAACCTACCCCCCAAGGTCTATGAGCACAGTCATGAGTGAGCTGATCGTGCGCCGTGCGCAGCCCTGGAG
AGGGATGCAATTTGCAAGAACAACAGAGGAGGGCTACCCGCAGTTGGCGACGAGCAGCTAGCGCGCT
GGCTTCAAACGCGCGAGCCTGCCGACTTGGAGGAGCGACGCAACTAATGATGGCCGAGTGCTCGTTAC
CGTGAGCTTGAGTGATGCAGCGGTCTTTGCTGACCCGAGATGCAGCGCAAGCTAGAGGAACATTG
CACTACACCTTTGACAGGGCTACGTACGCCAGGCTGCAAGATCTCCAACGTGGAGCTCTGCAACCTGG
TCTCTACCTTGAATTTTGACGAAAACCGCCTTGGGCAAAACGTGCTTCAATCCACGCTCAAGGGCGA
GGCGCGCCGCGACTACGTCGCGACTGCGTTTACTTATTTCTATGCTACACCTGGCAGACGGCCATGGGC
GTTTGGCAGCAGTCTTGGAGGAGTGCAACCTCAAGGAGCTGCAGAACTGCTAAAGCAAAACTGAAGG
ACCTATGGACGGCCTTCAACGAGCGCTCCGTGGCCGCGCACCTGGCGGACATCATTTTCCCCGAACGCCT
GCTTAAACCCCTGCAACAGGGTCTGCCAGACTTACCAGTCAAAGCATGTTGCAGAACTTTAGGAACCTTT
ATCCTAGAGCGCTCAGGAATCTTGGCCGCCACCTGCTGTGCACTTCTTAGCGACTTTGTGCCCATTAGT
ACCGCGAATGCCCTCCGCCGCTTTGGGGCCACTGCTACCTTCTGAGCTAGCCAACCTTGCCTACCA
CTCTGACATAATGGAAGACGTGAGCGGTGACGGTCTACTGGAGTGTCACTGTGCTGCAACCTATGCACC
CCGACCCGCTCCTGTTTGAATTCGAGCTGTCTAACGAAAGTCAAATTATCGGTACCTTTGAGCTGC
AGGGTCCCTCGCCTGACGAAAAGTCCGCGGCTCCGGGTTGAAACTCACTCCGGGGCTGTGGACGTCGCG
TTACCTTCGCAATTTGTACCTGAGGACTACCACGCCCACGAGATTAGGTTCTACGAAGACCAATCCCGC
CCGCCAATGCGGAGCTTACCGCCTGCGTCATTACCCAGGGCCACATTCTTGGCCAATTGCAAGCCATCA

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

ACAAAGCCCGCCAAGAGTTTCTGCTACGAAAGGGACGGGGGGTTTACTTGGACCCCCAGTCCGGCGAGGA
GCTCAACCCAATCCCCCGCCGCGCAGCCCTATCAGCAGCAGCCGCGGGCCCTTGCTTCCCAGGATGGC
ACCCAAAAGAAGCTGCAGCTGCCGCGCCACCCACGGACGAGGAGGAATACTGGGACAGTCAGGCAGAG
GAGGTTTTTGGACGAGGAGGAGGAGACATGATGGAAGACTGGGAGAGCCTAGACGAGGAAGCTTCCGAGG
TCGAAGAGGTGTGACAGCAAACACCGTCACCCCTCGGTGCGATTCCCCCTCGCCGGCGCCCCAGAAATCGGC
AACCAGTTCCAGCATGGCTACAACCTCCGCTCCTCAGGCGCCGCCGGCACTGCCCGTTCCGCCAGCCAAC
CGTAGATGGGACACCACTGGAACAGGGCCGGTAAGTCCAAGCAGCCGCCCGCCGTTAGCCCAAGAGCAAC
AACAGCGCCAAGGCTACCGCTCATGGCGCGGGCACAAGAACGCCATAGTTGCTTGCTTGCAAGACTGTGG
GGGCAACATCTCCTTCGCCCGCCGCTTTCTTCTCTACCATCACGGCGTGGCCTTCCCCCGTAACATCCTG
CATTACTACCGTCATCTCTACAGCCCATAGTGCACGGCGGCGAGCGGCGAGCAACAGCAGCGGGCC
ACACAGAAGCAAGGACACCGGATAGCAAGACTCTGACAAAGCCCAAGAAATCCACAGCGGGCGGAGCAG
CAGGAGGAGGAGCGCTGCGTCTGGCGCCCAACGAACCCGTATCGACCCGCGAGCTTAGAAACAGGATTTT
TCCCACTCTGTATGCTATATTTCAACAGAGCAGGGGCCAAGAACAAGAGCTGAAAATAAAAAACAGGTCT
CTGCGATCCCTACCCGCGAGCTGCCTGTATCACAAAAGCGAAGATCAGCTTTCGGCGCACGCTGGAAGACG
CGGAGGCTCTCTCAGTAAATACTGCGCGTGAATCTTAAGGACTAGTTTCGGCGCCCTTTCTCAAATTTA
AGCGCAAAACTACGTCTATCTCCAGCGGCCACACCCGGCGCCAGCACCTGTGTCAGCGCCATTATGAGC
AAGGAAATTTCCACGCCCCATATGTGGAGTTACCGCCACAAATGGGACTTGGCGCTGGAGCTGCCCAAG
ACTACTCAACCCGAATAAATACTACATGAGCGCGGGACCCCATGATATCCCGGGTCAACGGAATCCGCGC
CCACCGAAACCGAATTCTCTTGAACAGGCGGCTATTACCACCACACCTCGTAATAACCTTAATCCCCGT
AGTTGGCCCGCTGCCCTGGTGTACAGGAAAGTCCCGCTCCACCACCTGTGGTACTTCCAGAGACGCCC
AGGCCGAAGTTCAGATGACTAATCAGGGGCGCAGCTTGGCGGGCGGCTTTCGTACAGGGTGCGGTCCGC
CGGGCAGGGTATAACTCAGCTGACAATCAGAGGGCGAGGTATTAGCTCAACGACGAGTCGGTGAGCTCC
TCGCTTGGTCTCCGTCCGGACGGGACATTTAGATCGCGCGCGCCGGCGCTCCTTCATTACGCGCTCTG
AGGCAATCCTAATCTGCAGACCTCGTCTCTGAGCCGCGCTCTGGAGGCATTGGAACCTCTGCAATTTAT
TGAGGAGTTTGTGCCATCGGTCTACTTTAACCCCTTCTCGGGACCTCCCGGCCACTATCCGGATCAATTT
ATTCTTAATTTGACGCGGTAAAGGACTCGGCGGACGGCTACGACTGAATGTTAAGTGGAGAGGCAGAGC
AATGCGCCTGAAACACCTGGTCCACTGTGCGCGCCACAAGTGCTTTGCCCGGACTCCGGTGAGTTTTG
CTACTTTGAATTGCCCAGGATCATATCAGGGGCCCCGCGCACGGCGTCCGGCTTACCGCCCAGGGAGAG
CTTGCCCGTAGCCTGATTTCGGGAGTTTACCCAGCGCCCCCTGCTAGTTGAGCGGGACAGGGACCCCTGTG
TTCTCACTGTGATTTGCAACTGTCTAACCTTGGATTACATCAAGATCTTTGTTGCCATCTCTGTGCTGA
GTATAATAAATACAGAAATTAATAATATACTGGGCTCCTATCGCCATCCTGTAAACGCCACCGCTCTTAC
CCGCCAAGCAAAACCAAGGCGAACCTTACCTGGTACTTTTAACATCTCTCCCTCTGTGATTTACAACAGT
TTCAACCCAGACGGAGTGAGTCTACGAGAGAACCTCTCCGAGCTCAGCTACTCCATCAGAAAAACACCA
CCCTCCTTACCTGCCGGGAACGTACGAGTGCGTCACCGCGCGCTGCACCACACCTACCGCCTGACCGTAA
ACCAGACTTTTTCCGGACAGACCTCAATAACTCTGTTTACCAGAACAGGAGGTGAGCTTAGAAAAACCTT
AGGGTATTAGGCCAAAGGCGCAGCTACTGTGGGGTTTATGAACAATTCAAGCAACTCTACGGGCTATTCT
AATTCAGGTTTCTCTAGAAATGGACGGAATTATTACAGAGCAGCGCTGCTAGAAAGACGCGGCGAGCG
GCCGAGCAACAGCGCATGAATCAAGAGCTCCAAGACATGGTTAATCTGCACCAAGTGCAAAAGGGGTATCT
TTTGTCTGGTAAAGCAGGCGCAAGCTACCTACGACAGTAATACCACCGGACACCGCCTTAGCTACAAGTT
GCCAACCAAGCGTCAGAAATGGTGGTCTAGGTGGGAGAAAAGCCATTACCATAACTCAGCACTCGGTA
GAAACCGAAGGCTGCATTCACTCACCTTGTCAAGGACCTGAGGATCTCTGCACCCTTATTAAGACCCCTGT
GCGGTCTCAAAGATCTTATTCCTTTAATAAATAAATAAATAAAGCATCACTTACTTAAATCAG
TTAGCAAATTTCTGTCCAGTTTATTACGAGCACCTCCTTGCCCTCCTCCAGCTCTGGTATTGCAGCTT
CCTCCTGGCTGCAACTTTCTCCACAATCTAATGGAATGTCAGTTTCTCCTGTTCTCTGTCCATCCGCA
CCCACTATCTTCATGTTGTTGTCAGATGAAGCGCGCAAGACCGTCTGAAGATACCTTCAACCCCGTGTATC
CATATGACACGGAACCGGCTCCTCCAACGTGTCCTTTTCTTACTCCTCCCTTTGTATCCCCCAATGGGTT
TCAAGAGAGTCCCCCTGGGGTACTCTCTTTGCGCTATCCGAACCTCTAGTTACCTCCAATGGCATGCTT
GCGCTCAAATGGGCAACGGCCTCTCTCTGGACGAGGCGCGCAACCTTACCTCCCAAAATGTAACCACTG
TGAGCCACCTCTCAAAAAACCAAGTCAAAACATAAACCTGGAATATCTGCACCCCTCACAGTTACCTC
AGAAGCCCTAAGTGTGGCTGCCGCCGACCTCTAATGGTCGCGGGCAACACACTCACCATGCAATCAGAG
GCCCCGCTAACCGTGACGACTCCAACTTAGCATTTGCCACCAAGGACCCCTCAGGTGTCAGAAAGGAA
AGCTAGCCCTGCAAAACATCAGGCCCCCTCACCAACCGATAGCAGTACCCCTTACTATCACTGCCTCACC
CCCTCTAATCTAGTCCACTGGTAGCTTGGGCGATTGACTTGAAAGAGCCCATTTATACAAAAATGGAAGA
CTAGGACTAAAGTACGGGGCTCCTTTGCATGTAACAGACGACCTAAACACTTTGACCGTAGCACTGGTC
CAGGTGTGACTATTAAATAACTTCTTGTCAAACATAAGTTACTGGAGCCTTGGGTTTGTATTACAAGG
CAATATGCAACTTAATGTAGCAGGAGGACTAAGGATTGATTCTCAAAACAGACGCTTATACTTGATGTT
AGTTATCCGTTTGATGCTCAAAACCAACTAAATCTAAGACTAGGACAGGGCCCTCTTTTATAAATCAG
CCCACAACCTGGATATTAACATAACAAAGGCTTTACTTGTTTACAGCTTCAAAACAATTCCAAAAGCT
TGAGGTTAACCTAAGCACTGCCAAGGGTTGATGTTTGACGCTACAGCCATAGCCATTAATGCAGGAGAT

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

GGGCTTGAATTTGGTTACCTAATGCACCAACACAAATCCCCTCAAAACAAAATTTGGCCATGGCCCTAG
 AATTTGATTCAAACAAGGCTATGGTTCCTAACTAGGAAGTGGCCTTAGTTTTGACAGCACAGGTGCCAT
 TACAGTAGGAAACAAAATAATGATAAGCTAACTTTGTGGACCACACCAGCTCCATCTCTAACTGTAGA
 CTAAATGCAGAGAAAGATGCTAACTCACTTTGGTCTTAACAAAATGTGGCAGTCAAATACTTGCTACAG
 TTTCAGTTTTTGGCTGTTAAAGGCAGTTTGGCTCCAATATCTGGAACAGTTCAAAGTGCTCATCTTATTAT
 AAGATTTGACGAAATGGAGTGCTACTAAACAATTCCTTCTGGACCCAGAATATTGGAACCTTTAGAAAT
 GGAGATCTTACTGAAGGCACAGCCTATACAAACGCTGTGGATTATGCTTAACCTATCAGCTTATCCAA
 AATCTCACGGTAAACATGCCAAAAGTAACATTGTTCAGTCAAGTTTACTTAAACGGAGACAAAATAAACC
 TGTAACACTAACCATTACACTAAACGGTACACAGGAAACAGGAGACACAACCTCCAAGTGCATACTCTATG
 TCATTTTTCATGGGACTGGTCTGGCCACAACCTACATTAATGAAATATTTGCCACATCCTCTTACACTTTTT
 CATACATTGCCCAAGAATAAAGAATCGTTTGTGTTATGTTTCAACGTGTTATTTTTTCAATTGCAGAAAA
 TTTCGAATCATTTTTTCATTAGTAGTATAGCCCCACCACCATAGCTTATACAGATCACCCTACCTTAA
 TCAAACTCACAGAACCCTAGTATTCAACCTGCCACCTCCCTCCCAACACACAGAGTACACAGTCCCTTTCT
 CCCCCGCTGGCCTTAAAAAGCATCATATCATGGGTAACAGACATATTCTTAGGTGTTATATTTCCACACGG
 TTTCTGTGTCGAGCCAAACGCTCATCAGTGATATTAAATAAACTCCCCGGGCAGCTCACTTAAGTTCATGTC
 GCTGTCCAGCTGCTGAGCCACAGGCTGCTGTCCAACCTTGCGGTTGCTTAACGGGCGGCGAAGGAGAAGTC
 CACGCTACATGGGGGTAGAGTCATAATCGTGCATCAGGATAGGGCGGTGGTGTGTCAGCAGCGCGCGAA
 TAAACTGCTGCCGCCCGCTCCGTCTGCAGGAATACAACATGGCAGTGGTCTCCTCAGCGATGATTTCG
 CACGCCCCGAGCATAAGGCGCCTTGTCTCCGGGCACAGCAGCGCACCTGATCTCACTTAAATCAGCA
 CAGTAACTGCAGCACAGCACCACAATATTGTTCAAATCCCACAGTGCAGGCGCTGTATCCAAAGCTCA
 TGGCGGGGACCACAGAACCACGCTGGCCATCATACCACAAGCGCAGGTAGATTAAGTGGCGACCCCTCAT
 AAACACGCTGGACATAAACATTACCTCTTTTGGCATGTTGTAATTCACCACCTCCCGGTACCATATAAAC
 CTCTGATTAAACATGGCGCCATCCACCACCATCTAAACCAGCTGGCCAAAACCTGCCCGCCGCTATAC
 ACTGCAGGGAACCGGGACTGGAACAATGACAGTGGAGAGCCAGGACTCGTAACCATGGATCATCATGCT
 CGTCATGATATCAATGTTGGCACAACACAGGCACACGTGCATACACTTCTCAGGATTACAAGCTCCTCC
 CGCGTTAGAACCATATCCAGGGAACAACCCATTCTGAATCAGCGTAAATCCCACACTGCAGGGAAGAC
 CTCGACAGTAACTCAGCTTGTGTCATTGTCAAAGTGTTACATTCCGGCAGCAGCGGATGATCCTCCAGTAT
 GGTAGCGCGGGTTTCTGTCTCAAAGGAGGTAGACGATCCCTACTGTACGGAGTGCGCGGAGACAACCGA
 GATCGTGTGTCGTAGTGTATGTCATGCCAAATGGAACGCCGACGTAGTCATATTCTCTGAAGCAAAACAG
 GTGCGGCGGTGACAAACAGATCTGCGTCTCCGGTCTCGCGCTTAGATCGCTCTGTGTAGTAGTGTAGT
 ATATCCACTCTCTCAAAGCATCCAGGCGCCCCCTGGCTTCGGGTTCTATGTAACTCCTTCATGCGCCGC
 TGCCCTGATAACATCCACCACCGCAGAAATAGCCACACCCAGCCAACCTACACATTCTGTTCTGCGAGTCA
 CACACGGGAGGAGCGGGAAGAGCTGGAAGAACCATGTTTTTTTTTTTATCCAAAAGATTATCCAAAACC
 TCAAAATGAAGATCTATTAAGTGAACGCGCTCCCCTCCGGTGGCGTGGTCAAACCTACAGCCAAAGAAC
 AGATAATGGCATTGTAAGATGTTGCACAATGGCTTCCAAAAGGCAAAACGGCCCTCAGCTCCAAGTGGAC
 GTAAAGGCTAAACCCCTTCAGGCTGAATCTCTCTATAAACATTCCAGCACCTTCAACCATGCCCAATAA
 TTCTCATCTCGCCACCTTCTCAATATATCTCTAAGCAAATCCCGAATATTAAGTCCGGCCATTGTAAAAA
 TCTGCTCCAGAGCGCCCTCCACCTTCAGCCTCAAGCAGCGAATCATGATTGCAAAAATTACAGTTCTCTCA
 CAGACCTGTATAAGATTCAAAGCGGAACATTAACAAAAATACCGCGATCCCGTAGGTCCCTTCCGAGGG
 CCAGCTGAACATAATCGTGCAGGTCTGCACGGACAGCGCGGCCACTTCCCCGCCAGGAACCTTGACAAA
 AGAACCCACACTGATTATGACACGCATACTCGGAGCTATGCTAACCAGCGTAGCCCCGATGTAAGCTTTG
 TTGCATGGGCGCGATATAAAATGCAAGGTGCTGCTCAAAAAATCAGGCAAAAGCCTCGCGCAAAAAGAA
 AGCAGATCGTAGTCATGCTCATGCGAGATAAAGGCAGGTAAGCTCCGGAACACCACAGAAAAAGACACCA
 TTTTTCTCTCAAACATGCTGTCGGGTTTCTGCATAAACACAAAATAAAATAACAAAAAACATTTAAACA
 TTAGAAGCCTGTCTTACAACAGGAAAAACAACCTTATAAGCATAAGACGGACTACGGCCATGCCCGCGT
 GACCGTAAAAAACTGGTCACCGTGATTAAAAAGCACCACCGACAGCTCCTCGGTGATGTCGGGAGTCAT
 AATGTAAGACTCGGTAAACACATCAGGTTGATTACATCGGTGCTAAGGAGCGACCGAAATAGCCC
 GGGGAATACATACCCGAGGCGTAGAGACAACATTACAGCCCCCATAGGAGGTATAACAAATTAATAG
 GAGAGAAAAACATATAACACCTGAAAAACCTCCTGCTAGGCAAAATAGCACCTTCCCGCTCCAGAAC
 AACATACAGCGCTTCCACAGCGGCAGCCATACAGTCAAGCTTACCAGTAAAAAAGGAAACCTATTAAAA
 AAACACCACTCGACACGGCACAGCTCAATCAGTCACAGTGTAAAAAAGGGCCAAGTGCAGAGCGAGTAT
 ATATAGGACTAAAAATGACGTAACGGTTAAAGTCCACAAAAACACCCAGAAAACCGCACGCGAACCTA
 CGCCAGAAAACGAAAGCCAAAAACCCACAACCTTCTCAAATCGTCACTTCCGTTTTCACGTTTACGTC
 ACTTCCCATTTTAAAGAAAACATAATTCCCAACACATACAAGTTACTCCGCCCTAAACCTACGTCACCC
 GCCCGGTTCCACGCCCCGCGCCAGCTCACAACTCCACCCCTCATATCATATTGGCTTCAATCCAAA
 ATAAGGTATATTATTGATGATGTTAATTAAATCCGATCGGATATCGAGCTCTCCCGGGAATTCCG
 GATCTGCGACGCGAGGCTGGATGGCTTCCCTTATGATTCTTCTCGCTTCCGGCGGCATCGGGATGCC
 CGCGTTGCAGGCCATGCTGTCCAGGCAGGTAGATGACGACCATCAGGACAGCTTACGGCCAGCAAAAG
 GCCAGGAACCGTAAAAAGGCCGCTGCTGGCGTTTTTCCATAGGCTCCGCCCTGACGAGCATCACA

Table 10 (continued) Nucleotide sequence of pAd/PL-DEST™.

AAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGG
AAGCTCCCTCGTGCGCTCTCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCG
GGAAGCGTGGCGCTTTCTCAATGCTCACGCTGTAGGTATCTCAGTTCCGGTGTAGGTGCTTCGCTCCAAGC
TGGGCTGTGTGCACGAACCCCCCGTTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTC
CAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTAT
GTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTA
TCTGCGCTCTGCTGAAGCCAGTTACCTTCGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCAC
CGCTGGTAGCGGTGGTTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGAT
CCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGAACGAAAACTCACGTTAAGGGATTTTGGTCATGA
GATTATCAAAAAGGATCTTACCTAGATCCTTTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCT
GACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTTCGTTTCATCCATAGTTG
CCTGACTCCCCGTCGTGTAGATAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGAT
ACCGCGAGACCCACGCTCACCGGCTCCAGATTTATCAGCAATAAACAGCCAGCCGGAAGGGCCGAGCGC
AGAAGTGGTCTCGCAACTTTATCCGCCCTCCATCCAGTCTATTAATTGTTGCCGGAAGCTAGAGTAAGTA
GTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGNTGCAGGCATCGTGGTGTACGCTCGTCGTT
TGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAA
AAAGCGGTTAGCTCCTTCGGTCTCCGATCGTTGTGAGAAGTAAGTTGGCCGAGTGTTATCACTCATGG
TTATGGCAGCACTGCATAATTCTCTTACTGTTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTA
CTCAACCAAGTCATTCTGAGAATAGTGATGCGGCGACCGAGTTGCTCTTGCCCCGCGTCAACACGGGAT
AATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTTGGAACGTTCTTCGGGGCGAAAACTCT
CAAGGATCTTACCGCTGTTGAGATCCAGTTTCGATGTAACCACTCGTGCACCCAACTGATCTTCAGCATC
TTTTACTTTTACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAGGGGAATAAGG
GCGACACGGAAATGTTGAATACTCATACTCTTCTTTTCAATATTATTGAAGCATTTATCAGGGTTATT
GTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATAACAAATAGGGGTTCCGCGCACATTTCC
CCGAAAAGTGCCACCTGACGTCTAAGAAACATTATTATCATGACATTAACCTATAAAAATAGGCGTATC
ACGAGGCCCTTTCGTCTTCAAGGATCCGAATTCCTGGGAGAGCTCGATATCGCATGCGGATTTAAATTAA
TTAA

Table 11 Nucleotide sequence of pAd/CMV/V5-GW/lacZ.PL-DEST™.

CATCATCAATAATATACCTTATTTTGGATTGAAGCCAATATGATAATGAGGGGGTGGAGTTTGTGACGTG
 GCGCGGGGCGTGGGAACGGGGCGGGTGACGTAGTAGTGTGGCGGAAGTGTGATGTTGCAAGTGTGGCGGA
 ACACATGTAAGCGACGGATGTGGCAAAAGTGACGTTTGTGGTGTGCGCCGGTGTACACAGGAAGTGACAA
 TTTTCGCGCGGTTTATAGGCGGATGTTGTAGTAAATTTGGGCGTAACCGAGTAAGATTTGGCCATTTTCGC
 GGGAAAACCTGAATAAGAGGAAGTGAAATCTGAATAATTTGTGTTACTCATAGCGCGTAATATTTGTCTA
 GGGCCGCGGGGACTTTGACCGTTTACGTGGAGACTCGCCAGGTGTTTTCTCAGGTGTTTTCCGCGTTC
 CGGGTCAAAGTTGGCGTTTTTATTATTATAGTCAGTCGAAGCTTGGATCCGGTACCTCTAGAATTTCTCGAG
 CGGCCGCTAGCGACATCGGATCTCCCGATCCCTATGGTTCGACTCTCAGTACAATCTGCTCTGATGCCGC
 ATAGTTAAGCCAGTATCTGCTCCCTGCTTGTGTGTTGGAGGTGCTGAGTAGTGCAGCGAGCAAAATTTAA
 GCTACAACAAGGCAAGGCTTGACCGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGC
 TTCGCGATGTACGGGCGAGATATACGCGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTAC
 GGGGTCAATTAGTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCCGCTGGC
 TGACCGCCCAACGACCCCGCCCATTTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGA
 CTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTATCA
 TATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCTGGCATTATGCCCAGTACATG
 ACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGTATGCGGT
 TTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGAATCAGCGGATTTCCAAGTCTCCACCCCATTTGA
 CGTCAATGGGAGTTTGTGTTTGGCACCAGCAATCAACGGGACTTTCCAAAATGTGTAACAACTCCGCCCA
 TTGACGCAATGGGCGGTAGGCGGTGACGGTGGGAGGTCTATATAAGCAGAGCTCTCTGGCTAACTAGAG
 AACCCACTGCTTACTGGCTTATCGAAATTAATCAGCTCACTATAGGGAGACCCAAGCTGGCTAGTTAAG
 CTATCAACAAGTTTGTACAAAAAAGCAGGCTCCGCGGCCGCCCTTACCATGATAGATCCCGTCTGTTT
 TACAACGTGCTGACTGGGAAAACCTGGCGTTACCCAATTAATCGCTTGCAGCACATCCCCCTTTTCGC
 CAGCTGGCGTAATAGCGAAGAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAA
 TGGCGCTTTGCTGTTTCCGGCACCAGAGCGGTGCCGAAAGCTGGCTGGAGTGCATCTTCTCTGAGG
 CCGATACTGTCTGCTCCCTCAAACCTGGCAGATGACCGGTTACGATGCGCCCATCTACACCAACGTAAC
 CTATCCCATTACGGTCAATCCGCCGTTTGTTCACCGGAGAATCCGACGGGTGTTACTCGCTCACATTT
 AATGTTGATGAAAGCTGGCTACAGGAAGGCCAGACGCGAATTATTTTGTATGGCGTTAACTCGGCGTTT
 ATCTGTGGTGAACGGCGCTGGGTGCGTTACGGCAGGACAGTCGTTTGCCGCTCTGAATTTGACCTGAG
 CGCATTTTTTACCGCCCGGAGAAAACCGCCTCGCGGTGATGGTGTGCGTTGGAGTGACGGCAGTTATCTG
 GAAGATCAGGATATGTGGCGGATGAGCGGCATTTTCCGTGACGTCTGTTGCTGCATAAACCGACTACAC
 AAATCAGCGATTTCCATGTTGCCACTCGCTTTAATGATGATTTTACGCCGCTGTACTGGAGGCTGAAGT
 TCAGATGTGCGCGAGTTGCGTGACTACCTACGGGTAAACAGTTTCTTTATGGCAGGGTGAAACCGAGGTC
 GCCAGCGGCACCGCGCCTTTCCGGCGGTGAAATTATCGATGAGCGTGGTGGTTATGCCGATCGCGTCACAC
 TACGTCTGAACGTGCAAAACCCGAACTGTGGAGCGCGGAAATCCCGAATCTCTATCGTGGCGTGGTTGA
 ACTGCACACCGCCGACGCTGATTGAAGCAGAACGCTGCGATGTGCGTTTCCGCGAGGTGCGGATT
 GAAAAATGGTCTGCTGCTGCTGAACGCAAGCGTTGCTGATTGAGGCGTTAACCGTCACGAGCATCATC
 CTCTGCATGGTCAAGTCATGGATGAGCAGACGATGGTGCAGGATATCTGCTGATGAAGCAGAACAACTT
 TAACGCCGTGCGCTGTTTCGATTATCCGAACCATCCGCTGTGTTGATACGCTGTGCGACCGCTACGGCCTG
 TATGTGGTGGATGAAGCCAATATTGAAACCCACGGCATGGTGCCAATGAATCGTCTGACCGATGATCCGC
 GCTGGCTACCGGCGATGAGCGAACCGGTAACGCGAATGGTGCAGCGGATCGTAATCACCCGAGTGTGAT
 CATCTGGTCTGTTGGGAATGAATCAGGCCACGGCGTAATCAGCAGCGCTGTATCGCTGGATCAAACTCT
 GTCGATCTTTCCGCCCGGTGCGATGATGAAGCGCGGAGCCGACACCGCCACCGATATTATTTGCC
 CGATGTACGCGCGGTGGATGAAGACCAGCCCTTCCCGCTGTGCCGAAATGGTCCATCAAAAAATGGCT
 TTCGCTACCTGGAGAGACGCGCCCGCTGATCTTTTGGCAATACGCCACGCGATGGGTAACAGTCTTGGC
 GGTTCGCTAAATACTGGCAGGCGTTTCGTGATATCCCGTTTACAGGGCGGCTTCGTCTGGGACTGGG
 TGGATCAGTCGCTGATTAAATATGATGAAAACGGCAACCCGTGGTTCGCTTACGGCGGTGATTTTGGCGA
 TACGCCGAACGATCGCCAGTTCGTATGAACGGTCTGGTCTTTGCCGACCGCACGCCGATCCAGCGCTG
 ACGGAAGCAAAACACAGCAGCAGTTTTCAGTTCGTTTATCCGGGCAAACCATCGAAGTGACCCAGCG
 AATACCTGTTCCGTCATAGCGATAACGAGCTCCTGCACTGGATGGTGGCGCTGGATGGTAAGCCGCTGGC
 AAGCGGTGAAGTGCTCTGGATGTGCTCCACAAGGTAAACAGTTGATTGAACTGCCTGAACTACCGCAG
 CCGGAGAGCGCCGGCAACTCTGGCTCACAGTACGCGTAGTGCAACCGAACGCGACCGCATGGTCAGAAG
 CCGGGCACATCAGCGCTGGCAGCAGTGGCGTCTGGCGGAAACCTCAGTGTGACGCTCCCGCGCGCTC
 CCACGCCATCCCGCATCTGACCACCAGCGAAATGGATTTTGCATCGAGCTGGGTAATAAGCGTTGGCAA
 TTTAACCGCCAGTCAGGCTTTCTTTTACAGATGTGGATTGGCGATAAAAAACAAGTCTGACGCCGCTGC
 GCGATCAGTTTACCCGTGCACCGCTGGATAACGACATTGGCGTAAGTGAAGCGACCCGATTGACCTAA
 CGCTCGGTCGAACGCTGGAAGGCGCGGCCATTACAGGCCGAAGCAGCGTTGTTGCAAGTGACCGGCA
 GATACACTTGTGATGCGGTGCTGATTACGACCGCTCACGCGTGGCAGCATCAGGGGAAAACCTTATTTA
 TCAGCCGGAACCTACCGGATTGATGGTAGTGGTCAAATGGCGATTACCGTTGATGTTGAAGTGGCGAG
 CGATACACCGCATCCGGCGCGGATTGGCCTGAACTGCCAGCTGGCGCAGGTAGCAGAGCGGGTAAACTGG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/lacZ.PL-DEST™.

CTCGGATTAGGGCCGCAAGAAAACATATCCCGACCGCCTTACTGCCGCCTGTTTTGACCGCTGGGATCTGC
CATTGTGACACATGTATACCCCGTACGTCCTTCCCGAGCGAAAACGGTCTGCGCTGCGGGACGCGCAATT
GAATTATGGCCACACCACTGGCGCGGCGACTTCCAGTTCAACATCAGCCGCTACAGTCAACAGCAACTG
ATGGAACACAGCCATCGCCATCTGCTGCACGCGGAAGAAGGCACATGGTGAATATCGACGGTTTCCATA
TGGGGATTGGTGGCGACGACTCCTGGAGCCCGTCAGTATCGGCGGAGTTCCAGCTGAGCGCCGGTTCGCTA
CCATTACCAGTTGGTCTGGTGTCAAAAACTAAGGGTGGGCGCGCCGACCCAGCTTTCTTGTAACAAAGTG
GTTGATCTAGAGGGCCCGCGGTTTCAAGGTAAGCCTATCCCTAACCTCTCCTCGGTCTCGATTCTACGC
GTACCGGTTAGTAATGAGTTTAAACGGGGGAGGCTAAGTGAACACGGAAGGAGACAATACCGGAAGGAA
CCCGCGCTATGACGGCAATAAAAAGACAGAATAAAACGCGACGGGTGTTGGGTGCTTTGTTTATAAACGCG
GGGTTCCGGTCCAGGGCTGGCACTCTGTGATACCCACCGAGACCCCATTTGGGGCCAATACGCGCGCGT
TTCTTCTTTTCCCCACCCACCCCAAGTTCCGGTGAAGGCCAGGGCTCGCAGCCAACGTGCGGGCG
GCAGGCCCTGCCATAGCAGATCCGATTGACAGATCACTGAAATGTGTGGCGGTGGCTTAAGGGTGGGAA
AGAATATATAAGGTGGGGGTCTTATGTAGTTTGTATCTGTTTTGCAGCAGCCGCGCGCCATGAGCAC
CAACTCGTTTGATGGAAGCATTGTGAGCTCATATTTGACAACGCGCATGCCCCATGGGCGGGGTGCGT
CAGAATGTGATGGGCTCCAGCATTGATGGTCCGCCGCTCTGCCCGAACTCTACTACCTTGACCTACG
AGACCGTGTCTGGAACCGCGTTGGAGACTGCAGCCTCCGCGCGGCTTACGCGCTGCAGCCACCGCCG
CGGGATTGTGACTGACTTTGCTTTCTGAGCCCGCTTGCAAGCAGTGCAGCTTCCCGTTTCATCCGCGCG
GATGACAAGTTGACGGCTCTTTTGGCACAATTGGATTCTTTGACCCGGGAACCTAATGTGCTTTCTCAGC
AGCTGTTGGATCTGCGCCAGCAGGTTTCTGCCCTGAAGGCTTCTCCCTCCCAATGCGGTTTAAAAACAT
AAATAAAAAACAGACTCTGTTTGGATTGGATCAAGCAAGTGTCTTGCTGTCTTTATTTAGGGGTTTTG
CGCGCGCGGTAGGCCCGGGACCAGCGGTCTCGGTGCTTGAGGGTCTGTGTATTTTTCCAGGACGTGGT
AAAGGTGACTCTGGATGTTTTCAGATACATGGGCATAAGCCCGTCTCTGGGGTGGAGGTAGCACCCTGAG
AGCTTCATGCTGCGGGGTGGTGTGTGTAGATGATCCAGTCTGAGCAGGAGCGCTGGGCGTGGTGCCTAAAA
ATGTCTTTTTCAGTAGCAAGCTGATTGCCAGGGGCAGGCCCTTGGTGTAAAGTGTTTTCAAAGCGGTTAAGCT
GGGATGGGTGCATACGTGGGATATGAGATGCATCTTGGACTGTATTTTTAGGTTGGCTATGTTCCAGC
CATATCCCTCCGGGGATTTCATGTTGTGTCAGAACACCAGCAGTGTATCCGGTGCACCTGGGAAATTTG
TCATGTAGCTTAGAAGGAAATGCGTGGAAAGAACTTGGAGACGCCCTTGTGACCTCCAAGATTTTCCATGC
ATTCGTCCATAATGATGGCAATGGGCCACGGGCGCGGCTGGGCGAAGATATTTCTGGGATCACTAAC
GTCATAGTTGTGTTCCAGGATGAGATCGTCATAGGCCATTTTACAAAGCGCGGGCGAGGTTGCCAGAC
TGCGGTGCTGCTGCGGGGTGGT
GTTTCAGATGGGGGATCATGTCTACCTGCGGGGCGATGAAGAAAACGGTTTCCGGGTAGGGGAGATCAG
CTGGGAAGAAAGCAGGTTCTTGAGCAGCTGCGACTTACCGCAGCCGGTGGGCGGTAATCACACCTATT
ACCGGGTGCAACTGGTAGTTAAGAGAGCTGCAGCTGCCGTATCCCTGAGCAGGGGGGCCACTTCGTAA
GCATGTCCCTGACTCGCATGTTTTCCCTGACCAAATCCGCCAGAAGGCGCTCGCCGCCAGCGATAGCAG
TTCTTGAAGGAAGCAAAGTTTTTCAACGGTTTGGAGCCGTCCGCCGTAGGCATGCTTTTGAGCGTTTGA
CCAAGCAGTTCCAGGCGGTCCCACAGCTCGGTACCTGCTCTACGGCATCTCGATCCAGCATCTCTCTC
GTTTTCCGGGTTGGGGCGGCTTTTCGCTGTACGGCAGTAGTCCGTGCTCGTCCAGACGGGCCAGGTCATG
TCTTTCCAGGGGCGCAGGGTCTCGTCCAGCGTAGTCTGGGTACGGTGAAGGGGTGCGCTCCGGGCTGCG
CGCTGGCCAGGGTGCCTTGGGCTGGTCTGCTGCTGCTGAAGCGTGCCTGCTTCCGCTGCGCGT
GGCCAGGTAGCATTGACCATGGTGTCTAGTCCAGCCCCCTCCGCGCGTGGCCCTTGGCGCGCAGCTTG
CCCTTGAGGAGGGCGCCACAGAGGGGCGAGTGCAGACTTTTGAGGGCGTAGAGCTTGGGCGCGAGAAATA
CCGATTCCGGGGAGTAGGCATCCGCGCCGAGGCCCGCAGACGGTCTCGCATTCACAGCCAGGTGAG
CTCTGGCCGTTCCGGGTCAAAAACAGGTTTCCCCATGCTTTTGTATGCGTTTCTTACCTCTGGTTTCC
ATGAGCCGTTGTCACGCTCGGTGACGAAAAGGCTGTCCGTGTCCTCCGTATACAGACTTGAGAGGCTGT
CCTCGAGCGGTGTTCCGCGGTCTCTCTGTATAGAACTCGGACCACTCTGAGACAAAGGCTCGCGTCCA
GGCCAGCACGAAGGAGGCTAAGTGGGAGGGGTAGCGGTGCTTGTCCACTAGGGGGTCCACTCGTCCAGG
GTGTGAAGACACATGTGCCCTCTTCCGCATCAAGGAAGGTGATTGGTTTGTAGGTGTAGGCCACGTGAC
CGGGTGTCTTGAAGGGGGCTATAAAGGGGGTGGGGGCGGCTTCTCTCTACTCTCTTCCGCATCGCT
GTCCTGCGAGGGCCAGCTGTTGGGGTGAAGTACTCCCTCTGAAAAGCGGGCATGACTTCTGCGCTAAGATTG
TCAGTTTCAAAAACGAGGAGGATTTGATATTCACCTGGCCCGCGGTGATGCCCTTGGAGGGTGGCCGAT
CCATCTGGTCAGAAAAGACAATCTTTTTGTGTCAAGCTTGGTGGCAAACGACCCGTAGAGGGCGTTGGA
CAGCAACTTGGCGATGGAGCGCAGGGTTTGGTTTTTGTGCGGATCGGCGCGCTCTTGGCCGCGATGTTT
AGCTGCACGTATTGCGCGCAACGCACCGCCATTCCGGAAAGACGGTGGTGGCTCGTCCGGCACCAGGT
GCACGCGCAACCGCGGTGTGTGAGGGTGACAAGGTCAACGCTGGTGGCTACCTCTCCGCGTAGCGCTC
GTTGGTCCAGCAGAGGGCGGCCCTTGGCGGAGCAGAATGGCGGTAGGGGGTCTAGCTGCGTCTCGTCC
GGGGGGTCTGCGTCCACGGTAAAGACCCCGGCGAGCGCGCGCTCGAAGTAGTCTATCTTGATCTCTT
GCAAGTCTAGCGCTGCTGCCATGCGCGGGCGGCAAGCGCGCGCTCGTATGGGTTGAGTGGGGGACCCCA
TGGCATGGGGTGGGTGACGCGGAGGCGTACATGCCGCAAATGTCTGTAACGTAGAGGGGCTCTCTGAGT
ATTCCAAGATATGTAGGGTAGCATCTTCCACCGCGGATGCTGGCGCGCACGTAATCGTATAGTTCGTGCG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

AGGGAGCGAGGAGGTCGGGACCGAGGTTGCTACGGGCGGGCTGCTCTGCTCGGAAGACTATCTGCCTGAA
 GATGGCATGTGAGTTGGATGATATGTTGGACGCTGGAAGACGTTGAAGCTGGCGTCTGTGAGACCTACC
 GCGTCACGCACGAAGGAGGCGTAGGAGTCGCGCAGCTTGTGACCAGCTCGGCGGTGACCTGCACGTCTA
 GGGCGCAGTAGTCCAGGGTTTCCCTTGATGATGTCACTTATCCTGTCCCTTTTTTTTCCACAGCTCGCG
 GTTGAGGACAACTCTTCGCGGTCTTTCCAGTACTCTTGGATCGGAAACCCGTGCGCCTCCGAACGGTAA
 GAGCCTAGCATGTAGAAGTGGTTGACGGCCTGGTAGGCGCAGCATCCCTTTTCTACGGGTAGCGCGTATG
 CCTGCGCGGCTTCGCGAGCGAGGTGTGGGTGAGCGCAAAGGTGTCCCTGACCATGACTTTGAGGTACTG
 GTATTTGAAGTCAGTGTGTCGTCGCATCCGCCCTGCTCCAGAGCAAAAAGTCCGTGCGCTTTTGGGAACGC
 GGATTTGGCAGGCGAAGGTGACATCGTTGAAGAGTATCTTTCCGCGCGAGGCATAAAGTTGCGTGTGA
 TGCGGAAGGGTCCCGGCACCTCGGAACGGTTGTTAATTACCTGGGCGGCGAGCACGATCTCGTCAAAGCC
 GTTGATGTTGTGGCCACAATGTAAAGTTCCAAGAAGCGCGGATGCCCTTGATGGAAGGCAATTTTTTA
 AGTTCCCTCGTAGGTGAGCTCTTCAGGGGAGCTGAGCCCGTGTCTGAAAGGGCCAGTCTGCAAGATGAG
 GTTGGAAGCGACGAATGAGCTCCACAGGTACGCGGCCATTAGCATTTGACAGGTGGTCCGGAAGGTCCCT
 AAATGGCGACCTATGGCCATTTTTTCTGGGGTGATGAGTAGAAGGTAAGCGGGTCTTGTTCGAGCGG
 TCCCATCCAAGGTTCCGCGGTAGGTCTCGCGCGGCAGTCACTAGAGGCTCATCTCCGCCGAACTTCATGA
 CCAGCATGAAGGCGACGAGCTGCTTCCCAAAGGCCCCCATCAAGTATAGGTCTCTACATCGTAGGTGAC
 AAGAGACGCTCGGTGCGAGGATGCGAGCCGATCGGGAAGAACTGGATCTCCCGCCACCAATTGGAGGAG
 TGGCTATTGATGTGGTGAAGTAGAAGTCCCTGCGACGGGCGGAACACTCGTGCTGGCTTTGTAAAAAC
 GTGCGCAGTACTGCGCAGCGGTGCACGGGCTGTACATCTGACAGAGGTTGACCTGACGACCGCGCACAAAG
 GAAGCAGAGTGGGAATTTGAGCCCCCTGCGCTGGCGGGTTTGGCTGGTGGTCTTCTACTTCGGCTGCTTGT
 CCTTGACCGTCTGGCTGCTCGAGGGGAGTTACGGTGGATCGGACCACCAAGCGCGCGAGCCCAAAGTCC
 AGATGTCGCGCGCGCGCGGTGCGAGCTTGATGACAAACATCGCGCAGATGGGAGCTGTCCATCGTCTGGAG
 CTCCCGCGGCGTCAGGTACAGGCGGGAGCTCCTGCAGGTTTACCTCGCATAGACGGGTGAGGCGCGGGCT
 AGATCCAGGTGATACCTAATTTCCAGGGGCTGGTTGGTGGCGGCTCGATGGCTTGCAAGAGGCCGCATC
 CCCGCGGCGCGACTACGGTACCGCGCGCGCGGGCGGTGGGCCGCGGGGTGTCTTGGATGATGCATCTAA
 AAGCGGTGACGCGGCGCGAGCCCCCGAGGTAGGGGGGGCTCCGGACCCCGCGGGAGAGGGGGCAGGGGCA
 CGTCGGCGCGCGCGCGCGGGCAGGAGCTGGTGTGCGCGCGTAGGTTGCTGGCGAACGCGACGACGCGGCG
 GTTGATCTCTGAATCTGGCGCCTCTGCGTGAAGACGACGCGGCCGCTGAGCTTGAGCCTGAAAGAGAGT
 TCGACAGAATCAATTTTCGGTGTCTGACGCGCGCTGCGCGCAAAATCTCCTGCACGTCTCCTGAGTTGT
 CTTGATAGGCGATCTCGGCCATGAAGTCTGCTGATCTCTTCTCTGAGATCTCCGCGTCCGGCTCGCTC
 CACGGTGGCGGCGAGGTGCTTGGAAATGCGGGCCATGAGCTGCGAGAAGGCGTTGAGGCCTCCCTCGTTC
 CAGACGCGGCTGTAGACCACGCCCTTCGGCATCGCGGCGCGCATGACCACCTGCGCGAGATTGAGCT
 CCACGTGCCGGGCGAAGACGGCGTAGTTTCGCGAGGCGTGAAAGAGGTAGTTGAGGGTGGTGGCGGTGTG
 TTCTGCCACGAAGAAGTACATAACCCAGCGTCGCAACGTGGATTCTGTTGATATCCCCAAGGCCTCAAGG
 CGCTCCATGGCCTCGTAGAAGTCCACGGCGAAGTTGAAAACTGGGAGTTGCGCGCCGACACGGTTAACT
 CTTCTCCAGAAGACGGATGAGCTCGGCGACAGTGTGCGCGACCTCGCGCTCAAAGGCTACAGGGGCTC
 TTCTTCTTCTCAATCTCCTCTTCCATAAGGGCCTCCCTTCTTCTTCTTGGCGGCGGTGGGGGAGGG
 GGGACACGGCGGCGACGACGGCGCACCGGGAGGCGGTGCAAAAGCGCTCGATCATCTCCCGCGGCGAC
 GCGCATGTTCTCGGTGACGGCGCGGCGGCTTCTCGCGGGGCGCAGTTGGAAGACGCCGCCCGTCTATGTC
 CCGGTTATGGGTTGGCGGGGGGCTGCCATGCGGCAGGGATACGGCGCTAACGATGCATCTCAACAATTGT
 TGTGTAGGTACTCCGCCGCGAGGGACCTGAGCGAGTCCGCATCGACCGGATCGGAAAACCTCTCGAGAA
 AGGCGTCTAACCAGTCACAGTCGCAAGGTAGGCTGAGCACCGTGGCGGGCGGCGAGCGGGCGGCGGTGCGG
 GTTGTCTTGGCGGAGGTGCTGCTGATGATGAATTAAGTAGGCGGTCTTGAGACGGCGGATGGTTCGAC
 AGAAGCACCATGTCCTTGGGTCCGGCTGCTGAATGCGCAGGCGGTGCGCCATGCCCCAGGCTTCGTTTT
 GACATCGGCGCAGGTCTTTGTAGTAGTCTTGATGAGCCTTTCTACCGGCACTTCTTCTTCTCTCTCTC
 TTGTCTGTCATCTTGTGATCTATCGCTGCGGCGGCGGCGAGTTTGGCCGTAGGTGGCGCCCTCTTCTCT
 CCCATGCGTGTGACCCCGAAGCCCTCATCGGCTGAAGCAGGGCTAGGTGCGCGACAACGCGCTCGGCTA
 ATATGGCCTGCTGCACCTGCGTGAGGGTAGACTGGAAGTCATCCATGTCCACAAAGCGGTGGTATGCGCC
 CGTGTGATGGTGTAGTGCAGTTGGCCATAACCGGACAGTTAACGGTCTGGTGACCCGGTGGCGAGAGC
 TCGGTGTACCTGAGACGCGAGTAAGCCCTCGAGTCAAATACGTAGTCGTTGCAAGTCCGCACCAAGGTACT
 GGTATCCACCAAAAAGTGGCGGCGGCTGGCGGTAGAGGGGCCAGCGTAGGGTGGCGGGGCTCCGGG
 GGCGAGATCTTCCAACATAAGGCGATGATATCCGTAGATGTACCTGGACATCCAGGTGATGCCGCGGCG
 GTGGTGGAGGCGCGCGAAAGTCCGCGACGCGGTTCCAGATGTTGCGCAGCGGCAAAAAGTCTCCATGG
 TCGGGACGCTCTGGCCGGTCAAGCGCGCAATCGTTGACGCTCTAGACCGTGCAAAAGGAGAGCCTGTA
 AGCGGGCACTCTTCGTGGTCTGGTGGATAAATCGCAAGGGTATCATGGCGGACGACCGGGGTTTCGAGC
 CCCGATCCGGCCGTCCGCGGTGATCCATGCGGTTACCGCCCGGTGTCGAACCCAGGTGTGCGACGTC
 GACAACGGGGAGTGCTCTTTTGGCTTCTTCCAGCGCGGCGGCTGCTGCGCTAGCTTTTGGCCAC
 TGGCCGCGCGCAGCGTAAGCGGTTAGGCTGGAAGCGAAAGCATTAAGTGGCTCGCTCCCTGTAGCCGGA
 GGGTTATTTTCAAGGGTTGAGTCGCGGGACCCCGGTTGAGTCTCGGACCGGCGGACTGCGGCGAAG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

GGGGGTTTGCCTCCCCGTCATGCAAGACCCCGCTTGCAAATTCTCCGAAACAGGGACGAGCCCTTTT
 TTGCTTTTCCAGATGCATCCGGTGTGCGGCAGATGCGCCCCCTCTCAGCAGCGGCAAGAGCAAGAG
 CAGCGGCAGACATGCAGGGCACCTCCCCCTCTCTACCGCGTCAGGAGGGGCGACATCCGCGGTGACG
 CGGCAGCAGATGGTGTATTACGAACCCCGCGCGCCGGGCCCCGCACTACCTGGACTTGGAGGAGGGCGA
 GGGCCTGGCGCGGCTAGGAGCGCCCTCTCTGAGCGGTACCCAAGGGTGCAGCTGAAGCGTGATACGCGT
 GAGGCGTACGTGCCGCGGCAGAACCTGTTTCGCGACCGCGAGGGAGAGGAGCCGAGGAGATGCGGGATC
 GAAAGTTCCACGCAGGGCGCGAGCTGCGGCATGGCCTGAATCGCGAGCGGTTGCTGCGCGAGGAGACTT
 TGAGCCCGACGCGCGAACCCGGGATTAGTCCCGCGCGCACACGTTGGCGGGCCCGACCTGGTAACCGCA
 TACGAGCAGACGGTGAACAGGAGATTAACTTTCAAAAAAGCTTTAACAACACGTCGCTACGCTTGTGG
 CGCGCGAGGAGGTGGCTATAGGACTGATGCATCTGTGGGACTTTGTAAGCGCGCTGGAGCAAAACCCAAA
 TAGCAAGCCGCTCATGGCGCAGCTGTTCCCTTATAGTGCAGCACAGCAGGGACAACGAGGCATTACAGGGAT
 GCGCTGCTAAACATAGTAGAGCCCCGAGGGCCGCTGGCTGCTCGATTTGATAAACATCCTGCAGAGCATAG
 TGGTGCAGGAGCGCAGCTTGAGCCTGGCTGACAAGGTGGCCGCCATCAACTATTCCATGCTTAGCCTGGG
 CAAGTTTACGCCCCGCAAGATATACCATACCCCTTACGTTCCCATAGACAAGGAGGTAAAGATCGAGGGG
 TTCTACATGCGCATGGCGCTGAAGGTGCTTACCTTGAGCGACGACCTGGGCGTTTATCGCAACGAGCGCA
 TCCACAAGGCCGTGAGCGTGAGCCGGCGCGCGAGCTCAGCGACCGCGAGCTGATGCACAGCCTGCAAG
 GGCCCTGGCTGGCACGGGCAGCGCGATAGAGAGGCCGAGTCTACTTTGACGCGGGCGCTGACCTGCGC
 TGGGCCCCAAGCCGACGCGCCCTGGAGGCAGCTGGGGCCGGACCTGGGCTGGCGGTGGCACCCGCGCGCG
 CTGGCAACGTCGGCGCGCTGGAGGAATATGACGAGGACGATGAGTACGAGCCAGAGGACGGCGAGTACTA
 AGCGGTGATGTTTCTGATCAGATGATGCAAGACGCAACCGACCCGGCGGTGCGGGCGGCGCTGCAGAGCC
 AGCCGTCCGGCCTTAACCTCCACGGACGACTGGCGCCAGGTATGACCGCATCATGTGCTGACTGCGCGG
 CAATCCTGACGCGTTCCGGCAGCAGCCGACGCGCAACCGGCTCTCCGCAATTCTGGAAGCGGTGTTCCCG
 GCGCGCCCAAAACCCACGCGAGAGAGGTGCTGGCGATCGTAAACGCGCTGGCCGAAAAACAGGGCCATCC
 GGCCCGACGAGGCCGCGCTGGTCTACGACGCGCTGCTTACGCGCGTGGCTCGTTACAACAGCGGCAACGT
 GCAGACCAACCTGGACCGGCTGGTGGGGGATGTGCGCGAGGCCGTGGCGCAGCGTGAGCGCGCGCAGCAG
 CAGGGCAACCTGGGCTCCATGGTTGCACTAAACGCTTCTCTGAGTACACAGCCCGCAACGTGCCGCGGG
 GACAGGAGGACTACACCAACTTTGTGAGCGCACTGCGGCTAATGGTGACTGAGACACCGCAAAGTGAGGT
 GTACCACTCTGGGCCAGACTATTTTTTCCAGACAGTAGACAAGGCTGCGAGACCGTAACACCTGAGCCAG
 GCTTTTCAAAAACCTTGCGGGCTGTGGGGGTGCGGGCTCCACAGGCGACCGCGCGGAGCTGTCTAGCT
 TGCTGACGCCCCAATCGCGCCTGTGTGCTGCTAATAGCGCCCTTACGGACAGTGGCAGCGTGTCCCG
 GGACACATACCTAGGTCACTTGCTGACACTGTACCGCGAGGCCATAGGTACAGGCGCATGTGGACGAGCAT
 ACTTTCCAGGAGATTACAAGTGTGACCGCGCGCTGGGGCAGGAGACACGGGCAGCCTGGAGGCAACCC
 TAACTACCTGCTGACCAACCGCGCGCAGAAAGATCCCCCTCGTTGCACAGTTTAAACAGCGAGGAGGAGCG
 CATTTCGCGCTACGTGCAGCAGAGCGTGAGCCTTAACCTGATGCGCGACGGGGTAACGCCACGCGTGGCG
 CTGGACATGACCGCGCGCAACATGGAACCGGCATGTATGCTCAAACCGGCCGTTTATCAACCGCCCTAA
 TGGACTACTTTCATGCGCGCGCGCCGCTGAACCCGAGTATTTACCAATGCCATCTTGAACCCGCACTG
 GCTACCGCCCCCTGGTTTCTACACCGGGGATTGAGGTGCCCGAGGGTAACGATGGATTCTCTGGGAC
 GACATAGACGACAGCGTGTTTTCCCGCAACCGCAGACCTGCTAGAGTTGCAACAGCGCGAGCAGGCAG
 AGGGCGCGCTGCGAAAGGAAAGCTTCCGCGAGGCCAAGCAGCTTGTCCGATCTAGGCGCTGCGGCCCCGCG
 GTCAGATGCTAGTAGCCCATTTCCAAGCTTGATAGGGTCTCTTACCAGCACTCGCACCAACCCGCCCCGCG
 CTGCTGGGCGAGGAGGAGTACCTAAACAACCTCGCTGCTGCAGCCGACGCGCAAAAAACCTGCCCTCCGG
 CATTTCCCAACAACGGGATAGAGAGCCTAGTGGACAAGATGAGTAGATGGAAGACGTACGCGCAGGAGCA
 CAGGGACGTGCCAGGCCGCGCCCCGCCACCCGCTCGTCAAAGGCACGACCGTACGCGGGGTCTGGTGTGG
 GAGGACGATGACTCGGCAGACGACAGCAGCGTCCTGGATTGAGGAGGAGTGGCAACCCGTTTGGCGACC
 TTCGCCCCAGGCTGGGGAGAATGTTTTAAAAAAGCATGATGCAAAATAAAAACTCACCAAGGC
 CATGGCACCGAGCGTTGGTTTTCTTGATTTCCCTTAGTATGCGGCGCGCGCGATGTATGAGGAAGGTC
 CTCCTCCCTCTACGAGAGTGTGGTGAGCGCGCGCCAGTGGCGGCGCGCTGGGTTCTCCCTTCGATGC
 TCCCCGGACCCGCGCTTTGTGCTCCGCGTACCTGCGGCTACCGGGGGAGAAACAGCATCCGTTAC
 TCTGAGTTGGCACCCCTATTTCGACACACCCGTGTGTACCTGGTGGACAACAAGTCAACGGATGTGGCAT
 CCCTGCACTACAGAACGACACAGCAACTTTTGACACCGTCAATTCAAACAATGACTACAGCCCGGG
 GGAGGCAAGCACACAGACCATCAATCTTGACGACCGGTGCACTGGGGCGGCGACCTGAAAACCATCCTG
 CATACCAACATGCCAAATGTGAACGAGTTTATGTTTACCAATAAGTTTAAAGCGCGGGTGTATGGTGTGCG
 GCTTGCTTACTAAGGACAATCAGGTGGAGCTGAAATACGAGTGGGTGGAGTTTACGCTGCCCCGAGGGCAA
 CTACTCCGAGACCATGACCATAGACCTTATGAACAACCGCATCGTGGAGCACTACTTGAAAGTGGGCGA
 CAGAACGGGTTCTGGAAAGCGACATCGGGGTAAAGTTTGACACCCGCAACTTCAGACTGGGGTTTGACC
 CCGTCACTGGTCTTGTATGCTGCTGGGTATATACAAACGAAGCCTTCCATCCAGACATCATTTTGTGCGC
 AGGATGCGGGGTGGACTTACCCACAGCGCGCTGAGCAACTTGTGGGCATCCGCAAGCGGCAACCCCTC
 CAGGAGGGCTTTAGGATCACCTACGATGATCTGGAGGGTGGTAACATTCCCGCACTGTTGGATGTGGACG
 CCTACCAGGCGAGCTTGAAAGATGACACCGAACAGGGCGGGGTGGCGCAGGCGGCAACAGCAGTGG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

CAGCGGCGCGGAAGAGAACTCCAACGCGGCAGCCGCGGCAATGCAGCCGGTGGAGGACATGAACGATCAT
 GCCATTGCGGCGGACACCTTTGCCACACGGGCTGAGGAGAAGCGCGCTGAGGCCGAAGCAGCGGCCGAAG
 CTGCCGCCCCGCTGCGCAACCCGAGGTCGAGAAGCCTCAGAAGAAACCGGTGATCAAACCCCTGACAGA
 GGACAGCAAGAAACGCAGTTACAACCTAATAAGCAATGACAGCACCTTCACCCAGTACCGCAGCTGGTAC
 CTTGCATACAACTACGGCGACCCCTCAGACCGGAATCCGCTCATGGACCTGCTTTGCACTCCTGACGTAA
 CCTGCGGCTCGGAGCAGGTCTACTGGTTCGTTGCCAGACATGATGCAAGACCCCGTGACCTTCCGCTCCAC
 GCGCCAGATCAGCAACTTTCCGGTGGTGGGCGCCGAGCTGTTGCCCGTGCACTCCAAGAGCTTCTACAAC
 GACCAGGCGCTTACTCCCAACTCATCCGCCAGTTTACCTCTCTGACCCACGTGTTCAATCGCTTTCCCG
 AGAACAGATTTTGGCGCGCCCGCAGCCCCCACCCTACACCCTCAGTGAAAACGTTCTCTGCTCTCAC
 AGATCAGGGGACGCTACCGCTGCGCAACAGCATCGGAGGAGTCCAGCGAGTGACCATTACTGACGCCAGA
 CGCCGACCTGCCCTACGTTTACAAGGCCCTGGGCATAGTCTCGCCGCGCGTCTATCGAGCCGCACTT
 TTTGAGCAAGCATGTCCATCCTTATATCGCCAGCAATAACACAGGCTGGGGCCTGCGCTTCCCAAGCAA
 GATGTTTGGCGGGGCCAAGAAGCGCTCCGACCAACACCCAGTGCGCGTGCGCGGGCCTACCGCGCGCCC
 TGGGGCGCGCACAAACGCGGCGCGCACTGGGCGCACACCCGTCGATGACGCCATCGACGCGGTGGTGGAGG
 AGGCGCGCAACTACACGCCACGCGCCACCAGTGTCCACAGTGGACGCGGCCATTAGACCGGTGGTGGC
 CGGAGCCCGCGCTATGCTAAAAATGAAGAGACCGGAGGCGCGTAGCACGTGCGCACCCGCGCCGACCC
 GGCACCTGCCGCCCAACGCGCGCGCGCGGCCCTGCTTAACCGCGCACGTGCGACCGCGCGACGGGCGGCCA
 TGCGGGCGCGCTCGAAGGCTGGCCGCGGGTATTGTCACTGTGCCCCCAGGTCCAGGCGACGAGCGGCCG
 CGCAGCAGCCGCGGCCATTAGTGCTATGACTCAGGCTCGCAGGGGCAACGTGTATTGGGTGCGCGACTCG
 GTTAGCGGCTGCGCGTGCGCGTCCGCGACCCGCCCCCGCGCAACTAGATTGCAAGAAAAAATACTTAG
 ACTCGTACTGTTGTATGTATCCAGCGCGCGCGCGCGCAACGAAGCTATGTCCAAGCGCAAAATCAAGA
 AGAGATGCTCCAGGTATCGCGCCGAGATCTATGGCCCCCGAAGAAGGAAGAGCAGGATTACAAGCCC
 CGAAAGCTAAAGTCGGGTCAAAAAGAAAAAGAAAGATGATGATGATGAACCTTGACGACGAGGTGGAACCTG
 TGCACGCTACCGCGCCAGGCGACGGGTACAGTGGAAAGGTGACGCGTAAACGTTGTTTGGCAGCCCGG
 CACCACCGTAGTCTTTACGCCCGGTGAGCGCTCCACCCGACCTACAAGCGCGTGTATGATGAGGTGTAC
 GCGCAGGAGACCTGCTTGAGCAGGCCAACGAGCGCCTCGGGGAGTTTGCCTACGGAAGCGGCATAAGG
 ACATGCTGCGGTGCGCGTGGACGAGGGCAACCAACACCTAGCCTAAAGCCCGTAACACTGCAGCAGGT
 GCTGCCCGCGCTTGACCGTCCGAAGAAAGCGCGGCTAAAGCGCGAGTCTGGTGACTTGGCACCCACC
 GTGCAGCTGATGGTACCCAAGCGCCAGCGACTGGAAGATGTCTTGAAAAAATGACCGTGAACCTGGGC
 TGGAGCCCGAGCTCGCGCTGCGGCCAATCAAGCAGGTGGCGCGCGGACTGGGCGTGACAGCCGTGGACCT
 TCAGATACCCACTACAGTAGCACCAGTATTGCCACCGCCACAGAGGGCATGGAGACACAAACGTCCCCG
 GTTGCCCTCAGCGGTGGCGGATGCCGCGGTGCAGGCGGTGCTGCGGCCGCGTCCAAGACCTCTACGGAGG
 TGCAAACGGAACCGTGGATGTTTTCGCGTTTACGCCCCCGCGCGCCGCGCGGTTTCAGGAAGTACGGCGC
 CGCCAGCGCGCTACTGCCCCGAATATGCCCTACATCCTTCCATTGCGCCTACCCCCGCTATCGTGGGTAC
 ACCTACCGCCCCAGAAGACGAGCAACTACCCGACGCCGAACCACTGGAACCCGCGCGCGCGCTCGCC
 GTCGCCAGCCCGTGTGCCCCGATTTCGCTGCGCAGGGTGGCTCGCGAAGGAGGACAGGACCTGGTGCT
 GCCAACAGCGCGCTACCAACCCAGCATCGTTTAAAGCCGCTTGTGTTTCTTGAGATATGGCCCTC
 ACCTGCCGCTCCGTTTCCCGGTGCGCGGATTCCGAGGAAGAATGCACCGTAGGAGGGGCATGGCCGGCC
 ACGGCTGACGGGCGGCATGCGTCTGCGCACCAACCGCGCGCGCGCGTGCACCGTGCATGCGCGG
 CGGTATCTGCCCCCTCCTTATTCACCTGATCGCGCGGCGATTGGCGCCGTGCCCGGAATTGCATCCGTG
 GCCTTGACAGGCGCAGAGACACTGATTAAAAACAAGTTGCATGTGAAAAAATCAAAATAAAAGTCTGGAC
 TCTACGCTCGCTTGGTCTGTAACTATTTGTAGAATGGAAGACATCAACTTTGCGTCTCTGGCCCCCG
 GACACGGCTCGCGCCCGTTTATGGGAACTGGCAAGATATCGGCACCAGCAATATGAGCGGTGGCGCCTT
 CAGCTGGGGCTCGCTGTGGAGCGGCATTAAAAATTTTCGTTTCCACCGTTAAGAACTATGGCAGCAAGGCC
 TGGAACAGCAGCACAGGCCAGATGCTGAGGGATAAGTTGAAAGAGCAAAATTTCCAACAAAAGGTGGTAG
 ATGGCTGGCCTCTGGCATTAGCGGGGTGGTGGACCTGGCCAACAGGCAGTGCAAAATAAGATTAAACAG
 TAAGCTTGATCCCCGCCCTCCCGTAGAGGAGCTCCACCGGCCGTGGAGACAGTGTCTCCAGAGGGGCGT
 GCGGAAAAGCGTCCGCGCCCCGACAGGGAAGAACTCTGGTGACGCAATAGACGAGCCTCCCTCGTACG
 AGGAGGCACTAAAGCAAGGCCCTGCCACCACCCGTCCATCGCGCCCATGGCTACCGGAGTGTGGGCCA
 GCACACACCCGTAACGCTGGACCTGCCTCCCCCGCGACACCCAGCAGAAACCTGTGCTGCCAGGCCCG
 ACCGCCGTTGTTGTAACCCGTCCTAGCGCGCGTCCCTGCGCGCGCGCCGCGCAGCGTCCGCTGCTGCG
 GGCCCGTAGCCAGTGGCAACTGGCAAAGCACACTGAACAGCATCGTGGGTCTGGGGGTGCAATCCCTGAA
 GCGCCGACGATGCTTCTGAATAGCTAACGTGTCGTATGTGTGTCATGTATGCGTCCATGTGCGGCCAGA
 GGAGCTGTGAGCCGCGCGCGCGCGCTTTCCAAGATGGCTACCCCTTCGATGATGCGCGAGTGGTCTTA
 CATGCACATCTCGGGCCAGGACGCTCGGAGTACCTGAGCCCCGGGTGGTGCAGTTTGGCCGCGCCACC
 GAGAGTACTTCAGCTGAATAACAAGTTTAGAAACCCACCGTGGCGCCTACGCACGACGTGACCACAG
 ACCGTTCCCGAGCTTTGACGCTGCGGTTTCACTCTGTGGACCGTGAGGATACTGCGTACTCGTACAAGGC
 CGGTTTCAACCTAGCTGTGGTGATAACCGTGTGCTGGACATGGCTTCCACGTACTTTGACATCCCGCGG
 GTGCTGGACAGGGGCCCTACTTTAAGCCCTACTCTGGCACTGCCTACAACGCCCTGGCTCCCAAGGGTG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

CCCCAAATCCTTGGCAATGGGATGAAGCTGCTACTGCTCTTGAATAAACTAGAGAAGAGGACGATGA
 CAACGAAGACGAAGTAGACGAGCAAGCTGAGCAGCAAAAACTCACGTATTTGGGCAGGCGCCTTATTCT
 GGTATAAATATTACAAAGGAGGGTATTCAAATAGGTGTGCAAGGTCAAACACCTAAATATGCCGATAAAA
 CATTTCAACCTGAACCTCAAATAGGAGAATCTCAGTGGTACGAACTGAAATTAATCATGCAGCTGGGAG
 AGTCCTTAAAAAGACTACCCCAATGAAACCATGTTACGGTTTCATATGCAAAACCCACAAATGAAATGGA
 GGGCAAGGCATTTCTGTAAAGCAACAAATGGAAAGCTAGAAAGTCAAGTGGAAATGCAATTTTTCTCAA
 CTACTGAGGCGACCGCAGGCAATGGTGATAACTTGACTCCTAAAGTGGTATTGTACAGTGAAGATGTAGA
 TATAGAAACCCAGACACTCATATTTCTTACATGCCACTATTAAGGAAGGTAACCTACGAGAACTAATG
 GGCCAACAATCTATGCCCAACAGGCCAATTACATTTGCTTTTAGGGACAATTTTATTGGTCTAATGTATT
 ACAACAGCACGGGTAATATGGGTGTTCTGGCGGGCCAAGCATCGCAGTTGAATGCTGTTGTAGATTTGCA
 AGACAGAAACACAGAGCTTTTATACCAGCTTTTGCTTGATTCCATTGGTGATAGAACCAGGTACTTTTCT
 ATGTGGAATCAGGCTGTTGACAGCTATGATCCAGATGTTAGAATTATTGAAATCATGGAACCTGAAGATG
 AACTTCCAAATTAAGTCTTTCCACTGGGAGGTGTGATTAATACAGAGACTCTTACCAGGTAAAACCTAA
 AACAGGTGAGGAAATGGATGGGAAAAAGATGCTACAGAATTTTCAGATAAAAAATGAAATAAGAGTTGGA
 AATAATTTTGCCATGGAAATCAATCTAAATGCCAACCTGTGGAGAAATTTCTGTACTCCAACATAGCGC
 TGTATTTGGCCGACCAAGTAAAGTACAGTCTTCCACGTAAAAATTTCTGATAACCCAAACACCTACGA
 CTACATGAACAGCGAGTGGTGGCTCCCGGGTTAGTGGACTGCTACATTAACCTTGAGACAGCTGGTCC
 CTTGACTATATGGACAACGTCAACCCATTTAACCACCACCGCAATGCTGGCCTGCGCTACCGCTCAATGT
 TGCTGGGCAATGGTGGCTATGTGCCCTTCCACATCCAGGTGCGCTCAGAAGTTCTTTGCCATTAAAACT
 CCTTCTCCTGCCGGGCTCATACACCTACGAGTGAAGTTCAGGAAGGATGTTAACATGGTTCTGCAGAGC
 TCCCTAGGAAATGACCTAAGGGTTGACGGAGCCAGCATTAAGTTTGATAGCATTTGCCCTTTACGCCACCT
 TCTTCCCCATGGCCCAACACCGCCTCCACGCTTGAGGCCATGCTTAGAAACGACACCAACGACCATGCT
 CTTTAACGATATCTCTCCGCCGCAACATGCTTACCTTATACCGCCAACGCTACCAACGCTGCCCCATA
 TCCATCCCCCTCCGCAACTGGGCGCTTTCCGCCGCTGGGCTTCCAGCGCCTTAAGACTAAGGAAACCC
 CATCACTGGGCTCGGCTACGACCTTATTACACCTACTCTGGCTCTATACCTACCTAGATGGAACCTT
 TTACCTCAACCACACCTTTAAGAAGGTGGCCATTACCTTTGACTCTTCTGTGAGCTGGCCTGGCAATGAC
 CGCCTGCTTACCCCCAACGAGTTTGAATTAAGCGCTCAGTTGACGGGGAGGGTTACAACGTTGGCCAGT
 GTAACATGACCAAAGACTGGTTCCTGGTACAAATGCTAGCTAACTACAACATTGGCTACCAGGGCTTCTA
 TATCCCAGAGAGCTACAAGGACCGCATGTACTCTCTTTAGAACTTCCAGCCCATGAGCCGTCAGGTG
 TGAGGATGATACTAAAGACTACCAACAGGTGGGCATCTTACCAACACACAACAACTCTGGATTGTG
 TTGGCTACCTTTGCCCCACCATGCGCGAAGGACAGGCTACCTTGCTAACTTCCCCTATCCGCTTATAGG
 CAAGACCGCAGTTGACAGCATTACCCAGAAAAAGTTTCTTTGCGATCGCACCTTTGGCGCATCCCATT
 TCCAGTAACCTTTATGTCCATGGGCGCACTCACAGACCTGGGCCAAAACCTTCTCTACGCCAATCCGCCC
 ACGCGCTAGACATGACTTTTGAGGTGGATCCCATGGACGAGCCACCTTCTTTATGTTTGTGTTGAAGT
 CTTTGACGTGGTCCGTGTGCACCGCGCCACCGCGCGTCAATCGAAACCGTGTACCTGCGCACGCCCTT
 TCGGCGGCAACGCCACAACATAAAGAAGCAAGCAACATCAACAACAGCTGCCGCCATGGGCTCCAGTGA
 GCAGGAAGCTAAAGCCATTGTCAAAGATCTTGGTTGTGGGCCATATTTTTTGGGCACCTATGACAAGCGC
 TTTCCAGGCTTTGTTTCTCCACACAAGCTCGCCTGCGCCATAGTCAATACGGCCGCTGCGGAGACTGGGG
 GCGTACACTGGATGGCCTTTGCTTGAACCCGCACTCAAAAACATGCTACCTCTTTGAGCCCTTTGGCTT
 TTCTGACAGCGACTCAAGCAGGTTTACCAGTTTGAGTACGAGTCACTCCTGCGCCGTAGCGCCATTGCT
 TCTTCCCCGACCGCTGTATAACGCTGGAAGTCCACCCAAAGCGTACAGGGGCCAACTCGGCCGCT
 GTGGACTATTTCTGCTGCATGTTTCTCCAGCCTTTGCCAACTGGCCCCAACTCCCATGGATCACAACCC
 CACCATGAACCTTATTACCGGGGTACCAACTCCATGCTCAACAGTCCCCAGGTACAGCCACCTTGCCT
 CGCAACCAGGAACAGCTCTACAGCTTCTTGAGCGCCACTCGCCCTACTTCCGACGCCACAGTGCGCAGA
 TTAGGAGCGCCACTTCTTTTTGTCACTTGA AAAACATGTAAAAATAATGTACTAGAGACACTTTCAATAA
 AGGCAATGCTTTTATTTGTACTCTCGGGTGATTATTTACCCCCACCTTGCCGTCTGCGCGTTTAA
 AAATCAAAGGGGTTCTGCCGCGCATCGCTATGCGCCACTGGCAGGGACAGTTGCGATACTGGTGTTTAG
 TGCTCCACTTAAACTCAGGCACAACCATCCGCGCGAGCTCGGTGAAGTTTTCACTCCACAGGCTGCGCAC
 CATCACAACCGGTTTAGCAGGTGCGGCGCGGATATCTGAAGTCGAGTTGGGGCTCCGCCCTGCGCG
 CGCGAGTTGCGATACACAGGTTGACGACTGGAACACTATCAGCGCCGGGTGGTGACGCTGGCCAGCA
 CACTCTTGTGCGGAGATCAGATCCGCTCAGGTCTTCCGCGTTGCTCAGGGCGAACGAGTCAACTTTGG
 TAGCTGCCCTTCCCAAAAAGGGCGCGTGGCCAGGCTTTGAGTTGCACTCGCACCGTAGTGGCATCAAAAGG
 TGACCGTGCCCGGTCTGGGCGTTAGGATACAGCGCTGCATAAAAGCCTTGATCTGCTTAAAAGCCACCT
 GAGCCTTTGCGCCTTACAGAGAAGACATGCCGCAAGACTTGCCGGAAGTATGTTGCGCGACAGGCCG
 GTCGTGACGCGACGACCTTGCCTCGGTGTTGAGATCTGCACCACATTTCCGGCCCCACCGGTTCTTCAG
 ATCTTGGCCTTGCTAGACTGCTCCTTACGCGCGCGCTGCCGCTTTTCGCTCGTCACATCCATTTCAATCA
 CGTCTCTCTTATTTATCATAATGCTTCCGTGTAGACACTTAAGCTCGCCTTCGATCTCAGCGCAGCGGTG
 CAGCCACAACCGCGAGCCCGTGGGCTCGTGATGCTTGTAGGTACCTCTGCAACGACTGCAGGTACGCC
 TGCAGGAATCGCCCCATCATCGTCAAAAGGTCTTGTGCTGGTGAAGGTGAGTGCAGCTGCAACCGCGGTGCT

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

CCTCGTTTCAGCCAGGTCTTGCATACGGCCGCCAGAGCTTCCACTTGGTCAGGCAGTAGTTTGAAGTTCCG
 CTTTAGATCGTTATCCACGTGGTACTTGTCCATCAGCGCGCGCAGCCTCCATGCCCTTCTCCACGCA
 GACACGATCGGCACACTCAGCGGGTTCATCACCGTAATTTCACTTTCCGCTTCGCTGGGCTCTTCTCTT
 CCTCTTGGCTCCGCATACCACGCGCCACTGGGTCTCTTCACTTCCGCTTCGCTGGGCTCTTCTCTT
 TTTGCCATGCTTGTATAGCACCGGTGGGTGCTGAAACCCACCATTGTAGCGCCACATCTTCTCTTTCT
 TCCTCGCTGTCCACGATTACCTCTGGTGTATGGCGGGCGCTCGGGCTTGGGAGAAGGGCGCTTCTTTTCT
 TCTTGGGCGCAATGGCCAAATCCGCCGCCGAGGTGATGGCCGCCGGGCTGGGTGTGCGCGGCACCAGCGC
 GTCTTGTGATGAGTCTTCTCGTCTCGGACTCGATACGCCGCCCTCATCCGCTTTTTTGGGGCGCCCGG
 GGAGGCGGGCGGACGGGGACGGGACGACACGTCTCCATGGTTGGGGGACGTGCGCGCCGACCGCGTC
 CGCGCTCGGGGGTGGTTTTCGCGCTGCTCTTCCCGACTGGCCATTTCCTTCTCCTATAGGCAGAAAAA
 GATCATGGAGTCAGTCGAGAAGAAGGACAGCCTAACCGCCCCCTCTGAGTTCGCCACCACCGCCTCCACC
 GATGCCGCCAACCGCGCTACCACTTCCCGCTCGAGGCACCCCGCTTGAGGAGGAGGAAGTGATTATCG
 AGCAGGACCCAGGTTTTGTAAAGCAAGACGACGAGGACCGCTCAGTACCAACAGAGGATAAAAAGCAAGA
 CCAGGACAACGAGAGGCAAAACGAGGAACAAGTCGGGCGGGGGACGAAAGGATGGCGACTACCTAGAT
 GTGGGAGACAGCTGTGTTGAAGCATCTGCAGCGCCAGTGCGCCATTATCTGCAGCGCTTGAAGAGC
 GCAGCGATGTGCCCTCGCCATAGCGGATGTGACGCTTGCCTACGAACGCCACCTATTCTCACCGCGCT
 ACCCCCCAAACGCCAAGAAAACGGCACATGCGAGCCCAACCGCGCTCAACTTCTACCCCGTATTTGCC
 GTGCCAGAGGTGCTTGCCACCTATCACATCTTTTTTCCAAAACGCAAGATACCCCTATCTGCGGTGCCA
 ACCGCAGCCGAGCGGACAAGCAGCTGGCCTTGCGGCGAGGGCGCTGTCATACCTGATATCGCCTCGCTCAA
 CGAAGTGCCAAAAATCTTTGAGGGTCTTGGACGCGACGAGAAGCGCGCGGCAACGCTCTGCAACAGGAA
 AACAGCGAAAATGAAAGTCACCTGAGGTGTTGGTGGAACGAGGGTGACAACGCGCGCCTAGCCGTAC
 TAAACGCGAGCATCGAGGTCAACCACTTGGCTACCCGGCACCTAACCTACCCCAAGGTGATGAGCAC
 AGTCATGAGTGAGCTGATCGTGCCTGCGGACGCAAACTTGAAGGACCTATGAGACGGCTTCAAGAACAA
 GAGGAGGGCTACCCGAGTTGGCGACGAGCAGCTAGCGCGCTGGCTTCAAACGCGGAGCCTGCGGACT
 TGGAGGAGCGACGCAAACTAATGATGGCCGAGTGTCTGTTACCGTGAGCTTGAGTGCATGCAGCGGT
 CTTTGTGCTGACCCGAGATGCAGCGCAAGCTAGAGGAAACATTGCACTACACCTTTCGACAGGGCTACGTA
 CGCCAGGCTTGAAGATCTCCACGTGGAGCTTGCACCTGGTCTCCTACCTTGAATTTTGCACGAAA
 ACCGCTTGGGCAAAACGTGCTTCATTCCACGCTCAAGGGCGAGGCGCGCGGACTACGTCCGCGACTG
 CGTTTACTTATTTCTATGCTACACCTGGCAGACGGCCATGGGCGTTTGGCAGCAGTGTGTTGAGGAGTGC
 AACCTCAAGGAGTGAAGTCACTCCGGGGCTGTGGACGTGCGCTTACCTTCGCAAAATTTGTACCTGAGG
 CCGTGGCCGCGCACCTGGCGGACATCATTTTCCCGAACGCTGCTTAAACCTTGCAACAGGGTCTGCC
 AGACTTCACAGTCAAAGCATGTTGCAGAACTTTAGGAACCTTATCCTAGAGCGCTCAGGAATCTTGCCC
 GCCACCTGCTGTGCACTTCTTAGCGACTTTGTGCCCATTAAGTACCGCGAATGCCCTCCGCGCTTTGGG
 GCCACTGCTACCTTCTGCAGCTAGCCAACTACCTTGCCTACCACTCTGACATAATGGAAGACGTGAGCGG
 TGACGGTCTACTGGAGTGTCACTGTGCTGCAACCTATGCACCCCGCACCGCTCCCTGGTTTGCAATTG
 CAGCTGCTTAAAGAAAGTCAAATATCGGTACCTTTGAGCTGCAGGGTCCCTCGCCTGACGAAAAGTCCG
 CGGCTCCGGGTTGAAACTCACTCCGGGGCTGTGGACGTGCGCTTACCTTCGCAAAATTTGTACCTGAGG
 CTACCACGCCCCAGAGATTAGGTTCTACGAAGACCAATCCCGCCGCCAAATGCGGAGCTTACCGCTGC
 GTCATTACCCAGGGCCACATTCTTGGCCAATTGCAAGCCATCAACAAAGCCCGCAAGAGTTTCTGCTAC
 GAAAGGGACGGGGGGTTTACTTGGACCCCCAGTCCGGCGAGGAGCTCAACCCAAATCCCCCGCGCGCGCA
 GCCCTATCAGCAGCAGCCGCGGGCCCTTGTCTCCAGGATGGCACCCAAAAGAGCTGCAGCTGCGGCC
 GCCACCCACGGACGAGGAGGAATACTGGGACAGTCAGGCAGAGGAGGTTTTGGACGAGGAGGAGGAGGAC
 ATGATGGAAGACTGGGAGAGCCTAGACGAGGAAGCTTCCGAGGTGCAAGAGGTGTCAGACGAAACACCGT
 CACCTCGGTGCGATTCCCCCTCGCCGGCGCCCCAGAAATCGGCAACCGGTTCCAGCATGGCTACAACCTC
 CGCTCCTCAGGCGCGCGCGCACTGCCCGTTGCGCGACCCAACCGTAGATGGGACACCACTGGAACCAAGG
 GCCGGTAAGTCCAAGCAGCCGCGCGCTTAGCCCAAGAGCAACAACAGCGCCAAGGCTACCGCTCATGGC
 GCGGGCAAGAAGCCATAGTTGCTTGTGCAAGACTGTGGGGCAACATCTCTTCGCCCCGCGCTT
 TCTTCTTACCATCAGCGGTGGCCTTCCCCCGTAACATCTGCAATTACTACCGTCACTCTTACAGCCCA
 TACTGCACCGGCGGACGCGCAGCGGACGCAACAGCAGCGGCCACACAGAAGCAAAGGCGACCGGATAGC
 AAGACTTGAACAAGCCCAAGAAATCCACAGCGGCGGAGCAGCAGGAGGAGGAGCGCTGCGTCTGGCGC
 CCAACGAAACCGATTCGACCCGCGAGCTTAGAAACAGGATTTTCCCACTCTGTATGCTATATTAACA
 GAGCAGGGGCCAAGAACAAGAGCTGAAAATAAAAAACAGGTCTCTGCGATCCCTCACCCGAGCTGCCTG
 TATCAAAAAGCGAAGATCAGCTTCCGCGCACGCTGGAAGACGCGGAGGCTCTCTTCAAGTAAATACTGCG
 CGCTGACTCTTAAGGACTAGTTTTCGCGCCCTTCTCAAAATTAAGCGCGAAACTACGTCACTCTCCAGCG
 GCCACACCCGGCGCCAGCACCTGTGCTCAGCGCCATTATGAGCAAGGAAATTTCCACGCCCCATACATGTGG
 AGTTACAGCCACAAATGGGACTTGGCGCTGGAGCTGCCAAGACTACTCAACCCGAATAAATACTACATGA
 GCGCGGACCCACATGATATCCCGGTCAACGGAATCCGCGCCACCCGAAACCGAATTCTCTTGAACA
 GCGCGCTATTACCAACCACTCGTAATAACCTTAATCCCGTAGTTGGCCCGCTGCGCTGTGTACCAAG
 GAAAGTCCCGCTCCCACTGTGGTACTTCCAGAGACGCCAGGCCGAGTTTCAAGTACATGACTAACTCAG

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

GGGCGCAGCTTTCGGGGCGGCTTTCGTACAGGGTGCCTGCGCCGGGCGAGGTATAACTCACCTGACAAT
 CAGAGGGCGAGGTATTTCAGCTCAACGACGAGTCCGTGAGCTCCTCGCTTGGTCTCCGTCCGGACGGGACA
 TTTTCAGATCGGCGGCGCCGGCCCTTTCATTACGCTCTCGTCAGGCAATCCTAACTCTGCAGACCTCGT
 CCTCTGAGCCGCGCTCTGGAGGCATGGAACTCTGCAATTTATTGAGGAGTTTGTGCCATCGGTCTACTT
 TAACCCCTTCTCGGGACCTCCCGGCCACTATCCGGATCAATTTATTCCTAACTTTGACGCGGTAAAGGAC
 TCGGGCGAGCGGTACGACTGAATGTTAAGTGGAGAGGCAGAGCAACTGCGCCTGAAACACCTGGTCCACT
 GTCGCCGCCACAAGTGCTTTGCCCGGACTCCGGTGAGTTTGTGCTACTTTGAATTGCCCGAGGATCATAT
 CGAGGGCCCGGCGCAGGCGTCCGGCTTACCGCCGAGGAGAGCTTGCCCGTAGCCTGATTCCGGGAGTTT
 ACCCAGCGCCCCCTGTAGTTGAGCGGGACAGGGGACCCTGTGTTCTCACTGTGATTGTCAAACCTGTCTTA
 ACCTTGGATTACATCAAGATCTTTGTTGCCATCTCTGTGCTGAGTATAATAAATACAGAAATTTAAATAT
 ACTGGGGCTCCTATCGCCATCCTGTAAACGCCACCGTCTTACCCGCCCAAGCAAACCAAGCGGAACCTT
 ACCTGGTACTTTTAACATCTCTCCCTCTGTGATTACAACAGTTTCAACCCAGACGGAGTGAGTCTACGA
 GAGAACCTCTCCGAGCTCAGCTACTCCATCAGAAAAACACCACCCTCCTTACCTGCCGGGAACGTACGA
 GTGCGTCACCGCGCGTGCACCACACCTACCGCTGACCGTAACCCAGACTTTTTCGGACAGACCTCAA
 TACTCTGTTTACAGAAACAGGAGGTGAGCTTAGAAAAACCTTAGGGTATTAGGCCAAAGCGCAGCTAC
 TGTGGGGTTTATGAACAATTCAAGCAACTCTACGGGCTATTCTAATTCAGGTTTCTCTAGAAATGGACGG
 AATTATTACAGAGCAGCGCTGCTAGAAAGACGCAGGGCAGCGGCCGAGCAACAGCGCATGAATCAAGAG
 CTCCAAGACATGGTTAACTTGCACCACTGCAAAAGGGGTATCTTTTGTCTGGTAAAGCAGGCCAAAGTCA
 CCTACGACAGTAATACACCGGACACCGCCTTAGCTACAAGTTGCCAACCAAGCGTCAGAAATTTGGTGGT
 CATGGTGGGAGAAAAGCCCATTAACATAACTCAGCACTCGGTAGAAACCGAAGGCTGCATTCACTACCT
 TGTCAAGGACCTGAGGATCTCTGCACCCCTTATTAAGACCCTGTGCGGTCTCAAAGATCTTTATCCCTTTA
 ACTAATAAAAAAATAATAAAGCATCACTTACTTTAAATCAGTTAGCAAATTTCTGTCCAGTTTATTCA
 GCAGCACCTCCTTGCCCTCCTCCAGCTCTGGTATTGCAGCTTCTCTGGCTGCAAACCTTTCTCCACAA
 TCTAAATGGAATGTCAGTTTCTCTGTTCCTGTCCATCCGCACCCACTATCTTCATGTTGTGCAGATG
 AAGCGCGCAAGACCGTCTGAAGATACCTTCAACCCCGTGTATCCATATGACACGGAACCCGTCCTCCAA
 CTGTGCCTTTTCTTACTCCTCCCTTTGTATCCCCCAATGGGTTTCAAGAGAGTCCCCCTGGGGTACTCTC
 TTTGCGCCTATCCGAACCTCTAGTTACCTCCAATGGCATGCTTGCCTCAAAATGGGCAACGGCCTCTCT
 CTGGACGAGGCGGCAACCTTACCTCCCAAATGTAACCACTGTGAGCCACCTCTCAAAAAACCAAGT
 CAAACATAAACCTGGAATATCTGCACCCCTCAGTTTACCTCAGAAGCCCTAACTGTGGCTGCCGCCGC
 ACCTCTAATGGTCCGGGCAACACACTCACCATGCAATCAGAGGCCCGCTAACCGTGCACGACTCCAAA
 CTTAGCATTGCCACCCAAGGACCCCTCAGAGTGTGAGAAGAAAGCTAGCCCTGCAAACATCAGGCCCCC
 TCACCACCACCGATAGCAGTACCCTTACTATCACTGCCTCACCCTCTAACTACTGCCACTGGTAGCTT
 GGGCATTGACTTGAAGAGGCCATTATACACAAAATGGAAAAGTAGGACTAAAGTACGGGGCTCCTTTG
 CATGTAACAGACGACCTAAACACTTTGACCGTAGCAACTGGTCCAGGTGTGACTATTAATAAATCTCTCT
 TGCAAACTAAAGTTACTGGAGCCTTGGGTTTGTATTACAAGGCAATATGCAACTTAATGTAGCAGGAGG
 ACTAAGGATTGATTCTCAAAACAGACGCCCTTATACCTTGATGTTAGTTATCCGTTTGATGCTCAAAACCAA
 CTAATCTAAGACTAGGACAGGGCCCTCTTTTATAAACTCAGCCCACAACCTGGATATTAACATAACA
 AAGGCCCTTACTTTGTTTACAGCTTCAAAACAATTCCAAAAGCTTGAGGTTAACCTAAGCACTGCCAAGGG
 GTTGATGTTTACGCTACAGCCATAGCCATTAATGCAGGAGATGGGCTTGAATTTGGTTACCTAATGCA
 CCAACACAAATCCCTCAAAACAAAATTTGGCCATGGCCTAGAATTTGATTCAAAACAAGGCTATGGTTC
 CTAACCTAGGAAGTGGCCTTAGTTTGTGACAGCAGGTGCCATTACAGTAGGAAACAAAATAATGATAA
 GCTAAGTTTGTGGACCAACAGCTCCATCTCTAACTGTAGACTAAATGCAGAGAAAGATGCTAAACTC
 ACTTTGGTCTTAACAAAATGTGGCAGTCAAATACTTGTCTACAGTTTTCAGTTTGGCTGTTAAAGGCAGTT
 TGGCTCCAATATCTGGAACAGTTCAAAGTGCTCATCTTATTATAAGATTTGACGAAAATGGAGTGCTACT
 AAACAATTCCTTCTGGACCCAGAATATTGGAACCTTTAGAAATGGAGATCTTACTGAAGGCACAGCCTAT
 ACAACGCTGTTGGATTTATGCCTAACCTATCAGCTTATCCAAAATCTCAGGTAAAACCTGCCAAAAGTA
 ACATTGTGTCAGTCAAGTTTACTTAAACGGAGACAAAACCTGTAACACTAACCATTACACTAAACGG
 TACACAGGAAACAGGAGACACAACCTCAAGTGCATACTCTATGTCAATTTTCATGGGACTGGTCTGGCCAC
 AACTACATTAATGAAATATTGCCACATCCTCTTACACTTTTTCATACATTGCCCAAGAATAAAGAATCG
 TTTGTGTTATGTTTCAACGTGTTTATTTTCAATTGCGAGAAAATTTCAATCATTTTTTCATTCACTAGTA
 TAGCCCCACCACACATAGCTTATACAGATCACCCTACCTTAATCAAACCTCAGAGAACCTTAGTATTCAA
 CCTGCCACCTCCCTCCCAACACACAGAGTACACAGTCTTCTCCCCGGCTGGCCTTAAAAAGCATCATA
 TCATGGGTAACAGACATATTCTTAGGTGTATATTCCACACGGTTTCTGTGAGCCAAACGCTCATCAG
 TGATATTAATAAATCCCGGGCAGCTCACTTAAGTTTCATGTGCTGCTGCTGAGCTGCTGAGCCACAGGCTG
 CTGTCCAACCTTGCCTTAAACGGGCGGCGAAGGAGAAGTCCACGCCCTACATGGGGGTAGAGTCATAA
 TCGTGTCATCAGGATAGGGCGGTGGTGTGTCAGCAGCGCGGAATAAACTGCTGCGCGCGCGCTCCGTCC
 TGACGGAATACAACATGGCAGTGGTCTCCTCAGCGATGATTTCGACACCGCCCGCAGCATAAGGCGCTTGT
 CCTCCGGGCACAGCAGCGCACCTGATCTCACTTAATCAGCACAGTAAGTGCAGCACAGCACCAATA
 TTGTTCAAAATCCACAGTGAAGGCGCTGTATCCAAAGCTCATGGCGGGGACCACAGAACCACGCTGGC

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

CATCATACCACAAGCGCAGGTAGATTAAAGTGGCGACCCCTCATAAACACGCTGGACATAAACATTACCTC
 TTTTGGCATGTTGTAATTCACCACCTCCCGGTACCATATAAACCTCTGATTAAACATGGCGCCATCCACC
 ACCATCCTAAACCAGCTGGCCAAAACCTGCCGCCGGCTATACACTGCAGGGAACCGGGACTGGAACAAT
 GACAGTGGAGAGCCAGGACTCGTAACCATGGATCATCATGCTCGTCATGATATCAATGTTGGCACAACA
 CAGGCACACGTGCATACACTTCCTCAGGATTACAAGCTCCTCCCGCGTTAGAACCATATCCCAGGGAACA
 ACCCATTCCTGAATCAGCGTAAATCCACACTGCAGGGAAGACCTCGCACGTAACCTCACGTTGTGCATTG
 TCAAAGTGTTACATTCCGGGCAGCAGCGGATGATCCTCCAGTATGGTAGCGCGGGTTTCTGTCTCAAAAGG
 AGGTAGACGATCCCTACTGTACGGAGTGCGCCGAGACAACCGAGATCGTGTGGTTCGTAGTGTATGCCA
 AATGGAACGCGCGAGCTAGTCATATTTCTGAAGCCAAAACAGGTGCGGGCGTGACAAACAGATCTGCGT
 CTCCGGTCTCGCCGCTTAGATCGCTCTGTGTAGTAGTTGTAGTATATCCACTCTCTCAAAGCATCCAGGC
 GCCCCCTGGCTTCGGGTTCTATGTAACTCCTTCATGCGCCGCTGCCCTGATAACATCCACCACCGCAGA
 ATAAGCCACACCCAGCCAACCTACACATTCGTTCTGCGAGTCACACACGGGAGGAGCGGGAAGAGCTGGA
 AGAACCATGTTTTTTTTTTTATTCCAAAAGATTATCCAAAACCTCAAATGAAGATCTATTAAGTGAACG
 CGTCCCTCCGGTGGCGTGGTCAAACCTCTACAGCCAAAGAACAGATAATGGCATTGTGAAGATGTTGCA
 CAATGGCTTCCAAAAGGCAAACGGCCCTCACGTCAAAGTGGACGTAAAGGCTAAACCTTCAGGGTGAAT
 CTCTCTATAAACATTCCAGCACCTTCAACCATGCCCCAAATAATTCTCATCTCGCCACCTTCTCAATATA
 TCTCTAAGCAAATCCGAATATTAAGTCCGGCCATTGTAAAAATCTGCTCCAGAGCGCCCTCCACCTTCA
 GCCTCAAGCAGCGAATCATGATTGCAAAAATTCAGGTTCTCAGACCTGTATAAGATTCAAAAGCGGA
 ACATTAACAAAATACCGCATCCCGTAGGTCCCTTCGCGAGGCCAGCTGAACATAATCGTGCAGGTCTG
 CACGGACCAGCGCGGCCACTTCCCCGCCAGGAACCTTGACAAAAGAACCCCACTGATTATGACACGCAT
 ACTCGGAGCTATGCTAACAGCGTAGCCCCGATGTAAGCTTTGTTGCATGGGCGCGCATATAAAATGCAA
 GGTGCTGCTCAAAAATCAGGCAAAGCCTCGCGCAAAAAGAAAGCACATCGTAGTCATGCTCATGCAGA
 TAAAGCAGGTAGCTCCGGAACCCACAGAAAAAGACACCATTTTTCTCTCAAACATGTCTCGGGTT
 TCTGCATAAACACAAAATAAAATAACAAAAAACATTTAAACATTAGAAGCCTGTCTTACAACAGGAAAA
 ACAACCTTTATAAGCATAAGACGGACTACGGCCATGCCGGCGTGACCGTAAAAAACTGGTCACCGTGAT
 TAAAAAGCACCACCGACAGCTCCTCGGTATGTCCGGAGTCATAATGTAAGACTCGGTAAACACATCAGG
 TTGATTACATCGGTAGTGCTAAAAAGCGACCGAAATAGCCCCGGGGGAATACATACCCGCAGGCGTAGA
 GACAACATTACAGCCCCCATAGGAGGTATAACAAAATTAATAGGAGAGAAAAACACATAAACACCTGAAA
 AACCTCCTGCCTAGGCAAAATAGCACCTCCTCCGCTCCAGAACACATACAGCGCTTCCACAGCGGCAGC
 CATAACAGGTAGCTTCCAGTAAAGAAAGAAACCTATTAAAAAACACCACTCGACACGGCACCGCTC
 AATCAGTCACAGTGTAAGAAAGGGCCAGTGCAGAGCGAGTATATATAGGACTAAAAAATGACGTAACGG
 TTAAGTCCACAAAAAACCCAGAAAAACCGCACGCGAACCTACGCCAGAAACGAAAGCCAAAAACCC
 ACAACTTCTCAAATCGTCACTTCCGTTTTCACAGTTACGTCACTTCCCATTTTAAGAAACTACAATT
 CCCAACACATACAAGTTACTCCGCCCTAAAACCTACGTCAACCCGCCCGCTTCCACGCCCGCGCCACGT
 CACAACTCCACCCCTCATTATCATATTGGCTTCAATCCAAAATAAGGTATATTATTGATGATGTTAAT
 TAATTTAAATCCGCATCCGATATCGAGCTCTCCCGGGAATTCGATCTGCGACGCGAGGCTGGATGCCCT
 TCCCATTATGATTCTCTCGCTTCCGCGGCATCGGGATGCCCGCGTTGACGGCCATGCTGTCCAGGCA
 GGTAGATGACGACCATCAGGGACAGCTTCACGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTG
 CTGGCGTTTTTTCATAGGCTCCGCCCCCTGACGAGCATCACAAAATCGACGCTCAAGTCAGAGGTGGC
 GAAACCCGACAGGACTATAAGATACAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCTGTTCC
 GACCTGCGCGTTACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCAATGCTCA
 CGCTGTAGGTATCTCAGTTCGGTGTAGGTGCTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTC
 AGCCCGACCGCTGCGCCTTATCCGGTAACCTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCC
 ACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG
 TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCT
 TCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACACCGCTGGTAGCGGTGGTTTTTTTTGTTT
 CAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGTCTGAC
 GCTCAGTGAACGAAAACTCACGTTAAGGGATTTTGGTTCATGAGATTATCAAAAAGGATCTTACCTAGA
 TCCTTTTAAATCAATCTAAAGTATATATGAGTAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAG
 GCACCTATCTCAGCGATCTGTCTATTTCTGTCATCCATAGTTGCCTGACTCCCCGTCTGTGTAGATACTA
 CGATACGGGAGGGCTTACCATCTGGCCCGAGTGTGCAATGATAACCGGAGACCCACGCTCACCGGCTCC
 AGATTTATCAGCAATAAACAGCCAGCCGGAAGGGCCGAGCGCAGAAAGTGGTCTGCAACTTTATCCGCC
 TCCATCCAGTCTATTAATTGTTGCCGGAAGCTAGAGTAAGTAGTTCCGCACTTAATAGTTTGGCAACG
 TTGTTGCCATTGNTGACGGCATCGTGGTGTACGCTCGTCTTGGTATGGCTTCATTACGCTCCGGTTC
 CCAACGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAAAAGCGGTTAGCTCCTTCGGTCCCTCCG
 ATCGTTGTGCAAGTAAGTTGGCCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAATTTCTCTTA
 CTGTATGCCATCCGTAAGATGCTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTG
 TATGCGGCGAGGCTGTCTTGTGCCCGCGTCAACACGGGATAATACCGGCCACATAGCAGAACTTTA
 AAAGTGCTCATCATTTGGAACAGTTCTTCCGGGGCAAACTCTCAAGGATCTTACCGCTGTTGAGATCCA

Table 11 (continued) Nucleotide sequence of pAd/CMV/V5-GW/*lacZ*.PL-DEST™.

GTTCGATGTAACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTTACTTTCACCAGCGTTTCTGGGTG
AGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATA
CTCTTCCTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAAT
GTATTTAGAAAAATAACAAATAGGGGTCCGCGCACATTCCCCGAAAAGTGCCACCTGACGTCTAAGA
AACCATTATTATCATGACATTAACCTATAAAAAATAGGCGTATCACGAGGCCCTTTCGTCTTCAAGGATCC
GAATTCCTGGGAGAGCTCGATATCGCATGCGGATTTAAATTAATTAA

Table 12. Nucleotide sequence of pIB/V5-His-DEST.

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                                OpIE-2 pr
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1  CATGATGATA AACAAATGTAT GGTGCTAATG TTGCTTCAAC AACAAATCTG
   GTACTACTAT TTGTTACATA CCACGATTAC AACGAAGTTG TTGTTAAGAC
                                OpIE-2 pr
~~~~~
51 TTGAACTGTG TTTTCATGTT TGCCAACAAG CACCTTTATA CTCGGTGGCC
   AACTTGACAC AAAAGTACAA ACGGTTGTTC GTGGAAATAT GAGCCACCGG
                                OpIE-2 pr
~~~~~
101 TCCCCACCAC CAACTTTTTT GCACTGCAAA AAAACACGCT TTTGCACGCG
   AGGGGTGGTG GTTGAAAAAA CGTGACGTTT TTTTGTGCGA AAACGTGCGC
                                OpIE-2 pr
~~~~~
151 GGCCCATACA TAGTACAAAC TCTACGTTTC GTAGACTATT TTACATAAAT
   CCGGGTATGT ATCATGTTTG AGATGCAAAG CATCTGATAA AATGTATTTA
                                OpIE-2 pr
~~~~~
201 AGTCTACACC GTTGATACG CTCCAATAAC ACTACCACAC ATTGAACCTT
   TCAGATGTGG CAACATATGC GAGGTTTATG TGATGGTGTG TAACTTGGA
                                OpIE-2 pr
~~~~~
251 TTTGCAGTGC AAAAAAGTAC GTGTCGGCAG TCACGTAGGC CGGCCTTATC
   AAACGTCACG TTTTTCATG CACAGCCGTC AGTGCATCCG GCCGGAATAG
                                OpIE-2 pr
~~~~~
301 GGGTCGCGTC CTGTCACGTA CGAATCACAT TATCGGACCG GACGAGTGTT
   CCCAGCGCAG GACAGTGCAT GCTTAGTGTA ATAGCCTGGC CTGCTCACAA
                                OpIE-2 pr
~~~~~
351 GTCTTATCGT GACAGGACGC CAGCTTCCTG TGTTGCTAAC CGCAGCCGGA
   CAGAATAGCA CTGTCCTGCG GTCGAAGGAC ACAACGATTG GCGTCGGCCT
                                OpIE-2 pr
~~~~~
401 CGCAACTCCT TATCGGAACA GGACGCGCCT CCATATCAGC CGCGCGTTAT
   GCGTTGAGGA ATAGCCTTGT CCTGCGCGGA GGTATAGTCG GCGCGCAATA
                                OpIE-2 pr
~~~~~
451 CTCATGCACG TGACCGGACA CGAGGCGCCC GTCCCGCTTA TCGCGCCTAT
   GAGTACGTGC ACTGGCCTGT GCTCCGCGGG CAGGGCGAAT AGCGCGGATA
                                OpIE2FOR
~~~~~
                                OpIE-2 pr
~~~~~
501 AAATACAGCC CGCAACGATC TGGTAAACAC AGTTGAACAG CATCTGTTCC
   TTTATGTCGG GCGTTGCTAG ACCATTTGTG TCAACTTGTC GTAGACAAGC
551 AATTTAAAGC TTGATATCGA ATTCCTGCAG CCCAGCGCTG GATCCTCGAT
   TTAAATTTTCG AACTATAGCT TAAGGACGTC GGGTCGCGAC CTAGGAGCTA

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Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

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                                attR1
~~~~~
601 CACAAGTTTG TACAAAAAAG CTGAACGAGA AACGTAAAAT GATATAAATA
    GTGTTCAAAC ATGTTTTTTC GACTTGCTCT TTGCATTTTA CTATATTTAT
                                attR1
~~~~~
651 TCAATATATT AAATTAGATT TTGCATAAAA AACAGACTAC ATAATACTGT
    AGTTATATAA TTTAATCTAA AACGTATTTT TTGTCTGATG TATTATGACA
                                attR1
~~~~~
701 AAAACACAAC ATATCCAGTC ACTATGGCGG CCGCATTAGG CACCCCAGGC
    TTTTGTGTTG TATAGGTCAG TGATACCGCC GCGTAATCC GTGGGGTCCG
751 TTTACACTTT ATGCTTCCGG CTCGTATAAT GTGTGGATTT TGAGTTAGGA
    AAATGTGAAA TACGAAGGCC GAGCATATTA CACACCTAAA ACTCAATCCT
                                Cmr
~~~~~
801 TCCGTCGAGA TTTTCAGGAG CTAAGGAAGC TAAAATGGAG AAAAAAATCA
    AGGCAGCTCT AAAAGTCCTC GATTCCCTCG ATTTTACCTC TTTTTTTAGT
                                Cmr
~~~~~
851 CTGGATATAC CACCGTTGAT ATATCCCAAT GGCATCGTAA AGAACATTTT
    GACCTATATG GTGGCAACTA TATAGGGTTA CCGTAGCATT TCTTGTA AAA
                                Cmr
~~~~~
901 GAGGCATTTT AGTCAGTTGC TCAATGTACC TATAACCAGA CCGTTCAGCT
    CTCCGTAAAG TCAGTCAACG AGTTACATGG ATATTGGTCT GGCAAGTCGA
                                Cmr
~~~~~
951 GGATATTACG GCCTTTTAA AGACCGTAAA GAAAAATAAG CACAAGTTTT
    CCTATAATGC CGGAAAAATT TCTGGCATT TTTTTATTG GTGTTCAAAA
                                Cmr
~~~~~
1001 ATCCGGCCTT TATTCACATT CTTGCCCCGCC TGATGAATGC TCATCCGGAA
    TAGGCCGGAA ATAAGTGTA GAACGGGCGG ACTACTTACG AGTAGGCCTT
                                Cmr
~~~~~
1051 TTCCGTATGG CAATGAAAGA CCGTGAGCTG GTGATATGGG ATAGTGTTCA
    AAGGCATACC GTTACTTTCT GCCACTCGAC CACTATACCC TATCACAAGT
                                Cmr
~~~~~
1101 CCCTTGTTAC ACCGTTTTC ATGAGCAAAC TGAAACGTTT TCATCGCTCT
    GGGAACAATG TGGCAAAGG TACTCGTTTG ACTTTGCAA AGTAGCGAGA
                                Cmr
~~~~~
1151 GGAGTGAATA CCACGACGAT TTCCGGCAGT TTCTACACAT ATATTGCGAA
    CCTCACTTAT GGTGCTGCTA AAGGCCGTCA AAGATGTGTA TATAAGCGTT

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Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

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                                Cmr
~~~~~
1201 GATGTGGCGT GTTACGGTGA AAACCTGGCC TATTTCCCTA AAGGGTTTAT
    CTACACCGCA CAATGCCACT TTTGGACCGG ATAAAGGGAT TTCCCAAATA
                                Cmr
~~~~~
1251 TGAGAATATG TTTTTCGTCT CAGCCAATCC CTGGGTGAGT TTCACCAGTT
    ACTCTTATAC AAAAAGCAGA GTCGGTTAGG GACCCACTCA AAGTGGTCAA
                                Cmr
~~~~~
1301 TTGATTTAAA CGTGGCCAAT ATGGACAAC TCTTCGCCCC CGTTTTTCACC
    AACTAAATTT GCACCGGTTA TACCTGTTGA AGAAGCGGGG GCAAAAGTGG
                                Cmr
~~~~~
1351 ATGGGCAAAT ATTATACGCA AGGCGACAAG GTGCTGATGC CGCTGGCGAT
    TACCCGTTTA TAATATGCGT TCCGCTGTTT CACGACTACG GCGACCGCTA
                                Cmr
~~~~~
1401 TCAGGTTTCAT CATGCCGTTT GTGATGGCTT CCATGTCGGC AGAATGCTTA
    AGTCCAAGTA GTACGGCAAA CACTACCGAA GGTACAGCCG TCTTACGAAT
                                Cmr
~~~~~
1451 ATGAATTACA ACAGTACTGC GATGAGTGGC AGGGCGGGGC GTAAACGCGT
    TACTTAATGT TGTCATGACG CTACTCACCG TCCCGCCCCG CATTTGCGCA
1501 GGATCCGGCT TACTAAAAGC CAGATAACAG TATGCGTATT TGCGCGCTGA
    CCTAGGCCGA ATGATTTTCG GTCTATTGTC ATACGCATAA ACGCGCGACT
1551 TTTTTCGCGT ATAAGAATAT ATACTGATAT GTATACCCGA AGTATGTCAA
    AAAAACGCCA TATTCTTATA TATGACTATA CATATGGGCT TCATACAGTT
1601 AAAGAGGTAT GCTATGAAGC AGCGTATTAC AGTGACAGTT GACAGCGACA
    TTTCTCCATA CGATACTTCG TCGCATAATG TCACTGTCAA CTGTCGCTGT
1651 GCTATCAGTT GCTCAAGGCA TATATGATGT CAATATCTCC GGTCTGGTAA
    CGATAGTCAA CGAGTTCGGT ATATACTACA GTTATAGAGG CCAGACCATT
1701 GCACAACCAT GCAGAATGAA GCCCGTCGTC TCGGTGCCGA ACGCTGGAAA
    CGTGTGGGTA CGTCTTACTT CGGGCAGCAG ACGCACGGCT TGCGACCTTT
1751 GCGGAAAATC AGGAAGGGAT GGCTGAGGTC GCCCGGTTTA TTGAAATGAA
    CGCCTTTTAT TCCTTCCCTA CCGACTCCAG CGGGCCAAAT AACTTTACTT
                                ccdB
~~~~~
1801 CGGCTCTTTT GCTGACGAGA ACAGGGGCTG GTGAAATGCA GTTTAAGGTT
    GCCGAGAAAA CGACTGCTCT TGTCCCCGAC CACTTTACGT CAAATTCCAA
                                ccdB
~~~~~
1851 TACACCTATA AAAGAGAGAG CCGTTATCGT CTGTTTGTGG ATGTACAGAG
    ATGTGGATAT TTTCTCTCTC GGCAATAGCA GACAAACACC TACATGTCTC
                                ccdB
~~~~~
1901 TGATATTATT GACACGCCCC GCGACGGAT GGTGATCCCC CTGGCCAGTG
    ACTATAATAA CTGTGCGGGC CCGCTGCCTA CCACTAGGGG GACCGGTCAC

```

Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

```

                                ccdB
~~~~~
1951 CACGTCTGCT GTCAGATAAA GTCTCCCGTG AACTTTACCC GGTGGTGCAT
    GTGCAGACGA CAGTCTATTT CAGAGGGCAC TTGAAATGGG CCACCACGTA
                                ccdB
~~~~~
2001 ATCGGGGATG AAAGCTGGCG CATGATGACC ACCGATATGG CCAGTGTGCC
    TAGCCCCTAC TTTGACCGC GTACTACTGG TGGCTATACC GGTCACACGG
                                ccdB
~~~~~
2051 GGTCTCCGTT ATCGGGGAAG AAGTGGCTGA TCTCAGCCAC CGCGAAAATG
    CCAGAGGCAA TAGCCCCTTC TTCACCGACT AGAGTCGGTG GCGCTTTTAC
                                ccdB
~~~~~
2101 ACATCAAAAA CGCCATTAAC CTGATGTTCT GGGGAATATA AATGTCAGGC
    TGTAGTTTTT GCGGTAATTG GACTACAAGA CCCCTTATAT TTACAGTCCG
                                attR2
~~~~~
2151 TCCCTTATAC ACAGCCAGTC TGCAGGTCGA CCATAGTGAC TGGATATGTT
    AGGGAATATG TGTCGGTCAG ACGTCCAGCT GGTATCACTG ACCTATACAA
                                attR2
~~~~~
2201 GTGTTTTACA GTATTATGTA GTCTGTTTTT TATGCAAAAT CTAATTTAAT
    CACAAAATGT CATAATACAT CAGACAAAAA ATACGTTTTA GATTAAATTA
                                attR2
~~~~~
2251 ATATTGATAT TTATATCATT TTACGTTTCT CGTTCAGCTT TCTTGTAACA
    TATAACTATA AATATAGTAA AATGCAAAGA GCAAGTCGAA AGAACATGTT
    attR2                                V5 tag
~~~~~
2301 AGTGGTGATC GACCCGGGTC TAGAGGGCCC GCGGTTCGAA GGTAAGCCTA
    TCACCACTAG CTGGGCCCCAG ATCTCCCGGG CGCCAAGCTT CCATTCGGAT
                                V5 tag
~~~~~
                                Poly His 6
                                tag
~~~~~
2351 TCCCTAACCC TCTCCTCGGT CTCGATTCTA CGCGTACCGG TCATCATCAC
    AGGGATTGGG AGAGGAGCCA GAGCTAAGAT GCGCATGGCC AGTAGTAGTG
    Poly His 6 tag                                OpIE-2 PolyA
~~~~~
2401 CATCAACATT GAGTTTATCT GACTAAATCT TAGTTTGTAT TGTCATGTTT
    GTAGTGGTAA CTCAAATAGA CTGATTTAGA ATCAAACATA ACAGTACAAA
                                OpIE-2 PolyA
~~~~~
2451 TAATACAATA TGTTATGTTT AAATATGTTT TTAATAAATT TTATAAATA
    ATTATGTTAT ACAATACAAA TTTATACAAA AATTATTTAA AATATTTTAT

```

Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

OpIE-2 PolyA

~~~~~

2501 ATTTCAACTT TTATTGTAAC AACATTGTCC ATTTACACAC TCCTTTCAAG  
TAAAGTTGAA AATAACATTG TTGTAACAGG TAAATGTGTG AGGAAAGTTC

2551 CGCGTGGGAT CGATGCTCAC TCAAAGGCGG TAATACGGTT ATCCACAGAA  
GCGCACCCCTA GCTACGAGTG AGTTTCCGCC ATTATGCCAA TAGGTGTCTT  
pMB1 ori

~~~~~

2601 TCAGGGGATA ACGCAGGAAA GAACATGTGA GCAAAAGGCC AGCAAAAGGC
AGTCCCCTAT TCGTCCCTTT CTTGTACACT CGTTTTCCGG TCGTTTTCCG
pMB1 ori

~~~~~

2651 CAGGAACCGT AAAAAGGCCG CGTTGCTGGC GTTTTTCCAT AGGCTCCGCC  
GTCCTTGGCA TTTTCCGGC GCAACGACCG CAAAAAGGTA TCCGAGGCGG  
pMB1 ori

~~~~~

2701 CCCCTGACGA GCATCACAAA AATCGACGCT CAAGTCAGAG GTGGCGAAAC
GGGACTGCT CGTAGTGTTT TTAGCTGCGA GTTCAGTCTC CACCGCTTTG
pMB1 ori

~~~~~

2751 CCGACAGGAC TATAAAGATA CCAGGCGTTT CCCCCTGGAA GCTCCCTCGT  
GGCTGTCCTG ATATTCTAT GGTCCGCAA GGGGGACCTT CGAGGGAGCA  
pMB1 ori

~~~~~

2801 GCGCTCTCCT GTTCCGACCC TGCCGCTTAC CGGATACCTG TCCGCCTTTC
CGCGAGAGGA CAAGGCTGGG ACGGCGAATG GCCTATGGAC AGGCGGAAAG
pMB1 ori

~~~~~

2851 TCCCTTCGGG AAGCGTGGCG CTTTCTCATA GTCACGCTG TAGGTATCTC  
AGGGAAGCCC TTCGCACCGC GAAAGAGTAT CGAGTGCGAC ATCCATAGAG  
pMB1 ori

~~~~~

2901 AGTTCGGTGT AGGTCGTTTC CTCCAAGCTG GGCTGTGTGC ACGAACCCCC
TCAAGCCACA TCCAGCAAGC GAGGTTCGAC CCGACACACG TGCTTGGGGG
pMB1 ori

~~~~~

2951 CGTTCAGCCC GACCGCTGCG CCTTATCCGG TAACTATCGT CTTGAGTCCA  
GCAAGTCGGG CTGGCGACGC GGAATAGGCC ATTGATAGCA GAACTCAGGT  
pMB1 ori

~~~~~

3001 ACCCGGTAAG ACACGACTTA TCGCCACTGG CAGCAGCCAC TGGTAACAGG
TGGGCCATTC TGTGCTGAAT AGCGGTGACC GTCGTCGGTG ACCATTGTCC
pMB1 ori

~~~~~

3051 ATTAGCAGAG CGAGGTATGT AGGCGGTGCT ACAGAGTTCT TGAAGTGGTG  
TAATCGTCTC GCTCCATACA TCCGCCACGA TGTCTCAAGA ACTTCACCAC

Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

```

                                pMB1 ori
~~~~~
3101 GCCTAACTAC GGCTACACTA GAAGAACAGT ATTTGGTATC TCGCTCTGC
 CGGATTGATG CCGATGTGAT CTTCTTGTC TAAACCATAG ACGCGAGACG
 pMB1 ori
~~~~~
3151 TGAAGCCAGT TACCTTCGGA AAAAGAGTTG GTAGCTCTTG ATCCGGCAAA
    ACTTCGGTCA ATGGAAGCCT TTTTCTCAAC CATCGAGAAC TAGGCCGTTT
                                pMB1 ori
~~~~~
3201 CAAACCACCG CTGGTAGCGG TGGTTTTTTT GTTTGCAAGC AGCAGATTAC
 GTTGGTGGC GACCATCGCC ACCAAAAAAA CAAACGTTTC TCGTCTAATG
 pMB1 ori
~~~~~
3251 GCGCAGAAAA AAAGGATCTC AAGAAGATCC TTTGATCTTT TCTACGGGGT
    CGCGTCTTTT TTTCTTAGAG TTCTTCTAGG AAACCTAGAAA AGATGCCCCA
3301 CTGACGCTCA GTGGAACGAA AACTCACGTT AAGGGATTTT GGTTCATGCCC
    GACTGCGAGT CACCTTGCTT TTGAGTGCAA TTCCCTAAAA CCAGTACGGG
                                GP64 promoter
~~~~~
3351 TTGTTCCGAA GGGTTGTGTC ACGTAGGCCA GATAACGGTC GGGTATATAA
 AACAAAGGCTT CCCAACACAG TGCATCCGGT CTATTGCCAG CCCATATATT
 GP64 promoter
~~~~~
3401 GATGCCTCAA TGCTACTAGT AAATCAGTCA CACCAAGGCT TCAATAAGGA
    CTACGGAGTT ACGATGATCA TTTAGTCAGT GTGGTTCCGA AGTTATTCCT
    GP64 promoter                                EM7
~~~~~
3451 ACACACAAGC AAGCCCTTTG AGTCAAGGGC TGCCGGGCTG CAGCACGTGT
 TGTGTGTTTCG TTCGGGAAAC TCAGTTCCCG ACGGCCCGAC GTCGTGCACA
 EM7
~~~~~
3501 TGACAATTAA TCATCGGCAT AGTATATCGG CATAGTATAA TACGACAAGG
    ACTGTTAATT AGTAGCCGTA TCATATAGCC GTATCATATT ATGCTGTTCC
                                Blastcidin(r)
~~~~~
3551 TGAGGAACTA AACCATGGCC AAGCCTTTGT CTCAAGAAGA ATCCACCCTC
 ACTCCTTGAT TTGGTACCGG TTCGGAAACA GAGTTCTTCT TAGGTGGGAG
 Blastcidin(r)
~~~~~
3601 ATTGAAAGAG CAACGGCTAC AATCAACAGC ATCCCCATCT CTGAAGACTA
    TAACTTTCTC GTTGCCGATG TTAGTTGTCG TAGGGGTAGA GACTTCTGAT
                                Blastcidin(r)
~~~~~
3651 CAGCGTCGCC GCGCAGCTC TCTCTAGCGA CGGCCGCATC TTTACTGGTG
 GTCGCGCGG CCGCGTCGAG AGAGATCGCT GCCGGCGTAG AAGTGACCAC

```

Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

```

 Blastcidin(r)
~~~~~
3701 TCAATGTATA TCATTTTACT GGGGGACCTT GCGCAGAACT CGTGGTGCTG
    AGTTACATAT AGTAAAATGA CCCCCTGGAA CGCGTCTTGA GCACCACGAC
                                Blastcidin(r)
~~~~~
3751 GGGACTGCTG CTGCTGCGGC AGCTGGCAAC CTGACTTGTA TCGTCGCGAT
 CCGTGACGAC GACGACGCCG TCGACCGTTG GACTGAACAT AGCAGCGCTA
 Blastcidin(r)
~~~~~
3801 CGGAAATGAG AACAGGGGCA TCTTGAGCCC CTGCGGACGG TGCCGACAGG
    GCCTTTACTC TTGTCCCCGT AGAACTCGGG GACGCCTGCC ACGGCTGTCC
                                Blastcidin(r)
~~~~~
3851 TTCTTCTCGA TCTGCATCCT GGGATCAAAG CCATAGTGAA GGACAGTGAT
 AAGAAGAGCT AGACGTAGGA CCCTAGTTTC GGTATCACTT CCTGTCACTA
 Blastcidin(r)
~~~~~
3901 GGACAGCCGA CGGCAGTTGG GATTTCGTGAA TTGCTGCCCT CTGGTTATGT
    CCTGTGCGGT GCCGTCAACC CTAAGCACTT AACGACGGGA GACCAATACA
    Blastcidin(r)
~~~~~
3951 GTGGGAGGGC TAAGCACTTC GTGGCCGAGG AGCAGGACTG ACACGTCCCG
 CACCCTCCCG ATTTCGTGAAG CACCGGCTCC TCGTCCTGAC TGTGCAGGGC
4001 GGAGATCTGC ATGTCTACTA AACTCACAAA TTAGAGCTTC AATTTAATTA
 CCTCTAGACG TACAGATGAT TTGAGTGTTT AATCTCGAAG TTAAATTAAT
 Amp (r)
~~~~~
4051 TATCAGTTAT TACCCATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC
    ATAGTCAATA ATGGGTAAC TTTTCCTTCT CATACTCATA AGTTGTAAAG
                                Amp (r)
~~~~~
4101 CGTGTGCGCC TTATTCCCTT TTTTGCGGCA TTTTGCCTTC CTGTTTTTGC
 GCACAGCGGG AATAAGGGAA AAAACGCCGT AAAACGGAAG GACAAAAACG
 Amp (r)
~~~~~
4151 TCACCCAGAA ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG
    AGTGGGTCTT TGCGACCACT TTCATTTTCT ACGACTTCTA GTCAACCCAC
                                Amp (r)
~~~~~
4201 CACGAGTGGG TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG
 GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTCGCCATT CTAGGAACTC
 Amp (r)
~~~~~
4251 AGTTTTCGCC CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT
    TCAAAAGCGG GGCTTCTTGC AAAAGGTTAC TACTCGTGAA AATTTCAAGA

```



Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

```

                                Amp (r)
~~~~~
4301 GCTATGTGGC GCGGTATTAT CCCGTATTGA CGCCGGGCAA GAGCAACTCG
 CGATACACCG CGCCATAATA GGGCATAACT GCGGCCCGTT CTCGTTGAGC
 Amp (r)
~~~~~
4351 GTCGCCGCAT ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC
    CAGCGGCGTA TGTGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTCAG
                                Amp (r)
~~~~~
4401 ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC
 TGTCTTTTCG TAGAATGCCT ACCGTACTGT CATTCTCTTA ATACGTCACG
 Amp (r)
~~~~~
4451 TGCCATAACC ATGAGTGATA ACACTGCGGC CAACTTACTT CTGACAACGA
    ACGGTATTGG TACTCACTAT TGTGACGCCG GTTGAATGAA GACTGTTGCT
                                Amp (r)
~~~~~
4501 TCGGAGGACC GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT
 AGCCTCCTGG CTTCTCGAT TGGCGAAAAA ACGTGTTGTA CCCCCTAGTA
 Amp (r)
~~~~~
4551 GTAAC TCGCC TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAAA
    CATTGAGCGG AACTAGCAAC CCTTGGCCTC GACTTACTTC GGTATGGTTT
                                Amp (r)
~~~~~
4601 CGACGAGCGT GACACCACGA TGCCTGTAGC AATGGCAACA ACGTTGCGCA
 GCTGCTCGCA CTGTGGTGCT ACGGACATCG TTACCGTTGT TGCAACGCGT
 Amp (r)
~~~~~
4651 AACTATTAAC TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA
    TTGATAATTG ACCGCTTGAT GAATGAGATC GAAGGGCCGT TGTTAATTAT
                                Amp (r)
~~~~~
4701 GACTGGATGG AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT
 CTGACCTACC TCCGCCTATT TCAACGTCCT GGTGAAGACG CGAGCCGGGA
 Amp (r)
~~~~~
4751 TCCGGCTGGC TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT
    AGGCCGACCG ACCAAATAAC GACTATTTAG ACCTCGGCCA CTCGCACCCA
                                Amp (r)
~~~~~
4801 CTCGCGGTAT CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC
 GAGCGCCATA GTAACGTCGT GACCCCGGTC TACCATTCTG GAGGGCATAG
 Amp (r)
~~~~~
4851 GTAGTTATCT ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG
    CATCAATAGA TGTGCTGCCC CTCAGTCCGT TGATACCTAC TTGCTTTATC

```

Table 12 (continued). Nucleotide sequence of pIB/V5-His-DEST.

Amp (r)

~~~~~

4901	ACAGATCGCT	GAGATAGGTG	CCTCACTGAT	TAAGCATTGG	TAAGTGTGAG
	TGTCTAGCGA	CTCTATCCAC	GGAGTGACTA	ATTCGTAACC	ATTGACAGTC
4951	ACCAAGTTTA	CTCATATATA	CTTTAGATTG	ATTTAAAACT	TCATTTTAA
	TGGTTCAAAT	GAGTATATAT	GAAATCTAAC	TAAATTTTGA	AGTAAAAATT
5001	TTTAAAAGGA	TCTAGGTGAA	GATCCTTTTT	GATAATCT	
	AAATTTTCCT	AGATCCACTT	CTAGGAAAAA	CTATTAGA	

Table 13. Nucleotide sequence of the V5-His DEST cassette.

ph promoter
~~~~~

1 ATAAGTATTT TACTGTTTTTC GTAACAGTTT TGTAATAAAA AAACCTATAA  
TATTCATAAA ATGACAAAAG CATTGTCAAA ACATTATTTT TTTGGATATT  
51 ATATTCGCGG TTATTCATAC CGTCCCACCA TCGGGCGCGG ATCCCCGGGT  
TATAAGGCCT AATAAGTATG GCAGGGTGGT AGCCCCGCGC TAGGGGCCCA  
att R1  
~~~~~

101 ACCGATATCA CAAGTTTGTA CAAAAAGCT GAACGAGAAA CGTAAATGA
TGGCTATAGT GTTCAAACAT GTTTTTTCGA CTTGCTCTTT GCATTTTACT
att R1
~~~~~

151 TATAAATATC AATATATTAA ATTAGATTTT GCATAAAAAA CAGACTACAT  
ATATTTATAG TTATATAATT TAATCTAAAA CGTATTTTTT GTCTGATGTA  
att R1  
~~~~~

201 AATACTGTAA AACACAACAT ATCCAGTCAC TATGGCGGCC GCTCCCTAAC
TTATGACATT TTGTGTTGTA TAGGTCAGTG ATACCGCCGG CGAGGGATTG
251 CCACGGGGCC CGTGGCTATG GCAGGGCTTG CCGCCCCGAC GTTGGCTGCG
GGTGCCCCGG GCACCGATAC CGTCCCGAAC GCGGGGGCTG CAACCGACGC
301 AGCCCTGGGC CTTCACCAGA ACTTGGGGGT TGGGGTGGGG AAAAGGAAGA
TCGGGACCCG GAAGTGGGCT TGAACCCCCA ACCCCACCCC TTTTCCTTCT
351 AACGCGGGCG TATTGGTCCC AATGGGGTCT CGGTGGGGTA TCGACAGAGT
TTGCGCCCCG ATAACCAGGG TTACCCAGAG GCCACCCCAT AGCTGTCTCA
401 GCCAGCCCTG GGACCGAACC CCGCGTTTAT GAACAAACGA CCCAACACCC
CGGTCGGGAC CCTGGCTTGG GGCGCAAATA CTTGTTTGCT GGGTTGTGGG
451 GTGCGTTTTA TTCTGTCTTT TTATTGCCGT CATAGCGCGG GTTCCTTCCG
CACGCAAAAT AAGACAGAAA AATAACGGCA GTATCGCGCC CAAGGAAGGC
501 GTATTGTCTC CTTCCGTGTT TCAGTTAGCC TCCCCATCT CCCGGGCAAA
CATAACAGAG GAAGGCACAA AGTCAATCGG AGGGGGTAGA GGGCCCGTTT
~~~~~

tk gene  
N A E G M E R A F

551 CGTGCGCGCC AGGTCGCAGA TCGTCGGTAT GGAGCCTGGG GTGGTGACGT  
GCACGCGCGG TCCAGCGTCT AGCAGCCATA CCTCGGACCC CACCACTGCA  
~~~~~

tk gene
T R A L D C I T P I S G P T T V H .

601 GGGTCTGGAC CATCCCGGAG GTAAGTTGCA GCAGGGCGTC CCGGCAGCCG
CCCAGACCTG GTAGGGCCTC CATTCAACGT CGTCCCGCAG GGCCGTGCGC
~~~~~

tk gene  
. T Q V M G S T L Q L L A D R C G A .

651 GCGGGCGATT GGTCTGAATC CAGGATAAAG ACATGCATGG GACGGAGGCG  
CGCCCGCTAA CCAGCATTAG GTCCTATTTC TGTACGTACC CTGCCTCCGC  
~~~~~

tk gene
.. P S Q D Y D L I F V H M P R L R

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

701 TTTGGCCAAG ACGTCCAAAG CCCAGGCAAA CACGTTATAC AGGTCGCCGT
    AAACCGGTTT TGCAGGTTT GGGTCCGTTT GTGCAATATG TCCAGCGGCA
    ~~~~~
                                tk gene
      K A L V D L A W A F V N Y L D G N .
751 TGGGGGCCAG CAACTCGGGG GCCCGAAACA GGGTAAATAA CGTGTCCCCG
    ACCCCCGGTC GTTGAGCCCC CGGGCTTTGT CCCATTTATT GCACAGGGGC
    ~~~~~
                                tk gene
      . P A L L E P A R F L T F L T D G I .
801 ATATGGGGTC GTGGGCCCGC GTTGCTCTGG GGCTCGGCAC CCTGGGGCGG
    TATACCCAG CACCCGGGCG CAACGAGACC CCGAGCCGTG GGACCCCGCC
    ~~~~~
                                tk gene
      .. H P R P G A N S Q P E A G Q P P
851 CACGGCCGCC CCCGAAAGCT GTCCCCAATC CTCCCGCCAC GACCCGCCG
    GTGCCGGCGG GGGCTTTCGA CAGGGGTTAG GAGGGCGGTG CTGGGCGGCG
    ~~~~~
                                tk gene
      V A A G S L Q G W D E R W S G G G .
901 CCTGCAGATA CCGCACCGTA TTGGCAAGCA GCCCATAAAC GCGGCGAATC
    GGACGTCTAT GGC GTGGCAT AACCGTTCGT CGGGTATTTG CGCCGCTTAG
    ~~~~~
                                tk gene
      . Q L Y R V T N A L L G Y V R R I A .
951 GCGGCCAGCA TAGCCAGGTC AAGCCGCTCG CCGGGGCGCT GGCGTTTGGC
    CGCCGGTTCG ATCGGTCCAG TTCGGCGAGC GGCCCCGCGA CCGCAAACCG
    ~~~~~
                                tk gene
      .. A L M A L D L R E G P R Q R K A
1001 CAGGCGGTCG ATGTGTCTGT CCTCCGGAAG GGCCCCAAC ACGATGTTTG
    GTCCGCCAGC TACACAGACA GGAGGCCTTC CCGGGGGTTG TGCTACAAAC
    ~~~~~
                                tk gene
      L R D I H R D E P L A G L V I N T .
1051 TGCCGGGCAA GGTGGGCGGG ATGAGGGCCA CGAACGCCAG CACGGCCTGG
    ACGGCCCCGT CCAGCCGCCC TACTCCCGGT GCTTGCGGTC GTGCCGGACC
    ~~~~~
                                tk gene
      . G P L T P P I L A V F A L V A Q P .
1101 GGGGTCATGC TGCCATAAG GTATCGCGCG GCCGGGTAGC ACAGGAGGGC
    CCCAGTACG ACGGGTATTC CATAGCGCGC CGGCCCATCG TGTCCTCCCG
    ~~~~~
                                tk gene
      .. T M S G M L Y R A A P Y C L L A

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

1151 GCGGATGGGA TGGCGGTCGA AGATGAGGGT GAGGGCCGGG GCGGGGGCAT
    CCGCTACCCT ACCGCCAGCT TCTACTCCCA CTCCCGGCCC CCGCCCCGTA
    ~~~~~
                                tk gene
      A I P H R D F I L T L A P P P A H .
1201 GTGAGCTCCC AGCCTCCCCC CCGATATGAG GAGCCAGAAC GCGGTCGGTC
    CACTCGAGGG TCGGAGGGGG GGCTATACTC CTCGGTCTTG CCGCAGCCAG
    ~~~~~
                                tk gene
      . S S G A E G G I H P A L V A D T V .
1251 ACGGCATAAG GCATGCCCAT TGTATCTGG GCGCTTGTC TTACCACCGC
    TGCCGTATTC CGTACGGGTA ACAATAGACC CGCGAACAGT AATGGTGGCG
    ~~~~~
                                tk gene
      .. A Y P M G M T I Q A S T M V V A
1301 CGCGTCCCCG GCCGATATCT CACCCTGGTC GAGGCGGTGT TGTGTGGTGT
    GCGCAGGGGC CGGCTATAGA GTGGGACCAG CTCCGCCACA ACACACCACA
    ~~~~~
                                tk gene
      A D G A S I E G Q D L R H Q T T Y .
1351 AGATGTTTCG GATTGTCTCG GAAGCCCCCA ACACCCGCCA GTAAGTCATC
    TCTACAAGCG CTAACAGAGC CTCGGGGGGT TGTGGGCGGT CATTCACTAG
    ~~~~~
                                tk gene
      . I N A I T E S A G L V R W Y T M P .
1401 GGCTCGGGTA CGTAGACGAT ATCGTCGCGC GAACCCAGGG CCACCAGCAG
    CCGAGCCCAT GCATCTGCTA TAGCAGCGCG CTTGGGTCCC GGTGGTCGTC
    ~~~~~
                                tk gene
      .. E P V Y V I D D R S G L A V L L
1451 TTGCGTGGTG GTGGTTTTCC CCATCCCGTG GGGACCGTCT ATATAAACCC
    AACGCACCAC CACCAAAGG GGTAGGGCAC CCCTGGCAGA TATATTTGGG
    ~~~~~
                                tk gene
      Q T T T T K G M G H P G D I Y V R .
1501 GCAGTAGCGT GGGCATTTTC TGCTCCAGGC GGACTTCCGT GGCTTTTTGT
    CGTCATCGCA CCCGTAAAAG ACGAGGTCCG CCTGAAGGCA CCGAAAAACA
    ~~~~~
                                tk gene
      . L L T P M K Q E L R V E T A K Q Q .
1551 TGCCGGCGAG GCGCAACGC CGTACGTCGG TTGTTATGGC CGCGAGAACG
    ACGGCCGCTC CCGCGTTGCG GCATGCAGCC AACAATACCG GCGCTCTTGC
    ~~~~~
                                tk gene
      .. R R P R L A T R R N N H G R S R

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

1601 CGCAGCCTGG TCGAACGCAG ACGCGTGTG ATGGCAGGGG TACGAAGCCA
    GCGTCGGACC AGCTTGCGTC TGCGCACAAC TACCGTCCCC ATGCTTCGGT
    ~~~~~
                tk gene
    A A Q D F A S A H Q H C P Y S A M
1651 TAGATCCCGT TATCAATTAC TTATACTATC CGGCGCGCAA GCGAGCGTGT
    ATCTAGGGCA ATAGTTAATG AATATGATAG GCCGCGCGTT CGCTCGCACA
    ~~~~~
                ie-0 promoter
1701 GCGCCGGAGC ACAATTGATA CTGATTTACG AGTTGGGCAA ACGGGCTTTA
    CGCGGCCCTCG TGTTAAC TAT GACTAAATGC TCAACCCGTT TGCCCGAAAT
    ~~~~~
                ie-0 promoter
1751 TATAGCCTGT CCCCTCCACA GCCCTAGTGC CGTGCGCAAA GTGCCTACGT
    ATATCGGACA GGGGAGGTGT CGGGATCACG GCACGCGTTT CACGGATGCA
    ~~~~~
                ie-0 promoter
1801 GACCAGGCTC TCCTACGCAT ATACAATCTT ATCTCTATAG ATAAGGTTTC
    CTGGTCCGAG AGGATGCGTA TATGTTAGAA TAGAGATATC TATTCCAAAG
    ~~~~~
                ie-0 promoter
1851 CATATATAAA GCCTCTCGAT GGCTGAACGT GCACAGTATC GTGTTGATTT
    GTATATATTT CGGAGAGCTA CCGACTTGCA CGTGTCATAG CACAAC TAAA
    ~~~~~
                ie-0 promoter
1901 CTGAGTGCTA ACTAACAGTT ACAATGAACC GTTTTTTTTCG AGAGAATAAC
    GACTCACGAT TGATTGTCAA TGTTACTTGG CAAAAAAGC TCTCTTATTG
    ~~~~~
                ie-0 promoter
1951 ATTTTTGACG CGCCAAGGAC CGGGGGCAAG GGTCGTGCCA AATCTTTGCC
    TAAAAACTGC GCGGTTCTTG GCGGGCGTTC CCAGCACGGT TTAGAAACGG
    ~~~~~
                ie-0 promoter
2001 AGCGCCTGCC GCCAACTCGC CGCCGTGCGC TGTTGCTCCG CCGCCAAAAT
    TCGCGGACGG CGGTTGAGCG GCGGCAGCGG ACAAGCAGGC GGCGGTTTTA
    ~~~~~
                ie-0 promoter
2051 CTAACATCAA ACCACCTACG CGCATCTCTC CGCCTAAACA GCCTATGTGC
    GATTGTAGTT TGGTGGATGC GCGTAGAGAG GCGGATTTGT CGGATACACG
    ~~~~~
                ie-0 promoter
2101 ACCTCTCCGG CCAAGCCGTT GGAGCACAGC AGCATTGTAA GTAAAAAACC
    TGGAGAGGCC GGTTCGGCAA CCTCGTGTCTG TCGTAACATT CATTTTTTTG
    ~~~~~
                ie-0 promoter

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

2151 AGTCGTCAAC AGAAAAGATG GATATTTTGT GCCGCCCAGAG TTTGGGAACA
    TCAGCAGTTG TCTTTTCTAC CTATAAAACA CGGCGGGCTC AAACCCCTGT
    ~~~~~
                ie-0 promoter
2201 AGTTTGAAGG TTTGCCCCGCG TACAGCGACA AACTGGATTT CAAACAAGAG
    TCAAACCTTC AAACGGGCGC ATGTCGCTGT TTGACCTAAA GTTTGTTCTC
    ~~~~~
    ie-0 promoter
                                p10 promoter
2251 CGCGATCTAC GTACCTGCAG GCCCGGGCTC AACCCAACAC AATATATTAT
    GCGCTAGATG CATGGACGTC CGGGCCCCAG TTGGGTTGTG TTATATAATA
    ~~~~~
                p10 promoter
2301 AGTTAAATAA GAATTATTAT CAAATCATTT GTATATTAAT TAAAATACTA
    TCAATTTATT CTTAATAATA GTTTAGTAAA CATATAATTA ATTTTATGAT
    ~~~~~
    p10 promoter
                                lacZ
                                M T M I T
2351 TACTGTAAAT TACATTTTAT TTACAATTCA CTCTAGAATG ACCATGATTA
    ATGACATTTA ATGTAAAATA AATGTTAAGT GAGATCTTAC TGGTACTAAT
    ~~~~~
                lacZ
    . D S L A V V L Q R R D W E N P G
2401 CGGATTCACT GGCCGTCGTT TTACAACGTC GTGACTGGGA AAACCCCTGGC
    GCCTAAGTGA CCGGCAGCAA AATGTTGCAG CACTGACCCT TTTGGGACCG
    ~~~~~
                lacZ
    V T Q L N R L A A H P P F A S W R
2451 GTTACCCAAC TTAATCGCCT TGCAGCACAT CCCCCTTTTCG CCAGCTGGCG
    CAATGGGTTG AATTAGCGGA ACGTCGTGTA GGGGGAAAGC GGTGACCGC
    ~~~~~
                lacZ
    . N S E E A R T D R P S Q Q L R S L
2501 TAATAGCGAA GAGGCCCGCA CCGATCGCCC TTCCAACAG TTGCGCAGCC
    ATTATCGCTT CTCCGGGCGT GGCTAGCGGG AAGGGTTGTC AACGCGTCGG
    ~~~~~
                lacZ
    . N G E W R F A W F P A P E A V P
2551 TGAATGGCGA ATGGCGCTTT GCCTGGTTTC CGGCACCAGA AGCGGTGCCG
    ACTTACCGCT TACCGCGAAA CGGACCAAAG GCCGTGGTCT TCGCCACGGC

```

Table 13 (continued) Nucleotide sequence of the V5-His DEST cassette.

```

lacZ
-----
Bsu36I
-----
E S W L E C D L P E A D T V V V P .
2601 GAAAGCTGGC TGGAGTGC GA TCTTCCTGAG GCCGATACTG TCGTCGTCCC
CTTTCGACCG ACCTCAGCT AGAAGGACTC CGGCTATGAC AGCAGCAGGG
lacZ
-----
S N W Q M H G Y D A P I Y T N V T .
2651 CTCAAAGTGG CAGATGCACG GTTACGATGC GCCCATCTAC ACCAACGTAA
GAGTTTGACC GTCTACGTGC CAATGCTACG CGGGTAGATG TGGTTGCATT
lacZ
-----
Y P I T V N P P F V P T E N P T
2701 CCTATCCCAT TACGGTCAAT CCGCCGTTTG TTCCACGGA GAATCCGACG
GGATAGGGTA ATGCCAGTTA GGCGGCAAAC AAGGGTGCCT CTTAGGCTGC
lacZ
-----
G C Y S L T F N V D E S W L Q E G .
2751 GGTGTGTTACT CGCTCACATT TAATGTTGAT GAAAGCTGGC TACAGGAAGG
CCAACAATGA GCGAGTGTA ATTACAATA CTTTCGACCG ATGTCCTTCC
lacZ
-----
Q T R I I F D G V N S A F H L W C .
2801 CCAGACGCGA ATTATTTTGG ATGGCGTTAA CTCGGCGTTT CATCTGTGGT
GGTCTGCGCT TAATAAAAC TACCGCAATT GAGCCGCAA GTAGACACCA
lacZ
-----
N G R W V G Y G Q D S R L P S E
2851 GCAACGGGCG CTGGGTCGGT TACGGCCAGG ACAGTCGTTT GCCGTCTGAA
CGTTGCCCCG GACCCAGCCA ATGCCGCTCC TGTCAGCAA CGGCAGACTT
lacZ
-----
F D L S A F L R A G E N R L A V M .
2901 TTTGACCTGA GCGCATTTTT ACGCGCCGGA GAAAACCGCC TCGCGGTGAT
AAACTGGACT CGCGTAAAAA TGCGCGGCCT CTTTGGCGG AGCGCCACTA
lacZ
-----
V L R W S D G S Y L E D Q D M W R .
2951 GGTGCTGCGT TGGAGTGACG GCAGTTATCT GGAAGATCAG GATATGTGGC
CCACGACGCA ACCTCACTGC CGTCAATAGA CTTCTAGTC CTATACACCG
lacZ
-----
M S G I F R D V S L L H K P T T
3001 GGATGAGCGG CATTTTCCGT GACGTCTCGT TGCTGCATAA ACCGACTACA
CCTACTCGCC GTAAAAGGCA CTGCAGAGCA ACGACGTATT TGGCTGATGT

```


Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

                                lacZ
~~~~~
      Q  I  S  D  F  H  V  A  T  R  F  N  D  D  F  S  R  .
3051 CAAATCAGCG ATTTCCATGT TGCCACTCGC TTTAATGATG ATTTCAGCCG
      GTTTAGTCGC TAAAGGTACA ACGGTGAGCG AAATTACTAC TAAAGTCGGC
                                lacZ
~~~~~
      . A  V  L  E  A  E  V  Q  M  C  G  E  L  R  D  Y  L  .
3101 CGCTGTACTG GAGGCTGAAG TTCAGATGTG CGGCGAGTTG CGTGACTACC
      GCGACATGAC CTCCGACTTC AAGTCTACAC GCCGCTCAAC GCACTGATGG
                                lacZ
~~~~~
      . R  V  T  V  S  L  W  Q  G  E  T  Q  V  A  S  G
3151 TACGGGTAAC AGTTTCTTTA TGGCAGGGTG AAACGCAGGT CGCCAGCGGC
      ATGCCCATTTG TCAAAGAAAT ACCGTCCCAC TTTGCGTCCA GCGGTCGCCG
                                lacZ
~~~~~
      T  A  P  F  G  G  E  I  I  D  E  R  G  G  Y  A  D  .
3201 ACCGCGCCTT TCGGCGGTGA AATTATCGAT GAGCGTGGTG GTTATGCCGA
      TGGCGCGGAA AGCCGCCACT TTAATAGCTA CTCGCACCAC CAATACGGCT
                                lacZ
~~~~~
      . R  V  T  L  R  L  N  V  E  N  P  K  L  W  S  A  E  .
3251 TCGCGTCACA CTACGTCTGA ACGTCGAAAA CCCGAAACTG TGGAGCGCCG
      AGCGCAGTGT GATGCAGACT TGCAGCTTTT GGGCTTTGAC ACCTCGCGGC
                                lacZ
~~~~~
      . I  P  N  L  Y  R  A  V  V  E  L  H  T  A  D  G
3301 AAATCCCGAA TCTCTATCGT GCGGTGGTTG AACTGCACAC CGCCGACGGC
      TTTAGGGCTT AGAGATAGCA CGCCACCAAC TTGACGTGTG GCGGCTGCCG
                                lacZ
~~~~~
      T  L  I  E  A  E  A  C  D  V  G  F  R  E  V  R  I  .
3351 ACGCTGATTG AAGCAGAAGC CTGCGATGTC GGTTTCCGCG AGGTGCGGAT
      TGCGACTAAC TTCGTCTTCG GACGCTACAG CCAAAGGCGC TCCACGCCTA
                                lacZ
~~~~~
      . E  N  G  L  L  L  L  N  G  K  P  L  L  I  R  G  V  .
3401 TGAAAATGGT CTGCTGCTGC TGAACGGCAA GCCGTTGCTG ATTCGAGGCG
      ACTTTTACCA GACGACGACG ACTTGCCGTT CGGCAACGAC TAAGCTCCGC
                                lacZ
~~~~~
      . N  R  H  E  H  H  P  L  H  G  Q  V  M  D  E  Q
3451 TTAACCGTCA CGAGCATCAT CCTCTGCATG GTCAGGTCAT GGATGAGCAG
      AATTGGCAGT GCTCGTAGTA GGAGACGTAC CAGTCCAGTA CCTACTCGTC

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

                                lacZ
~~~~~
      T M V Q D I L L M K Q N N F N A V .
3501 ACGATGGTGC AGGATATCCT GCTGATGAAG CAGAACAAC TTAACGCCGT
      TGCTACCACG TCCTATAGGA CGACTACTTC GTCTTGTTGA AATTGCGGCA
                                lacZ
~~~~~
      . R C S H Y P N H P L W Y T L C D R .
3551 GCGCTGTTTCG CATTATCCGA ACCATCCGCT GTGGTACACG CTGTGCGACC
      CGCGACAAGC GTAATAGGCT TGGTAGGCGA CACCATGTGC GACACGCTGG
                                lacZ
~~~~~
      . Y G L Y V V D E A N I E T H G M
3601 GCTACGGCCT GTATGTGGTG GATGAAGCCA ATATTGAAAC CCACGGCATG
      CGATGCCGGA CATAACCAC CTACTTCGGT TATAACTTTG GGTGCCGTAC
                                lacZ
~~~~~
      V P M N R L T D D P R W L P A M S .
3651 GTGCCAATGA ATCGTCTGAC CGATGATCCG CGCTGGCTAC CGGCGATGAG
      CACGGTTACT TAGCAGACTG GCTACTAGGC GCGACCGATG GCCGCTACTC
                                lacZ
~~~~~
      . E R V T R M V Q R D R N H P S V I .
3701 CGAACGCGTA ACGCGAATGG TGCAGCGCGA TCGTAATCAC CCGAGTGTGA
      GCTTGCGCAT TCGGCTTACC ACGTCGCGCT AGCATTAGTG GGCTCACACT
                                lacZ
~~~~~
      . I W S L G N E S G H G A N H D A
3751 TCATCTGGTC GCTGGGGAAT GAATCAGGCC ACGGCGCTAA TCACGACGCG
      AGTAGACCAG CGACCCCTTA CTTAGTCCGG TGCCGCGATT AGTGCTGCGC
                                lacZ
~~~~~
      L Y R W I K S V D P S R P V Q Y E .
3801 CTGTATCGCT GGATCAAATC TGTCGATCCT TCCCGCCCGG TGCAGTATGA
      GACATAGCGA CCTAGTTTAG ACAGCTAGGA AGGGCGGGCC ACGTCATACT
                                lacZ
~~~~~
      . G G G A D T T A T D I I C P M Y A .
3851 AGGCGGCGGA GCCGACACCA CGGCCACCGA TATTATTGTC CCGATGTACG
      TCCGCCGCTT CGGCTGTGGT GCCGGTGGCT ATAATAAACG GGCTACATGC
                                lacZ
~~~~~
      . R V D E D Q P F P A V P K W S I
3901 CGCGCGTGGA TGAAGACCAG CCCTTCCCGG CTGTGCCGAA ATGGTCCATC
      GCGCGCACCT ACTTCTGGTC GGGAAGGGCC GACACGGCTT TACCAGGTAG

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

                                lacZ
~~~~~
      K K W L S L P G E T R P L I L C E .
3951 AAAAAATGGC TTTCGCTACC TGGAGAGACG CGCCCGCTGA TCCTTTGCGA
      TTTTTTACCG AAAGCGATGG ACCTCTCTGC GCGGGCGACT AGGAAACGCT
                                lacZ
~~~~~
      . Y A H A M G N S L G G F A K Y W Q .
4001 ATACGCCAC GCGATGGGTA ACAGTCTTGG CGGTTTCGCT AAATACTGGC
      TATGCGGGTG CGCTACCCAT TGTCAGAACG GCCAAAGCGA TTTATGACCG
                                lacZ
~~~~~
      . A F R Q Y P R L Q G G F V W D W
4051 AGGCGTTTTG TCAGTATCCC CGTTTACAGG GCGGCTTCGT CTGGGACTGG
      TCCGCAAAGC AGTCATAGGG GCAAATGTCC CGCCGAAGCA GACCCTGACC
                                lacZ
~~~~~
      V D Q S L I K Y D E N G N P W S A .
4101 GTGGATCAGT CGCTGATTAA ATATGATGAA AACGGCAACC CGTGGTCGGC
      CACCTAGTCA GCGACTAATT TATACTACTT TTGCCGTTGG GCACCAGCCG
                                lacZ
~~~~~
      . Y G G D F G D T P N D R Q F C M N .
4151 TTACGGCGGT GATTTTGGCG ATACGCCGAA CGATCGCCAG TTCTGTATGA
      AATGCCGCCA CTAAAACCGC TATGCGGCTT GCTAGCGGTC AAGACATACT
                                lacZ
~~~~~
      . G L V F A D R T P H P A L T E A
4201 ACGGTCTGGT CTTTGCCGAC CGCACGCCGC ATCCAGCGCT GACGGAAGCA
      TGCCAGACCA GAAACGGCTG GCGTGCGGCG TAGGTCGCGA CTGCCTTCGT
                                lacZ
~~~~~
      K H Q Q Q F F Q F R L S G Q T I E .
4251 AAACACCAGC AGCAGTTTTT CCAGTTCCGT TTATCCGGGC AAACCATCGA
      TTTGTGGTCG TCGTCAAAAA GGTCAAGGCA AATAGGCCCG TTTGGTAGCT
                                lacZ
~~~~~
      . V T S E Y L F R H S D N E L L H W .
4301 AGTGACCAGC GAATACCTGT TCCGTCATAG CGATAACGAG CTCCTGCACT
      TCACTGGTCG CTTATGGACA AGGCAGTATC GCTATTGCTC GAGGACGTGA
                                lacZ
~~~~~
      . M V A L D G K P L A S G E V P L
4351 GGATGGTGGC GCTGGATGGT AAGCCGCTGG CAAGCGGTGA AGTGCCTCTG
      CCTACCACCG CGACCTACCA TTCGGCGACC GTTCGCCACT TCACGGAGAC

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

                                lacZ
~~~~~
  D V A P Q G K Q L I E L P E L P Q .
4401 GATGTCGCTC CACAAGGTAA ACAGTTGATT GAACTGCCTG AACTACCGCA
     CTACAGCGAG GTGTTCCATT TGTCAACTAA CTTGACGGAC TTGATGGCGT
                                lacZ
~~~~~
  . P E S A G Q L W L T V R V V Q P N .
4451 GCCGGAGAGC GCCGGGCAAC TCTGGCTCAC AGTACGCGTA GTGCAACCGA
     CGGCCTCTCG CGGCCCGTTG AGACCGAGTG TCATGCGCAT CACGTTGGCT
                                lacZ
~~~~~
  . A T A W S E A G H I S A W Q Q W
4501 ACGCGACCGC ATGGTCAGAA GCCGGGCACA TCAGCGCCTG GCAGCAGTGG
     TCGCTGGCG TACCAGTCTT CGGCCCGTGT AGTCGCGGAC CGTCGTCACC
                                lacZ
~~~~~
  R L A E N L S V T L P A A S H A I .
4551 CGTCTGGCGG AAAACCTCAG TGTGACGCTC CCCGCCGCGT CCCACGCCAT
     GCAGACCGCC TTTTGGAGTC ACACTGCGAG GGGCGGCGCA GGGTGGCGTA
                                lacZ
~~~~~
  . P H L T T S E M D F C I E L G N K .
4601 CCCGCATCTG ACCACCAGCG AAATGGATTT TTGCATCGAG CTGGGTAATA
     GGGCGTAGAC TGGTGGTTCG TTTACCTAAA AACGTAGCTC GACCCATTAT
                                lacZ
~~~~~
  . R W Q F N R Q S G F L S Q M W I
4651 AGCGTTGGCA ATTTAACCGC CAGTCAGGCT TTCTTTCACA GATGTGGATT
     TCGCAACCGT TAAATTGGCG GTCAGTCCGA AAGAAAGTGT CTACACCTAA
                                lacZ
~~~~~
  G D K K Q L L T P L R D Q F T R A .
4701 GGCGATAAAA AACAACTGCT GACGCCGCTG CGCGATCAGT TCACCCGTGC
     CCGCTATTTT TTGTTGACGA CTGCGGCGAC GCGTAGTCA AGTGGGCACG
                                lacZ
~~~~~
  . P L D N D I G V S E A T R I D P N .
4751 ACCGCTGGAT AACGACATTG GCGTAAGTGA AGCGACCCGC ATTGACCCTA
     TGGCGACCTA TTGCTGTAAC CGCATTCACT TCGCTGGGCG TAACTGGGAT
                                lacZ
~~~~~
  . A W V E R W K A A G H Y Q A E A
4801 ACGCCTGGGT CGAACGCTGG AAGGCGGCGG GCCATTACCA GGCCGAAGCA
     TGCGGACCCA GCTTGCGACC TTCCGCCGCC CGGTAATGGT CCGGCTTCGT

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

lacZ
~~~~~
  A L L Q C T A D T L A D A V L I T .
4851 GCGTTGTTGC AGTGACGGC AGATACTT GCTGATGCGG TGCTGATTAC
    CGCAACAACG TCACGTGCCG TCTATGTGAA CGACTACGCC ACGACTAATG
      lacZ
~~~~~
  . T A H A W Q H Q G K T L F I S R K .
4901 GACCGCTCAC GCGTGGCAGC ATCAGGGGAA AACCTTATTT ATCAGCCGGA
    CTGGCGAGTG CGCACCCTCG TAGTCCCCTT TTGGAATAAA TAGTCGGCCT
      lacZ
~~~~~
  . T Y R I D G S G Q M A I T V D V
4951 AAACCTACCG GATTGATGGT AGTGGTCAAA TGGCGATTAC CGTTGATGTT
    TTTGGATGGC CTAACCTACCA TCACCAGTTT ACCGCTAATG GCAACTACAA
      lacZ
~~~~~
  E V A S D T P H P A R I G L N C Q .
5001 GAAGTGGCGA GCGATACACC GCATCCGGCG CGGATTGGCC TGAACCTCCA
    CTTACCCGCT CGCTATGTGG CGTAGGCCGC GCCTAACCGG ACTTGACGGT
      lacZ
~~~~~
  . L A Q V A E R V N W L G L G P Q E .
5051 GCTGGCGCAG GTAGCAGAGC GGGTAAACTG GCTCGGATTA GGGCCGCAAG
    CGACCGCGTC CATCGTCTCG CCCATTTGAC CGAGCCTAAT CCCGGCGTTC
      lacZ
~~~~~
  . N Y P D R L T A A C F D R W D L
5101 AAACTATCC CGACCGCCTT ACTGCCGCCT GTTTTGACCG CTGGGATCTG
    TTTTGATAGG GCTGGCGGAA TGACGGCGGA CAAACTGGC GACCCTAGAC
      lacZ
~~~~~
  P L S D M Y T P Y V F P S E N G L .
5151 CCATTGTCAG ACATGTATAC CCCGTACGTC TTCCCGAGCG AAAACGGTCT
    GGTAACAGTC TGTACATATG GGGCATGCAG AAGGGCTCGC TTTTGCCAGA
      lacZ
~~~~~
  . R C G T R E L N Y G P H Q W R G D .
5201 GCGCTGCGGG ACGCGCGAAT TGAATTATGG CCCACACCAG TGGCGCGGCG
    CGCGACGCCC TGCGCGCTTA ACTTAATACC GGGTGTGGTC ACCGCGCCGC
      lacZ
~~~~~
  . F Q F N I S R Y S Q Q Q L M E T
5251 ACTTCCAGTT CAACATCAGC CGCTACAGTC AACAGCAACT GATGGAAACC
    TGAAGGTCAA GTTGTAGTCG GCGATGTCAG TTGTCGTTGA CTACCTTTTG

```

Table 13 (continued). Nucleotide sequence of the V5-His DEST cassette.

```

                                lacZ
~~~~~
      S  H  R  H  L  L  H  A  E  E  G  T  W  L  N  I  D  .
5301 AGCCATCGCC ATCTGCTGCA CGCGGAAGAA GGCACATGGC TGAATATCGA
      TCGGTAGCGG TAGACGACGT GCGCCTTCTT CCGTGTACCG ACTTATAGCT
                                lacZ
~~~~~
      . G  F  H  M  G  I  G  G  D  D  S  W  S  P  S  V  S  .
5351 CGGTTTCCAT ATGGGGATTG GTGGCGACGA CTCCTGGAGC CCGTCAGTAT
      GCCAAAGGTA TACCCCTAAC CACCGCTGCT GAGGACCTCG GGCAGTCATA
                                lacZ
~~~~~
      . A  E  F  Q  L  S  A  G  R  Y  H  Y  Q  L  V  W
5401 CGGCGGAATT CCAGCTGAGC GCCGGTCGCT ACCATTACCA GTTGGTCTGG
      GCCGCCTTAA GGTCGACTCG CGGCCAGCGA TGGTAATGGT CAACCAGACC
      lacZ                               Attr2
~~~~~
      C  Q  K
5451 TGTCAAAAAT AATGACTGCA GGTCGACCAT AGTGA CTGGA TATGTTGTGT
      ACAGTTTTTA TTA CTGACGT CCAGCTGGTA TCA CTGACCT ATACAACACA
                                Attr2
~~~~~
5501 TTTACAGTAT TATGTAGTCT GTTTTTTATG CAAAATCTAA TTTAATATAT
      AAATGTCATA ATACATCAGA CAAAAAATAC GTTTTAGATT AAATTATATA
                                Attr2
~~~~~
5551 TGATATTTAT ATCATTTTAC GTTTCTCGTT CAGCTTTCTT GTACAAAGTG
      ACTATAAATA TAGTAAAATG CAAAGAGCAA GTCGAAAGAA CATGTTTCAC
      Attr2                               V5/His
~~~~~
                                G  K  P  I  P  N  P  L  L  G  .
5601 GTGAGAATGA ATGAAGATCT GGGGAAGCCT ATCCCTAACC CTCTCCTCGG
      CACTCTTACT TACTTCTAGA CCCCTTCGGA TAGGGATTGG GAGAGGAGCC
                                V5/His
~~~~~
      . L  D  S  T  R  T  G  H  H  H  H  H  H
5651 TCTCGATTCT ACGCGTACCG GTCATCATCA CCATCACCAT TGA
      AGAGCTAAGA TGCGCATGGC CAGTAGTAGT GGTAGTGGTA ACT

```

Table 14. Nucleotide sequence of the Mel/V5-His DEST cassette.

```

ph promoter
~~~~~
1 ATAAGTATTT TACTGTTTTTC GTAACAGTTT TGTAATAAAA AAACCTATAA
  TATTCATAAA ATGACAAAAG CATTGTCAAA ACATTATTTT TTTGGATATT
51 ATATTCCGGA TTATTCATAC CGTCCCACCA TCGGGCGCGG ATCCTATAAA
  TATAAGGCCT AATAAGTATG GCAGGGTGGT AGCCCGCGCC TAGGATATTT
      Melittin signal
      ~~~~~
      M K F L V N V A L V F M V V Y I S .
101 TATGAAATTC TTAGTCAACG TTGCCCTTGT TTTTATGGTC GTATACATTT
  ATACTTTAAG AATCAGTTGC AACGGGAACA AAAATACCAG CATATGTAAA
  Melittin signal                                attR1
  ~~~~~
  . Y I Y A
151 CTTACATCTA TGCGGCATGG TCGAATCAAA CAAGTTTGTA CAAAAAGCT
  GAATGTAGAT ACGCCGTACC AGCTTAGTTT GTTCAAACAT GTTTTTTCGA
      attR1
      ~~~~~
201 GAACGAGAAA CGTAAAATGA TATAAATATC AATATATTAA ATTAGATTTT
  CTTGCTCTTT GCATTTTACT ATATTATAG TTATATAATT TAATCTAAAA
      attR1
      ~~~~~
251 GCATAAAAAA CAGACTACAT AATACTGTAA AACACAACAT ATCCAGTCAC
  CGTATTTTTT GTCTGATGTA TTATGACATT TTGTGTTGTA TAGGTCAGTG
301 TATGGCGGCC GCTCCCTAAC CCACGGGGCC CGTGGCTATG GCAGGGCTTG
  ATACCGCCCGG CGAGGGATTG GGTGCCCCGG GCACCGATAC CGTCCCGAAC
351 CCGCCCCGAC GTTGGCTGCG AGCCCTGGGC CTTACCCGA ACTTGGGGGT
  GGCGGGGCTG CAACCGACGC TCGGGACCCG GAAGTGGGCT TGAACCCCA
401 TGGGGTGGGG AAAAGGAAGA AACGCGGGCG TATTGGTCCC AATGGGGTCT
  ACCCCACCCC TTTTCCTTCT TTGCGCCCGC ATAACCAGGG TTACCCAGA
451 CGGTGGGGTA TCGACAGAGT GCCAGCCCTG GGACCGAACC CCGCGTTTAT
  GCCACCCCAT AGCTGTCTCA CGGTCGGGAC CCTGGCTTGG GGCGCAAATA
501 GAACAAACGA CCCAACACCC GTGCGTTTTA TTCTGTCTTT TTATTGCCGT
  CTTGTTTGCT GGGTTGTGGG CACGCAAAAT AAGACAGAAA AATAACGGCA
551 CATAGCGCGG GTTCCTTCCG GTATTGTCTC CTTCCGTGTT TCAGTTAGCC
  GTATCGCGCC CAAGGAAGGC CATAACAGAG GAAGGCACAA AGTCAATCGG
      ~~~
      tk gene
      N A E .
601 TCCCCCATCT CCCGGGCAAA CGTGC GCGCC AGGTCGCAGA TCGTCGGTAT
  AGGGGGTAGA GGGCCCGTTT GCACGCGCGG TCCAGCGTCT AGCAGCCATA
      ~~~~~
      tk gene
      .. G M E R A F T R A L D C I T P I

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

651 GGAGCCTGGG GTGGTGACGT GGGTCTGGAC CATCCCGGAG GTAAGTTGCA
    CCTCGGACCC CACCACTGCA CCCAGACCTG GTAGGGCCTC CATTCAACGT
    ~~~~~
          tk gene
    S G P T T V H T Q V M G S T L Q L .
701 GCAGGGCGTC CCGGCAGCCG GCGGGCGATT GGTCGTAATC CAGGATAAAG
    CGTCCCGCAG GGCCGTCGGC CGCCCGCTAA CCAGCATTAG GTCCTATTTT
    ~~~~~
          tk gene
    . L A D R C G A P S Q D Y D L I F V .
751 ACATGCATGG GACGGAGGCG TTTGGCCAAG ACGTCCAAAG CCCAGGCAAA
    TGTACGTACC CTGCCTCCGC AAACCGGTTT TGCAGGTTTC GGGTCCGTTT
    ~~~~~
          tk gene
    . . H M P R L R K A L V D L A W A F
801 CACGTTATAC AGGTCGCCGT TGGGGGCCAG CAACTCGGGG GCCCGAAACA
    GTGCAATATG TCCAGCGGCA ACCCCCGGTC GTTGAGCCCC CGGGCTTTGT
    ~~~~~
          tk gene
    V N Y L D G N P A L L E P A R F L .
851 GGGTAAATAA CGTGTCCCCG ATATGGGGTC GTGGGCCCCG GTTGCTCTGG
    CCCATTATTT GCACAGGGGC TATACCCAG CACCCGGGCG CAACGAGACC
    ~~~~~
          tk gene
    . T F L T D G I H P R P G A N S Q P .
901 GGCTCGGCAC CCTGGGGCGG CACGGCCGCC CCCGAAAGCT GTCCCCAATC
    CCGAGCCGTG GGACCCCGCC GTGCCGGCGG GGGCTTTCGA CAGGGGTTAG
    ~~~~~
          tk gene
    . . E A G Q P P V A A G S L Q G W D
951 CTCCCGCCAC GACCCGCCGC CCTGCAGATA CCGCACCGTA TTGGCAAGCA
    GAGGGCGGTG CTGGGCGGCG GGACGTCTAT GGCGTGGCAT AACCGTTCGT
    ~~~~~
          tk gene
    E R W S G G G Q L Y R V T N A L L .
1001 GCCCATAAAC GCGGCGAATC GCGGCCAGCA TAGCCAGGTC AAGCCGCTCG
    CGGGTATTTG CGCCGCTTAG CGCCGTCGT ATCGGTCCAG TTCGGCGAGC
    ~~~~~
          tk gene
    . G Y V R R I A A L M A L D L R E G .
1051 CCGGGGCGCT GGCCTTTGGC CAGGCGGTCT ATGTGTCTGT CCTCCGGAAG
    GGCCCCGCGA CCGCAAACCG GTCCGCCAGC TACACAGACA GGAGGCCTTC
    ~~~~~
          tk gene
    . . P R Q R K A L R D I H R D E P L

```


Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

1101 GGGCCCCAAC ACGATGTTTG TGCCGGGCAA GGTCGGCGGG ATGAGGGCCA
    CCGGGGGTTG TGCTACAAAC ACGGCCCGTT CCAGCCGCCC TACTCCCGGT
    ~~~~~
                                tk gene
    A G L V I N T G P L T P P I L A V .
1151 CGAACGCCAG CACGGCCTGG GGGGTCATGC TGCCATAAG GTATCGCGCG
    GCTTGCGGTC GTGCCGGACC CCCAGTACG ACGGGTATTC CATAGCGCGC
    ~~~~~
                                tk gene
    . F A L V A Q P T M S G M L Y R A A .
1201 GCCGGGTAGC ACAGGAGGGC GGCGATGGGA TGGCGGTCGA AGATGAGGGT
    CGGCCCATCG TGTCTCCCG CCGCTACCCT ACCGCCAGCT TCTACTCCCA
    ~~~~~
                                tk gene
    . . P Y C L L A A I P H R D F I L T
1251 GAGGGCCGGG GGCGGGGCGT GTGAGCTCCC AGCCTCCCCC CCGATATGAG
    CTCCCGGCCC CCGCCCCGTA CACTCGAGGG TCGGAGGGGG GGCTATACTC
    ~~~~~
                                tk gene
    L A P P P A H S S G A E G G I H P .
1301 GAGCCAGAAC GGCGTCGGTC ACGGCATAAG GCATGCCCAT TGTTATCTGG
    CTCGGTCTTG CCGCAGCCAG TGCCGTATTC CGTACGGGTA ACAATAGACC
    ~~~~~
                                tk gene
    . A L V A D T V A Y P M G M T I Q A .
1351 GCGCTTGTC TTACCACCGC CGCGTCCCCG GCCGATATCT CACCCTGGTC
    CGCGAACAGT AATGGTGGCG GCGCAGGGGC CGGCTATAGA GTGGGACCAG
    ~~~~~
                                tk gene
    . . S T M V V A A D G A S I E G Q D
1401 GAGGCGGTGT TGTGTGGTGT AGATGTTTCG GATTGTCTCG GAAGCCCCCA
    CTCCGCCACA ACACACCACA TCTACAAGCG CTAACAGAGC CTTGCGGGGT
    ~~~~~
                                tk gene
    L R H Q T T Y I N A I T E S A G L .
1451 ACACCCGCCA GTAAGTCATC GGCTCGGGTA CGTAGACGAT ATCGTCGCGC
    TGTGGGCGGT CATTAGTAG CCGAGCCCAT GCATCTGCTA TAGCAGCGCG
    ~~~~~
                                tk gene
    . V R W Y T M P E P V Y V I D D R S .
1501 GAACCCAGGG CCACCAGCAG TTGCGTGGTG GTGGTTTTC CCATCCCGTG
    CTTGGGTCCC GGTGGTCGTC AACGCACCAC CACCAAAAGG GGTAGGGCAC
    ~~~~~
                                tk gene
    . . G L A V L L Q T T T T K G M G H

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

1551 GGGACCGTCT ATATAAACCC GCAGTAGCGT GGGCATTTTC TGCTCCAGGC
    CCCTGGCAGA TATATTTGGG CGTCATCGCA CCCGTAAAAG ACGAGGTCCG
    ~~~~~
                                tk gene
      P G D I Y V R L L T P M K Q E L R .
1601 GGACTTCCGT GGCTTTTGT TGCCGGCGAG GGC GCAACGC CGTACGTCGG
    CCTGAAGCA CCGAAAAACA ACGGCCGCTC CCGCGTTGCG GCATGCAGCC
    ~~~~~
                                tk gene
      . V E T A K Q Q R R P R L A T R R N .
1651 TTGTTATGGC CGCGAGAACG CGCAGCCTGG TCGAACGCAG ACGCGTGTTC
    AACAATACCG GCGCTCTTGC GCGTCGGACC AGCTTGCGTC TGCGCACAAC
    ~~~~~
                                tk gene
      . . N H G R S R A A Q D F A S A H Q
1701 ATGGCAGGGG TACGAAGCCA TAGATCCCGT TATCAATTAC TTATACTATC
    TACCGTCCCC ATGCTTCGGT ATCTAGGGCA ATAGTTAATG AATATGATAG
    ~~~~~
                                tk gene
      pr
      H C P Y S A M
1751 CGGCGCGCAA GCGAGCGTGT GCGCCGGAGC ACAATTGATA CTGATTTACG
    GCCGCGCGTT CGCTCGCACA CGCGGCCTCG TGTAACTAT GACTAAATGC
    ~~~~~
                                ie-0 pr
1801 AGTTGGGCAA ACGGGCTTTA TATAGCCTGT CCCCTCCACA GCCCTAGTGC
    TCAACCCGTT TGCCCGAAAT ATATCGGACA GGGGAGGTGT CGGGATCACG
    ~~~~~
                                ie-0 pr
1851 CGTGCGCAAA GTGCCTACGT GACCAGGCTC TCCTACGCAT ATACAATCTT
    GCACGCGTTT CACGGATGCA CTGGTCCGAG AGGATGCGTA TATGTTAGAA
    ~~~~~
                                ie-0 pr
1901 ATCTCTATAG ATAAGGTTTC CATATATAAA GCCTCTCGAT GGCTGAACGT
    TAGAGATATC TATTCCAAAG GTATATATTT CGGAGAGCTA CCGACTTGCA
    ~~~~~
                                ie-0 pr
1951 GCACAGTATC GTGTTGATTT CTGAGTGCTA ACTAACAGTT ACAATGAACC
    CGTGTCATAG CACAACATAA GACTCACGAT TGATTGTCAA TGTTACTTGG
    ~~~~~
                                ie-0 pr
2001 GTTTTTTTTCG AGAGAATAAC ATTTTGTACG CGCCAAGGAC CGGGGGCAAG
    CAAAAAAGC TCTCTTATTG TAAAACTGC GCGGTTCTCTG GCCCCCGTTC
    ~~~~~
                                ie-0 pr
2051 GGTCTGCGCA AATCTTTGCC AGCGCCTGCC GCCAACTCGC CGCCGTCGCC
    CCAGCACGGT TTAGAAACGG TCGCGGACGG CGGTTGAGCG GCGGCAGCGG
    ~~~~~
                                ie-0 pr

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```
2101 TGTTCGTCCG CCGCCAAAAT CTAACATCAA ACCACCTACG CGCATCTCTC
    ACAAGCAGGC GCGGGTTTTA GATTGTAGTT TGGTGGATGC GCGTAGAGAG
    ~~~~~~
                ie-0 pr
2151 CGCCTAAACA GCCTATGTGC ACCTCTCCGG CCAAGCCGTT GGAGCACAGC
    GCGGATTTGT CGGATACACG TGGAGAGGCC GGTTCCGGCAA CCTCGTGTCTG
    ~~~~~~
                ie-0 pr
2201 AGCATTGTAA GTAAAAAACC AGTCGTCAAC AGAAAAGATG GATATTTTGT
    TCGTAACATT CATTTTTTGG TCAGCAGTTG TCTTTTCTAC CTATAAAACA
    ~~~~~~
                ie-0 pr
2251 GCCGCCCCGAG TTTGGGAACA AGTTTGAAGG TTTGCCCCGCG TACAGCGACA
    CGGCGGGCTC AAACCCTTGT TCAAACTTCC AAACGGGCGC ATGTCGCTGT
    ~~~~~~
```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

                                ie-0 pr
                                p10
pr
2301 AACTGGATTT CAAACAAGAG CGCGATCTAC GTACCTGCAG GCCCGGGGCTC
    TTGACCTAAA GTTTGTTCTC GCGCTAGATG CATGGACGTC CGGGCCCGAG
    ~~~~~
                                ie-0 pr
                                p10 pr
2351 AACCCAACAC AATATATTAT AGTTAAATAA GAATTATTAT CAAATCATTT
    TTGGGTTGTG TTATATAATA TCAATTTATT CTTAATAATA GTTTAGTAAA
    p10 pr
2401 GTATATTAAT TAAAATACTA TACTGTAAAT TACATTTTAT TTACAATTCA
    CATATAATTA ATTTTATGAT ATGACATTTA ATGTAAAATA AATGTTAAGT
    lacZ
    ~~~~~
    M T M I T D S L A V V L Q R R
2451 CTCTAGAATG ACCATGATTA CGGATTCACT GGCCGTCGTT TTACAACGTC
    GAGATCTTAC TGGTACTAAT GCCTAAGTGA CCGGCAGCAA AATGTTGCAG
    lacZ
    ~~~~~
    D W E N P G V T Q L N R L A A H
2501 GTGACTGGGA AAACCCTGGC GTTACCCAAC TTAATCGCCT TGCAGCACAT
    CACTGACCCT TTTGGGACCG CAATGGGTTG AATTAGCGGA ACGTCGTGTA
    lacZ
    ~~~~~
    P P F A S W R N S E E A R T D R P
2551 CCCCTTTTCG CCAGCTGGCG TAATAGCGAA GAGGCCCGCA CCGATCGCCC
    GGGGGAAAGC GGTCGACCGC ATTATCGCTT CTCCGGGCGT GGCTAGCGGG
    lacZ
    ~~~~~
    S Q Q L R S L N G E W R F A W F P
2601 TTCCCAACAG TTGCGCAGCC TGAATGGCGA ATGGCGCTTT GCCTGGTTTC
    AAGGGTTGTC AACGCGTCGG ACTTACCGCT TACCGCGAAA CGGACCAAAG
    lacZ
    ~~~~~
                                Bsu36I
                                ~~~~~
    A P E A V P E S W L E C D L P E
2651 CGGCACCAGA AGCGGTGCCG GAAAGCTGGC TGGAGTGCGA TCTTCCTGAG
    CCCGTGGTCT TCGCCACGGC CTTTCGACCG ACCTCACGCT AGAAGGACTC

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

lacZ
~~~~~
Bsu36I
~
  A D T V V V P S N W Q M H G Y D A .
2701 GCCGATACTG TCGTCGTCCC CTCAAAC TGG CAGATGCACG GTTACGATGC
    CGGCTATGAC AGCAGCAGGG GAGTTTGACC GTCTACGTGC CAATGCTACG
          lacZ
~~~~~
  . P I Y T N V T Y P I T V N P P F V .
2751 GCCCATCTAC ACCAACGTAA CCTATCCCAT TACGGTCAAT CCGCCGTTTG
    CGGGTAGATG TGGTTGCATT GGATAGGGTA ATGCCAGTTA GCGCGCAAAC
          lacZ
~~~~~
  . P T E N P T G C Y S L T F N V D
2801 TTCCACGGA GAATCCGACG GGTGTTACT CGCTCACATT TAATGTTGAT
    AAGGGTGCCT CTTAGGCTGC CCAACAATGA GCGAGTGTA ATTACAATA
          lacZ
~~~~~
  E S W L Q E G Q T R I I F D G V N .
2851 GAAAGCTGGC TACAGGAAGG CCAGACGCGA ATTATTTTGT ATGGCGTTAA
    CTTTCGACCG ATGTCCTTCC GGTCTGCGCT TAATAAAAAC TACCGCAATT
          lacZ
~~~~~
  . S A F H L W C N G R W V G Y G Q D .
2901 CTCGGCGTTT CATCTGTGGT GCAACGGGCG CTGGGTCGGT TACGGCCAGG
    GAGCCGCAAA GTAGACACCA CGTTGCCCGC GACCCAGCCA ATGCCGGTCC
          lacZ
~~~~~
  . S R L P S E F D L S A F L R A G
2951 ACAGTCGTTT GCCGTCTGAA TTTGACCTGA GCGCATTTTT ACGCGCCGGA
    TGTCAGCAAA CGGCAGACTT AAAC TGGACT CGCGTAAAAA TGCGCGGCCT
          lacZ
~~~~~
  E N R L A V M V L R W S D G S Y L .
3001 GAAAACCGCC TCGCGGTGAT GGTGCTGCGT TGGAGTGACG GCAGTTATCT
    CTTTTGGCGG AGCGCCACTA CCACGACGCA ACCTCACTGC CGTCAATAGA
          lacZ
~~~~~
  . E D Q D M W R M S G I F R D V S L .
3051 GGAAGATCAG GATATGTGGC GGATGAGCGG CATTTTCCGT GACGTCTCGT
    CCTTCTAGTC CTATACACCG CCTACTCGCC GTAAAAGGCA CTGCAGAGCA
          lacZ
~~~~~
  . L H K P T T Q I S D F H V A T R
3101 TGCTGCATAA ACCGACTACA CAAATCAGCG ATTTCCATGT TGCCACTCGC
    ACGACGTATT TGGCTGATGT GTTAGTCGC TAAAGGTACA ACGGTGAGCG

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

lacZ
~~~~~
      F N D D F S R A V L E A E V Q M C .
3151 TTTAATGATG ATTTTCAGCCG CGCTGTACTG GAGGCTGAAG TTCAGATGTG
      AAATTACTAC TAAAGTCGGC GCGACATGAC CTCCGACTTC AAGTCTACAC
lacZ
~~~~~
      . G E L R D Y L R V T V S L W Q G E .
3201 CGGCGAGTTG CGTGACTACC TACGGGTAAAC AGTTTCTTTA TGGCAGGGTG
      GCCGCTCAAC GCACTGATGG ATGCCCATTTG TCAAAGAAAT ACCGTCCCAC
lacZ
~~~~~
      . T Q V A S G T A P F G G E I I D
3251 AAACGCAGGT CGCCAGCGGC ACCGCGCCTT TCGGCGGTGA AATTATCGAT
      TTTGCGTCCA GCGGTCGCCG TGGCGCGGAA AGCCGCCACT TTAATAGCTA
lacZ
~~~~~
      E R G G Y A D R V T L R L N V E N .
3301 GAGCGTGGTG GTTATGCCGA TCGCGTCACA CTACGTCTGA ACGTCGAAAA
      CTCGCACCAC CAATACGGCT AGCGCAGTGT GATGCAGACT TGCAGCTTTT
lacZ
~~~~~
      . P K L W S A E I P N L Y R A V V E .
3351 CCCGAAACTG TGGAGCGCCG AAATCCCGAA TCTCTATCGT GCGGTGGTTG
      GGGCTTTGAC ACCTCGCGGC TTTAGGGCTT AGAGATAGCA CGCCACCAAC
lacZ
~~~~~
      . L H T A D G T L I E A E A C D V
3401 AACTGCACAC CGCCGACGGC ACGCTGATTG AAGCAGAAGC CTGCGATGTC
      TTGACGTGTG GCGGCTGCCG TCGGACTAAC TTCGTCTTCG GACGCTACAG
lacZ
~~~~~
      G F R E V R I E N G L L L L N G K .
3451 GGTTCGCGC AGGTGCGGAT TGAATGGT CTGCTGCTGC TGAACGGCAA
      CCAAAGGCGC TCCACGCCTA ACTTTTACCA GACGACGACG ACTTGCCGTT
lacZ
~~~~~
      . P L L I R G V N R H E H H P L H G .
3501 GCCGTTGCTG ATTCGAGGCG TTAACCGTCA CGAGCATCAT CCTCTGCA TG
      CGGCAACGAC TAAGCTCCGC AATTGGCAGT GCTCGTAGTA GGAGACGTAC
lacZ
~~~~~
      . Q V M D E Q T M V Q D I L L M K
3551 GTCAGGTCAT GGATGAGCAG ACGATGGTGC AGGATATCCT GCTGATGAAG
      CAGTCCAGTA CCTACTCGTC TGCTACCACG TCCTATAGGA CGACTACTTC

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

lacZ

~~~~~

Q N N F N A V R C S H Y P N H P L .  
 3601 CAGAACAAC TTAACGCCGT GCGCTGTTTCG CATTATCCGA ACCATCCGCT  
 GTCTTGTTGA AATTGCGGCA CGCGACAAGC GTAATAGGCT TGGTAGGCGA  
 lacZ

~~~~~

· W Y T L C D R Y G L Y V V D E A N .
 3651 GTGGTACACG CTGTGCGACC GCTACGGCCT GTATGTGGTG GATGAAGCCA
 CACCATGTGC GACACGCTGG CGATGCCGGA CATAACCAC CTACTTCGGT
 lacZ

~~~~~

· I E T H G M V P M N R L T D D P  
 3701 ATATTGAAAC CCACGGCATG GTGCCAATGA ATCGTCTGAC CGATGATCCG  
 TATAACTTTG GGTGCCGTAC CACGGTTACT TAGCAGACTG GCTACTAGGC  
 lacZ

~~~~~

R W L P A M S E R V T R M V Q R D .
 3751 CGCTGGCTAC CGGCGATGAG CGAACGCGTA ACGCGAATGG TGCAGCGCGA
 GCGACCGATG GCCGCTACTC GCTTGCGCAT TGCCTTACC ACGTCGCGCT
 lacZ

~~~~~

· R N H P S V I I W S L G N E S G H .  
 3801 TCGTAATCAC CCGAGTGTGA TCATCTGGTC GCTGGGGAAT GAATCAGGCC  
 AGCATTAGTG GGCTCACACT AGTAGACCAG CGACCCCTTA CTTAGTCCGG  
 lacZ

~~~~~

· G A N H D A L Y R W I K S V D P
 3851 ACGGCGCTAA TCACGACGCG CTGTATCGCT GGATCAAATC TGTCGATCCT
 TGCCGCGATT AGTGCTGCGC GACATAGCGA CCTAGTTTAG ACAGCTAGGA
 lacZ

~~~~~

S R P V Q Y E G G G A D T T A T D .  
 3901 TCCGCCCCGG TGCAGTATGA AGGCGGCGGA GCCGACACCA CGGCCACCGA  
 AGGGCGGGCC ACGTCATACT TCCGCCGCCT CGGCTGTGGT GCCGGTGGCT  
 lacZ

~~~~~

· I I C P M Y A R V D E D Q P F P A .
 3951 TATTATTTGC CCGATGTACG CGCGCGTGGA TGAAGACCAG CCCTTCCCGG
 ATAATAAACG GGCTACATGC GCGCGCACCT ACTTCTGGTC GGGAAGGGCC
 lacZ

~~~~~

· V P K W S I K K W L S L P G E T  
 4001 CTGTGCCGAA ATGGTCCATC AAAAAATGGC TTTCGCTACC TGGAGAGACG  
 GACACGGCTT TACCAGGTAG TTTTACC G AAGCGATGG ACCTCTCTGC

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

                                lacZ
~~~~~
 R P L I L C E Y A H A M G N S L G .
4051 CGCCCGCTGA TCCTTTGCGA ATACGCCCAC GCGATGGGTA ACAGTCTTGG
 GCGGGCGACT AGGAAACGCT TATGCGGGTG CGCTACCCAT TGTCAGAACC
 lacZ
~~~~~
  . G F A K Y W Q A F R Q Y P R L Q G .
4101 CGGTTTCGCT AAATACTGGC AGGCGTTTCG TCAGTATCCC CGTTTACAGG
    GCCAAAGCGA TTTATGACCG TCCGCAAAGC AGTCATAGGG GCAAATGTCC
                                lacZ
~~~~~
 . G F V W D W V D Q S L I K Y D E
4151 GCGGCTTCGT CTGGGACTGG GTGGATCAGT CGCTGATTAA ATATGATGAA
 CGCCGAAGCA GACCCTGACC CACCTAGTCA GCGACTAATT TATACTACTT
 lacZ
~~~~~
  N G N P W S A Y G G D F G D T P N .
4201 AACGGCAACC CGTGGTCGGC TTACGGCGGT GATTTTGGCG ATACGCCGAA
    TTGCCGTTGG GCACCAGCCG AATGCCGCCA CTAAAACCGC TATGCGGCTT
                                lacZ
~~~~~
 . D R Q F C M N G L V F A D R T P H .
4251 CGATCGCCAG TTCTGTATGA ACGGTCTGGT CTTTGCCGAC CGCACGCCGC
 GCTAGCGGTC AAGACATACT TGCCAGACCA GAAACGGCTG GCGTGCGGCG
 lacZ
~~~~~
  . P A L T E A K H Q Q Q F F Q F R
4301 ATCCAGCGCT GACGGAAGCA AAACACCAGC AGCAGTTTTT CCAGTTCCGT
    TAGGTCGCGA CTGCCTTCGT TTTGTGGTCG TCGTCAAAA GGTCAAGGCA
                                lacZ
~~~~~
 L S G Q T I E V T S E Y L F R H S .
4351 TTATCCGGGC AAACCATCGA AGTGACCAGC GAATACCTGT TCCGTCATAG
 AATAGGCCCC TTTGGTAGCT TCACTGGTCG CTTATGGACA AGGCAGTATC
 lacZ
~~~~~
  . D N E L L H W M V A L D G K P L A .
4401 CGATAACGAG CTCCTGCACT GGATGGTGGC GCTGGATGGT AAGCCGCTGG
    GCTATTGCTC GAGGACGTGA CCTACCACCG CGACCTACCA TTCGGCGACC
                                lacZ
~~~~~
 . S G E V P L D V A P Q G K Q L I
4451 CAAGCGGTGA AGTGCCTCTG GATGTCGCTC CACAAGGTAA ACAGTTGATT
 GTTCGCCACT TCACGGAGAC CTACAGCGAG GTGTTCCATT TGTCAACTAA

```



Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

 lacZ
~~~~~
      E L P E L P Q P E S A G Q L W L T .
4501 GAACTGCCTG AACTACCGCA GCCGGAGAGC GCCGGGCAAC TCTGGCTCAC
      CTTGACGGAC TTGATGGCGT CGGCCTCTCG CGGCCCCTTG AGACCGAGTG
                                lacZ
~~~~~
 . V R V V Q P N A T A W S E A G H I .
4551 AGTACGCGTA GTGCAACCGA ACGCGACCGC ATGGTCAGAA GCCGGGCACA
 TCATGCGCAT CACGTTGGCT TGCCTGGCG TACCAGTCTT CGGCCCCTGT
 lacZ
~~~~~
      . S A W Q Q W R L A E N L S V T L
4601 TCAGCGCCTG GCAGCAGTGG CGTCTGGCGG AAAACCTCAG TGTGACGCTC
      AGTCGCGGAC CGTCGTCACC GCAGACCGCC TTTTGGAGTC AACTGCGAG
                                lacZ
~~~~~
 P A A S H A I P H L T T S E M D F .
4651 CCCGCCGCGT CCCACGCCAT CCCGCATCTG ACCACCAGCG AAATGGATTT
 GGGCGGCGCA GGGTGCGGTA GGGCGTAGAC TGGTGGTCGC TTTACCTAAA
 lacZ
~~~~~
      . C I E L G N K R W Q F N R Q S G F .
4701 TTGCATCGAG CTGGGTAATA AGCGTTGGCA ATTTAACCGC CAGTCAGGCT
      AACGTAGCTC GACCCATTAT TCGCAACCGT TAAATTGGCG GTCAGTCCGA
                                lacZ
~~~~~
 . L S Q M W I G D K K Q L L T P L
4751 TTCTTTCACA GATGTGGATT GGCATAAAA AACAACTGCT GACGCCGCTG
 AAGAAAGTGT CTACACCTAA CCGCTATTTT TTGTTGACGA CTGCGGCGAC
 lacZ
~~~~~
      R D Q F T R A P L D N D I G V S E .
4801 CGCGATCAGT TCACCCGTGC ACCGCTGGAT AACGACATTG GCGTAAGTGA
      GCGCTAGTCA AGTGGGCACG TGGCGACCTA TTGCTGTAAC CGCATTCACT
                                lacZ
~~~~~
 . A T R I D P N A W V E R W K A A G .
4851 AGCGACCCGC ATTGACCCTA ACGCCTGGGT CGAACGCTGG AAGGCGGCGG
 TCGCTGGGCG TAACTGGGAT TGCGGACCCA GCTTGCGACC TTCCGCCGCC
 lacZ
~~~~~
      . H Y Q A E A A L L Q C T A D T L
4901 GCCATTACCA GGCCGAAGCA GCGTTGTTGC AGTGCACGGC AGATACACTT
      CCGTAATGGT CCGGCTTCGT CGCAACAACG TCACGTGCCG TCTATGTGAA

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

                                lacZ
~~~~~
 A D A V L I T T A H A W Q H Q G K .
4951 GCTGATGCGG TGCTGATTAC GACCGCTCAC GCGTGGCAGC ATCAGGGGAA
 CGACTACGCC ACGACTAATG CTGGCGAGTG CGCACCGTCG TAGTCCCCTT
 lacZ
~~~~~
      . T L F I S R K T Y R I D G S G Q M .
5001 AACCTTATTT ATCAGCCGGA AAACCTACCG GATTGATGGT AGTGGTCAAA
      TTGGAATAAA TAGTCGCGCT TTTGGATGGC CTAACCTACCA TCACCAGTTT
                                lacZ
~~~~~
 . A I T V D V E V A S D T P H P A
5051 TGGCGATTAC CGTTGATGTT GAAGTGGCGA GCGATACACC GCATCCGGCG
 ACCGCTAATG GCAACTACAA CTTACCGCT CGCTATGTGG CGTAGGCCCG
 lacZ
~~~~~
      R I G L N C Q L A Q V A E R V N W .
5101 CGGATTGGCC TGAAGTCCA GCTGGCGCAG GTAGCAGAGC GGGTAAACTG
      GCCTAACCGG ACTTGACGGT CGACCGCGTC CATCGTCTCG CCCATTGAC
                                lacZ
~~~~~
 . L G L G P Q E N Y P D R L T A A C .
5151 GCTCGGATTA GGGCCGCAAG AAACTATCC CGACCGCCTT ACTGCCGCTT
 CGAGCCTAAT CCCGGCGTTC TTTTGATAGG GCTGGCGGAA TGACGGCGGA
 lacZ
~~~~~
      . F D R W D L P L S D M Y T P Y V
5201 GTTTTGACCG CTGGGATCTG CCATTGTCAG ACATGTATAC CCCGTACGTC
      CAAACTGGC GACCCTAGAC GGTAACAGTC TGTACATATG GGGCATGCAG
                                lacZ
~~~~~
 F P S E N G L R C G T R E L N Y G .
5251 TTCCCGAGCG AAAACGGTCT GCGCTGCGGG ACGCGCGAAT TGAATTATGG
 AAGGGCTCGC TTTTGCCAGA CGCGACGCCC TCGCGGCTTA ACTTAATACC
 lacZ
~~~~~
      . P H Q W R G D F Q F N I S R Y S Q .
5301 CCCACACCAG TGGCGCGGCG ACTTCCAGTT CAACATCAGC CGCTACAGTC
      GGGTGTGGTC ACCGCGCCGC TGAAGGTCAA GTTGTAGTCG GCGATGTCAG
                                lacZ
~~~~~
 . Q Q L M E T S H R H L L H A E E
5351 AACAGCAACT GATGGAAACC AGCCATCGCC ATCTGCTGCA CGCGGAAGAA
 TTGTCGTTGA CTACCTTTGG TCGGTAGCGG TAGACGACGT GCGCCTTCTT

```

Table 14 (continued). Nucleotide sequence of the Mel/V5-His DEST cassette.

```

 lacZ
~~~~~
      G T W L N I D G F H M G I G G D D .
5401 GGCACATGGC TGAATATCGA CGGTTTCCAT ATGGGGATTG GTGGCGACGA
      CCGTGTACCG ACTTATAGCT GCCAAAGGTA TACCCCTAAC CACCGCTGCT
                                lacZ
~~~~~
 . S W S P S V S A E F Q L S A G R Y .
5451 CTCCTGGAGC CCGTCAGTAT CGGCGGAATT CCAGCTGAGC GCCGGTCGCT
 GAGGACCTCG GGCAGTCATA GCCGCCTTAA GGTCGACTCG CGGCCAGCGA
 lacZ Attr2
~~~~~
      . H Y Q L V W C Q K
5501 ACCATTACCA GTTGGTCTGG TGTCAAAAAT AATGACTGCA GGTCGACCAT
      TGGTAATGGT CAACCAGACC ACAGTTTTTA TTAGTGACGT CCAGCTGGTA
                                Attr2
~~~~~
5551 AGTGA CTGGA TATGTTGTGT TTTACAGTAT TATGTAGTCT GTTTTTTATG
 TCACTGACCT ATACAACACA AAATGTCATA ATACATCAGA CAAAAAATAC
 Attr2
~~~~~
5601 CAAAATCTAA TTAAATATAT TGATATTTAT ATCATTTTAC GTTCTCTCGTT
      GTTTTAGATT AAATTATATA ACTATAAATA TAGTAAAATG CAAAGAGCAA
                                Attr2                               V5/His
~~~~~
 G K P
5651 CAGCTTTCTT GTACAAAGTG GTGAGAATGA ATGAAGATCT GGGGAAGCCT
 GTCGAAAGAA CATGTTTCAC CACTCTTACT TACTTCTAGA CCCCTTCGGA
 V5/His
~~~~~
      I P N P L L G L D S T R T G H H H .
5701 ATCCCTAACC CTCTCCTCGG TCTCGATTCT ACGCGTACCG GTCATCATCA
      TAGGGATTGG GAGAGGAGCC AGAGCTAAGA TGCGCATGGC CAGTAGTAGT
                                stop codon
                                ~~~
 V5/His
~~~~~
      . H H H
5751 CCATCACCAT TGA
      GGTAGTGGTA ACT

```

Table 15. Baculoviral promoter sequences.

## AcMNPV ORF 25 promoter sequence

Ggtgtcttcattagatgccaatcacgtacgcaacagtcgcaaaagaacacacagtttcgtctccgcgacccgtgtaaaaaagtcg  
 ctccgcaatgtttgtaatcatgtcacgcaatcgccgagccaaaggttaacaaacgtatccatacgcgactgtaaatggacatgcac  
 gtacacacacttggtttgccttcttctactagtagcagcggtgatggaatgtgtgcgcaaacgattcacgctcggcgatctgttagcata  
 cgcgcaatacggcgacaaggttacgtgtgcataatcaatacactcgtctcggaccaatttttatttctgcttcgcaatactcgcacaca  
 cgtgatcgtcaacttgattgtattaaacccgttaacgatcaagctgttaataaacgccgtgtttcaatgggataatttcaaacgaactatg  
 tcttctattaacatgtcgaatacgtgttcggcggtgtgtgcgaaagttgtcacacacgctgataaaataaacggggcggtcctcg  
 ttcatttagctcgttaaagttacgggtcaaaatgagcacgtttgcgtcgtttggtttagcgacacgtttatatggccagtttggtttgttc  
 ggcgttaatgacgtgactgtggacaaatcgtgttctaaatacacaactcgtactcgaatgttgatatgtagttggttagccgatcta  
 tcttaaaatcaacttttgcaactcgtgatagagcacacgtccacatactgtcgataaacccgttgctcaaccgctcaaacgggtgaa  
 ttttagcttgaaagggcgcatgttggaatgactaaaaggaatattttcaataaagtcagtagtgacgcaaacggtgtctacgca  
 catgctggcaacagagtcgtccatattattatatactttatctgtgaaacacttcaattagacttgaaccacagcagacagcgacgt  
 cggtagc

## AcMNPV lef 3 promoter

Ccgagaagaaggcggtttgtataaaacccattttcgaatggtaacaaactgttttagcatttggatcgtttcgtgtcaaacgcgtcga  
 aaacttttaaacgcaattgccggcgagcagcagcaaatataatagctgcgtctgcacatgatcaaatcaagttgagacgttctt  
 gttcgtttcgcgtccattaacgtcaaccgagccatctgccaacaccagatcgcacgcgttgccacacttgatgctaatacacaac  
 attttatcaaacacgtcgctgactgtcgggccccgtaatggtgtgaaattttgcgtttgcgcactgtcggttgtacacgcacaccga  
 gttgtttgtcaacgtgacgccatacgtttgcaaacgggttcaacgacatggtatagttggcaaacgcgccgggtccgccacaaat  
 ccaaaaacgtgtcaacgtgtcggcaaacgtgaaacttttgcgtcgtctgatagtttcgcaaacatctaggtctgcgcgttggcggttg  
 tcaataatatttgagcgagcgcaaacaccgactgtcgtcgaacgtgttcaaacatcttgagttatttattttgtcgaacatttttac  
 tctcgtgtcgtcgaatgtttgtcgtcaaaaagacggccaacacgctcagcaaaactatacaataaagaacaaaatcgtacgca  
 atattaacattgaccgtttgatcgttaaacggacgggtcgttcagagccgccttattctctcgttgatattgttaaagttttgttttaatt  
 gtacacaatcgccgtgttgatcgtgaaatttcaaatcggttttgaacattgttctgaacgtgtgtgcgagcggtgtgtcggcca  
 cgtttataatcaactccctccacgctaacgaacgggtgctcggcgacacttcgatttcgtcgccattcagatttgcctcggatagattcc  
 cacatatcgacaacgaat

## AcMNPV TLP promoter

tgctagcccaattggccactgtgtacgaaatcgtcgtcaacgtgttgaatacatgttgcccgtaccgttgggtaaatctatgcac  
 ggagtcgccggaacactcgtactggtgtcagagtttctgatccggtgatgcacgttatcagttgtgactcgttatttcaaacatttgaa  
 atattgcgtgtcgcgcatatcgccgttatgtacgtgtgtccggcgccgttaaacgcgcacggatgcgctccacgcacgacattaagtt  
 gcgatcaaatattttatcgcggggcattcggccaccacgtggcgccatttacgcactgcataaactggttgacgagcaaatggagg  
 gaaagtagatagtagtagccgtcgtcgtttcacacaattcgttaactttactggccggtttccggtcaaacgtgaattatctg  
 gacattctcgtcgtcgtcgtcgtttgcaaacacctaagatagaacgtgggatgatacaagtcgcggttggtagaataatctttgtc  
 caaggttgggtcaacaccaacgtgtccagcaaacgctcgtccatgggataaagaccggcagactgtgtcgcacggcgccacggg  
 aacacattttagtgtcgtaatcaagttaaaatcgggggcatttcaggtgcacgtcggcctgtcggcgtcaaaataaactcgttgg  
 gattttcatcattgtcctaacgcgatcgtgtacgattcgtacacaggttgaatttttgatttaagaaatcaaaaattcaatccggtcatca  
 tgcacgcttctgtataggtggaaggtcgacggtgtgaaccacgttacaataagttttgcataatccgacacgtagccattac  
 gtcgggtgtgggtcgtcgtcgttgcgtcgttcacataatcagtcacacttggagccgcttgggtgaaagtcgttcgtcaaatcaaat  
 aaattgccaataacattaaagtaaacgtattataagaaaaagctt

Table 15. (continued) Baculoviral promoter sequences.

## AcMNPV hr5 sequence

Gttttacgcgtagaattctaccgtaaagcgagtttagttatgagccatgtgcaaaacatgacatcagctttttttataacaaatgacat  
catttcttgattgtgttttacgtagaattctactcgtaaagcgagttcagtttgaaaaacaaatgacatcatcttttgattgtgctttacaa  
gtagaattctaccgtaaataagttcggtttgaaaaacaaatgagtcattgtatgatcatattgcaacaaatgactcatcaatcga  
tcgtgcgtacacgtagaattctactcgtaaagcgagtttagagccgtgtgcaaaacatgacatcatctcgattgaaaaacaaatgacat  
catccactgatcgtgcgttacaagtagaattctactcgtaaagccagttcggtttagagccgtgtgcaaaacatgacatcagcttatgact  
cgtacttgattgtgttttacgcgtagaattctactcgtaaagc

Table 16. IE-1 promoter, coding, and polypeptide sequence.

## AcMNPV IE-1 promoter

Gttttacgcgtagaattctacccgtaaagcgagtttagttatgagccatgtgcaaaacatgacatcagctttttttataacaaatgacat  
catttcttgattgtgttttacacgtagaattctactcgtaaagcgagttcagtttgaaaaacaaatgacatcatcttttgattgtgctttacaa  
gtagaattctacccgtaaatacaagttcggtttgaaaaacaaatgagtcataattgtatgatcatattgcaaacaaatgactcatcaatcga  
tcgtgcgtacacgtagaattctactcgtaaagcgagtttagagccgtgtgcaaaacatgacatcatctcgattgaaaaacaaatgacat  
catccactgatcgtgcgttacaagtagaattctactcgtaaagccagttcggttatgagccgtgtgcaaaacatgacatcagcttatgact  
cgtacttgattgtgttttacgcgtagaattctactcgtaaagc

## AcMNPV IE-1 coding sequence

atgacgcaaattaatttaacgcgtcgtacaccagcgcttcgacgccgtcccgagcgtcgttcgacaacagctattcagagttttgtgata  
aacaacccaacgactatttaagttattataacctcccaccccggtatggagccgacacggtgatctgacagcgagactgcggcagc  
ttcaaaacttttggcaagcgtaactgtaactgataatgatttagtggaatgttgctcaagaccactgataatctgaagaagcagttag  
ttctgcttattattcggaaatcccttgagcagcctgtgtggagcaaccatcgccagttctgcttatcatcggaatctttgagcattctgct  
gggtgtgaaccaaccatcggaactggaactaaacggaagctggacgaatacttgacaattcacaaggtgtgtgtggccagtttaac  
aaaattaaattgaggcctaataacaagaaaagcacaattcaaaagctgtgcaaccctgaacagacaattaatcacacacgaacatttg  
cacggctcgttcaactcaagaaattacgcattattttactaatgattttgcgccgtatttaaatcggttcgacgacaacgactacaattccaa  
caggttctccgaccatatgtccgaaactggttattacatgtttgtgttaaaaaaagtgaagtgaagccgttgaaattatatttgccaagta  
cgtgagcaatgtgtttacgaataatacaacaattattacatggttagataatcgcgtgtttgtggaactttgataaaattaggtttatgatt  
cgtacaatttggttaagaaccggcatagaaattcctcattctcaagatgtgtgcaacgacgagacggctgcacaaaattgtaaaaaat  
gccatttcgtcgtatgtgcaccacacgtttaaagctgctctgacttcatatttaatttagatatgtattacgcgcaaacacatttgtagctttg  
ttacaatcgttggcgaaaagaaaatgtgggttcttttgagcaagttgtacgaaatgtatcaagataaaaaattatttactttgcctattatgct  
tagtcgtaaaagagagtaaatgaaattgagactgcatttaataatttctgtatcgccgtatgtgagtcacaataataaagtattcggaaagtgt  
gcagttcccgaacaaccccaacaataatgtggtggacaatttaatttaattgtaacaaaaaaagtagcgtcacgtacaataacagc  
agcgtcgtaatctttgtttaataattataaatacatgacaataattgcgagtaataataacgcagaaaaattaaaaaagggttaagaaggag  
gacggcagcatgcacattgtcgaacagatttgactcagaatgtagataatgtaaagggtcacaaattttatagattgtctttcaaaaacga  
ggagcgattgactatagctaagaaaaacaaagattttattgatttctggcgaaattaaagttagacgttagtcaagtaattcaaaaa  
tataatagatttaagcatcacatgtttgtaatcggtaaagtgaaccgaagagagcactacattgcacaataattgttaaaattgttagct  
ttaatttacagggtctgttccgtgttcgacgctataacgtttcggaacaaaaactaaattgtaaatataaaaaattcgaatttaatt

## AcMNPV IE-1 protein sequence

Mtqinfnasysastpsrasfdnsysefcdkqpnndylsyynhptpdgadvtsdsetaaasnflasvnsltndnlvecllkttdnlee  
avssayysesleqpvrveqpspsayhaesfehsagvnpqsatgtrkldeyldnsqgvvgqfnkiklrpkkykkstiqscatleqti  
nhntnictvastqeithyftndfapylmrfdndynsrfsdhmsetgyymfvvkksevkpfeiiakysvsnvyeytnnym  
vdnrvfvvtfdkirfmisynlvketgieiphsqdvndetaaqnckkchfvdvhhtfkaaltsyfuldmyyaqttfvtllqslgerk  
cgflsklyemyqdknlftlpimlsrkesneietasnnffvspyvsqilkyesvqfpdnppnkyvvdnlhnlivnkstltkyssv  
anllfnnykyhdniasnnaenlkkvkkedgsmhiveqyltqnvdnkvghnfivlsfkneerltiakknkefywisgeikdvdv  
sqviqkynrfkhmfvigkvnresttlhnnllklallqlglvplsdaitfaeqklncykkfefn

Table 17. Nucleotide sequence of plasmid pLenti6/V5-DEST.

AATGTAGTCTTATGCAATACTCTTGTAGTCTTGCAACATGGTAACGATGAGTTAGCAACATGCCTTACAA  
 GGAGAGAAAAAGCACCGTGCATGCCGATTGGTGGAAAGTAAGGTGGTACGATCGTGCCTTATTAGGAAGGC  
 AACAGACGGGTCTGACATGGATTGGACGAACCACTGAATTGCCGCATTGCAGAGATATTGTATTTAAGTG  
 CCTAGCTCGATACATAAACGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTA  
 GGAACCCACTGCTTAAGCCTCAATAAAGCTTGCCCTTGAGTGCTTCAAGTAGTGTGTGCCGTCTGTTGT  
 GTGACTCTGGTAACATAGAGATCCCTCAGACCCCTTTTAGTCAGTGTGGAAAATCTCTAGCAGTGGCGCCCG  
 AACAGGGACTTGAAAGCGAAAGGGAAACCAGAGGAGCTCTCTCGACGCAGGACTCGGCTTGCTGAAGCGC  
 GCACGGCAAGAGGCGAGGGGCGGCGACTGGTGAGTACGCCAAAAATTTGACTAGCGGAGGCTAGAAGGA  
 GAGAGATGGGTGCGAGAGCGTCAGTATTAAGCGGGGAGAATTAGATCGCGATGGGAAAAAATTCGGTTA  
 AGGCCAGGGGGAAGAAAAAATATAAATTAACATATAGTATGGGCAAGCAGGGAGCTAGAACCATTTCG  
 CAGTTAATCCTGGCTGTGTAGAAACATCAGAAGGCTGTAGACAAATACTGGGACAGCTACAACCATCCCT  
 TCAGACAGGATCAGAAGAACTTAGATCATTATATATACAGTAGCAACCCCTCTATTGTGTGCATCAAAGG  
 ATAGAGATAAAAGACACCAAGGAAGCTTTAGACAAGATAGAGGAAGAGCAAAAACAAAAGTAAGACCACCG  
 CACAGCAAGCGGCCGTGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAGTGAATTAT  
 ATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCCACCAAGGCAAGAGAAGAGTGGTGCA  
 GAGAGAAAAAGAGCAGTGGGAATAGGAGCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATG  
 GGCGCAGCGTCAATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAGAACA  
 ATTTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCAGTCTGGGGCATCAAGCAGCTCCA  
 GGCAAGAATCCTGGCTGTGGAAAGATACCTAAGGATCAACAGCTCCTGGGGATTGGGGTGTGCTCTGGA  
 AAACCTCATTGTCACCACTGCTGTGCTTGGAAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTGGGA  
 ATCACACGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTAATACACTCCTTAATTGA  
 AGAATCGCAAAACCAGCAAGAAAGAATGAACAAGAATTATTGGAATTAGATAAATGGGCAAGTTTGTGG  
 AATTGGTTTTAACATAACAAATTGGCTGTGGTATATAAATATTTCATAATGATAGTAGGAGGCTTGGTAG  
 GTTTAAGAATAGTTTTTGTGTACTTTCTATAGTGAATAGAGTTAGGCAGGGATATTCACCATTATCGTT  
 TCAGACCCACCTCCCAACCCGAGGGGACCCGACAGGCCGAAGGAATAGAAGAAGAAGGTGGAGAGAGA  
 GACAGAGATCGTCCGATTAGTGAAACGGATCTCGACGGTATCGATAAGCTTGGGAGTTCCGCGTTA  
 CATAACTTACGTTAAATGGCCCGCTGGCTGACCGCCCAACGACCCCGCCCATTGACGTCAATAATGAC  
 GTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGAGTATTTACGGTAACT  
 GCCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCTTATTGACGTCAATGACGGTAAAT  
 GGCCCGCCTGGCATATGCCCAGTACATGACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATT  
 AGTCATCGCTATTACCATGGTGTATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGACTCA  
 CCGGGATTTCGAAGTCTCCACCCCATTGACGTCAATGGGAGTTTGTTTTGGCACCAAAATCAACGGGACT  
 TTCCAAAATGTCTGAACAACTCCGCCCCATTGACGCAAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTA  
 TATAAGCAGAGCTCGTTTAGTGAACCGTCAGATCGCTGGAGACGCCATCCACGCTGTTTTGACCTCCAT  
 AGAAGACACCGACTCTAGAGGATCCACTAGTCCAGTGTGGTGAATTTCTGCAGATATCAACAAGTTTGTA  
 CAAAAAGCTGAACGAGAAACGTAAATGATATAAATATCAATATATTAATTAGATTGTGCATAAAAAA  
 CAGACTACATAAATACTGTAAACACAACATATCCAGTCACTATGGCGGCCGATTAGGCACCCCAAGGCTT  
 TACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTTTGTAGTTAGGATCCGGCGAGATTTTCAGGAGCT  
 AAGGAAGCTAAATGGAGAAAAAATCACTGGATATACCACGTTGTATATCCCAATGGCATCGTAAAG  
 AACATTTTGGAGGCAATTCAGTCAGTTGCTCAATGTGACCTATAACCAGACCGTTTCACTGGATATTACGGC  
 CTTTTTAAAGACCGTAAAGAAAAATAAGCACAAAGTTTTATCCGGCCTTATTACATTCTTGCCCGCCTG  
 ATGAATGCTCATCCGAATTCCGTATGGCAATGAAAGACGGTGAGCTGGTGTATGGGATAGTGTTCACC  
 CTTGTTACACCGTTTTCCATGAGCAAACGTAAACGTTTTTCATCGCTCTGGAGTGAATACCACGACGATTT  
 CCGGCAGTTTTCTACACATATATTGCAAGATGTGGCGTGTACGGTGAACCTGGCCTATTTCCCTAAA  
 GGGTTTATTGAGAATATGTTTTTCGTCTCAGCCAATCCCTGGGTGAGTTTACCAGTTTTGATTTAAACG  
 TGGCCAATATGGACAACTTCTTCGCCCCGTTTTTACCATGGGCAAAATTATACGCAAGGCGACAAGGT  
 GCTGATGCCGCTGGCGATTTCAGTTTCATCATGCCGTCTGTGATGGCTTCCATGTCCGCGAGAATGCTTAAT  
 GAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGCGTAAAGATCTGGATCCGGCTTACTAAAAGCCA  
 GATAACAGTATGCGTATTTGCGCGCTGATTTTTGCGGTATAAGAATATATACTGATATGTATACCCGAAG  
 TATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGACAGCTATCAGTTGC  
 TCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAGCACAAACATGCAGAATGAAGCCCGTCTGTG  
 CGTGCCGAACGCTGGAAAGCGGAAAATCAGGAAGGGATGGCTGAGGTGCGCCCGTTTTATTGAAATGAACG  
 GCTCTTTTGTGACGAGAACAGGACTGGTGAAATGCAGTTTAAAGTTTTACACCTATAAAGAGAGAGCC  
 GTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATTGACACGCCCGGGCGACGGATGGTGATCCCCCT  
 GGCCAGTGCACGCTGCTGTGTCAGATAAAGTCTCCCGTGAACCTTACCCGGTGGTGCATATCGGGGATGAA  
 AGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGTCTCCGTTATCGGGGAAGAAGTGGCTGATC  
 TCAGCCACCGCGAAAATGACATCAAAAACGCCATTAACCTGATGTTCTGGGGAATATAAATGTGAGGCTC  
 CGTTATACACAGCCAGTCTGCAGTGCACCATAGTGACTGGATATGTTGTGTTTTACAGTATTATGTAGT

Table 17 (continued). Nucleotide sequence of plasmid pLenti6/V5-DEST.

CTGTTTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTACGTTTCTCGTTTCAGCTTTTC  
 TTGTACAAAGTGGTTGATATCCAGCACAGTGGCGGCCGCTCGAGTCTAGAGGGCCCGCGTTTCGAAGGTA  
 AGCCTATCCCTAACCCCTCTCCTCGGTCTCGATTCTACGCGTACCGGTTAGTAATGAGTTTGGAAATTAATT  
 CTGTGGAATGTGTGTCAGTTAGGGTGTGGAAAGTCCCCAGGCTCCCCAGGCAGGCAGAAGTATGCAAAAGC  
 ATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAAGTCCCCAGGCTCCCCAGGCAGGCAGAAGTATGCAAA  
 GCATGCATCTCAATTAGTCAGCAACCAGTATGTCGCCCTAACTCCGCCCATCCCGCCCTAACTCCGCC  
 CAGTTCCGCCCATTTCTCCGCCCATGGCTGACTAATTTTTTTTTTATTTATGTCAGAGGCCGAGGCCGCTCT  
 GCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGGCCCTAGGCTTTTGCAAAAAGCTCCCGG  
 GAGCTTGTATATCCATTTTCGGATCTGATCAGCACGTGTGACAATTAATCATCGGCATAGTATATCGGC  
 ATAGTATAATACGACAAGGTGAGGAACATAAACCAGTGGCCAAAGCCTTTGTCTCAAGAAGAATCCACCCTCA  
 TTGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAAGACTACAGCGTCGCCAGCGCAGCTCT  
 CTCTAGCGACGGCCGCATCTTCACTGGTGTCAATGTATATCATTTTACTGGGGACCTTGTGCAGAATC  
 GTGGTGTCTGGGCACTGCTGCTGCTCGGCAGCTGGCAACCTGACTTGTATCGTCGCGATCGGAAATGAGA  
 ACAGGGGCATCTTGAGCCCCCTGCCGACGGTGCCGACAGGTGCTTCTCGATCTGCATCTGGGATCAAAGC  
 CATAGTGAAGGACAGTGATGGACAGCCGACGGCAGTTGGGATTCGTGAATTGCTGCCCTCTGGTTATGTG  
 TGGGAGGGCTAAGCACAATTCGAGCTCGGTACCTTTAAGACCAATGACTTACAAGGCAGCTGTAGATCTTT  
 AGCCATCTTTTAAAGAAAAGGGGGGAGTGGAAAGGCTAATTCACCTCCAACGAAGACAAGATCTGCTTT  
 TTGCTTGTACTGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTAGGGAACCC  
 ACTGCTTAAGCCTCAATAAAGCTTGCTTGTAGTGCTTCAAGTAGTGTGTGCCGCTCTGTTGTGTGACTCT  
 GGTAAGTAGAGATCCCTCAGACCCTTTTAGTCAGTGTGGAAAATCTCTAGCAGTAGTAGTTTCATGTCATC  
 TTATTATTCACTATTTATAACTTGCAAAGAAATGAATATCAGAGAGTGAGAGGAACCTGTTTATTGTCAGC  
 TTATAATGGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATTTTTTTCACTGCATCTCT  
 AGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGCTGGCTCTAGCTATCCCGCCCTAACTCC  
 GCCCATCCCGCCCCCTAACTCCGCCAGTTCCGCCCATTTCTCCGCCCATGGCTGACTAATTTTTTTTATT  
 TATGCAGAGGCCGAGGCCGCCCTCGGCCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGGCC  
 TAGGGACGTACCCAATTCCGCCCTATAGTGAGTCGTATTACGCGCGCTCACTGGCCGTCGTTTTACAACGT  
 CGTGACTGGGAAAACCTTGGCGTTACCCAACCTTAATCGCCTTGACGACATCCCCCTTTGCCAGCTGGC  
 GTAATAGCGAAGAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCCGAGCCTGAATGGCGAATGGGACGC  
 GCCCTGTAGCGCGCATTAAGCGCGCGGGTGTGGTGGTTACGCGCAGCGTGACCGCTACACTTGCCAGC  
 GCCCTAGCGCCCGCTCTTTTCGCTTTCTTCCCTTCTTCTCGCCACGTTCCGCCGCTTTCCCGCTCAAG  
 CTCTAAATCCGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCGACCCCAAAAACCTTGA  
 TTAGGGTGATGGTTACAGTAGTGGGCCATCGCCCTGATAGACGGTTTTTTCGCCCTTTGACGTTGGAGTCC  
 ACGTTCTTTAATAGTGGACTCTTGTTCCAACTGGAACAACACTCAACCCTATCTCGGTCTATTCTTTTG  
 ATTTATAAGGGATTTTGCCGATTTCCGCCCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTTAACGC  
 GAATTTTAAACAAAATATTAACGCTTACAATTTAGGTGGCATTTTTCGGGGAAATGTGCGCGGAACCCCTA  
 TTTGTTTATTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCCGTATAAATGCTTCA  
 ATAATATTGAAAAGGAAGATATGAGTATTCAACATTTCCGTGTGCGCCTTATTCCTTTTTTGGCGCA  
 TTTTGCTTCTCTGTTTTTGTCTACCCAGAAACGCTGGTGAAAGTAAAAGATGCTGAAGATCAGTTGGGTG  
 CACGAGTGGGTTACATCGAACTGGATCTCAACAGCGGTAAGATCCTTGAGAGTTTTTCGCCCGGAAGAACG  
 TTTTCCAATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATTATCCCGTATTGACGCCGGGCAA  
 GAGCAACTCGGTGCGCGCATACACTATTCTCAGAATGACTTGGTTGAGTACTCACCAGTCACAGAAAAGC  
 ATCTTACGGATGGCATGACAGTAAGAGAATTATGCAGTGCTGCCATAACCATGAGTGATAACACTGCGGC  
 CAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCCTTTTTTGCACAACATGGGGGATCAT  
 GTAACCTCGCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAAACGACGAGCGTGACACCAGCA  
 TGCCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAACTGGCGAACTACTTACTCTAGCTTCCCGGCA  
 ACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGCG  
 TGGTTTTATTGCTGATAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTCAGCACTGGGGCCAG  
 ATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAACGAAATAG  
 ACAGATCGCTGAGATAGGTGCTCACTGATTAAAGCATTGGTAACTGTGAGACCAAGTTTACTCATATATA  
 CTTTAGATTGATTTAAACTTCATTTTTAATTTAAAAGGATCTAGGTGAAGATCCTTTTTTGATAATCTCA  
 TGACCAAAATCCCTTAAACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATC  
 TTCTTGAGATCCTTTTTTTTCTGCGCGTAATCTGCTGCTTGCAAAACAAAAAACACCCGCTACCGCGGTG  
 GTTTGTTTGGCGGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGATAC  
 CAAATACTGTTCTTCTAGTGTAGCCGTAGTTAGGCCACCCTTCAAGAACTCTGTAGCACCGCCTACATA  
 CCTCGCTCTGCTAATCCTGTTACAGTGGCTGCTGCCAGTGGCGATAAGTCGTGCTTACCGGGTTGGAC  
 TCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGGGGCTGAACGGGGGGTTCTGTCACACAGCCCACT  
 TGGAGCGAAGCCTACACCGAAGTGAATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCCGA  
 AGGGAGAAAGCGGACAGGTATCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCA



Table 17 (continued). Nucleotide sequence of plasmid pLenti6/V5-DEST.

GGGGGAAACGCCTGGTATCTTTATAGTCCTGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGT  
GATGCTCGTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTTCCTGGCCTT  
TTGCTGGCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCTGTGGATAACCGTATTACGCC  
TTTGAGTGAGCTGATACCGCTCGCCGAGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGG  
AAGAGCGCCCAATACGCAAACCGCCTCTCCCGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAG  
GTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTAGCTCACTCATTAGGCACCC  
CAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGTGAGCGGATAACAATTTACACAG  
GAAACAGCTATGACCATGATTACGCCAAGCGCGCAATTAACCTCACTAAAGGGAACAAAAGCTGGAGCT  
GCAAGCTT

Table 18. Nucleotide sequence of plasmid pLenti6/V5-D-TOPO™.

AATGTAGTCTTATGCAATACTCTTGTAGTCTTGCAACATGGTAACGATGAGTTAGCAACATGCCTTACAA  
 GGAGAGAAAAAGCACCGTGCATGCCGATTGGTGGAAAGTAAGGTGGTACGATCGTGCCTTATTAGGAAGGC  
 AACAGACGGGTCTGACATGGATTGGACGAACCACTGAATTGCCGATTGCAGAGATATTGTATTAAAGTG  
 CCTAGCTCGATACATAAACGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTA  
 GGGAAACCCACTGCTTAAAGCCTCAATAAAGCTTGCCTTGAGTGCTTCAAGTAGTGTGTGCCCCGTCTGTTGT  
 GTGACTCTGGTAACTAGAGATCCCTCAGACCTTTTAGTCAGTGTGGAAAACTCTTAGCAGTGGCGCCCG  
 AACAGGCACTTGAAGCGAAAGGGAAACACAGAGGAGCTCTCTGACGCGAGGACTCGGCTTGCTGAAGCGC  
 GCACGGCAAGAGGCGAGGGGCGGCGACTGGTGAGTACGCCAAAAATTTTGAAGTAGCGGAGGCTAGAAGGA  
 GAGAGATGGGTGCGAGAGCGTCAGTATTAAAGCGGGGAGAATTAGATCGCGATGGGAAAAAATTCGGTTA  
 AGGCCAGGGGAAAGAAAAATATAAATTAAACATATAGTATGGGCAAGCAGGGAGCTAGAACCATTTCG  
 CAGTTAATCTGGCCTGTAGAAACATCAGAAGGCTGTAGACAAATACTGGGACAGCTACAACCATCCCT  
 TCAGACAGGATCAGAAGAACTTAGATCATTATATAATACAGTAGCAACCCCTCTATTGTGTGCATCAAAGG  
 ATAGAGATAAAAGACACCAAGGAAGCTTTAGACAAGATAGAGGAAGAGCAAAACAAAGTAAGACCACCG  
 CACAGCAAGCGGCCGTGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAGTGAATTAT  
 ATAAATATAAAGTAGTAAAAATTGAACCTTAGGAGTAGCACCCACCAAGGCAAAGAGAAGAGTGGTGCA  
 GAGAGAAAAAGAGCAGTGGGAATAGGAGCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATG  
 GCGCAGCGCTCAATGACGCTGACGGTACAGGCCACAATATTGTCTGGTATAGTGCAGCAGAGAACA  
 ATTTGTCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTCTGGGGCATCAAGCAGCTCCA  
 GGCAAGAACTCTGGCTGTGGAAAGATACCTAAAGGATCAACAGCTCTGGGGATTGGGGTTGCTCTGGA  
 AAACCTCATTTGCACCACTGCTGTGCCTTGGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTGGGA  
 ATCACACGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTAATACACTCTTAATTGA  
 AGAATCGCAAAACCAGCAAGAAAGAATGAACAAGAATTATTGGAATTAGATAAATGGGCAAGTTTGTGG  
 AATTGGTTTAACATAACAAATTGGCTGTGGTATATAAAATTATTACATAATGATAGTAGGAGGCTTGGTAG  
 GTTTAAGAATAGTTTTGCTGTACTTTCTATAGTGAATAGAGTTAGGCAGGGATATTACCAATTATCGTT  
 TCAGACCCACTCCCAACTCCCGAGGGGACCCGACAGGCCGAGGAATAGAAGAAGAAGGTGGAGAGAGA  
 GACAGAGACAGATCCATTTCGATTAGTGAACGGATCTCGACGGTATCGATAAGCTTGGGAGTTCGCGCTTA  
 CATAACTTACGGTAAATGGCCCGCTGGCTGACGCCCAACGACCCCGCCCATTGACGTCAATAATGAC  
 GTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGAGTATTTACGGTAACT  
 GCCCACCTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAAT  
 GGCCCGCTTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATT  
 AGTCATCGCTATTACCATGGTGTGCGGTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGCTCA  
 CGGGGATTTCGAAGTCCACCCCATTTGACGTCAATGGGAGTTTGTGTTTGGCACCAAAATCAACGGGACT  
 TTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTA  
 TATAAGCAGAGCTCGTTTGTAGTGAACCGTCAGATCGCCTGGAGACGCCATCCACGCTGTTTTGACCTCCAT  
 AGAAGACACCGACTCTAGAGATCCACTAGTCCAGTGTGGTGAATTGATCCCTTACCAAGGGCTCGAG  
 TCTAGAGGCGCCGCGGTTTGAAGGTAAGCCTATCCCTAACCCCTCTCTCGGTCTCGATTCTACGCGTACC  
 GGTTAGTAATGAGTTTGAATTAATTCTGTGGAATGTGTGTCAGTTAGGGTGTGGAAAGTCCCCAGGCTC  
 CCCAGGCAGGCAGAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAAGTCCCCAGG  
 CTCCCCAGCAGGCAGAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGTATCCCGCCCCCTAACT  
 CCGCCCATCCGCCCCCTAACTCCGCCCCAGTTCGCCCCATTCTCCGCCCCATGGCTGACTAATTTTTTTTA  
 TTTATGCAGAGGCGGAGGCGCCTCTGCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGG  
 CCTAGGCTTTTGCAAAAGCTCCCGGGAGCTTGTATATCCATTTTCGGATCTGATCAGCAGGTGTTGACA  
 ATTAATCATCGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAACATAACCATGGCCAAGCC  
 TTTGTCTCAAGAAGAATCCACCCTCATTGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAA  
 GACTACAGCGTCGCCAGCGCAGCTCTCTAGCGACGGCCGCATCTTCACTGGTGTCAATGTATATCATTT  
 TTAAGTGGGGACCTTGTGCGAAGCTCGTGGTGTGGGCACTGTGCTGCTGCGGCAGCTGGCAACCTGAC  
 TTGTATCGTCGCGATCGGAAATGAGAACAGGGGCATCTTGAGCCCTGCGGACGGTGGCGACAGGTGCTT  
 CTCGATCTGCATCTGGGATCAAAGCCATAGTGAAGGACAGTGTGACAGCCGACGGCAGTTGGGATTC  
 GTGAATTGCTGCCCTCTGTTATGTGTGGGAGGGCTAAGCACAAATTCGAGCTCGGTACCTTTAAGACCAA  
 TGACTTACAAGGCAGCTGTAGATCTTAGCCACTTTTAAAGAAAAGGGGGGACTGGAAGGGCTAATTCA  
 CTCCCAACGAAGACAAGATCTGCTTTTGGCTTGTACTGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGG  
 GAGCTCTCTGGCTAACTAGGGAACCCACTGCTTAAGCCTCAATAAAGCTTGCCTTGAAGTGTCTCAAGTAG  
 TGTGTGCCCTCTGTTGTGTGACTCTGGTAACTAGAGATCCCTCAGACCTTTTAGTCAGTGTGGAAAT  
 CTCTAGCAGTAGTAGTTTCATGTCTATTTATTCAGTATTTATAACTTGCAAAGAAATGAATATCAGAG  
 AGTGAGAGGAACCTGTTTATTCAGCTTATAATGGTTACAAATAAAGCAATAGCATCACAATTTTCACAA  
 ATAAAGCATTTTTCCTGCAATCTAGTTGTGGTTGTCCAACTCATCAATGTATCTTATCATGTCTG  
 GCTCTAGCTATCCCGCCCCCTAACTCCGCCCCATCCGCCCCCTAACTCCGCCCCAGTTCGCCCCATTCTCCGC  
 CCCATGGCTGACTAATTTTTTTTATTTATGACAGAGGCGGAGGCCCTCGGCCTCTGAGCTATTCCAGAA

Table 18 (continued). Nucleotide sequence of plasmid pLenti6/V5-D-TOPO™.

GTAGTGAGGAGGCTTTTTTGGAGGCCCTAGGGACGTACCCAATTGCGCCCTATAGTGAGTCGTATTACGCGC  
 GCTCACTGGCCGTCGTTTTACAACGTCGTGACTGGGAAAACCTGGCGTTACCCAACCTTAATCGCCTTGC  
 AGCACATCCCCCTTTTCGCCAGCTGGCGTAATAGCGAAGAGGCCCGACCGATCGCCCTTCCCAACAGTTG  
 CGCAGCCTGAATGGCGAATGGGACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGTGTGGTGGTTACGC  
 GCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTTCGCTTTCTTCCCTTCCTTTCTCGC  
 CACGTTTCGCCGGCTTTCCCGTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGTTCGGATTAGTGCTTTA  
 CGGCACCTCGACCCCAAAAACCTTGATTAGGGTGATGGTTACGTAAGTGGGCCATCGCCCTGATAGACGG  
 TTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACTCTTGTTCCAAACTGGAACAACACT  
 CAACCCATCTCGGTCTATTCTTTTGATTATATAAGGGATTGTCGGATTTCGGCCTATTGGTTAAAAAAT  
 GAGCTGATTAAACAAAAATTAACGCGAATTTAAACAAATATTAACGCTTACAATTTAGGTGGCACTTT  
 TCGGGGAAATGTGCGCGGAACCCCTATTGTTTATTCTTAAATACATTCAAATATGTATCCGCTCATG  
 AGACAATAACCCGTGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTG  
 TCGCCCTTATTCCCTTTTTCGCGGCATTTTGCCTTCCTGTTTTGCTCAGCCAGAAACGCTGGTGAAGT  
 AAAAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTACATCGAACTGGATCTCAACAGCGGTAAAGATC  
 CTTGAGAGTTTTTCGCCCGAAGAAGCTTTTCCAATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCG  
 TATTATCCCGTATTGACGCCGGGCAAGAGCAACTCGGTGCGCGCATACACTATTCTCAGAATGACTTGGT  
 TGAGTACTCAGCTACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAATTATGCACTGCTGCC  
 ATAACCATGAGTGATAACACTGCGGCCAACTTACTTTTGACAACGATCGGAGGACCGAAGGAGCTAACCG  
 CTTTTTTGCACAACATGGGGGATCATGTAACCTGCGCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCAT  
 ACCAAACGACGAGCGTGACACCAGATGCCCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAACCTGGC  
 GAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCAGGACCAC  
 TTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTATTGCTGATAAATCTGGAGCCGGTGAGCGTGGGTCTCG  
 CGGTATCATGTCAGCACTGGGGCCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACAGACGGGGAGT  
 CAGGCAACTATGGATGAACGAAATAGACAGATCGCTGAGTAGGTGCCTCACTGATTAAAGCATTGGTAAC  
 TGTGAGACCAAGTTTACTCATATATACTTTAGATTGATTTAAAACTTCATTTTAAATTTAAAAGGATCTA  
 GGTGAAGATCCTTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCA  
 GACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTTCTGCGCGTAATCTGCTGCTTGCAAA  
 CAAAAAACCACCGCTACCAGCGGTGGTTTTGTTGCGGGATCAAGAGCTACCAACTCTTTTTCCGAAGGT  
 AACTGGCTTCAGCAGAGCGCAGATACCAATACTGTTCTTCTAGTGAGCCGTAGTTAGGCCACCACTTC  
 AAGAACTCTGTAGCACCAGCTACATACCTCGCTCTGCTAATCCTGTTACCAAGTGGCTGCTGCCAGTGGCG  
 ATAAGTCGTGCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAAC  
 GGGGGGTTCTGTGCACACAGCCCAGCTTGAGCGAAGACCTACACCGAAGTGAATACCTACAGCGTGAG  
 CTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTTCGGAA  
 CAGGAGAGCGCACGAGGGAGCTTCCAGGGGAAACGCTGGTATCTTTATAGTCCTGTGCGGTTTTCCGCA  
 CCTCTGACTTGAGCGTCGATTTTTGTGATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAAC  
 GCGGCCCTTTTACGGTTCTTGCCCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCTGCGTTATCCCTTG  
 ATTCTGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGACGCCGAACGACCGAGCG  
 CAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCCAATACGCAACCGCCTCTCCCGCGCGTTGGCQG  
 ATTCATTAAATGCAGCTGGCACGACAGTTTTCCCGACTGGAAGCGGGCAGTGAGCGCAACGCAATTAATG  
 TGAGTTAGCTCACTCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAAT  
 TGTGAGCGGATAACAATTTACACAGGAAACAGCTATGACCATGATTACGCCAAGCGCGCAATTAACCCCT  
 CACTAAAGGGAACAAAAGCTGGAGCTGCAAGCTT

Table 19. Nucleotide sequence of pLenti4/V5-DEST.

AATGTAGTCTTATGCAATACTCTTGTAGTCTTGCAACATGGTAACGATGAGTTAGCAACATGCCTTACAA  
 GGAGAGAAAAAGCACCGTGCATGCCGATTGGTGGAAAGTAAGGTGGTACGATCGTGCCCTTATTAGGAAGGC  
 AACAGACGGGTCTGACATGGATTGGACGAACCACTGAATTGCCGCATTGCAGAGATATTGTATTTAAGTG  
 CCTAGCTCGATACATAAACGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTA  
 GGGAAACCACTGCTTAAGCCTCAATAAAGCTTGCTTGAGTGCTTCAAGTAGTGTGTGCCCGTCTGTGT  
 GTGACTCTGGTAAGTAGAGATCCCTCAGACCCCTTTAGTCAGTGTGGAAAATCTCTAGCAGTGGCGCCCG  
 AACAGGGACTTGAAGCGAAAGGGAAACAGAGGAGCTCTCTGACGCAGGACTCGGCTTGTCTGAAGCGC  
 GCACGGCAAGAGCGAGGGCGGCGACTGGTGTAGTACGCCAAAAATTTTACTAGCGGAGGCTAGAAGGA  
 GAGAGATTGGGTGCGAGAGCGTCACTATTAAAGCGGGGAGAATTAGATCGCGATGGGAAAAAATTCGGTTA  
 AGGCCAGGGGAAAGAAAAATATAAATTAAACATATAGTATGGGCAAGCAGGGAGCTAGAACGATTTCG  
 CAGTTAATCTGGCCTGTTAGAAACATCAGAAGCTGTAGACAAATACTGGGACAGCTACAACCATCCCT  
 TCAGACAGGATCAGAAGAACCTTAGATCATTATATAATACAGTAGCAACCCCTCTATTGTGTGCATCAAAGG  
 ATAGAGATAAAGACACCAAGGAAGCTTTAGACAAGATAGAGGAAGAGCAAAACAAAAGTAAGACCACCG  
 CACAGCAAGCGCGCGCTGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAGTGAATTAT  
 ATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCCACCAAGGCAAGAGAGAAGAGTGGTGCA  
 GAGAGAAAAAGAGCAGTGGGAAATAGGAGCTTTGTTCCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATG  
 GGCGCAGCGTCAATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAGAACA  
 ATTTGCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCACAGTCTGGGGCATCAAGCAGCTCCA  
 GGCAAGAATCCTGGCTGTGGAAAGATACCTAAAGGATCAACAGCTCCTGGGGATTGGGGTTGCTCTGGA  
 AAATCATTTCACCACTGCTGTGCTTGGAAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTGGGA  
 ATCACACGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTAATACACTCCTTAATTGA  
 AGAATCGCAAAACAGCAGAAAGAAATGAACAAGAATTATTGGAAATTAGATAAATGGGCAAGTTTGTGG  
 AATTGGTTTAACATAACAATTTGGCTGTGGTATATAAATTTATTCATAATGATAGTAGGAGGCTTGGTAG  
 GTTTAAGAAATAGTTTTTGTCTGTACTTTCTATAGTGAATAGAGTTAGGCAGGGATATTCACCATATCGTT  
 TCAGACCCACCTCCCAACCCGAGGGGACCCGACAGGCCCGAAGGAATAGAAGAAGAAGGTGGAGAGAGA  
 GACAGAGACAGATCCATTGATTAAGTGAACGGATCTCGACGGTATCGATAAGCTTGGGAGTTCCGCGTTA  
 CATAACTTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCCGCCCATTTGACGTCAATAATGAC  
 GTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGAGTATTTACGGTAAACT  
 GCGCCGCTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAAT  
 AGTCATCGCTATTACCATGGTGTATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGACTCA  
 CGGGGATTTCCAAGTCTCCACCCCATTTGACGTCAATGGGAGTTTGTGTTTGGCACCAAAATCAACGGGACT  
 TTCCAAAATGTGTAACAACCTCCGCCCCATTGACGCAAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTA  
 TATAAGCAGAGCTCGTTTTAGTGAACCGTCAGATCGCCTGGAGACGCCATCCACGCTGTTTTGACCTCCAT  
 AGAAGACACCGACTCTAGAGGATCCACTAGTCCAGTGTGGTGAATTTCTGCAGATATCAACAAGTTTGTA  
 CAAAAAGCTGAACGAGAAACGTAAAAATGATATAAATATCAATATATTAAATTAGATTTTGCATAAAAAA  
 CAGACTACATAATACTGTAAACACACATATCCAGTCACTATGGCGGCCGCGATTAGGCACCCAGGCTT  
 TACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTTTGAGTTAGGATCCGGCGAGATTTTCAGGAGCT  
 AAGGAAGCTAAATGGAGAAAAAATCACTGGATATACCACCGTTGATATATCCCAATGGCATCGTAAAG  
 AACATTTTGGAGCATTTTCACTGAGTTCATGTAACCTATAACAGACCGTTTCACTGAGTATTTACGGC  
 CTTTTTAAAGACCGTAAAGAAAAATAGCACAAAGTTTATCCGGCCTTTATTCACATTCTTGCCCGCCTG  
 ATGAATGCTCATCCGAATTCCGTATGGCAATGAAGACCGTGAGCTGGTGTATGGGATAGTGTTCACC  
 CTTGTTACACCGTTTTCCATGAGCAAACTGAAACGTTTTTCATCGCTCTGGAGTGAATACCACGACGATTT  
 CCGGCAGTTTTCTACACATATATTTCGCAAGATGTGGCGTGTACGGTGAAAACCTGGCCTATTTCCCTAAA  
 GGGTTTATTGAGAAATATGTTTTTCTGCTCAGCCAATCCCTGGGTGAGTTTCACCAAGTTTTGATTAAACG  
 TGGCCAATATGGACAACTTCTTCGCCCCGTTTTTACCATGGGCAAAATATATACGCAAGGCGACAAGGT  
 GCTGATGCCGCTGGCGATTCAAGTTTCATCATGCCGCTGTGATGGCTTCCATGTCCGCAGAAATGCTTAAT  
 GAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGCGGTAAAGATCTGGATCCGGCTTACTAAAAGCCA  
 GATAACAGTATGCGTATTTGCGCGCTGATTTTGGCGTATAAGAATATATACTGATATGTATACCCGAAG  
 TATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTGACAGTTGACAGCGACAGCTATCAGTTGC  
 TCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAAGCAACCATGCAGAAATGAAGCCCGTCTG  
 CGTGCCGAACGCTGGAAAGCGGAAATCAGGAAGGGATGGCTGAGGTGCCCCGTTTTATTGAAATGAACG  
 GCTCTTTTGTGACGAGAACAGGGACTGGTGAAATGCAGTTTAAAGTTTACACCTATAAAAGAGAGAGCC  
 GTTATCGTCTGTTGTGGATGTACAGAGTATATTATGACACGCCCGGGCGACGGATGGTGTATCCCCCT  
 GGCCAGTGCACGCTGCTGTGTAGATAAAGTCTCCCGTGAACCTTTACCCGGTGGTGCATATCGGGGATGAA  
 AGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGCTCTCCGTTATCGGGGAAGAAGTGGCTGATC  
 TCAGCCACCGCGAAAATGACATCAAAAACGCCAATTAAACCTGATGTTTCTGGGGAATATAAATGTCAAGCTC  
 CGTTATACACAGCCAGTCTGCAGGTCGACCATAGTACTGGATATGTTGTGTTTTACAGTATTATGTAGT

Table 19 (continued). Nucleotide sequence of pLenti4/V5-DEST.

CTGTTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTTACGTTTCTCGTTCAGCTTTC  
 TTGTACAAAGTGGTTGATATCCAGCACAGTGGCGGCCGCTCGAGTCTAGAGGGCCCGCGGTTTCAAGGTA  
 AGCCTATCCCTAACCCTCTCCTCGGTCTCGATTCTACGCGTACCGGTTAGTAATGAGTTTGGAAATTAATT  
 CTGTGGAATGTGTGTGTCAGTTAGGGTGTGGAAAGTCCCAGGCTCCCAGGCAGGCAGAGTATGCAAAAGC  
 ATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAAGTCCCAGGCTCCCAGGCAGGCAGAGTATGCAAAAGC  
 GCATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAAGTCCCAGGCTCCCAGGCAGGCAGAGTATGCAAAAGC  
 CAGTTCCGCGCATTCTCCGCCCCATGGCTGACTAATTTTTTTTTATTTATGCAAGGCCGAGGCCGCTCT  
 GCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGGCCTAGGCTTTTGCAAAAGCTCCCCC  
 TGTTGACAATTAATCATCGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAATAAACCATG  
 GCCAAGTTGACCAGTGCCGTTCCGGTGCTACCGCGCGACGTCGCCGGAGCGGTTCGAGTTCTGGACCG  
 ACCGGCTCGGGTTCTCCCGGGACTTCGTGGAGGACGACTTCGCCGGTGTGGTCCGGGACGACGTGACCCCT  
 GTTCATCAGCGCGGTCCAGGACCAGGTGGTGGCGGACAACACCCTGGCCTGGGTGTGGGTGCGCGGCTG  
 GACGAGCTGTACGCCGAGTGGTGGGAGGTCGTGTCCACGAACTTCGGGGACGCTCCGGGCGGCGCATGA  
 CAGGATCGGGCAGCAGCGGTGGGGCGGGAGTTCCGCCCTGCGCGACCCGGCCGGCAACTGCGTGCACCTT  
 CGTGGCCGAGGAGCAGGACTGACACGTGCTACGAGATTTAAATGGTACCTTTAAGACCAATGACTTACAA  
 GGCAGCTGTAGATCTTAGCCACTTTTTAAAGAAAAGGGGGGACTGGAAGGGCTAATCACTCCCAACGA  
 AGACAAGATCTGCTTTTTGCTTGTACTGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTG  
 GCTAATCTAGGGAACCCACTGCTTAAGCCTCAATAAGCTTGCCCTTGAGTGCTTCAAGTAGTGTGTGCCCG  
 TCTGTTGTGTGACTCTGGTAACTAGAGATCCCTCAGACCCTTTTAGTCAGTGTTGAAAATCTCTAGCAGT  
 AGTAGTTCATGTCATCTTATTATTCAGTATTTATAACTTGCAAGAAATGAATATCAGAGAGTGAGAGGA  
 ACTTGTTTATTGCACTTTATAATGGTTACAAATAAGCAATAGCATCACAAATTTACAAATAAAGCATTT  
 TTTTTCATCTGAGTTCTAGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGTCTGGCTCTAGCTA  
 TCCCGCCCCCTAACTCCGCCCATCCCGCCCCCTAACTCCGCCCAGTTCGCCCATTTCTCCGCCCATGGCTG  
 ACTAATTTTTTTTATTTATGCAAGGCCGAGGCGCCTCGGCCCTTGAGCTATTCCAGAAGTAGTGAGGA  
 GGCTTTTTTTGGAGGCCTAGGGACGTACCCAATTCGCCCTATAGTGAGTCTGATTACCGCGCTCACTGGC  
 CGTCTGTTTTACAACGTCTGACTGGGAAAACCTGGCGTTACCCAACCTAATCGCCTTGACGACATCCC  
 CTTTTCGCCAGCTGGCGTAATAGCGAAGAGGCCCGACCCGATCGCCCTTCCCAACAGTTGCGCAGCCTGA  
 ATGGCGAATGGGACGCGCCCTGTAGCGCGCATTAAGCGCGCGCGGTGTGGTGGTTACGCGCAGCGTGAC  
 CGCTACACTTGTCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTCTCGCCACGTTCCGCC  
 GGCTTTCCCGCTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCG  
 ACCCCAAAAAATTGATTAGGGTGATGGTTACAGTAGTGGGCCATCGCCCTGATAGACGGTTTTTTCGCC  
 TTTGACGTTGGAGTCCACGTTCTTAAATAGTGACTCTTGTTCAAAACCTGGAACAACACTCAACCCTATC  
 TCGGCTCTATTCTTTTGATTTATAAGGGATTTTGGCGATTTCCGCCCTATTGGTTAAAAAATGAGCTGATT  
 AACAAAAATTTAACCGGAATTTTAACAAAATATTAACGCTTACAATTTAGGTGGCACTTTTCGGGGAAT  
 GTGCGCGAACCCTATTGTTTATTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAAC  
 CTTGATAAATGCGCTTCAATAATATGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTGCGCCCTTAT  
 TCCCTTTTTTGGCGCATTTTGCCCTTCTGTTTTTGTCTACCCAGAAACGCTGGTGAAAGTAAAGATGCT  
 GAAGATCAGTTGGGTGCACGAGTGGGTACATCGAAGTGGATCTCAACAGCGGTAAGATCCTTGAGAGTT  
 TTCGCCCGAAGAACGTTTTTCAATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATTATCCCG  
 TATTGACGCCGGGAAGAGCAACTCGGTGCGCCGATACACTATTCTCAGAATGACTTGGTTGAGTACTCA  
 CCAGTCACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAATTATGCAGTGCTGCCATAACCATGA  
 GTGATAACACTGCGGCCAATTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCA  
 CAACATGGGGGATCATGTAACCTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAACGAC  
 GAGCGTGACACCACGATGCCGTGTAGCAATGGCAACAACGTTGCGCAAACTATTAACTGGCGAACTACTTA  
 CTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGACAGGACCACTTCTGCGCTC  
 GGCCCTTCCGGCTGGCTGGTTTATGCTGATAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATT  
 GCAGCACTGGGGCCAGATGGTAAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGAGTCAGGCAACTA  
 TGGATGAACGAAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATTGGTAAGTGTGACACCA  
 AGTTTACTCATATATACTTTAGATTGATTTAAAACTTCATTTTTAATTTAAAGGATCTAGGTGAAGATC  
 CTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCGTAG  
 AAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAAACAAAAAACCC  
 ACCGCTACCAGCGGTGGTTTGTGTTGCCGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTC  
 AGCAGAGCCGAGATACCAATACTGTTCTTCTAGTGTAGCCGTAGTTAGGCCACCCTTCAAGAACTCTG  
 TAGCACCGCTACATACCTCGCTCTGCTAATCCTGTTACAGTGGCTGCTGCCAGTGGCGATAAGTCGTG  
 TCTTACCGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAACGGGGGTTTCG  
 TGCACACAGCGCTTGGAGCGAAGCAACGACTACCGGAAGTGAAGTACCTACAGCGTGAGCTGAGTGAAGAA  
 CGGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTGGGAACAGGAGAGCG  
 CACGAGGGAGCTTCCAGGGGGAAACGCTGGTATCTTTATAGTCTGTGCGGTTTTGCCACCTCTGACTT  
 GAGCGTCGATTTTTGTGATGCTCGTCAGGGGGCGGAGCCTATGAAAAACGCCAGCAACGCGGCTTTT

Table 19 (continued). Nucleotide sequence of pLenti4/V5-DEST.

TACGGTTCCTGGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCTGTGGA  
TAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTCA  
GTGAGCGAGGAAGCGGAAGAGCGCCCAATACGCAAACCGCCTCTCCCCGCGCGTTGGCCGATTCAATTAAT  
GCAGCTGGCACGACAGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTAGCT  
CACTCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGTGAGCGGA  
TAACAATTTACACAGGAAACAGCTATGACCATGATTACGCCAAGCGCGCAATTAACCCTCACTAAAGGG  
AACAAAAGCTGGAGCTGCAAGCTT

Table 20. Nucleotide sequence of pLenti6/UbC/V5-DEST.

AATGTAGTCTTATGCAATACTCTTGTAGTCTTGCACATGGTAACGATGAGTTAGCAACATGCCTTACAA  
 GGAGAGAAAAAGCACCGTGCATGCCGATTGGTGGAAGTAAGGTGGTACGATCGTGCCCTTATTAGGAAGGC  
 AACAGACGGGTCTGACATGGATTGGACGAACCACTGAATTGCCGCATTGCAGAGATATTGTATTTAAGTG  
 CCTAGCTCGATACATAAACGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTA  
 GGGAAACCCACTGCTTAAGCCTCAATAAAGCTTGCCTTGAGTGCTTCAAGTAGTGTGTGCCCGTCTGTGT  
 GTGACTCTGGTAACTAGAGATCCCTCAGACCCCTTTAGTCAGTGTGGAATCTCTAGCAGTGGCGCCCCG  
 AACAGGGACTTGAAAGCGAAAGGGAAACCAGAGGAGCTCTCTCAGCGCAGGACTCGGCTTGCTGAAGCGC  
 GCACGGCAAGAGGCGAGGGGCGGCGACTGGTGAGTACGCCAAAAATTTTACTAGCGGAGGCTAGAAAGGA  
 GAGAGATGGGTGCGAGAGCGTCAGTATTAAGCGGGGAGAATTAGATCGCGATGGGAAAAAATTCGGTTA  
 AGGCCAGGGGAAAGAAAAATATAAATTTAAACATATAGTATGGGCAAGCAGGGAGCTAGAACCATTCCG  
 CAGTTAATCCTGGCCTTTAGAAACATCAGAAGGCTGTAGACAAATACTGGGACAGCTACAACCATCCCT  
 TCAGACAGGATCAGAAGAACTTAGATCATTATATAATACAGTAGCAACCCCTCTATTGTGTGCATCAAAGG  
 ATAGAGATAAAAGACACCAAGGAAGCTTTAGACAAGATAGAGGAAGAGCAAAACAAAAGTAAGACCACCG  
 CACAGCAAGCGGCCGCTGATCTTCAGACCTGGAGGAGGAGATATGAGGGACAATTGGAGAAGTGAATTAT  
 ATAAATATAAAGTAGTAAAAATTGAACCATTAGGAGTAGCACCCACCAAGGCAAGAGAAGAGTGGTGCA  
 GAGAGAAAAAGAGCAGTGGGAATAGGAGCTTTGTTCTTGGGTTCTTGGGAGCAGCAGGAAGCACTATG  
 GGCGCAGCGTCAATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGCAGCAGCAGAACA  
 ATTTGTCTGAGGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCAGAGTCTGGGGCATCAAGCAGCTCCA  
 GGCAAGAACTCTGGCTGTGGAAGATACCTAAAGGATCAACAGCTCCTGGGGATTGGGGTTGCTCTGGA  
 AAATCATTGTGACCACTGCTGTGCCTTGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTGGGA  
 ATCACACGACCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTAATACACTCCTTAATTGA  
 AGAATCCGAAACAGCAAGAAAGAATGAACAAGAATTATTGGAATTAGATAAATGGGCAAGTTTGTGG  
 AATTGGTTTAACATAACAAATTGGCTGTGGTATATAAAATTATTATATAGTAGTAGGAGGCTTGGTAG  
 GTTTAAGAATAGTTTTGTCTGACTTTCTATAGTGAATAGAGTTAGGCAGGGATATTACCATTTATCGTT  
 TCAGACCCACCTCCGACCCCGAGGGGACCCGACAGGCCGAAGGAATAGAAGAAGAAGGTGGAGAGAGA  
 GACAGAGACAGATCCATTTCGATTAGTGAACGGATCTCGACGGTATCGGATCTGGCCTCCGCGCCGGTTT  
 TGGCGCTCCCGCGGGCGCCCCCTCTCACGGCGAGCGCTGCCACGTGACAGCAAGGGCGCAGGAGCGT  
 CCTGATCTTCCGCCCGGACGCTCAGGACAGCGGCCGCTGCTCATAAGACTCGGCTTAGAACCCAGT  
 ATCAGCAGAAGGACATTTTAGGACGGGACTTGGGTGACTCTAGGGCACTGGTTTCTTTCCAGAGAGCGG  
 AACAGGCGAGGAAAAGTAGTCCCTTCTCGGCGATTCTGCGGAGGGATCTCCGTGGGGCGGTGAACGCCGA  
 TGATTATATAAGGACGCGCCGGGTGTGGCACAGCTAGTTCCGTGCGAGCCGGGATTGGGTGCGGTTCT  
 TGTGTTGGATCGTGTGATCGTCACTTGGTGAGTAGCGGGCTGCTGGGCTGGCCGGGGCTTTCTGTGGC  
 GCCGGGCCGCTCGGTGGGACGGAAGCGTGTGGAGAGACCGCAAGGGCTGTAGTCTGGGTCCGCGAGCAA  
 GGTGCCCCGAACTGGGGGTTGGGGGAGCGCAGCAAAATGGCGGCTGTTCCCGAGTCTTGAATGGAAGA  
 CGCTTGAGGCGGGCTGTGAGGTGTTGAAACAAGGTGGGGGCGATGGTGGGCGGCAAGAACCCAGGT  
 CTTGAGGCCTTCGCTAATGCGGAAAGCTCTTATTCCGGTGAGATGGGTGGGGCACCATCTGGGGACCC  
 TGACGTGAAGTTGTCACTGACTGGAGAATCGGTTTGTCTGTGCGGGGGCGGCAGTTATGCGGTG  
 CCGTTGGGCAGTGACCCGTACCTTTGGGAGCGCGCCCTCGTCTGTGCTGACGTACCCGTTCTGTT  
 GGCTTATAATGTCAGGTGGGGCCACCTGCCGTTAGGTGTGCGGTAGGCTTTCTCCGTGCGAGGACGAG  
 GGTTCGGGCCTAGGGTAGGCTCTCTGAATCGACAGGCGCGGACCTCTGGTGAGGGGAGGGATAAGTGA  
 GGCGTCAGTTCTTTGGTGGTCTTTATGTACCTATCTTCTTAAGTAGCTGAAGCTCCGGTTTGAACAT  
 GCGCTCGGGGTTGGCGAGTGTGTTTTGTGAAGTTTTTTAGGCACCTTTTGAATGTAATCATTGGGTCA  
 ATATGTAATTTTCAGTGTAGACTAGTAAATTGTCCGCTAAATCTGGCCGTTTTTGGCTTTTTGTAG  
 ACGAAGCTTGGTACCGAGCTCGGATCCACTAGTCCAGTGTGGTGAATCTGCGAGATATCAACAAGTTTG  
 TACAAAAAGCTGAACGAGAAACGTAAATGATATAAATATCAATATATAAATTAGATTTTGCATAAAA  
 AACAGACTACATAAATACTGTAAAACACAACATATCCAGTCACTATGGCGGCCGATTAGGCACCCAGGC  
 TTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTTTGAAGTTAGGATCCGGCGAGATTTTCAGAG  
 CTAAGGAAGCTAAATGGAGAAAAAATCACTGGATATACCACGTTGATATATCCCAATGGCATCGTAA  
 AGAACATTTTGGGCATTTTCAGTCAGTTGCTCAATGTACCTATAACCAGACGTTTCAGCTGGATATTACG  
 GCCTTTTTAAAGACCGTAAAGAAAAATAAGCACAAGTTTATCCGGCTTTTATTACATTTCTGCCCGCC  
 TGATGAATGCTCATCCGGAATTCGATGGCAATGAAAGACGGTGAGCTGGTGATATGGGATAGTGTTCA  
 CCCTTGTTACACCGTTTTCATGAGCAAACTGAAACGTTTTCATCGCTCTGGAGTGAATACCACGACGAT  
 TTTCCGGCAGTTTCTACAGATATATTGCAAGATGTGGCGGTGTTACGGTGAAACCTGGCCATTTCCTTAA  
 AAGGGTTTATTGAGAATATGTTTTCTGTCTCAGCCAATCCCTGGGTGAGTTTACCAGTTTGTATTAA  
 CGTGGCCAATATGGACAACCTTCTCGCCCCGTTTTCACCATGGGCAAAATATTATACGCAAGGCGACAAG  
 GTGCTGATGCCGTGGCGATTTCAGTTTCATCATGCCGTCTGTGATGGCTTCCATGTCGGCAGAATGCTTA  
 ATGAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGCGTAAAGATCTGGATCCGGCTTACTAAAAGC  
 CAGATAACAGTATGCGTATTTGCGCGCTGATTTTTGCGGTATAAGAATATATACTGATATGTATACCCGA  
 AGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGACAGCTATCAGTT

Table 20 (continued). Nucleotide sequence of pLenti6/UbC/V5-DEST.

GCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAGCACAACCATGCAGAATGAAGCCCGTCGTC  
 TGGCTGCCGAACGCTGGAAAGCGGAAAATCAGGAAGGGATGGCTGAGGTCGCCCGGTTTATTGAAATGAA  
 CGGCTCTTTTGTGACGAGAACAGGGACTGGTGAATGCAGTTTAAAGTTTACACCTATAAAAGAGAGAG  
 CCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATTGACACGCCCGGGCGACGGATGGTGATCCCC  
 CTGGCCAGTGCACGTCTGCTGTGATATAAAGTCTCCCGTGAACCTTACCCGGTGGTGCATATCGGGGATG  
 AAAGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGGTCTCCGTTATCGGGGAAGAAGTGGCTGA  
 TCTCAGCCACCGGAAAATGACATCAAAAACGCCATTAACTGATGTTCTGGGGAATATAAAATGTCAAGC  
 TCCGTTATACACAGCCAGTCTGCAGGTGACCATAGTGACTGGATATGTTGTGTTTACAGTATTATGTA  
 GTCTGTTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTACGTTTCTCGTTCAGCTT  
 TCTGTACAAAGTGGTTGATATCCAGCACAGTGGCGGCCGCTCGAGTCTAGAGGGCCCGCGTTTCGAAGG  
 TAAGCCTATCCCTAACCTCTCCTCGGTCTCGATTCTACGCGTACCGGTTAGTAATGAGTTTGAATTAA  
 TTCTGTGGAATGTGTGTGATAGGTAGGTGTGGAAGTCCCAGGCTCCCAGGCAGGCAGAAGTATGCAAA  
 GTCATGCTCAATTAGTCAGCAACCATAGTCCCGCCCTAACTCCGCCCATCCGCCCCCTAACTCCG  
 AAGCATGCATCTCAATTAGTCAGCAACCATAGTCCCGCCCTAACTCCGCCCATCCGCCCCCTAACTCCG  
 CCCAGTTCGCGCCATTCTCGCCCCATGGCTGACTAATTTTTTTTATTTATGCAGAGGCCGAGGCCGCCT  
 CTGCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGGCCCTAGGCTTTTGCAAAAAGCTCCC  
 GGGAGCTTGTATATCCATTTTCGGATCTGATCAGCAGTGTGACAAATAATCATCGGCATAGTATATCG  
 GCATAGTATAATACGACAAGGTGAGGAACTAAACCATGGCCAAGCCTTTGTCTCAAGAAGAAATCCACCT  
 CATTGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAAGACTACAGCGTCGCCAGCGCAGCT  
 CTCTCATGCAGCCCGCATCTTCACTGGTGTCAATGTATATCATTTTACTGGGGACCTTGTGCAGAAC  
 TCGTGGTGTGCGGCATGCTGCTGCTGCGGCAGTGTGCAACCTGACTTGTATCGTCGCGATCGGAAATGA  
 GAACAGGGGCATCTTGAGCCCCGCGGACGGTGCCGACAGGTGCTTCTCGATCTGCATCCTGGGATCAAA  
 GCCATAGTGAAGGACAGTGTGACAGCCGACGGCAGTTGGGATTCTGTGAATTGCTGCCCTCTGGTTATG  
 TGTGGGAGGCTAAGCACAAATTCGAGCTCGGTACCTTTAAGACCAATGACTTACAAGGCAGCTGTAGATC  
 TTAGCCACTTTTTAAAGAAAAGGGGGGACTGGAAGGGCTAATCACTCCCAACGAAGACAAAGATCTGCT  
 TTTTGCTTGTACTGGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGGCTAACTAGGGAAC  
 CCATGGCTTAAGCCTCAATAAAGCTTGCCCTGAGTGTCTCAAGTAGTGTGTGCCGTCTGTTGTGTGACT  
 CTGGTAACCTAGAGATCCCTCAGACCCTTTTAGTCAGTGTGGAATACTCTAGCAGTAGTATTCATGTCA  
 TCTTATTATTCAGTATTTATAACTTGCAAGAAATGAATATCAGAGAGTGAGAGGAACCTGTTTATTGCA  
 GCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATTTTTTTTCACTGCATT  
 CTAGTTGTGTTTGTCCAAACTCATCAATGTATCTTATCATGTCTGGCTCTAGCTATCCCGCCCCTAACT  
 CCGCCCATCCCGCCCCTAACTCCGCCCAGTTCGCGCCATTCTCGCCCCATGGCTGACTAATTTTTTTTA  
 TTTATGCAGAGGCCGAGGCCGCTCGGCCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGG  
 CCTAGGGACCTCAATTCGCCCTATAGTGAGTCTGATTACGCGCGCTCACTGGCCGTCTGTTTACAAC  
 GTCGTGACTGGGAAAACCTTGGCGTTACCCAATTAATCGCCTTGACGACATCCCTTTTCGCCAGCTG  
 GCGTAATAGCGAAGAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGGAC  
 GCGCCCTGTAGCGGCGCATTAAGCGCGCGGGTGTGGTGTACGCGCAGCGTGACCGCTACACTTGCCA  
 GCGCCCTAGCGCCCGCTCCTTTTCGCTTTCTTCCCTTCCTTTCTCGCCACGTTTCGCCGCTTTCCCGCTCA  
 AGCTCTAAATCGGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCGACCCCAAAAACTT  
 GATTAGGGTGATGGTTACGTAAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGT  
 CCACGTTCTTTAATAGTGGACTCTGTTTCCAACTGGAACAACACTCAACCTATCTCGGTCTATTCTTT  
 TGATTTATAAGGGATTTTGGCGATTTTCGGCCTATTGGTTAAAAAATGAGCTGATTTAACAATAAATTAAC  
 GCGAATTTTAACAATAATTAACGCTTACAATTTAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCC  
 TATTTGTTTATTTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCTGATAAATGCTT  
 CAATAATATTGAAAAAGGAAGAGTATGAGTATTAACATTTCCGTGTCGCCCTTATTCCTTTTTTTGCGG  
 CATTTTGCCCTTCTGTTTTTGTCTACCCAGAAACGCTGGTGAAAGTAAAGATGCTGAAGATCAGTTGGG  
 TGCACGAGTGGGTTACATCGAAGTGGATCTCAACAGCGGTAAGATCCTTGAGAGTTTTTCGCCCGAAGAA  
 CGTTTTTCCAATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATTATCCCGTATTGACGCCGGGC  
 AAGAGCAACTCGGTGCGCCGATACACTATTCTCAGAATGACTTGGTTGAGTACTACCCAGTCACAGAAAA  
 GCATCTTACGGATGGCATGACAGTAAGAGAATTATGCAGTGTGCCATAACCATGAGTGATAAAGTGGC  
 GCCAATTACTTCTGACAAAGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCACAACATGGGGGATC  
 ATGTAACCTGCGCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATAACAAACGACGAGCGTGACACCAC  
 GATGCCGTGATGAATGGCAACAACGTTGCGCAAACTATTAAGTGGCGAACTACTTACTCTAGCTTCCCGG  
 CAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCAGGACCATTCTGCGCTCGGCCCTTCCGGCTG  
 CTTGGTTTATTGCTGATAAATCTGGAGCCGTTAGCGTGGGTCTCGCGGTATCATTCAGCAGTGAACGAAAT  
 AGATGGTAAGCCCTCCCGTATCGTAGTTATCTACAGCAGGGGAGTCAGGCAACTATGGATGAACGAAAT  
 AGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATTTGGTAACTGTGAGACCAAGTTTACTCATATA  
 TACTTTAGATTGATTTAAACTTCAATTTTAAATTTAAAGGATCTAGGTGAAGATCCTTTTTTGATAATCT  
 CATGACCAAAATCCCTTAAAGTGTGTTTTCGTTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGA



Table 20 (continued). Nucleotide sequence of pLenti6/UbC/V5-DEST.

TCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAACCACCGCTACCAGCGG  
TGGTTTGTGTTGCCGGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGAT  
ACCAAATACTGTTCTTCTAGTGTAGCCGTAGTTAGGCCACCACCTCAAGAACTCTGTAGCACCGCTTACA  
TACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCTTACCGGGTTGG  
ACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAACGGGGGGTTCGTGCACACAGCCCAG  
CTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCC  
GAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTC  
CAGGGGGAAACGCCTGGTATCTTTATAGTCTGTGCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTT  
GTGATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTTCCTGGCC  
TTTTGCTGGCCTTTTGTCTACATGTTCTTTCCCTGCGTTATCCCCTGATTCTGTGGATAACCGTATTACCG  
CCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGC  
GGAAGAGCGCCCAATACGCAAACCGCCTCTCCCCGCGCGTTGGCCGATTCAATTAATGCAGCTGGCACGAC  
AGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTAGCTCACTCATTAGGCAC  
CCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGTGAGCGGATAACAATTTACAC  
AGGAAACAGCTATGACCATGATTACGCCAAGCGCGCAATTAACCTCACTAAAGGGAACAAAAGCTGGAG  
CTGCAAGCTT

Table 21. Nucleotide sequence of plasmid pLP1.

TTGGCCCATTCGCATACGTTGTATCCATATCATAATATGTACATTTATATTGGCTCATGTCCAACATTACC  
 GCCATGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCA  
 TATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCCGCC  
 CATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGT  
 GGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATT  
 GACGTCAATGACGGTAAATGGCCCGCTGGCATTATGCCAGTACATGACCTTATGGGACTTTCTTACTT  
 GGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGCGGTTTGGCAGTACATCAATGGGCG  
 TGGATAGCGGTTTGA CTACGGGGATTTC AAGTCTCCACCCCATGACGTCAATGGGAGTTTGT TTTGG  
 CACCAAAATCAACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAATGGGCGGTAGGC  
 GTGTACGGTGGGAGGTCTATATAAGCAGAGCTCGTTTAGTGAAACCGTCAGATCGCTGGAGACGCCATCC  
 ACGCTGTTTTTGACCTCCATAGAAGACCGGGACCGATCCAGCCTCCCTCGAAGCTTACATGTGGTACC  
 GAGCTCGGATCCTGAGAACTTCAGGGTGAGTCTATGGGACCTTGATGTTTTCTTTCCCTTCTTTTCTA  
 TGGTTAAGTTCATGT CATAGGAAGGGGAGAAGTAACAGGGTACACATATTGACCAATCAGGGTAATTTT  
 GCATTTGTAATTTTAAAAAATGCTTTCTTTCTTTTAAATACTTTTTTGT TTTATCTTATTTCTAATACTTT  
 CCTAATCTCTTTCTTTT CAGGGCAATAATGATACAATGTATCATGCCTCTTTGCACCATTTCTAAAGAATA  
 ACAGTGATAATTTCTGGGTTAAGGCAATAGCAATATTTCTGCATATAAATATTTCTGCATATAAATTTCT  
 ACTGATGTAAGAGGTTTCATATTGCTAATAGCAGTACAATCCAGCTACCATTTCTGCTTTTATTTTATGG  
 TTGGGATAAGGCTGGATTATTCTGAGTCCAAGCTAGGCCCTTTTGCTAATCATGTTTCATACCTCTTATCT  
 TCCTCCCAAGCTCTCTGGGCAACGTGCTGGTCTGTGTGCTGGCCCATCACTTTGGCAAAGCACGTGAGAT  
 CTGAATTCGAGATCTGCCGCCGCCATGGGTGCGAGAGCGTCAGTATTAAGCGGGGAGAATTAGATCGAT  
 GGGAAAAAATTCGGTTAAGGCCAGGGGGAAGAAAAAATATAAATTAAAACATATAGTATGGGCAAGCAG  
 GGAGCTAGAACGATTCCGAGTTAATCCTGGCCTGTTAGAAAACATCAGAAGGCTGTAGACAAATACTGGGA  
 CAGCTACAACCATCCCTTCAGACAGGATCAGAAGAACTTAGATCATTATATAATACAGTAGCAACCTCT  
 ATTGTGTGCATCAAAGGATAGAGATAAAAGACACCAAGGAAGCTTTAGACAAGATAGAGGAAGAGCAAAA  
 CAAAAGTAAGAAAAAAGCACAGCAAGCAGCAGCTGACACAGGACACAGCAATCAGGTGAGCCAAAATTAC  
 CCTATAGTGACAGAACATCCAGGGGCAATGGTACATCAGGCCATATCACCTAGAACTTTAAATGCATGGG  
 TAAAAGTAGTAGAAGAGAAGGCTTT CAGCCAGAAGTGATACCCATGTTTT CAGCATTATCAGAAGGAGC  
 CACCCCAAGATTTAAACACCATGCTAAACACAGTGGGGGGACATCAAGCAGCCATGCAATGTTAAAA  
 GAGACCATCAATGAGGAAGCTGCAGAAATGGGATAGAGTGATCCAGTGATGACAGGGCCTATTGCACAG  
 GCCAGATGAGAGAACCAAGGGGAAGTGACATAGCAGGAACCTACTAGTACCCTTCAGGAACAAATAGGATG  
 GATGACACATAATCCACCTATCCCAGTAGGAGAAATCTATAAAAGATGGATAATCCTGGGATTAAATAAA  
 ATAGTAAGAAATGTATAGCCCTACAGCATTTCTGGACATAAGACAAGGACCAAGGAACCTTTAGAGACT  
 ATGTAGACCGATTCTATAAACTCTAAGAGCCGAGCAAGCTTCACAAGAGGTAAAAAATTGGATGACAGA  
 AACCTTGTTGGTCCAAAATGCGAACCCAGATTGTAAGACTATTTTAAAGCATTGGGACCAGGAGCGACA  
 CTAGAAGAAATGATGACAGCATGT CAGGGAGTGGGGGGACCCGGCCATAAAGCAAGAGTTTGGCTGAAG  
 CAATGAGCCAAGTAACAAATCCAGCTACCAATATGATACAGAAAGGCAATTTAGGAACCAAGAAAGAC  
 TGTTAAGTGTTTCAATTGTGGCAAAGAAGGGCACATAGCCAAAAATTGCAGGGCCCTAGGAAAAAGGGC  
 AGATCTGGCCTTCCCACAAGGGGAAGGCCAGGGAATTTCTTCAGAGCAGACCAGAGCCAACAGCCCCACC  
 AGAAGAGAGCTTCAGGTTTGGGGAAGAGACAACAACCTCCCTCTCAGAAGCAGGAGCCGATAGACAAGGAA  
 CTGTATCCTTTAGCTTCCCTCAGATCACTCTTTGGCAGCGACCCCTCGTCACAATAAAGATAGGGGGCA  
 ATTAAGGAAGCTCTATTAGATACAGGAGCAGATGATACAGTATTAGAAGAAATGAATTTGCCAGGAAGA  
 TGGAAACCAAAAATGATAGGGGGAATTGGAGGTTTTATCAAAGTAAGACAGTATGATCAGATACTCATAG  
 AAATCTGCGGACATAAAGCTATAGGTACAGTATTAGTAGGACCTACACCTGTCAACATAATTGGAAGAAA  
 TCTGTTGACTCAGATTGGCTGCATTTAAATTTTCCCATTAGTCCTATTGAGACTGTACCAGTAAATTA  
 AAGCCAGGAATGGATGGCCCAAAAGTTAAACAATGGCCATTGACAGAAGAAAAATAAAGCATTAGTAG  
 AAATTTGTACAGAAATGGAAGGAAGGAAAAATTTCAAAAATGGGCCTGAAAATCCATACAATACTCC  
 AGTATTTGCCATAAAGAAAAAGACAGTACTAAATGGAGAAAAATTAGTAGATTTTCAGAGAACTTAATAAG  
 AGAACTCAAGATTTCTGGGAAGTTCAATTAGGAATACCACATCCTGCAGGGTTAAACAGAAAAATCAG  
 TAACAGTACTGGATGTGGGCGATGCATATTTTCAGTTCCCTTAGATAAAGACTTCAGGAAGTATACTGC  
 ATTTACCATACTAGTATAAACAATGAGACACCAGGATTAGATATCAGTACAATGTGCTTCCACAGGGA  
 TGGAAGGATCACCAGCAATTTCCAGTGTAGCATGACAAAAATCTTAGAGCCTTTTAGAAAAACAAATC  
 CAGACATAGTCTATCATCAATACATGGATGATTTGTATGTAGGATCTGACTTAGAAATAGGGCAGCATAG  
 AACAAAAATAGAGGAACCTGAGACAACATCTGTTGAGGTGGGGATTACCACACCAGACAAAAACATCAG  
 AAAGAACCTCCATTCTTTGGATGGGTTATGAACTCCATCCTGATAAATGGACAGTACAGCCTATAGTGC  
 TGCCAGAAAAGGACAGCTGGACTGTCAATGACATACAGAAATTAGTGGGAAAAATTGAATTGGGCAAGTCA  
 GATTTATGCAGGGATTAAAGTAAGGCAATTATGTAACTTCTTAGGGGAACCAAGCACTAACAGAAGTA  
 GTACCACTAACAGAAGAAGCAGAGCTAGAAGTGGCAGAAAACAGGGAGATTCTAAAAGAACCGGTACATG

Table 21 (continued). Nucleotide sequence of plasmid pLP1.

GAGTGTATTATGACCCATCAAAAGACTTAATAGCAGAAATACAGAAGCAGGGGCAAGGCCAATGGACATA  
TCAAAATTTATCAAGAGCCATTTAAAAATCTGAAAACAGGAAAGTATGCAAGAATGAAGGGTGCCCACT  
AATGATGTGAAACAATTACAGAGGCAGTACAAAAATAGCCACAGAAAGCATAGTAATATGGGGAAAGA  
CTCCTAAATTTAAATTACCCATACAAAAGGAACATGGGAAGCATGGTGGACAGAGTATTGGCAAGCCAC  
CTGGATTCTGAGTGGGAGTTTGTCAATACCCCTCCCTTAGTGAAGTTATGGTACCAGTTAGAGAAAGAA  
CCCATAAATAGGAGCAGAACTTTCTATGTAGATGGGGCAGCCAATAGGGAACTAAATTAGGAAAAGCAG  
GATATGTAACAGAGAGGAAGACAAAAAGTTGTCCCTTAACGGACACAACAAATCAGAAGACTGAGTT  
ACAAGCAATTCATCTAGCTTTGCAGGATTCGGGATTAGAAGTAAACATAGTGACAGACTCACAATATGCA  
TTGGGAATCATTCAAGCACAACCAGATAAGAGTGAATCAGAGTTAGTCAGTCAAATAATAGAGCAGTTAA  
TAAAAAGGAAAAAGTCTACCTGGCATGGGTACCAGCACACAAAGGAATTGGAGGAAATGAACAAGTAGA  
TAAATTGGTCAGTGTGGAATCAGGAAAGTACTATTTTATAGTGAATAGATAAGGCCCAAGAAGAACAT  
GAGAAATATCAGCTAATAGCAACAGACATAGGCTAGTGATTTTAACTTACCACCTGTAGTAGCAAGAAA  
TAGTAGCCAGCTGTGATAAATGTGAGCTAAAAGGGGAAGCCATGCATGGACAAGTAGACTGTAGCCAGG  
AATATGGCAGCTAGATTGTACACATTTAGAAGGAAAAGTTATCTTGGTAGCAGTTTATGTAGCCAGTGGA  
TATATAGAAGCAGAAGTAATTCAGCAGAGACAGGGCAAGAAACAGCATACTTCCTCTTAAATTTAGCAG  
GAAGATGGCCAGTAAAAACAGTACATACAGACATGGCAGCAATTTACCAGTACTACAGTTAAGGCCGC  
CTGTTGGTGGGCGGGGATCAAGCAGGAATTTGGCATTCCCTACAATCCCCAAAGTCAAGGAGTAATAGAA  
TCTATGAATAAAGAATTAAAGAAAATTATAGGACAGGTAAGAGATCAGGCTGAACATCTTAAGACAGCAG  
TACAAATGGCAGTATTCATCCACAATTTTAAAGAAAAGGGGGGATTGGGGGGTACAGTGCAGGGGAAAG  
AATAGTAGACATAATAGCAACAGACATACAACTAAAGAAATTACAAAAACAAATTACAAAAATTCAAAAT  
TTTCGGGTTTATTACAGGGACAGCAGAGATCCAGTTTGGAAAGGACCAGCAAAGCTCCTCTGGAAAGGTG  
AAGGGGCAGTAGTAATACAAGATAATAGTGACATAAAAGTAGTGCCAAGAAGAAAAGCAAAGATCATCAG  
GGATTATGGAAGACAGATGGCAGGTGATGATTGTGTGGCAAGTAGACAGGATGAGGATTAAACACATGGAA  
TTCCGGAGCGGCCGAGGAGCTTTGTTCCCTGGGTTCTTGGGAGCAGCAGGAAGCACTATGGGCGCAGCG  
TCAATGACGCTGACGGTACAGGCCAGACAATTATTGTCTGGTATAGTGACAGCAGCAGAAACAATTTGTCTGA  
GGGCTATTGAGGCGCAACAGCATCTGTTGCAACTCAGCTCTGGGGCATCAAGCAGCTCCAGGCAAGAAT  
CTGGCTGTGGAAGATACTTAAAGGATCAACAGCTCCTGGGGATTGGGGTTGCTCTGGAAACCTCAAT  
TGCACCACTGCTGTGCCCTTGAATGCTAGTTGGAGTAATAAATCTCTGGAACAGATTGGAATCACACGA  
CCTGGATGGAGTGGGACAGAGAAATTAACAATTACACAAGCTTCGGCGGAATTCACCCACCAGTGCAGG  
CTGCCATCAGAAAGTGGTGGCTGGTGTGGCTAATGCCCTGGCCCAAGTATCACTAAGCTCGCTTTCT  
TGCTGTCCAATTTCTATTAAAGGTTCCCTTTGTTCCCTAAGTCCAACCTACTAACTGGGGGATATTATGAA  
GGGCCCTTGAGCATCTGGATTCTGCCTAATAAAAAACATTTATTTTCATTGCAATGATGTATTTAAATAT  
TTCTGAATATTTTACTAAAAAGGGAATGTGGGAGGTCAGTGCATTTAAACATAAAGAAATGAAGAGCTA  
GTTCAAACCTTGGGAAATAACACTATATCTTAACTCCATGAAAGAAGGTGAGGCTGCAACACAGCTAATG  
CACATTGGCAACAGCCCTGATGCCTATGCCCTTATTCATCCCTCAGAAAAGGATTCAAGTAGAGGCTTGA  
TTTGGAGGTTAAAGTTTGTCTATGCTGTATTTACATTACTTATTGTTTTAGCTGTCTCATGAATGTCT  
TTTCACTACCCATTTGCTTATCCTGCATCTCTCAGCCTTGACTCCACTCAGTTCTCTTGCTTAGAGATAC  
CACCTTTCCCTGAAAGTGTTCCTTCCATGTTTACGGCGAGATGGTTTCTCCTCGCCTGGCCACTCAGCC  
TTAGTTGTCTCTGTTGCTTATAGAGGCTACTTTGAAGAAGGAAAAACAGGGGGCATGGTTTGACTGTCC  
TGTGAGCCCTTCTCCCTGCCTCCCCACTCAGAGTGACCCGGAATCCCTCGACATGGCAGCTCAGCAGT  
AGTGCAGCCGAGTCTGCTTCTCGCTCAGTACGCTCGCTCGCTCGCTCGTTCCGCTGCGGCGAGCGGT  
ATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAATATGTGA  
GCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCTTGTGGCGTTTTCATAGGCTCCGCC  
CCCCTGACGAGCATCAAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAGATA  
CCAGGCGTTTCCCTCGGAAGCTCCCTCGTGCCTCTCCTGTTCCGACCTGCGCTTACCGGATACCTG  
TCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCAGCTGTAGGTATCTCAGTTCCGTTGT  
AGGTCGTTCTGCTCCAAGCTGGGCTGTGTGACGAACCCCGCTTCAGCCGACCGCTGCGCTTATCCGG  
TAACATATCGCTTGTAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAACAGG  
ATTAGCAGAGCGAGGTATGTAGGCGGTGTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTACACTA  
GAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTG  
ATCCGGCAAAACAAACACCGCTGGTAGCGGTGGTTTTTTTGTGTTGCAAGCAGCAGATTACGCGCAGAAAA  
AAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGAACGAAAACCTCAGGTT  
AAGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTTAAATGAAGTTT  
TAAATCAATCTAAAGTATATATGAGTAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCT  
TCTCAGCGATCTGTCTATTTCGTTTCATCCATAGTTGCTGACTCCCGCTCGTGTAGATAACTACGATAC  
GGGAGGGCTTACCATCTGGCCCCAGTGTGCAATGATACCGCGAGACCCACGCTCACCGGCTCCAGATTT  
ATCAGCAATAAACAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGGTCTGCAACTTTATCCGCTCCATC  
CAGTCTATTAATTTGTGCGGGAAGCTAGAGTAAGTAGTTCCGCGATTAATAGTTTGGCGAACGTTGTTG  
CCATTGCTACAGGCATCGTGGTGTACGCTCGTCTGTTTGGTATGGCTTCATTAGCTCCGGTTCCCAACG

Table 21 (continued). Nucleotide sequence of plasmid pLP1.

ATCAAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCCTCCGATCGTT  
GTCAGAAGTAAGTTGGCCGCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAATTCTTTACTGTCA  
TGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGATGCG  
GCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGTG  
CTCATCATTGAAAAACGTTCTTCGGGGCGAAAACCTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGA  
TGTAACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTCTGGGTGAGCAAA  
AACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTC  
CTTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTT  
AGAAAAATAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTGACGGGATCCCCTGAGG  
GGGCCCCCATGGGCTAGAGGATCCGGCCTCGGCCTCTGCATAAATAAAAAAATTAGTCAGCCATGAGC

Table 22. Nucleotide sequence of plasmid pLP2.

AATGTAGTCTTATGCAATACTCTTGTAGTCTTGCAACATGGTAACGATGAGTTAGCAACATGCCTTACAA  
 GGAGAGAAAAAGCACCGTGCATGCCGATTGGTGGAGTAAGGTGGTACGATCGTGCCTTATTAGGAAGGC  
 AACAGACGGGTCTGACATGGATTGGACGAACCACTGAATTCCGCATTGCAGAGATATTGTATTTAAGTGC  
 CTAGCTCGATACAATAAACGCCATTTGACCATTACCACATTGGTGTGCACCTCCAAGCTCGAGCTCGTT  
 TAGTGAACCGTCAGATCGCCTGGAGACGCCATCCACGCTGTTTTGACCTCCATAGAAGACACCGGGACCG  
 ATCCAGCTCCCTCGAAGCTAGTCGATTAGGCATCTCCTATGGCAGGAAGAAGCGGAGACAGCGACGAA  
 GACCTCCTCAAGCAGTCAGACTCATCAAGTTTCTCTATCAAAGCAACCCACCTCCCAATCCCCAGGGGA  
 CCGGACAGGCCCGAAGGAATAGAAGAAGAAGGTGGAGAGAGAGACAGAGACAGATCCATTGATTAGTGA  
 ACGGATCCTTAGCACTTATCTGGGACGATCTGCGGAGCCTGTGCCTCTTCAGCTACCACCGCTTGAGAGA  
 CTTACTCTTGATTGTAACGAGGATTGTGGAACCTCTGGGACGCAAGGGGTGGGAAGCCCTCAAATATTGG  
 TGGAAATCTCTACAATATTGGAGTCAGGAGCTAAAGAATAGTGTCTGTAGCTTGCTCAATGCCACAGCTA  
 TAGCAGTAGCTGAGGGGACAGATAGGGTTATAGAAGTAGTACAAGAAGCTTGGCACTGGCCGTCGTTTTTA  
 CAACGTCGTGATCTGAGCCTGGGAGATCTCTGGCTAACTAGGGAACCCACTGCTTAAGCCTCAATAAAGC  
 TTGCCTTGAGTGTCTCAAGTAGTGTGTGCCCGTCTGTGTGTGACTCTGGTAAC TAGAGATCAGGAAAAC  
 CCTGGCGTTACCAACTTAATCGCCTTGACGACATCCCCCTTTCCGCGAGCTGGCGTAATAGCGAAGAGG  
 CCCGACCGGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGCGCCTGATGCGGTATTTCT  
 CCTTACGCATCTGTGCGGTATTTACACCGCATCTCAAGCAACCATAGTACGCGCCCTGTAGCGCGG  
 CATTAAAGCGCGGCGGGTGTGGTGTACGCGCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGC  
 TCCTTTTCGCTTTCTTCCCTTCTTCTCGCCACGTTGCGCGGCTTTCCCGCTCAAGCTCTAAATCGGGGG  
 CTCCCTTTAGGGTTCCGATTTAGTGTCTTACGGCACCTCGACCCCAAAAACTTGATTGGGTGATGGTT  
 CACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAG  
 TGGACTCTTGTTCAAAACCTGGAACAACACTCAACCCTATCTCGGGCTATTCTTTTGATTATAAGGGATT  
 TTGCCGATTTTCGGCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTTAACGCGAATTTTAACAAAA  
 TATTAACGTTTACAATTTTATGGTGCATCTCAGTACAATCTGCTCTGATGCCGATAGTTAAGCGCGC  
 CCGACACCCGCGCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGTCTCCCGCATCCGCTTACAGACAA  
 GCTGTGACCGTCTCCGGGAGCTGCATGTGTGACAGGTTTTTACCCTCATCACCGAAACGCGGAGACGAA  
 AGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTATGATAATAATGGTTTTCTTAGACGTGAGGTGG  
 CACTTTTTCGGGGAATGTGCGCGGAACCCCTATTTGTTTATTTTTCTAAATACATTCAAATATGTATCCG  
 CTCATGAGACAATAACCCCTGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATT  
 TCCGTGTGCGCCCTTATTTCCCTTTTTCGGGCATTTTGCCCTTCTGTTTTGCTCACCCAGAAACGCTGGT  
 GAAAGTAAAGATGTGTAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAAGTGGATCTCAACGCGGT  
 AAGATCCTTGAGAGTTTTTCGCCCCGAAGAACGTTTTTCCAATGATGAGCACTTTTAAAGTTCTGCTATGTG  
 GCGCGGTATTATCCCGTATTGACGCGCGGCAAGAGCAACTCGGTGCGCGCATACACTATTCTCAGAATGA  
 CTTGGTTGAGTACTACCAAGTCACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAAATTATGCAGT  
 GCTGCCATAACCATGAGTGATAACACTGCGGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGC  
 TAACCGCTTTTTTGCACAACATGGGGGATCATGTAACTCGCCTTGATCGTTGGGAACCGGAGCTGAATGA  
 AGCCATACCAACGACGAGCGTGACACCAAGATGCCCTGTAGCAATGGCAACAACGTTGCGCAAACCTATTA  
 ACTGGCGAAGTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCAG  
 GACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATAAATCTGGAGCCGGTGAGCGTGG  
 GTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACG  
 GGGAGTCAGGCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATT  
 GGTAACGTGTCAGACCAAGTTTACTCATATATACTTTAGATTGATTTAAACTTCATTTTAAATTTAAAG  
 GATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTTCGTTCCACTGA  
 GCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCT  
 TGCAAAACAAAAAACCAACCGCTACCAGCGGTGGTTTTGTTGCGGATCAAGAGCTACCAACTCTTTTTCC  
 GAAGGTAAGTGGCTTCAGCAGAGCGCAGATACCAATACTGTTCTTCTAGTGTAGCCGTAGTTAGGCCAC  
 CACTTCAAGAAGTCTGTAGCAGCGCTACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCA  
 GTGGCGATAAGTCGTGTCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGG  
 CTGAACGGGGGGTTCGTGCACACAGCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAG  
 CGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGACGGG  
 TCGGAACAGGAGAGCGCAGGAGGAGCTTCCAGGGGGAACGCTGGTATCTTTATAGTCTCTGCGGTT  
 TCGCCACCTCTGACTTGAGCGTCTGATTTTGTGATGCTGCTCAGGGGGGCGGAGCCTATGGAAAAACGCC  
 AGCAACGCGCGCTTTTACGGTTCTTGGCTTTTGTGCGCTTTTGTCTCACATGTTCTTTCTGCGTTAT  
 CCCCTGATTCTGTGGATAACCGTATTACCGCTTTGAGTGAGCTGATACCGCTCGCCGACGCCGAACGAC  
 CGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCCAATACGCAACCGCCTCTCCCGCGCGT  
 TGGCCGATTCAATTAATGCAGCTGGCACGACAGGTTTCCGACTGGAAAGCGGGCAGTGAGCGCAACGCA  
 TTAATGTGAGTTAGTCACTCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTG

Table 22 (continued). Nucleotide sequence of plasmid pLP2.

TGGAATTGTGAGCGGATAACAATTTACACAGGAAACAGCTATGACATGATTACGAATTCGATGTACGGG  
CCAGATATACGCGTATCTGAGGGGACTAGGGTGTGTTTAGGCGAAAAGCGGGGCTTCGGTTGTACGCGGT  
TAGGAGTCCCCTCAGGATATAGTAGTTTCGCTTTTGCATAGGGAGGGGGA

Table 23. Nucleotide sequence of plasmid pLP/VSVG.

TTGGCCCATTCGATACGTTGTATCCATATCATAATATGTACATTTATATTGGCTCATGTCCAACATTACC  
 GCCATGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCA  
 TATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCCCGCC  
 CATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGT  
 GGAGTATTACGGTAACTGCCCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATT  
 GACGTCAATGACGGTAAATGGCCCGCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCTACTT  
 GGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGGGTTTTGGCAGTACATCAATGGGCG  
 TGGATAGCGGTTTACTCAGCGGATTTCAGTCTCCACCCATTGACGTCAATGGGAGTTTGTTTTGG  
 CACCAAAATCAACGGGACTTTCCAAAATGTCGTAACAACCTCCGCCCCATTGACGCAAAATGGGCGGTAGGC  
 GTGTACGTTGGGAGGTCTATATAAGCAGAGCTCGTTTGTAGAACCGTCAGATCGCTGGAGACGCCATCC  
 ACGCTGTTTTGACCTCCATAGAAGACACCGGGACCGATCCAGCCTCCCCTCGAAGCTTACATGTGGTACC  
 GAGCTCGGATCCTGAGAACTTCAGGGTGAGTCTATGGGACCTTGATGTTTTCTTTCCCTTCTTTTCTA  
 TGGTTAAGTTCATGTCTATAGGAAGGGGAGAAGTAACAGGGTACACATATTGACCAAAATCAGGGTAATTTT  
 GCATTTGTAATTTTAAAAAATGCTTTCTTCTTTAATATACTTTTTGTATTATCTTATTTCTAATACTTT  
 CCCTAATCTCTTTCTTTTCAAGGCAATAATGATACAATGTATCATGCCTCTTTGCACCATTTCTAAGAATA  
 ACAGTGATAAATTTCTGGGTTAAGGCAATAGCAATATTTCTGCATATAAATATTTCTGCATATAAATTTGTA  
 ACTGATGTAAAGGTTTTCATATTGCTAATAGCAGCTACAATCCAGCTACCATTCTGCTTTTATTTTATGG  
 TTGGGATAAGGCTGGATTATTCTGAGTCCAAGCTAGGCCCTTTTGCTAATCATGTTTCATACCTCTTATCT  
 TCCTCCACAGCTCCTGGGCAACGTGCTGGTCTGTGTGCTGGCCCATCACTTTGGCAAAGCAGTGAGAT  
 CTGAATCTGACACTATGAAGTGCCCTTTTGTACTTAGCCTTTTATTATTGTTGGGTGAATTGCAAGTTCA  
 CCATAGTTTTTCCACACAACCAAAAAGGAACTGGAAAAATGTTCTTCTAATTACCATTATTGCCCCGT  
 AAGCTCAGATTTAAATTGGCATAATGACTTAATAGGCACAGCCTTACAAGTCAAAAATGCCCAAGAGTCAC  
 AAGGCTATTCAAGCAGACGGTGGATGTGTCTATGCTTTCCAAATGGGTCACTACTTGTGATTTCCGCTGGT  
 ATGGACCGAAGTATATAACACATTCCATCCGATCCTTCACTCCATCTGTAGAACAATGCAAGGAAAGCAT  
 TGAACAAACGAAACAAGGAACCTGGCTGAATCCAGGCTTCCCTCCTCAAAGTTGTGGATATGCAACTGTG  
 ACGGATGCCGAAGCAGTGATTGTCCAGGTGACTCCTCACCATGTCTGGTTGATGAATACACAGGAGAAT  
 GGGTTGATTACAGTTCATCAACGGAAATGCAGCAATTACATATGCCCCACTGTCCATAACTCTACAAC  
 CTGGCATTCTGACTATAAGGTCAAAGGGCTATGTGATTCTAACCTCATTTCCATGGACATCACCTTCTTC  
 TCAGAGGACGGAGAGCTATCATCCCTGGGAAAGGAGGCGACAGGGTTCAGAAGTAACCTACTTTGCTTATG  
 AAATCGGAGGCAAGGCCTGCAAAAATGCAATACTGCAAGCATTTGGGGAGTCAGACTCCCATCAGGTGTCTG  
 TTTCTGAGATGCTGATAAGGATCTCTTTGCTGCAGCCAGATTCCCTGAATGCCAGAGGGTCAAGTATC  
 TCTGCTCCATCTCAGACCTCAGTGGATGTAAGTCTAATTCAAGGACGTTGAGAGGATCTTGGATTATTCCC  
 TCTGCCAAGAAACCTGGAGCAAAATCAGAGCGGGTCTTCCAATCTCTCCAGTGGATCTCAGCTATCTTGC  
 TCCTAAAAACCCAGGAACCGGTCTGCTTTTACCATAATCAATGGTACCCTAAAATACCTTTGAGACCAGA  
 TACATCAGAGTCGATATTGCTGCTCCAATCCTCTCAAGAATGGTCGGAATGATCAGTGGAACTACCACAG  
 AAAGGGAACCTGTGGGATGACTGGGCACCATATGAAGACGTGGAAATTTGGACCCAATGGAGTTCTGAGGAC  
 CAGTTCAGGATATAAGTTTCCCTTATACATGATTGGACATGGTATGTTGGACTCCGATCTTCATCTTAGC  
 TCAAAGGCTCAGGTGTTTGAACATCCTCACATTCAAGACGCTGCTTCGCAACTTCTGATGATGAGAGTT  
 TATTTTTTGGTGATACTGGGCTATCCAAAATCCAATCGAGCTTGTAGAAGGTTGGTTCACTAGTTGGAA  
 AAGCTCTATTGCCTCTTTTTTCTTTATCATAGGGTTAATCATTGGACTATTCTTGGTTCTCCGAGTTGGT  
 ATCCATCTTTGCATTAAATTAAGACACACCAAGAAAAGACAGATTTATACAGACATAGAGATGAACCGAC  
 TTGGAAAGTAACCTCAATCCTGCACAACAGATTCTTCATGTTTGGACCAATCAACTTGTGATACCATGC  
 TCAAAGAGGCCTCAATTATATTGAGTTTTTAATTTTATGAAAAAAGGAAATTTGAGGAAATTTGAGGAC  
 CACCAGTGCAGGCTGCCTATCAGAAAGTGGTGGCTGGTGGCTAATGCCCTGGCCCAAGTATCACTA  
 AGCTCGCTTTCTTGCTGTCCAATTTCTATTAAAGGTTTCTTTGTTCCCTAAGTCCAATACTACTAACTGGG  
 GGATATTATGAAGGGCCTTGAGCATCTGGATTCTGCCTAATAAAAAACATTTATTTTTCATTGCAATGATG  
 TATTTAAATTATTTCTGAATATTTTACTAAAAAGGGAATGTGGGAGGTCACTGCAATTTAAACATAAAGA  
 AATGAAGAGCTAGTTCAAACCTTGGGAAAATACACTATATCTTAACTCCATGAAAGAAGGTGAGGCTGC  
 AAACAGCTAATGCATTTGGCAACAGCCCCCTGATGCCTATGCCTTATTTCATCCCTCAGAAAAGGATTCAA  
 GTAGAGGCTTGATTGGAGGTTAAAGTTTTGCTATGCTGTATTTTACATTACTTATTGTTTATAGCTGCTC  
 TCATGAATGCTTTTTCACTACCCATTTGCTTATGCTCATCTCTCAGCCTTGACTCCACTCAGTTCTCTT  
 GCTTAGAGATACCACCTTTCCCTGAAGTGTTTCTTCCATGTTTTACGGCGAGATGGTTTTCTCTCGCT  
 GGCCACTCAGCCTTAGTTGTCTCTGTGCTTATAGAGGTCTACTTGAAGAAGGAAAAACAGGGGGCATG  
 GTTTGACTGTCTGTGAGCCCTTCTTCCCTGCCTCCCCCACTCACAGTGACCCGGAATCCCTCGACATGG  
 CAGTCTAGCACTAGTGCGGCCGAGATCTGCTTCTCGCTCACTGACTCGCTGCGCTCGGTCTGGCT  
 GCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGA  
 AAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCTTGCTGGCGTTTTTCC

Table 23 (continued). Nucleotide sequence of plasmid pLP/VSVG.

ATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGG  
ACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCCGACCCTGCCGCTT  
ACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCTGTAGGTATC  
TCAGTTCGGTGTAGGTCTGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTCAGCCCGACCGCTG  
CGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCC  
ACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAATC  
ACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGT  
TGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTGGTTTGTGTTTGCAGCAGCAGATT  
ACGCGCAGAAAAAAGGATCTCAAGAAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACG  
AAAATCAGCTTAAGGGATTTTGGTTCATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTA  
AAAATGAAGTTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATC  
AGTGAGGCACCTATCTCAGCGATCTGTCTATTTCTGTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGA  
TAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACC  
GGCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGGTCTTCAACTTTA  
TCCGCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCCGCCAGTTAATAGTTTGC  
GCAACGTTGTTGCCATTGCTACAGGCATCGTGGTGTACGCTCGTCTGTTGGTATGGCTTCATTAGCTC  
CGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGT  
CCTCCGATCGTTGTCAGAAGTAAGTTGGCCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAATT  
CTCTTACTGTATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGA  
ATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAATACCGCGCCACATAGCAGA  
ACTTTAAAGTGCTCATATTGGAAAACGTTCTTCGGGGCGAAAACTCTCAAGGATCTTACCGCTGTTGA  
GATCCAGTTCGATGTAACCCACTCGTGCAACCAACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTC  
TGGGTGAGCAAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATA  
CTCATACTCTTCCTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGATACATAT  
TTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCGAAAAGTGCCACCTGACGG  
GATCCCCTGAGGGGGCCCCCATGGGCTAGAGGATCCGGCCTCGGCCTCTGCATAAATAAAAAAATTAGT  
CAGCCATGAGC



Table 28. Nucleotide sequence of plasmid pcDNA<sup>TM</sup>6.2/V5-DEST.

GACGGATCGGGAGATCTCCCGATCCCTATGGTGCCTCTCAGTACAATCTGCTCTGATGCCGCATAGTT  
 AAGCCAGTATCTGCTCCCTGCTTGTGTGTTGGAGTTCGCTGAGTAGTGCGCGAGCAAAATTTAAGCTACA  
 ACAAGGCAAGGCTTGACCGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTTGCCTGCTTCGCG  
 ATGTACGGGCCAGATATACGCGTTGACATTGATTATGACTAGTTATTAATAGTAATCAATTACGGGGTC  
 ATTAGTTCATAGCCCATATATGGAGTTCGCGTTACATAAATTACGGTAAATGGCCCCGCTGGCTGACCG  
 CCCAACGACCCCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCC  
 ATTGACGTCAATGGGTGGAGTATTTACGGTAACTGCCCACTTGGCAGTACATCAAGTGTATCATATGCC  
 AAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCCGCTGGCATTATGCCAGTACATGACCTTA  
 TGGGACTTTCTTATGGCAGTACATCTACGTATTACAAATGTTCTTCGGGTGATGCTGCCAATTAGTGA  
 AGTACATCAATGGGCGTGGATAGCGGTTTGACTCACGGGGATTTCGAAGTCTCCACCCCATGACGTCAA  
 TGGGAGTTTGTTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCTGAACAACCTCCGCCCCATTGACG  
 CAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTATATAAGCAGAGCTCTCTGGCTAACTAGAGAACCCA  
 CTGCTTACTGGCTTATCGAAATTAATACGACTCACTATAGGGAGACCCAAGCTGGCTAGTTAAGCTATCA  
 ACAAGTTTGTACAAAAAGCTGAACGAGAAACGTAAATGATATAAATATCAATATATTAAATTAGATTT  
 TGCATAAAAAACAGACTACATAAATACTGTAAACACAACATATCCAGTCACTATGAATCAACTAGCTTA  
 TGGTATTAGTGACCTGTAGTCGACCGACAGCCTTCCAAATGTTCTTCGGGTGATGCTGCCAATTAGTGA  
 ACCGACAGCCTTCCAAATGTTCTTCTCAAAACGGAATCGTCGTATCCAGCCTACTCGCTATTGTCCTCAAT  
 GCCGTATTAAATCATAAAAAGAAATAAGAAAAAGAGGTGCGAGCCTCTTTTTTGTGTGACAAAATAAAAA  
 CATCTACCTATTTCATATACGCTAGTGTCTATAGTCTCTGAAAATCATCTGCATCAAGAACAATTTACAACT  
 CTTTACTTTTTCTTACAAAGTCGTTTCGGCTTCATCTGGATTTTTCAGCCTCTATACTTACTAAACGTGAT  
 AAAGTTTCTGTAATTTCTACTGTATCGACCTGCAGACTGGCTGTGTATAAGGGAGCCTGACATTTATATT  
 CCCCAGAACATCAGGTTAATGGCGTTTTTGTATGTCATTTTCGCGGTGGCTGAGATCAGCCACTTCTTCCC  
 CGATAACGGAGACCGGCACACTGGCCATATCGGTGTTTCATCATGCGCCAGCTTTTCATCCCCGATATGCAC  
 CACCGGGTAAAGTTTACGGGAGACTTTATCTGACAGCAGACGTGCCTGGCCAGGGGGATCACCATCCGT  
 CGCCCCGGCGTGTCAATAATATCACTCTGTACATCCACAAACAGACGATAACGGCTCTCTCTTTTATAGG  
 TGTAAACCTTAACTGCATTTTACCAGTCCCTGTTCTCGTCAGCAAAAGAGCCGTTTCATTTCAATAAAC  
 GGGGACCTCAGCCATCCCTTCCCTGATTTTCCGCTTTCCAGCGTTTCGGCACGCAGACGACGGGCTTCATT  
 CTGCATGGTTGTGCTTACCAGACCGGAGATATTGACATCATATATGCCCTTGAGCAACTGATAGCTGTGCG  
 TGTCAACTGTCACTGTAATACGCTGCTTCATAGCACACCTCTTTTGGACATACTTCGGGTATACATATCA  
 GTATATATTCTTATACGCAAAAATCAGCGCGCAAAATACGCATACTGTTATCTGGCTTTTATAGCCGG  
 ATCCACGCGATTACGCCCCCGCTGCCACTCATCGCAGTACTGTTGTAATTCATTAAGCATTTCTGCCGAC  
 ATGGAAGCCATCACAGACGGCATGATGAACCTGAATCGCCAGCGGCATCAGCACCTTGTGCGCTTGCCTA  
 TAATATTTGCCCATGGTGAACCGGGGCGAAGAAGTTTCCATATTGGCCACGTTTAAATCAAAACTGG  
 TGAAACTCACCCAGGGATTGGCTGAGACGAAAAACATATTTCTCAATAAACCTTTAGGGAATAGGCCAG  
 GTTTTACCGTAACACGCCACATCTTCCGAATATATGTGTAGAACTGCCGGAATCGTCGTGGTATTCA  
 CTCAGAGCGATGAAAACGTTTTCAGTTTGCTCATGGAAAACGGTGTAAACAGGGTGAACACTATCCCATA  
 TCACCAGCTCACCCTCTTTTATTGCCATACGGAATTCGGATGAGCATTTCATCAGGCGGGCAAGAATGTG  
 AATAAAGGCGGGATAAAACCTTGTGCTTATTTTTCTTTACGGTCTTTAAAAAGGCCGTAATATCCAGCTGA  
 ACGGTCTGGTTATAGGTACATTGAGCAACTGACTGAAATGCCTCAAAATGTTCTTTACGATGCCATTGGG  
 ATATATCAACGGTGGTATATCCAGTGATTTTTTCTCCATTTTAGCTTCTTAGCTCCTGAAAATCTCGA  
 TAACTCAAAAATACGCCCCGGTAGTGATCTTATTTTCATTATGGTGAAAGTTGGAACCTCTTACGTGCCGA  
 TCAACGTCTCATTTTCGCCAAAAGTTGGCCCCAGGGCTTCCCGGTATCAACAGGGACACCAGGATTTATTT  
 ATTCGCGAAGTGATCTTCCGTACAGGTATTTATTCGGCGCAAAGTGCGTCGGGTGATGCTGCCAATT  
 AGTCGACTACAGGTCACTAATACCATCTAAGTAGTTGATTTCATAGTGACTGGATATGTTGTGTTTACAG  
 TATTATGTAGTCTGTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTACGTTTCTC  
 GTTCAGCTTTCTTGTACAAAGTGGTTGATCTAGAGGGCCCGGTTTCGAAGGTAAGCCTATCCCTAACCC  
 TCTCCTCGGTCTCGATTCTACGCGTACCGGTTAGTAATGAGTTTAAACGGGGGAGGCTAACTGAAACACG  
 GAAGGAGACAATACCGGAAGGAACCCGCGCTATGACGGCAATAAAAAGACAGAATAAAACGCACGGGTGT  
 TGGGTCTGTTGTTTATAAACGCGGGGTTCCGTCACAGGGCTGGCACTCTGTGATACCCACCGAGACCC  
 CATTTGGGGCCAATACGCCCCGCTTTCTTCTTTTCCCCACCCACCCCAAGTTTCGGGTGAAGGCCAG  
 GGCTCGCAGCCAACGTGCGGGCGGCAGGCCCTGCCATAGCAGATCTGCGCAGCTGGGGCTCTAGGGGGTA  
 TCCCCACGCGCCCTTAGCGCGCATTAAGCGCGGCGGGTGTGGTGGTTACGCGCAGCGTGACCGCTACA  
 CTTGCCAGCGCCCTAGCGCCCGCTCTTTCTGCTTTCTTCCCTTCTTTCTCGCCACGTTTCGCCGCTTTC  
 CCCGTCAAGCTCTAAATCGGGGCATCCCTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCGACCCCAA  
 AAACTTTGATTAGGGTGATGGTTTACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTTCGCCCTTTGACG  
 TTGGAGTCCACGTTCTTTAATAGTGGACTCTTGTTCAAAACCTGGAACAACACTCAACCCTATCTCGGTCT  
 ATTCCTTTGATTTATAAGGGATTTTGGGGATTTTCGGCTATTGGTTAAAAAATGAGCTGATTTTAAACAAA  
 ATTTAACGCGAATTAATCTGTGGAATGTGTGTCAGTTAGGGTGTGGAAAGTCCCCAGGCTCCCCAGCAG

Table 28 (continued). Nucleotide sequence of plasmid pcDNA<sup>TM</sup>6.2/V5-DEST.

GCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAAGTCCCCAGGCTCCCCAGC  
AGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCATAGTCCCGCCCCCTAACTCCGCCCATC  
CCGCCCTAACTCCGCCCAGTTCGCCCATTTCTCCGCCCATGGCTGACTAATTTTTTTTATTTATGTCAG  
AGGCCGAGGCCGCTCTGCCTCTGAGCTATTCAGAAAGTAGTGAGGAGGCTTTTTTGGAGGCCCTAGGCTT  
TTGCAAAAAGCTCCCGGGAGCTTGTATATCCATTTTCGGATCTGATCAGCACGTGTTGACAATTAATCAT  
CGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAATAAACCATGGCCAAGCCTTTGTCTCA  
AGAAGAATCCACCCTCATTGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAAGACTACAGC  
GTCCGCAGCGCAGCTCTCTCTAGCGACGGCCGCATCTTCACTGGTGTCAATGTATATCATTCTACTGGGG  
GACCTTGTGTCAGAACTCGTGGTGTCTGGGCACTGCTGCTGCTGCGGCAGCTGGCAACCTGACTTGTATCGT  
CGCGATCGGAAATGAGAACAGGGGCATCTTGAGCCCCTGCGACGGTGCCGACAGGTGCTTCTCGATCTG  
CATCCTGGGATCAAAGCCATAGTGAAGGACAGTGATGGACAGCCGACGGCAGTTGGGATTCTGTAATTGC  
TGCCCTCTGGTTATGTGTGGGAGGGCTAAGCACTTCGTGGCCGAGGAGCAGGACTGACACGTGCTACGAG  
ATTTTCGATTCCACCGCCGCTTCTATGAAAGGTTGGGCTTCGGAATCGTTTTCCGGGACGCGCGCTGGAT  
GATCTCCAGCGCGGGGATCTCATGCTGGAGTTCTTCGCCCAACCCAACTGTTTTATTGCAGCTTATAAT  
GGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATTTTTTTCACTGCATTCTAGTTGTG  
GTTTTGTCCAACTCATCAATGTATCTTATCATGTCTGTATACCGTCGACCTCTAGCTAGAGCTTGGCGTA  
ATCATGGTCATAGCTGTTTTCTGTGTGAAATTGTTATCCGCTCACAAATCCACACAACATACGAGCCGGA  
AGCATAAAGTGTAAGCCTGGGGTGCCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGC  
CCGCTTTTCAGTCCGGGAAACCTGTCTGTCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAGAGCGG  
TTTTGCGTATTGGGCGCTCTTCCGCTTCTCTGCTCACTGACTCGCTGCGCTCGGTGCTTCCGCTGCGGCGA  
GCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACA  
TGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCT  
CCGCCCCCTGACGAGCATCACAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAA  
AGATACCAGGCGTTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCTGTTCCGACCCCTGCCGCTTACCGGAT  
ACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGCGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTT  
GGTGTAGGTGCTGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTACGCCCGACCGCTGCGCCTTA  
TCCGGTAACCTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTA  
ACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCTTAACACGGCTA  
CACTAGAAGAACAGTATTTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGC  
TCTTGATCCGGCAAAACAAACACCGCTGGTAGCGGTTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAA  
AAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGAAAACCTCACG  
TTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAATGAAGT  
TTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCAC  
CTATCTCAGCGATCTGTCTATTTCTGTTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAAATACGAT  
ACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTACCCGGCTCCAGAT  
TTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGGTCTGCAACTTTATCCGCTCCA  
TCCAGTCTATTAAATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCCGCCAGTTAATAGTTTGCGCAACGTTGT  
TGCCATTGCTACAGGCATCGTGGTGTACGCTCGTCGTTTGGTATGGCTTCATTACAGCTCCGGTTCCCAA  
CGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAAAAGCGGTTAGCTCCTTCGGTCTTCCGATCG  
TTGTGAGAAGTAAGTTGGCCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAATTCTTACTGT  
CATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAAGTCAACCAAGTCATTCTGAGAATAGTGTATG  
CGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAATACCGCGCCACATAGCAGAACTTTAAAG  
TGCTCATCATTGAAAAACGTTCTTCGGGGCGAAAACCTCAAGGATCTTACCGCTGTTGAGATCCAGTTC  
GATGTAACCCACTCGTGACCCAACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTCTGGGTGAGCA  
AAAAAGGAAGGCAAAATGCCGCAAAAAGGGAATAAGGGCGACACGAAATGTTGAATACTCATACTCT  
TCCTTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTAT  
TTAGAAAAATAAACAAATAGGGGTTCGCGCACATTTCCCGGAAAAGTGCCACCTGACGTC

Table 29. Nucleotide sequence of plasmid pcDNA™6.2/GFP-DEST.

GACGGATCGGGAGATCTCCCGATCCCTATGGTGCACCTCTCAGTACAATCTGCTCTGATGCCGCATAGTT  
 AAGCCAGTATCTGCTCCCTGCTTGTGTGTTGGAGGTCGCTGAGTAGTGCGCGAGCAAAATTTAAGCTACA  
 ACAAGGCAAGGCTTGACCGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGCTTCGCG  
 ATGTACGGGCCAGATATACGCGTTGACATTGATTATGACTAGTTATTAATAGTAATCAATTACGGGGTC  
 ATTAGTTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCGCTGGCTGACCG  
 CCCAACGACCCCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCC  
 ATTGACGTCAATGGGTGGAGTATTTACGGTAAATGCCCACTTGGCAGTACATCAAGTGTATCATATGCC  
 AAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCTGGCATATTGCCAGTACATGACCTTA  
 TGGGACTTTCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGTGCGGTTTTGGC  
 AGTACATCAATGGGCGTGGATAGCGGTTTGACTCACGGGGATTTCGAAGTCTCCACCCCATTGACGTCAA  
 TGGGATTTTGTGTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCGTAACAACCTCCGCCCCATTGACG  
 CAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCCTATAAGCAGAGCTCTCTGGCTAACTAGAGAACCA  
 CTGCTTACTGGCTTATCGAAATTAATACGACTCACTATAGGGAGACCCAAAGCTGGCTAGTTAAGCTATCA  
 ACAAGTTTGTACAAAAAGCTGAACGAGAAACGTAAAATGATATAAAATATCAATATATTAAATTAGATTT  
 TGCATAAAAAACAGACTACATAACTGTAAAACACAACATATCCAGTCACTATGAATCAACTACTTAGA  
 TGGTATTAGTGACCTGTAGTCGACCGACAGCCTTCCAAATGTTCTTCGGGTGATGCTGCCAACTTAGTCG  
 ACCGACAGCCTTCCAAATGTTCTTCTCAAACGGAATCGTCGTATCCAGCCTACTCGCTATTGTCTCTCAAT  
 CGCGTATTAAATCATAAAAAGAAATAGAAAAAGAGGTGCGAGCCTCTTTTTGTGTGACAAAAATAAAAA  
 CATCTACCTATTTCATATACGCTAGTGTCTATAGTCTGAAAATCATCTGCATCAAGAACAAATTTCAACAA  
 CTTTATACTTTTCTCTTACAAGTCGTTTCGGCTTTCATCTGGATTTTTCAGCCTCTATACTTACTAAACGTGAT  
 AAAGTTTCTGTAATTTCTACTGTATCGACCTGCAGACTGGCTGTGTATAAGGGAGCCTGACATTTATATT  
 CCCCAGAACATCAGGTTAATGGCGTTTGTGATGTCATTTTCGCGGTGGCTGAGATCAGCCACTTCTTCCC  
 CGATAACGGAGACCGGCACACTGGCCATATCGGTGGTCATCATGCCCAGCTTTCATCCCCGATATGCAC  
 CACCGGGTAAAGTTCACGGGAGACTTTATCTGACAGCAGACGTGCACTGGCCAGGGGGATCACCATCCGT  
 CGCCCGGGCGTGTCAATAATATCACTCTGTACATCCACAACAGACGATAACGGCTCTCTCTTTTATAGG  
 TGTAACCTTTAAACTGCATTTCCACAGTCCCTGTTCTCTCGTCAGCAAAAGAGCCGTTCAATTTCAAAACC  
 GGGCGACCTCAGCCATCCCTTCTGATTTTCCGCTTTCAGCGTTCGGCACGCAGACGACGGGCTTCATT  
 CTGCATGGTTGTGCTTACCAGACCGGAGATATTGACATCATATATGCCCTTGAGCAACTGATAGCTGTGCG  
 TGTCAACTGTCACTGTAATACGTGCTTCATAGCACACCTCTTTTGGACATACTTCGGGTATACATATCA  
 GTATATATTCTTATACCGCAAAATCAGCGCGCAAAATACGCATACTGTTATCTGGCTTTTAGTAAGCCCG  
 ATCCACGCGATTACGCCCCGCTTCCACTCATCGCAGTACTGTTGTAATTCATTAAGCATTCTGCCGAC  
 ATGGAAGCCATCAGACGGCATGATGAACCTGAATCGCCAGCGGCATCAGCACCTTGTGCGCTTTCGCTG  
 TAATATTTGCCCATGGTGAAGAACGGGGCGAAGAGTTGTCCATATTGGCCACGTTTAAATCAAAACCTG  
 TGAACTCACCCAGGGATTGGCTGAGACGAAAAACATATTTCTCAATAAAACCTTTAGGGAAATAGGCCAG  
 GTTTTTCACCGTAACACGCCACATCTTGCGAATATATGTGTAGAAACTGCCGGAATCGTCGTGGTATTCA  
 CTCCAGAGCGATGAAAACGTTTCAGTTTGTCTCATGGAAGACGGTGTAAACAAGGGTGAACACTATCCCAT  
 TCACCAGCTCACCGTCTTTTCAATGCCATACGGAATTCGGGATGAGCATTTCATCAGCGGGCAAGAATGTG  
 AATAAAGCCGGATAAACTTGTGCTTATTTTCTTTACGGTCTTTAAAAAGGCCGTAATATCCAGCTGA  
 ACGTCTGCTTATAGGTACATTGAGCAACTGACTGAAATGCCTCAAAATGTTCTTTACGATGCCATTGGG  
 ATATATCAACGGTGGTATATCCAGTGATTTTCTTCCATTTTAGCTTCTTAGCTCCTGAAAATCTCCGA  
 TAACTCAAAAAATACGCCCCGGTAGTGATCTTATTTTCAATATGGTGAAAGTTGGAACCTCTTACGTGCCGA  
 TCAACGTCTCATTTTCGCCAAAAGTTGGCCAGGGCTTCCCGGTATCAACAGGGACACCAGGATTTATTT  
 ATTCTGCGAAGTGATCTTCCGTACAGGTATTTATTCCGGCGCAAAGTGCGTCGGGTGATGCTGCCAACTT  
 AGTCGACTACAGGTCACTAATACCATCTAAGTAGTTGATTCATAGTGACTGGATATGTTGTGTTTTACAG  
 TATTATGTAGTCTGTTTTTTATGCAAAATCTAATTTAATATATGATATTTATATCATTTTTACGTTTCTC  
 GTTCAGCTTTCTTGTACAAAGTGGTTGATCTAGAGGGCCCCGGCTAGCAAAGGAGAAGAACTTTTAC  
 TGGAGTTGTCCCAATCTTGTGTAATTAGATGGTGATGTTAATGGGCACAAATTTCTGTGAGTGGAGAG  
 GGTGAAGGTGATGCTACATACGGAAGCTTACCCTTAAATTTATTTGCACTACTGGAAAACCTACCTGTTT  
 CATGGCCAACACTTGTCACTACTTTCTCTTATGGTGTTCATGCTTTTCCCGTTATCCGGATCATATGAA  
 ACGGCATGACTTTTTTCAAGAGTGCCATGCCGAAGGTTATGTACAGGAACGCATATATCTTTCAAAGAT  
 GACGGGAACCTACAAGACGCGTGCTGAAGTCAAGTTTGAAGGTGATACCTTGTTAATCGTATCGAGTTAA  
 AAGGTATTGATTTTAAAGAAGATGGAACATTTCTCGGACACAACTCGAGTACAACATAACTCACACAA  
 TGTATACATCAGGCAGACAAACAAAGATGGAATCAAAGCTAACTTCAAAATTCGTACAAACATTGAA  
 GATGGATCCGTTCAACTAGCAGACCATTTCAACAAATACTCCAATTGGCGATGGCCCTGTCTCTTTAC  
 CAGACAACCATTTACCTGTGACACAACTCTGCCCTTTCGAAAGATCCCAACGAAAAGCGTGACCACATGGT  
 CCTTCTTGAGTTTGTAACTGCTGCTGGGATTACACATGGCATGGATGAATAGTAATGAGTCCACGTTTAA  
 ACGGGGGAGGCTAACTGAAACACGGAAGGAGACAATACCGGAAGGAACCCGCGCTATGACGGCAATAAAA  
 AGACAGAATAAAACGCACGGGTGTTGGGTGCTTTGTTTCAAAACGCGGGGTTCCGTCCCAGGGCTGGCAC  
 TCTGTGATACCCACCCAGACCCCATTTGGGGCAATACGCCCGCGTTTCTTCTTTTCCCCACCCACCC

Table 29. Nucleotide sequence of plasmid pcDNA™6.2/GFP-DEST.

CCCCAGTTTCGGGTGAAGGCCAGGGCTCGCAGCCAACGTCGGGGCGGCAGGCCCTGCCATAGCAGATCT  
 GCGCAGCTGGGGCTCTAGGGGTATCCCCACGCGCCCTGTAGCGCGCATTAAGCGCGCGGGTGTGGTG  
 GTTACGCGCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCCT  
 TTCTCGCCACGTTTCGCGGGCTTTCCTCCGTCAGCTCTAAATCGGGGCATCCCTTTAGGGTTCCGATTTAG  
 TGCTTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTACAGTAGTGGGCCATCGCCCTGA  
 TAGACGGTTTTTCGCCCCCTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACTCTTGTTCAAACTGGAA  
 CAACACTCAACCCCTATCTCGGTCTATTCTTTTGATTTATAAGGGATTTTGGGGATTTCGGCCCTATTGGTT  
 AAAAAATGAGCTGATTTTAAACAAAAATTAACGCGAATTAATTCTGTGGAATGTGTGTCAAGTTAGGGTGTG  
 GAAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGGTG  
 TGGAAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCATA  
 GTCCCGCCCTAACTCCGCCCCTCCGCCCCCTAACTCCGCCCAGTTCCGCCCATTCTCCGCCCCATGGCT  
 GACTAATTTTTTTTATTTATGAGAGGCCGAGCCCGCTCTGCCTCTGAGCTATTCCAGAAGTAGTGAGG  
 AGGCTTTTTTGGAGGCCTAGGCTTTTGCAAAAGCTCCCGGGAGCTTGTATATCCATTTTCGGATCTGAT  
 CAGCAGTGTTGACAATTAATCATCGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAATA  
 AACCATGGCCAGCCTTTGTCTCAAGAAGAATCCACCTCATTTGAAAGAGCAACGGCTACAATCAACAGC  
 ATCCCCATCTCTGAAGACTACAGCGTCGCCAGCGCAGCTCTCTTAGCGACGGCCGCATCTTCACTGGTG  
 TCAATGTATATCATTTTTACTGGGGGACCTTGTGCAGAACTCGTGGTGCTGGGCACTGCTGCTGCTGCGGC  
 AGCTGGCAACCTGACTTGTATCGTCGCGATCGGAAATGAGAACAGGGGCATCTTGAGCCCTGCGGACGG  
 TGCCGACAGGTGCTTCTCGATCTGCATCTCGGATCAAAGCCATAGTGAAGGACAGTGATGGACAGCCGA  
 CGGCAGTTGGGATTCTGTAATTGCTGCCCTCTGGTTATGTGTGGGAGGGCTAAGCACTTCGTGGCCGAGG  
 AGCAGGACTGACACGTGCTACGAGATTTGATTCACCGCCGCTTCTATGAAAGGTTGGGCTTCGGAAT  
 CGTTTTCCGGGACGCCGCTGGATGATCCTCCAGCGCGGGATCTCATGCTGGAGTTCTTCGCCCAACCC  
 AACTTGTATTATGACGCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCAT  
 TTTTTTCACTGCATTCTAGTTGTGGTTGTCCAACTCATCAATGTATCTTATCATGTCTGTATACCGTC  
 GACCTCTAGCTAGAGCTTGGCGTAATCATGGTCATAGCTGTTTCTGTGTGAAATTGTATCCGCTCACA  
 ATTCCACACAACATACGAGCCGAAGCATAAAGTGTAAGCCCTGGGGTGCCTAATGAGTGAGCTAACTCA  
 CATTAAATTGCGTTGCGCTCACTGCCCGCTTCCAGTCGGGAAACCTGTGCTGCCAGCTGCATTAATGAAT  
 CGGCCAACGCGCGGGGAGAGCGGTTTGCATATTGGGCGCTCTTCCGCTTCTCGCTCACTGACTCGCTG  
 CGTTCGGTTCGGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGCGGTAATACGGTTATCCACAGAAT  
 CAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGC  
 GTTGTGCGGCTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGG  
 TGGCGAAACCCGACAGGACTATAAGATACCAGCGGTTTCCCTGGAAGCTCCCTCGTGCCTCTCCTG  
 TTCCGACCTGCGCTTACCGGATACCTGTCCGCTTCTCCTTTCGGGAAGCGTGGCGCTTCTCATAG  
 CTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTGCTTCCGCTCCAAGCTGGGCTGTGTGCACGAACCCCC  
 GTTCAGCCCGACCGCTGCGCTTATCCGGTAACTATCGTCTTGAGTCCAACCGGTAAAGACAGCACTTAT  
 CGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTT  
 GAAGTGTGGCCTAACTACGGCTACACTAGAAGAACAGTATTGGTATCTGCGCTCTGCTGAAGCCAGTT  
 ACCTTCGGAAGAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTTTTTTTGT  
 GCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTACGGGGTCTGA  
 CGCTCAGTGGAACGAAAACCTACGTTAAGGGATTTTGGTTCATGAGATTATCAAAAAGGATCTTACCTAG  
 ATCCTTTTAAATTAATAATGAAGTTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTT  
 ACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTCGTTTCATCCATAGTTGCGCTGACT  
 CCCCCTCGTGTAGATAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATAACCGGA  
 GACCCACGCTCACCGGCTCCAGATTATCAGCAATAAACAGCCAGCCGGAAGGGCCGAGCGCAGAAGTG  
 GTCCTGCAACTTTATCCGCCCTCCATCCAGTCTATTAATTGTTGCCGGAAGCTAGAGTAAGTAGTTCGCC  
 AGTTAATAGTTTGCACAACGTTGTTGCCATTGCTACAGGCATCGTGGTGTACGCTCGTCTGTTGTTATG  
 GCTTCATTCAGCTCCGTTTCCCAACGATCAAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGG  
 TTAGCTCCTTCGGTCTCCGATCGTTGTGAGAAGTAAGTTGGCCGAGTGTTATCACTCATGGTTATGGC  
 AGCACTGCATAATTCTTACTGTCTATGCCATCCGTAAGATGCTTTCTGTGACTGGTGAGTACTCAACC  
 AAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGGCCGCGCTCAATACGGGATAATACCG  
 CGCCACATAGCAGAACTTTAAAAGTGCTCATCATTTGGAAAACGTTCTTCGGGGCGAAAACCTCTCAAGGAT  
 CTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTTACT  
 TTCACACAGCTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAGGGGAATAAGGGCGACAC  
 GGAAATGTTGAATACTCATACTCTTCTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCAT  
 GAGCGGATACATATTGAATGTATTTAGAAAAATAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAA  
 GTGCCACCTGACGTC

Table 30. Amino acid sequence of a polypeptide having  $\beta$ -lactamase activity.

|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |  |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|--|
| Met | Gly | His | Pro | Glu | Thr | Leu | Val | Lys | Val | Lys | Asp | Ala | Glu | Asp | Gln |  |
| 1   |     |     |     | 5   |     |     |     |     | 10  |     |     |     |     | 15  |     |  |
| Leu | Gly | Ala | Arg | Val | Gly | Tyr | Ile | Glu | Leu | Asp | Leu | Asn | Ser | Gly | Lys |  |
|     |     |     | 20  |     |     |     |     | 25  |     |     |     |     | 30  |     |     |  |
| Ile | Leu | Glu | Ser | Phe | Arg | Pro | Glu | Glu | Arg | Phe | Pro | Met | Met | Ser | Thr |  |
|     |     | 35  |     |     |     |     | 40  |     |     |     |     | 45  |     |     |     |  |
| Phe | Lys | Val | Leu | Leu | Cys | Gly | Ala | Val | Leu | Ser | Arg | Asp | Asp | Ala | Gly |  |
|     | 50  |     |     |     | 55  |     |     |     |     |     | 60  |     |     |     |     |  |
| Gln | Glu | Gln | Leu | Gly | Arg | Ile | His | Tyr | Ser | Gln | Asn | Asp | Leu | Val |     |  |
| 65  |     |     |     | 70  |     |     |     |     | 75  |     |     |     |     | 80  |     |  |
| Glu | Tyr | Ser | Pro | Val | Thr | Glu | Lys | His | Leu | Thr | Asp | Gly | Met | Thr | Val |  |
|     |     |     |     | 85  |     |     |     |     | 90  |     |     |     |     | 95  |     |  |
| Arg | Glu | Leu | Cys | Ser | Ala | Ala | Ile | Thr | Met | Ser | Asp | Asn | Thr | Ala | Ala |  |
|     |     |     | 100 |     |     |     | 105 |     |     |     |     |     | 110 |     |     |  |
| Asn | Leu | Leu | Leu | Thr | Thr | Ile | Gly | Gly | Pro | Lys | Glu | Leu | Thr | Ala | Phe |  |
|     |     | 115 |     |     |     |     | 120 |     |     |     |     | 125 |     |     |     |  |
| Leu | His | Asn | Met | Gly | Asp | His | Val | Thr | Arg | Leu | Asp | His | Trp | Glu | Pro |  |
|     | 130 |     |     |     |     | 135 |     |     |     |     | 140 |     |     |     |     |  |
| Glu | Leu | Asn | Glu | Ala | Ile | Pro | Asn | Asp | Glu | Arg | Asp | Thr | Thr | Met | Pro |  |
| 145 |     |     |     |     | 150 |     |     |     |     | 155 |     |     |     |     | 160 |  |
| Val | Ala | Met | Ala | Thr | Thr | Leu | Arg | Lys | Leu | Leu | Thr | Gly | Glu | Leu | Leu |  |
|     |     |     |     | 165 |     |     |     |     | 170 |     |     |     |     | 175 |     |  |
| Thr | Leu | Ala | Ser | Arg | Gln | Gln | Leu | Ile | Asp | Trp | Met | Glu | Ala | Asp | Lys |  |
|     |     |     | 180 |     |     |     | 185 |     |     |     |     |     | 190 |     |     |  |
| Val | Ala | Gly | Pro | Leu | Leu | Arg | Ser | Ala | Leu | Pro | Ala | Gly | Trp | Phe | Ile |  |
|     | 195 |     |     |     |     |     | 200 |     |     |     |     | 205 |     |     |     |  |
| Ala | Asp | Lys | Ser | Gly | Ala | Gly | Glu | Arg | Gly | Ser | Arg | Gly | Ile | Ile | Ala |  |
|     | 210 |     |     |     |     | 215 |     |     |     |     | 220 |     |     |     |     |  |
| Ala | Leu | Gly | Pro | Asp | Gly | Lys | Pro | Ser | Arg | Ile | Val | Val | Ile | Tyr | Thr |  |
| 225 |     |     |     | 230 |     |     |     |     |     | 235 |     |     |     |     | 240 |  |
| Thr | Gly | Ser | Gln | Ala | Thr | Met | Asp | Glu | Arg | Asn | Arg | Gln | Ile | Ala | Glu |  |
|     |     |     |     | 245 |     |     |     |     | 250 |     |     |     |     | 255 |     |  |
| Ile | Gly | Ala | Ser | Leu | Ile | Lys | His | Trp |     |     |     |     |     |     |     |  |
|     | 260 |     |     |     | 265 |     |     |     |     |     |     |     |     |     |     |  |

Table 31. Nucleotide sequence of pLenti4TO/V5-DEST.

aatgtagtcttatgcaataactctttagtcttgcacatggtaacgatgagttagcaacatgccttacaaggaga  
 gaaaaagcaccgtgcatgccgattgggtggaagtaaggtggtagcatcggtgccttatttaggaaggcaacagacggg  
 tctgacatggattggacgaaccactgaattgccgcattgcagagatattgtatttaagtgccctagctcgatacat  
 aaacgggtctctctgggttagaccagatctgagcctgggagctctctggctaactaggaaccactgcttaagcc  
 tcaataaagcttgcccttagtgcttcaagtagtggtgcccgtctgttgtgtgactctggtaactagagatccct  
 cagacccttttagtcagtggtgaaaaatctctagcagtgccgcgccgaacagggacttgaaagcgaaagggaaacca  
 gaggagctctctcgacgcaggactcggtctgtgaagcgcgacggcaagaggcgagggggcgagctgggtgagt  
 acgcaaaaaattttagctagcggaggctagaaggagagagatgggtgagagagcgtcagatttaagcgggggaga  
 attagatcgcgatgggaaaaaattcggttaaggccaggggggaaagaaaaatataaattaaaacatatagtatgg  
 gcaagcaggagcgtagaacgattctgcagtttaactcctggcctgttagaaacatcagaaggctgtagacaaatactg  
 ggacagctacaaccatcccttcagacaggatcagaagaacttagatcattatataatacagtagcaaccctctat  
 tgtgtgcatcaaaggatagagataaaagacaccaaggaagctttagacaagatagaggaagagcaaaacaaagt  
 aagaccaccgcacagcaagcgccgctgatcttcagacctggaggaggagatatgagggacaattggagaagtga  
 attatataataataaagtagtaaaaaattgaaccattaggagtagcaccaccaaaggcaagagaagagtggtgca  
 gagagaaaaaagagcagtggaataggagctttgttccttgggttcttgggagcagcaggaagcactatgggcgc  
 agcgtcaatgacgctgacggtacaggccagacaattattgtctggtatagtgacgagcagaacaatttgcgtgag  
 ggctattgaggagcgtagaacgattctgtgcaactcctggcctgttagaaacatcagaaggctgtagacaaatactg  
 tgtggaagatacctaaaggatcaacagctcctggggatttgggggtgtctctggaaaactcatttgcaccactgc  
 tgtgccttgggaatgctagtggagtaataaatctctggaacagatttgggaatcacacgacctggatggagtggga  
 cagagaaattacaattacacaagcttaatacactccttaattgaagaatcgcaaaaccagcaagaaagaatga  
 acaagaattatttgggaattagataaatgggcaagtttgttgggaattgggttaacataacaaattggcgtgtggtatat  
 aaaattattcataatgatagtaggaggttggtaggtttaagaatagtttttgcgtgactttctatagtgaaatag  
 agttaggcagggaattaccattatcgtttcagaccacccctccaaccccgaggggacccgacaggcccggaagg  
 aatagaagaaggaaggtggagagagacagagacagatccattcgattagtgaacggatctcgacggatctcgata  
 agcttgggagttccgcgttacataaacttacggtaaatggcccgctggctgaccgcccacgaccccgccatt  
 gacgtcaataatgacgtatgttcccatagtaacgccaatagggaactttccattgacgtcaatgggtggagtattt  
 acggtaaaactgccacttggcagtagcatcaagtgtatcatatgccaaagtacgcccctatttgacgtcaatgacgg  
 taaatggcccgctggcatatgccagtagacgttatgggactttcctacttggcagtagatctacgtatt  
 agtcatcgctattaccatgggtgatgagggttttggcagtagatcaatgggctggatagcgggttgactcacgggg  
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Table 31 (continued). Nucleotide sequence of pLenti4TO/V5-DEST.

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Table 32. Nucleotide sequence of pLenti6/TR.

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Table 33. Nucleotide sequence of pLenti6/V5.

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Table 33 (continued). Nucleotide sequence of pLenti6/V5.

cgcagcgtgaccgctacacttgccagcgccctagcgcccgctcctttcgctttcttcccttcctttctcgccacg  
ttcgccggctttcccgctcaagctctaaatcgggggctccctttaggggtccgatttagtgctttacggcacctc  
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acgttggagtcacggttctttaatagtggaactcttggtccaaactggaacaacactcaaccctatctcggtctat  
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Table 34. Nucleotide sequence of pLenti3/V5-TREx.

aatgtagtctttagcaataactctttagtcttgcacatggtaacgatgagtttagcaacatgccttacaaggaga  
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Table 34 (continued). Nucleotide sequence of pLenti3/V5-TREx.

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gaaacagctatgacctgattacgccaagcgcgcaattaaccctcactaaagggaacaaaagctggagctgcaag  
ctt

Table 35. Nucleotide sequence of a nucleic acid fragment containing the tetracycline repressor coding sequence.

```

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tgaggtcgggaatcgaaggtttaacaaccgtaaaactcgccagaagctaggtgtagagcagcctacattgtattg
gcatgtaaaaaataagcgggcttttgctcgacgccttagccattgagatgttagataggcaccatactcacttttg
ccctttagaaggggaaagctggcaagattttttacgtaataacgctaaaagttttagatgtgctttactaagtca
tcgcgatggagcaaaaagtacatttaggtacacggcctacagaaaaacagtatgaaactctcgaaaatcaattagc
ctttttatgccacaagggtttttcactagagaatgcattatatgcactcagcgtgtggggcattttacttttagg
ttgcgtattggaagatcaagagcatcaagtcgctaaagaagaaagggaaacacctaactactgatagtatgccgcc
attattacgacaagctatcgaattatttgatcaccaagggtgcagagccagccttcttattcggccttgaattgat
catatgcggattagaaaaacaacttaaattgtgaaagtgggtccgcgtacagcggatcccgggaattctagagggc
ccgcggttcgaacaaaaactcatctcagaagaggatctgaatatgcata

```

Table 36. Nucleotide sequence of pRRL6/V5 also referred to as pLenti6/V5.

|      |             |            |              |             |             |             |
|------|-------------|------------|--------------|-------------|-------------|-------------|
| 1    | aatgtagtct  | tatgcaatac | tctttagtagtc | ttgcaacatg  | gtaacgatga  | gttagcaaca  |
| 61   | tgccttacaa  | ggagagaaaa | agcacctgtc   | atgccgattg  | gtggaagtaa  | ggtaggtacga |
| 121  | tctgtcctta  | ttaggaaggc | aacagacggg   | tctgacatgg  | attggacgaa  | ccactgaatt  |
| 181  | gccgcattgc  | agagatattg | tatttaagtg   | cctagctcga  | tacaataaac  | gggtctctct  |
| 241  | ggtttagacca | gatctgagcc | tgggagctct   | ctggctaact  | agggaaacca  | ctgcttaagc  |
| 301  | ctcaataaag  | cttgccctga | gtgcttcaag   | tagtgtgtgc  | ccgtctgttg  | tgtgactctg  |
| 361  | gtaactagag  | atccctcaga | cccttttagt   | cagtgtggaa  | aatctctagc  | agtggcgccc  |
| 421  | gaacagggac  | ctgaaagcga | aagggaaacc   | agagctctct  | cgacgcagga  | ctcggcttgc  |
| 481  | tgaagcgcgc  | acggcaagag | gcgagggcg    | gcgactggtg  | agtacgccaa  | aaattttgac  |
| 541  | tagcggaggc  | tagaaggaga | gagatgggtg   | cgagagcgtc  | agtattaagc  | gggggagaat  |
| 601  | tagatcgca   | tgggaaaaaa | tccggttaag   | gccaggggga  | aagaaaaaat  | ataaatataa  |
| 661  | acatatagta  | tgggcaagca | gggagctaga   | acgattcgca  | gttaatcctg  | gcctgttaga  |
| 721  | aacatcagaa  | ggctgtagac | aaatactggg   | acagctacaa  | ccatcccttc  | agacaggatc  |
| 781  | agaagaactt  | agatcattat | ataatacagt   | agcaaccctc  | tattgtgtgc  | atcaaaggat  |
| 841  | agagataaaa  | gacaccaagg | aagctttaga   | caagatagag  | gaagagcaaa  | acaaaagtaa  |
| 901  | gaccaccgca  | cagcaagcgg | ccgctgatct   | tcagacctgg  | aggaggagat  | atgaggggaca |
| 961  | attggagaag  | tgaattatat | aaatataaag   | tagtaaaaat  | tgaaccatta  | ggagtagcac  |
| 1021 | ccaccaaggc  | aaagagaaga | gtggtgcaga   | gagaaaaaag  | agcagtggga  | ataggagctt  |
| 1081 | tactccttgg  | gttcttggga | gcagcaggaa   | gcactatggg  | cgcagcctca  | atgacgttga  |
| 1141 | cggtagcaggc | cagacaatta | ttgtctggta   | tagtgcagca  | gcagaacaat  | ttgtgagggg  |
| 1201 | ctattgaggc  | gcaacagcat | ctgttgcaac   | tcacagtctg  | gggcatcaag  | cagctccagg  |
| 1261 | caagaatcct  | ggctgtggaa | agatacctaa   | aggatcaaca  | gctcctgggg  | atttgggggt  |
| 1321 | gctctggaaa  | actcatttgc | accactgctg   | tgccttggaa  | tgctagttag  | agtaataaat  |
| 1381 | ctctggaaca  | gattggaatc | acacgacctg   | gatggagtgg  | gacagagaaa  | ttaacaatta  |
| 1441 | cacaagctta  | atacactcct | taattgaaga   | atcgcaaaac  | cagcaagaaa  | agaatgaaca  |
| 1501 | agaattattg  | gaattagata | aatgggcaag   | tttgtggaat  | tgggttaaca  | taacaaattg  |
| 1561 | gctgtggtat  | ataaaattat | tcataatgat   | agtaggaggc  | ttggtagggt  | taagaatagt  |
| 1621 | ttttgctgta  | ctttctatag | tgaatagagt   | taggcaggga  | tattcaccat  | tatcgtttca  |
| 1681 | gaccacctc   | ccaaccccga | ggggaccoga   | caggcccga   | ggaatagaag  | aagaagggtg  |
| 1741 | agagagagac  | agagacagat | ccattcgatt   | agtgaacgga  | tctcgacggg  | atcgataagc  |
| 1801 | ttgggagttc  | cgcgttacat | aacttacggg   | aaatggcccg  | cctggctgac  | cgcccaacga  |
| 1861 | cccccgccca  | ttgacgtcaa | taatgacgta   | tgttcccata  | gtaacgccaa  | tagggacttt  |
| 1921 | ccattgacgt  | caatgggtgg | agtatttacg   | gtaaaactgcc | cacttggcag  | tacatcaagt  |
| 1981 | gtatcatatg  | ccaagtacgc | cccctattga   | cgtcaatgac  | ggtaaatggc  | ccgcctggca  |
| 2041 | ttatgccag   | tacatgacct | tatgggactt   | tcctacttgg  | cagtacatct  | acgtattagt  |
| 2101 | catcgctatt  | accatggtga | tgcggttttg   | gcagtacatc  | aatgggctgt  | gatagcgggt  |
| 2161 | tgactcacgg  | ggatttccaa | gtctccaccc   | cattgacgtc  | aatgggagtt  | tgttttggca  |
| 2221 | ccaaaatcaa  | cgggactttc | caaaatgtcg   | taacaactcc  | gccccattga  | cgcaaatggg  |
| 2281 | cggtaggcgt  | gtacggtggg | aggtctatat   | aagcagagct  | cgtttagtga  | accgtcagat  |
| 2341 | cgctggagga  | gcctatccac | gctgttttga   | cctccataga  | agacaccgac  | cttagaggat  |
| 2401 | ccactagtcc  | agtgtggtgg | aattctgcag   | atatccagca  | cagtggcggc  | cgtctgagtc  |
| 2461 | tagagggccc  | gcggttcgaa | ggtaagccta   | tccttaaccc  | tctcctcggt  | ctcgattcta  |
| 2521 | cgcgtaccgg  | ttagtaatga | gtttggcctg   | ctgccggctc  | tgccggcctct | tccgcgtctt  |
| 2581 | cgccttcgcc  | ctcagacgag | tccgatctcc   | ctttgggcgg  | cctccccgcc  | tggaaattaat |
| 2641 | tctgtggaat  | gtgtgtcagt | tagggtgtgg   | aaagtcccca  | gggtccccag  | gcaggcagaa  |
| 2701 | gtatgcaaag  | catgcatctc | aattagtcag   | caaccagggt  | tggaaagtcc  | ccaggctccc  |
| 2761 | cagcaggcag  | aagtatgcaa | agcatgcata   | tcaattagtc  | agcaaccata  | gtcccgcccc  |
| 2821 | taactccgcc  | catcccgcgc | ctaactccgc   | ccagttccgc  | ccattctccg  | ccccatggct  |
| 2881 | gactaatttt  | ttttatttat | gcagaggccg   | aggccgcctc  | tgctctgag   | ctattccaga  |
| 2941 | agtagtgagg  | aggctttttt | ggaggcctag   | gcttttgcaa  | aaagctcccg  | ggagcttgta  |
| 3001 | tatccatttt  | cggatctgat | cagcacgtgt   | tgacaattaa  | tcatcggcac  | agtatatcgg  |
| 3061 | catagtataa  | tacgacaagg | tgaggaaacta  | aacctatggc  | aagcctttgt  | ctcaagaaga  |
| 3121 | atccaccctc  | attgaaagag | caacggctac   | aatcaacagc  | atccccatct  | ctgaagacta  |
| 3181 | cagcgtcgcc  | agcgcagctc | tctctagcga   | cggccgcata  | ttcactgggtg | tcaatgtata  |
| 3241 | tcattttact  | gggggacctt | gtgcagaact   | cgtggtgctg  | ggcactgctg  | ctgctgcggc  |
| 3301 | agctggcaac  | ctgacttgta | tcgtcgcgat   | cggaaatgag  | aacaggggca  | tcttgagccc  |
| 3361 | ctgcggacgg  | tgccgacagg | tgcttctcga   | tctgcatcct  | gggatcaaag  | ccatagtga   |
| 3421 | ggacagtgat  | ggacagccga | cggcagttgg   | gattcgtgaa  | ttgctgccct  | ctggttatgt  |
| 3481 | gtgggagggc  | taagcacaat | tcgagctcgg   | tacctttaag  | accaatgact  | tacaaggcag  |

Table 36 (continued). Nucleotide sequence of pRRL6/V5 also referred to as pLenti6/V5.

```

3541 ctgtagatct tagccacttt ttaaagaaa aggggggact ggaagggcta attcactccc
3601 aacgaagaca agatctgctt ttgtcttgta ctgggtctct ctggttagac cagatctgag
3661 cctgggagct ctctggctaa ctagggaacc cactgcttaa gcctcaataa agcttgccct
3721 gagtgcttca agtagtggtg gcccgctctg tgtgtgactc tggtaactag agatccctca
3781 gaccctttta gtcagtgtgg aaaatctcta gcagtagtag ttcatgtcat cttattatct
3841 agtattttata acttgcaaag aaatgaatat cagagagtga gaggaacttg tttattgcag
3901 cttataatgg ttacaaataa agcaatagca tcacaaatct cacaataaaa gcattttttt
3961 cactgcattc tagttgtggg ttgtccaaac tcatcaatgt atcttatcat gtctggctct
4021 agctatcccg cccctaactc cgcccagttc cgcccattct ccgccccatg gctgactaat
4081 tttttttatt tatgcagagg ccgaggccgc ctccggcctc gagctattcc agaagtagtg
4141 aggagccttt ttggaggcc taggcttttg cgtcgagacg tacccaattc gccctatagt
4201 gagtcgtatt acgcgcgctc actggccgctc gttttacaac gtcgtgactg ggaaaaccct
4261 ggcgttaccc aacttaatcg ccttgacgca catccccctt tcgccagctg gcgtaatagc
4321 gaagaggccc gcaccgatcg cccttcccaa cagttgcgca gcctgaatgg cgaatggcgc
4381 gacgcgccct gtagcggcgc attaagcgcg gcgggtgtgg tggttacgcg cagcgtgacc
4441 gctacacttg ccagcgcctc agcgcgcgct cctttcgctt tcttcccttc ctttctcgcc
4501 acgttcgcgc gctttccccc tcaagctcta aatcgggggc tccctttagg gttccgattt
4561 agtgctttac ggcacctcga ccccaaaaaa cttgattagg gtgatgggtc acgtatggg
4621 ccatcgccct gatagacggg ttttcgcctt ttgacgttgg agtccacgtt ctttaatagt
4681 ggactcttgt tccaaactgg aacaacactc aaccctatct cgggtctattc ttttgattta
4741 taagggattt tgccgatttc ggctatttgg ttaaaaaatg agctgattta acaaaaattt
4801 aacgcgaatt ttaacaaaat attaacgttt acaatttccc aggtggcact tttcggggaa
4861 atgtgcgcgg aacccctatt tgtttatttt tctaaataca ttcaaatatg tatccgctca
4921 tgagacaata accctgataa atgcttcaat aatattgaaa aaggaagagt atgagtattc
4981 aacatttccg tgtcgccctt attccctttt ttgcggcatt ttgccttctc gttttgtctc
5041 acccagaaaac ctgggtgaaa gtaaaagatg ctgaagatca gttgggtgca cgagtgggtt
5101 acatcgaact ggatctcaac agcggtaaga tccttgagag ttttcgcccc gaagaacgtt
5161 ttccaatgat gagcactttt aaagtctctg tatgtggcgc ggtattatcc cgtattgacg
5221 ccgggcaaga gcaactcggg ccgcgcatac actattctca gaatgacttg gttgagtact
5281 caccagtcac agaaaagcat cttacggatg gcatgacagt aagagaatta tgcaagtctg
5341 ccataacctc gagtgataac actgcggcca acttacttct gacaacgatc ggaggaccga
5401 aggagctaac agcttttttg cacaacattg gggatcatgt aactcgccct gatcgttggg
5461 aaccggagct gaatgaagcc ataccaaagc acgagcgtga caccacgatg cctgtagcaa
5521 tggcaacaac gttgcgcaaa ctattaactg gcgaactact tactctagct tcccggcaac
5581 aattaataga ctggatggag gcggataaag ttgcaggacc acttctgcgc tcggcccttc
5641 cggctggctg gtttatttgc gataaatctg gagccgggtg gcgtgggtct cgcggtatca
5701 ttgcagcact ggggccagat ggtaagccct ccgctatcgt agttatctac acgacgggga
5761 gtcaggcaac tatggatgaa cgaaatagac agatcgctga gatagggtgc tcaactgatta
5821 agcatttgta actgtcagac caagtttact catatatact ttagattgat ttaaaacttc
5881 atttttaatt taaaaggatc taggtgaaga tcctttttga taatctcatg accaaaatcc
5941 cttaacgtga gttttcgttc cactgagcgt cagaccccggt agaaaagatc aaaggatctt
6001 cttgagatcc tttttttctg cgcgtaatct gctgcttgca aacaaaaaaa ccaccgctac
6061 cagcggtggt ttgtttgccc gatcaagagc taccaactct ttttccgaag gtaactggct
6121 tcagcagagc gcagatacca aatactgtcc ttctagtgtg gccgtagtta ggccaccact
6181 tcaagaactc tgtagcaccg cctacatacc tcgctctgct aatcctgtta ccagtggctg
6241 ctgccagtcg cgataagtcg tgtcttaccg ggttggactc aagacgatag ttaccggata
6301 aggcgcagcg gtcgggctga acggggggtt cgtgcacaca gccagcgttg gagcgaacga
6361 cctacaccga actgagatac ctacagcgtg agctatgaga aagcgccacg ctcccgaag
6421 ggagaaaggc ggacaggtat ccggtaaagc gcagggtcgg aacaggagag cgcacgaggg
6481 agcttccagg gggaaacgcc tggatctttt atagtcctgt cgggtttcgc cacctctgac
6541 ttgagcgtcg atttttctga tgctcgtcag gggggcggag cctatggaaa aacgccagca
6601 acgcggcctt ttacgggttc ctggcctttt gctggccttt tgctcacatg ttcttctctg
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6781 tacgcaaacc gcctctcccc gcgcgttggc cgattcatta atgcagctgg cacgacaggt
6841 tccccgactg gaaagcgggc agtgagcgca acgcaattaa tgtgagttag ctactcatt
6901 aggcacccca ggctttacac tttatgcttc cggctcgtat gttgtgtgga attgtgagcg
6961 gataacaatt tcacacagga aacagctatg accatgatta cgccaagcgc gcaattaacc
7021 ctactaaaag ggaacaaaag ctggagctgc aagctt

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## WHAT IS CLAIMED IS:

1. A nucleic acid molecule comprising all or a portion of at least one viral genome and further comprising at least two recombination sites that do not substantially recombine with each other, wherein at least one recombination site is capable of undergoing recombination with a compatible recombination site in the presence of at least one protein active in lambda recombination and wherein the nucleic acid molecule replicates in prokaryotic and eukaryotic cells.

2. A nucleic acid molecule according to claim 1, wherein the nucleic acid molecule comprises all or a portion of at least one viral genome selected from the group consisting of an adenovirus genome and an adeno-associate virus genome.

3. A nucleic acid molecule according to claim 1, wherein the nucleic acid molecule comprises all or a portion of at least one retroviral genome.

4. A nucleic acid molecule according to claim 3, wherein the retroviral genome is a lentiviral genome.

5. A nucleic acid molecule according to claim 1, wherein the nucleic acid molecule comprises all or a portion of at least one viral genome selected from the group consisting of a herpesvirus genome and a pox virus genome.

6. A nucleic acid molecule according to claim 1, wherein the nucleic acid molecule comprises all or a portion of at least one RNA virus genome.

7. A nucleic acid molecule according to claim 6, wherein the RNA virus is selected from a group consisting of a flavivirus genome, a togavirus genome, and an alphavirus genome.

8. A nucleic acid molecule according to claim 1, wherein the nucleic acid molecule is a plasmid or a bacmid comprising a prokaryotic origin of replication and a selectable marker.

9. A nucleic acid molecule according to claim 1, further comprising one or more features selected from the group consisting of a promoter, a viral terminal repeat, a splice site, a packaging signal, a nucleic acid sequence responsive to one or more viral proteins, a recognition site, a recombination site, a sequences encoding one or more marker proteins or polypeptides, a sequence encoding one or more epitopes recognizable by an antibody, an origin of replication, an intervening sequence, an internal ribosome entry sequences, and a polyadenylation signal.

10. A nucleic acid molecule according to claim 1, further comprising a nucleotide sequence of interest between the two recombination sites.

11. A nucleic acid molecule according to claim 10, wherein the sequence of interest comprises one or more sequences selected from the group consisting a sequence encoding one or more polypeptides, a sequence encoding one or more tRNA sequences, a sequence encoding one or more ribozyme sequences, one or more promoter sequences, one or more enhancer sequences, and one or more repressor sequences.

12. A nucleic acid molecule comprising all or a portion of a baculoviral genome and further comprising one or more recombination sites, wherein at least one recombination site is capable of undergoing recombination with a compatible recombination site in the presence of at least one protein active in lambda recombination.

13. A nucleic acid molecule according to claim 12, wherein the molecule comprises two recombination sites that do not substantially recombine with each other.

14. A nucleic acid molecule according to claim 13, wherein the sequence of interest comprises one or more sequences selected from the group consisting a sequence encoding one or more polypeptides, a sequence encoding one or more tRNA sequences, a sequence encoding one or more ribozyme sequences, one or more promoter sequences, one or more enhancer sequences, and one or more repressor sequences.

15. A composition comprising the nucleic acid molecule of claim 1.

16. A composition comprising the nucleic acid molecule of claim 12.

17. A method of constructing a recombinant virus, comprising:
  - (a) providing a first nucleic acid molecule comprising all or a portion of at least one viral genome and at least a first and a second recombination site that do not substantially recombine with each other;
  - (b) contacting the first nucleic acid molecule with a second nucleic acid molecule comprising a sequence of interest flanked by at least a third and a fourth recombination site under conditions such that recombination occurs between the first and third recombination site and between the second and fourth recombination site; and
  - (c) introducing the nucleic acid molecule of step (b) into a cell that packages the nucleic acid molecule of step (b).
18. A method according to claim 17, wherein the first nucleic acid molecule comprises all or a portion of at least one viral genome selected from the group consisting of an adenovirus genome and an adeno-associate virus genome.
19. A method according to claim 17, wherein the first nucleic acid molecule comprises all or a portion of at least one retroviral genome.
20. A method according to claim 19, wherein the retroviral genome is a lentiviral genome.
21. A method according to claim 17, wherein the first nucleic acid molecule comprises all or a portion of at least one viral genome selected from the group consisting of a herpesvirus genome and a pox virus genome.
22. A method according to claim 17, wherein the first nucleic acid molecule comprises all or a portion of at least one RNA virus genome.
23. A method according to claim 22, wherein the RNA virus is selected from a group consisting of a flavivirus genome, a togavirus genome, and an alphavirus genome.
24. A method according to claim 17, wherein the first nucleic acid molecule is a plasmid or a bacmid comprising an origin of replication and a selectable marker.

25. A method according to claim 17, wherein the portion of the second nucleic acid between the recombination site comprises a nucleotide sequence of interest.

26. A method according to claim 25, wherein the sequence of interest comprises one or more sequences selected from a group consisting a sequence encoding one or more polypeptides, a sequence encoding one or more tRNA sequences, a sequence encoding one or more ribozyme sequences, one or more promoter sequences, one or more enhancer sequences, and one or more repressor sequences.

27. A method according to claim 17, further comprising digesting the first nucleic acid molecule with a restriction enzyme that cleaves the first nucleic acid at a site between the recombination sites.

28. A recombinant virus produced by the method of claim 17.

29. A composition comprising the recombinant virus of claim 28.

30. A method of producing a fusion polypeptide, comprising:  
providing a host cell comprising a first nucleic acid sequence encoding the fusion polypeptide, wherein the sequence comprises at least a first coding region and a second coding region separated by a sequence comprising a stop codon;  
expressing in the cell a second nucleic acid sequence comprising one or more suppressor tRNAs that suppress the stop codon; and  
incubating the host cell under conditions sufficient to express the fusion polypeptide comprising the first coding region and the second coding region, wherein at least one of the first and/or second nucleic acid sequences is present on a nucleic acid molecule comprising all or a portion of at least one viral genome.

31. A method according to claim 30, further comprising introducing nucleic acid molecules comprising the first and the second nucleic acid sequences into the host cell.

32. A method according to claim 30, wherein the nucleic acid molecule comprising the second nucleic sequence comprises all or a portion of an adenoviral genome.

33. A method according to claim 30, wherein the first coding region and stop codon are flanked by recombination sites that do not substantially recombine with each other.

34. A method of expressing a polypeptide, comprising:  
contacting a cell with a nucleic acid molecule comprising a sequence encoding the polypeptide operably linked to a promoter and a repressor sequence, wherein the nucleic acid molecule comprises all or a portion of a viral genome;  
contacting the cell with a nucleic acid molecule encoding a protein that binds to the repressor sequence; and  
incubating the cell under conditions sufficient to express the polypeptide.

35. A method according to claim 34, wherein the viral genome is a lentivirus genome.

36. The method according to claim 34, wherein the viral genome is an HIV-1 genome.

37. The method according to claim 34, wherein the repressor sequence is the tetracycline operator sequence and the protein is the tetracycline repressor protein.

38. The method according to claim 37, wherein conditions sufficient to express the polypeptide comprise incubating the cell in the presence of tetracycline.

39. A method of expressing a polypeptide, comprising:  
contacting a cell with a nucleic acid molecule comprising a sequence encoding the polypeptide operably linked to a promoter and a repressor sequence, wherein the nucleic acid molecule comprises all or a portion of a viral genome and wherein the cell expresses a protein that binds to the repressor sequence; and  
incubating the cell under conditions sufficient to express the polypeptide.

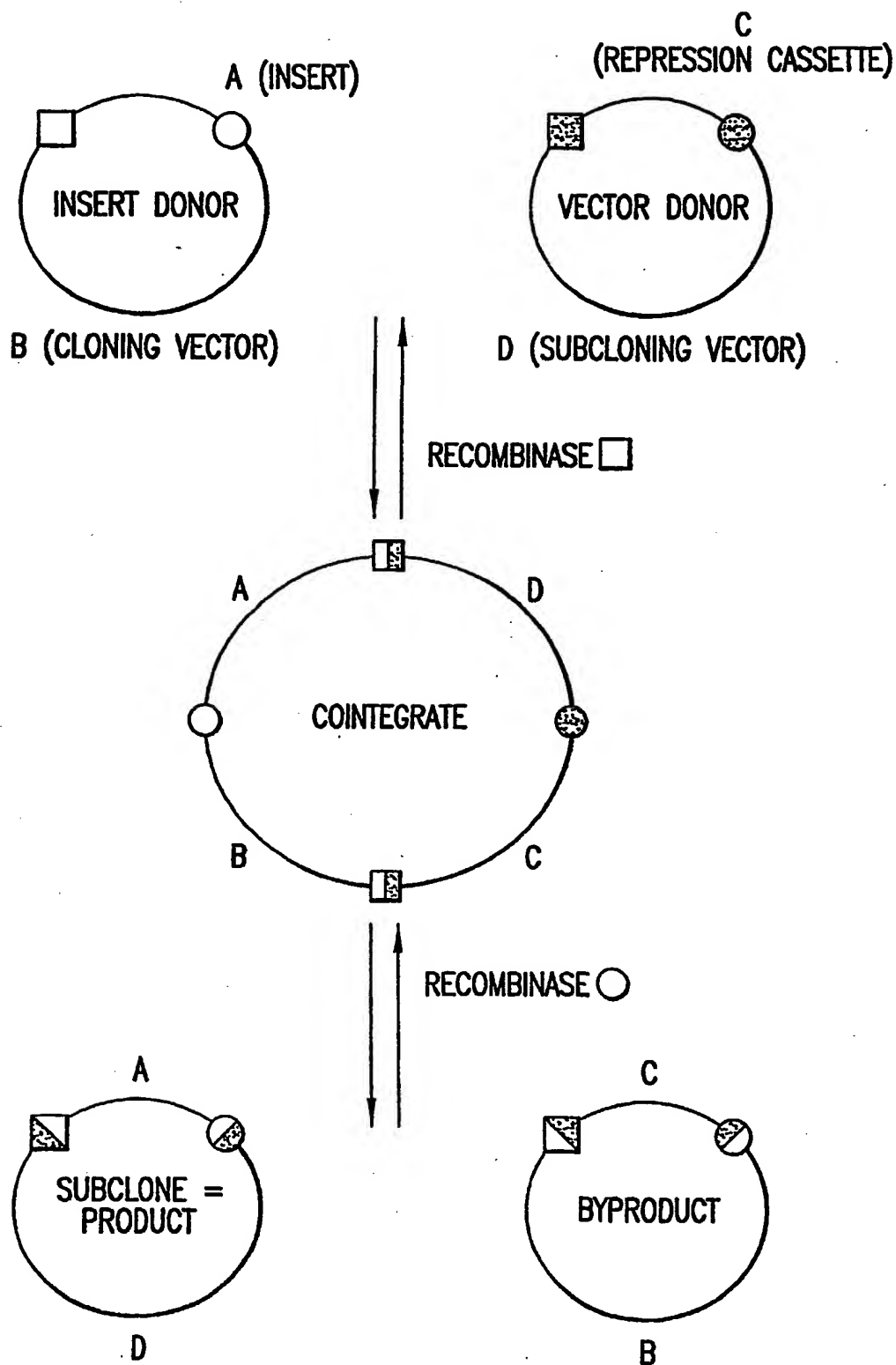
40. The method according to claim 39, wherein the viral genome is a lentiviral genome.

41. The method according to claim 39, wherein the viral genome is an HIV-1 genome.

42. The method according to claim 39, wherein the repressor sequence is the tetracycline operator sequence and the protein is the tetracycline repressor protein.

43. The method according to claim 42, wherein conditions sufficient to express the polypeptide comprise incubating the cell in the presence of tetracycline.

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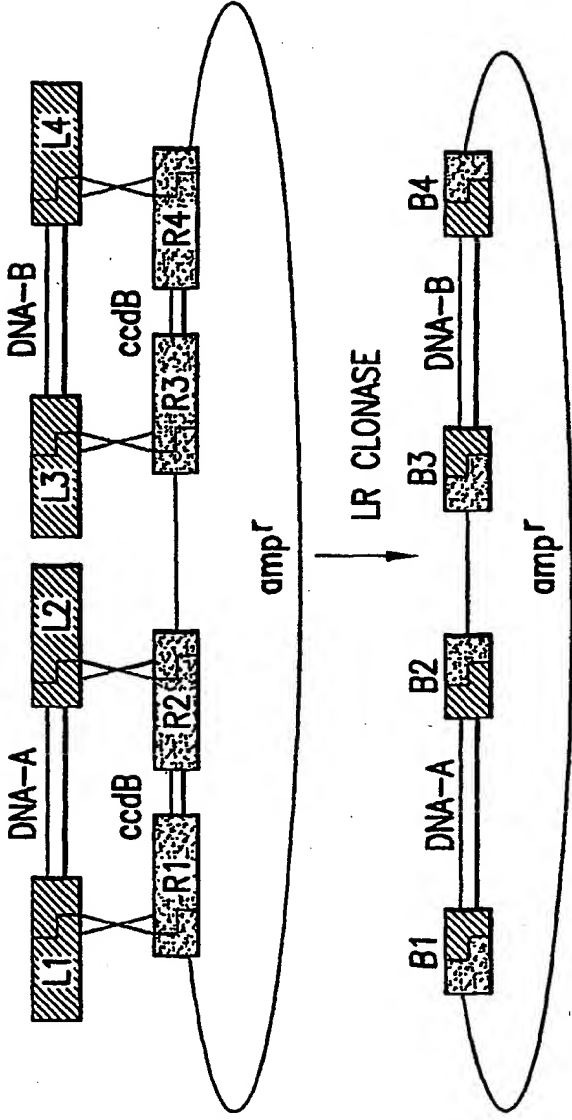


FIG.2



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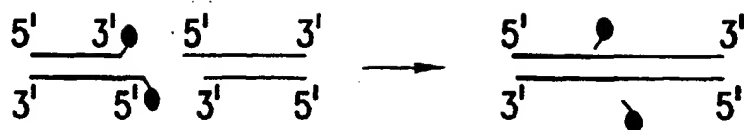


FIG. 3A

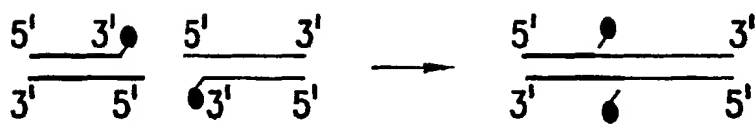


FIG. 3B

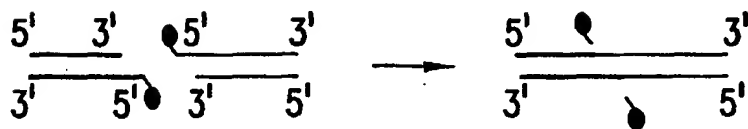


FIG. 3C

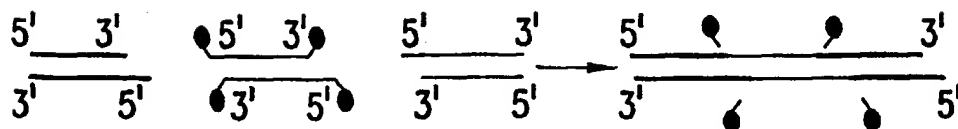


FIG. 3D

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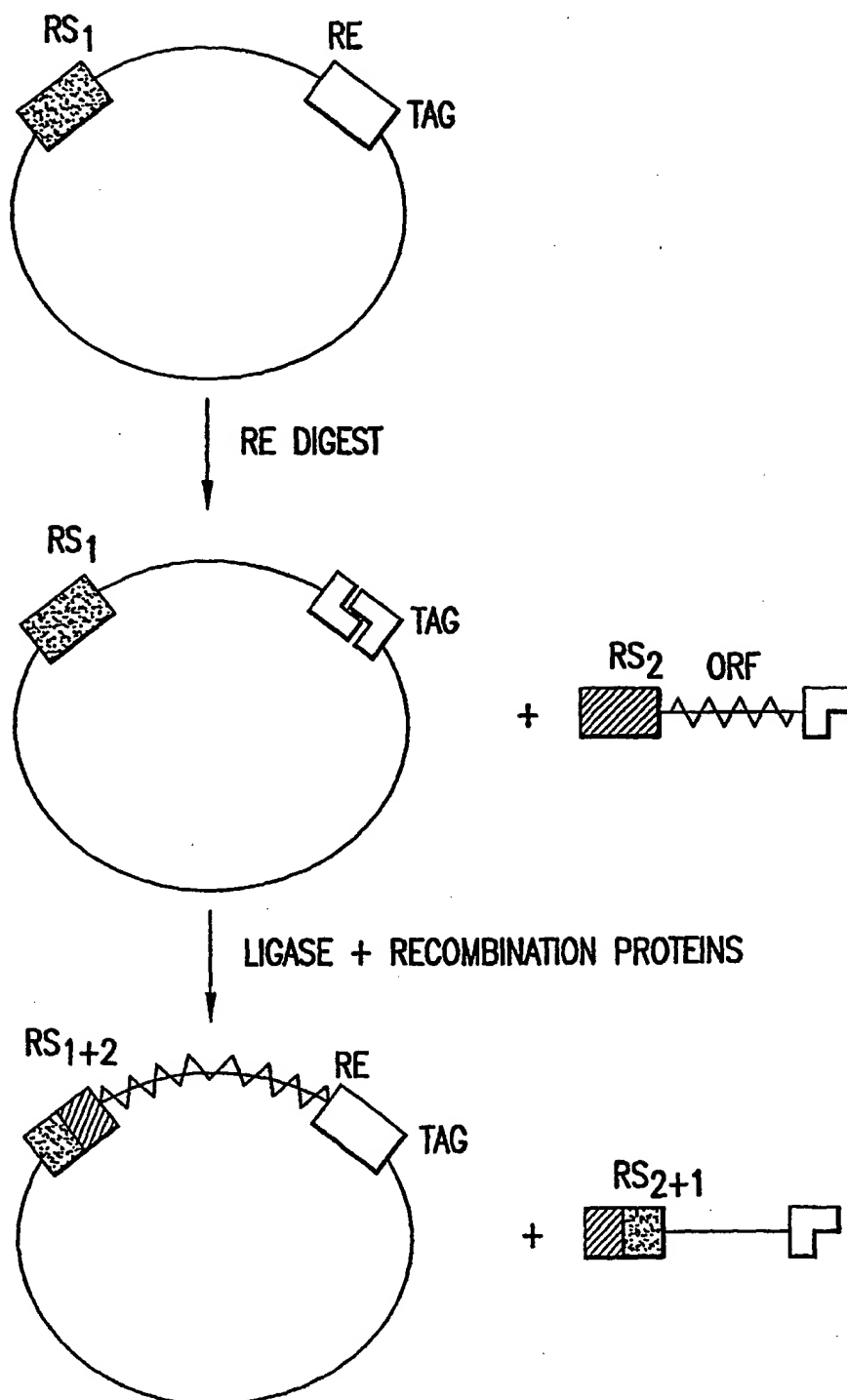


FIG.4

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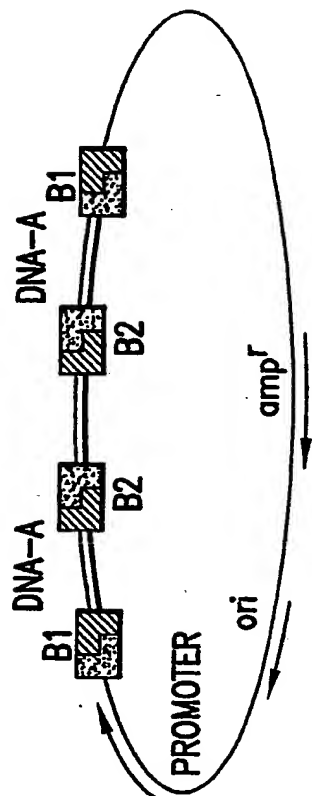


FIG. 5A

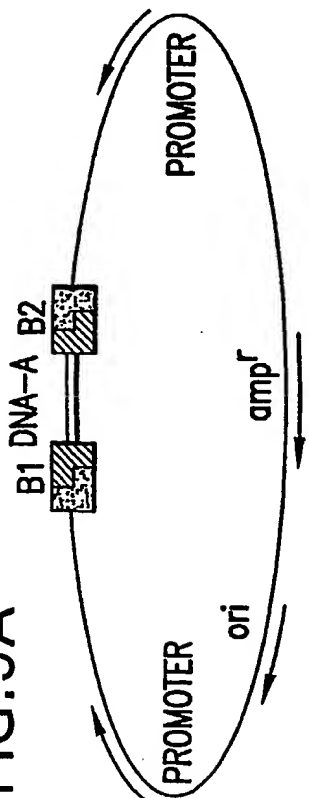


FIG. 5B

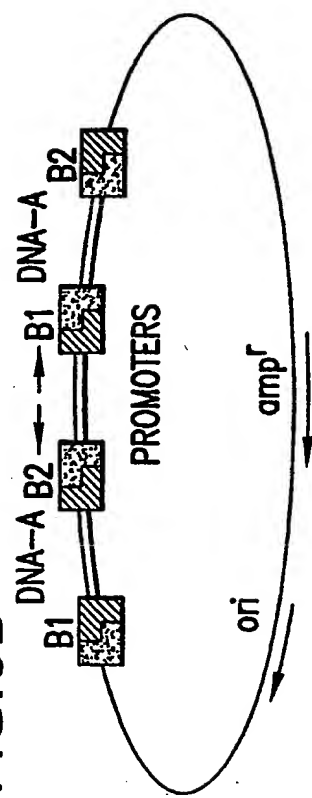


FIG. 5C

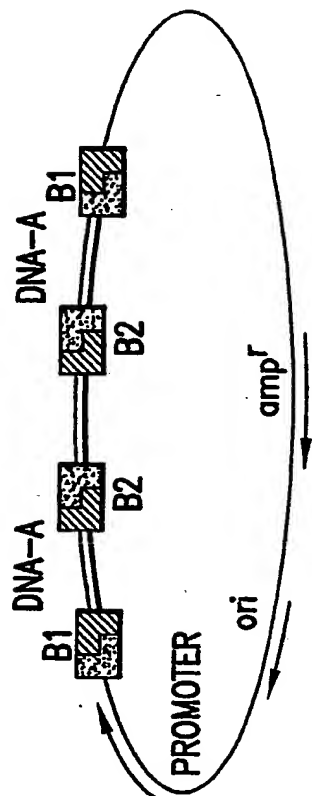


FIG. 5D

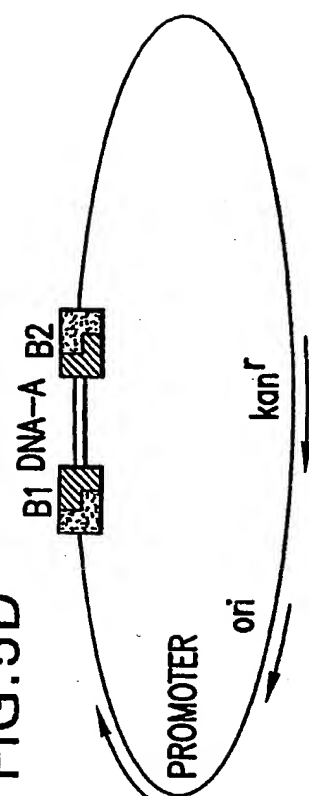


FIG. 5E

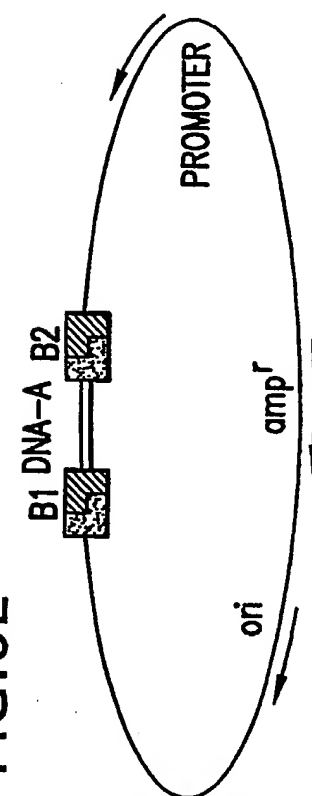


FIG. 5F

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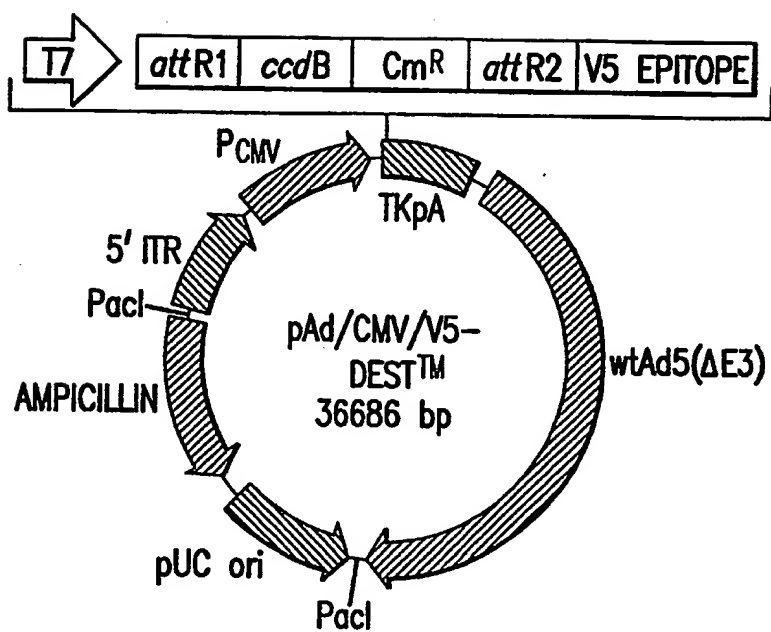


FIG.6

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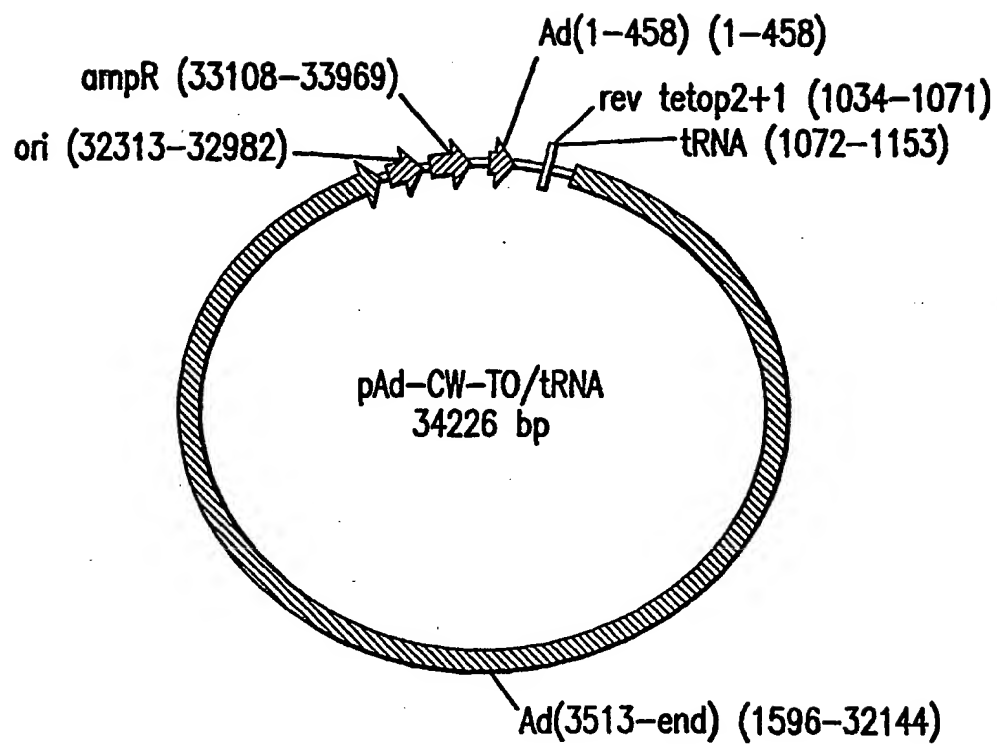


FIG. 7

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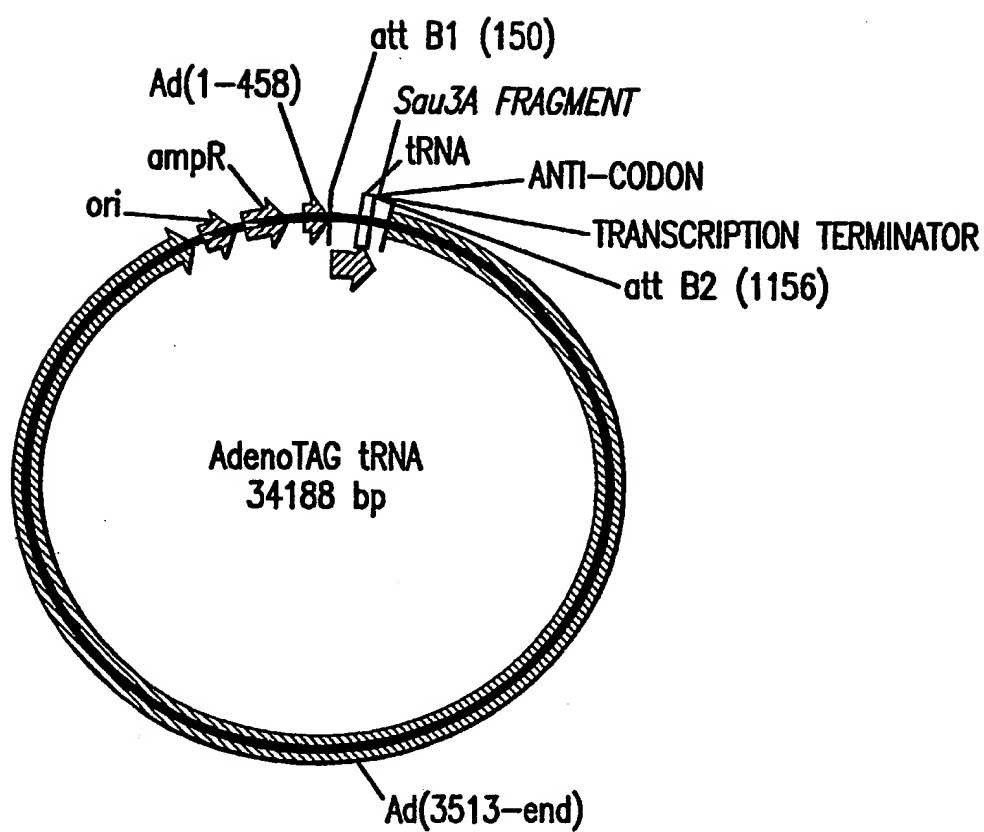


FIG.8

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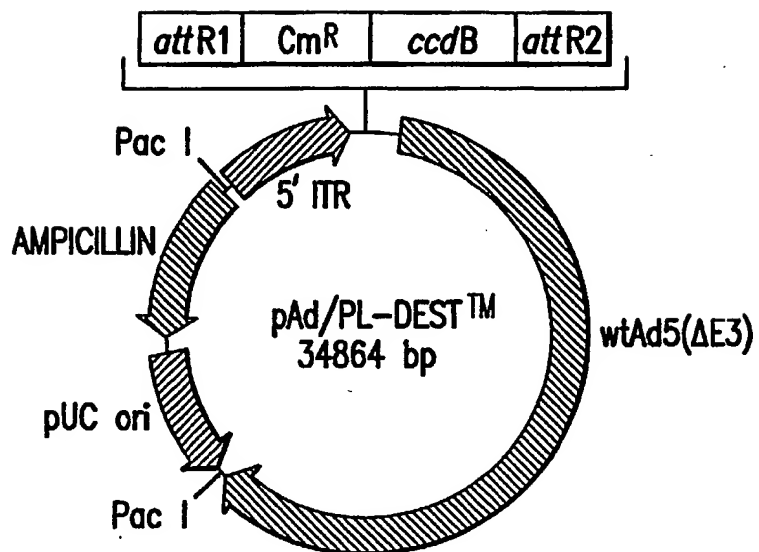


FIG.9

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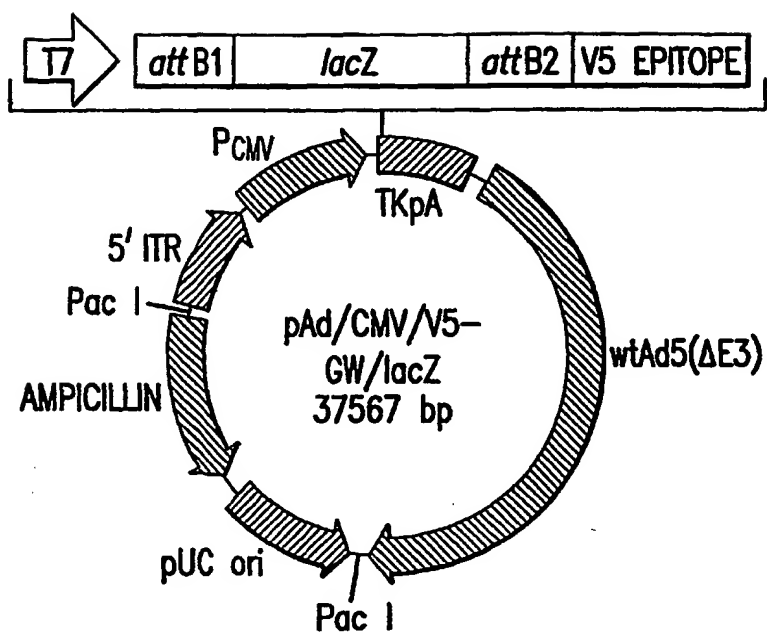


FIG.10



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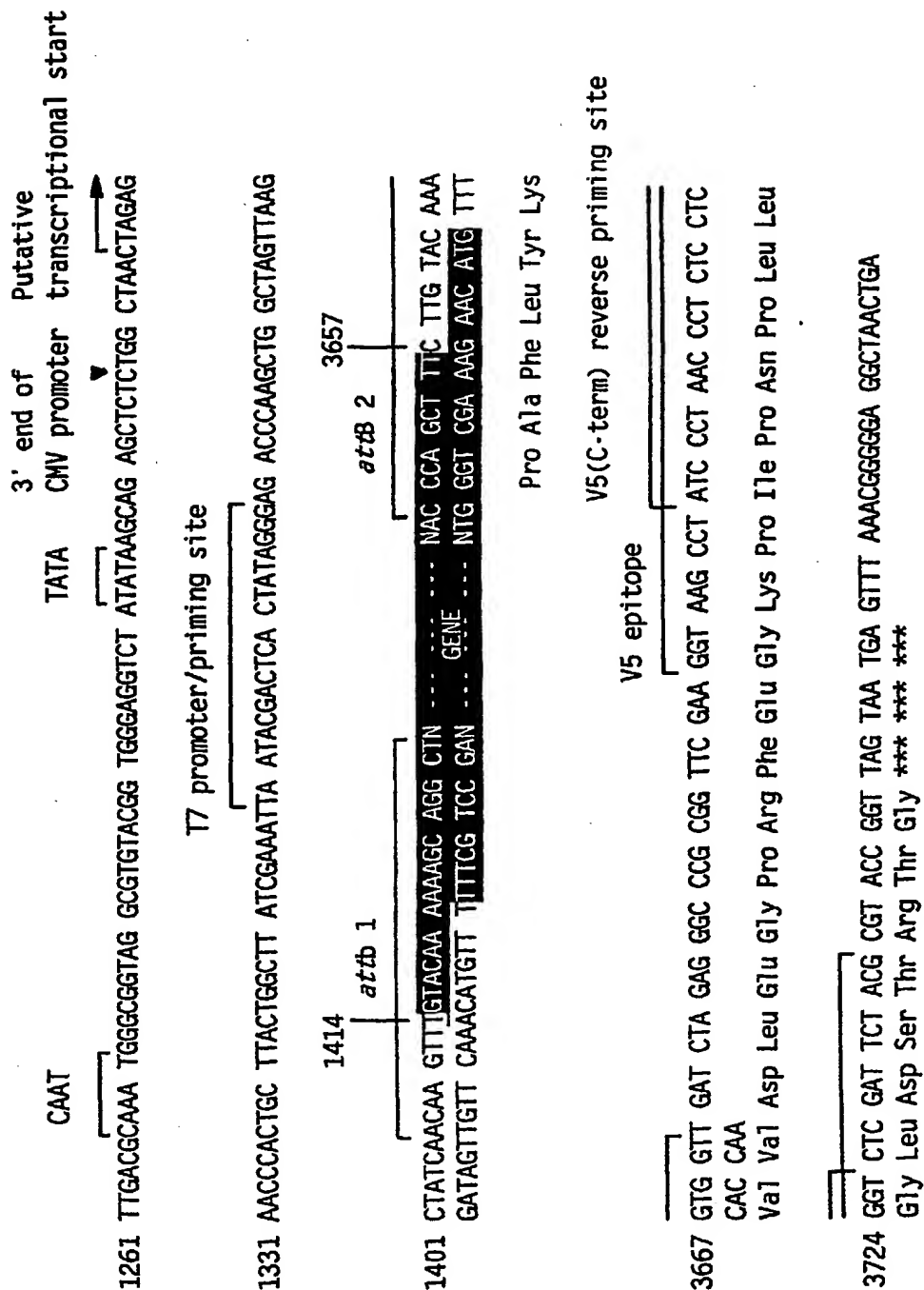


FIG.11

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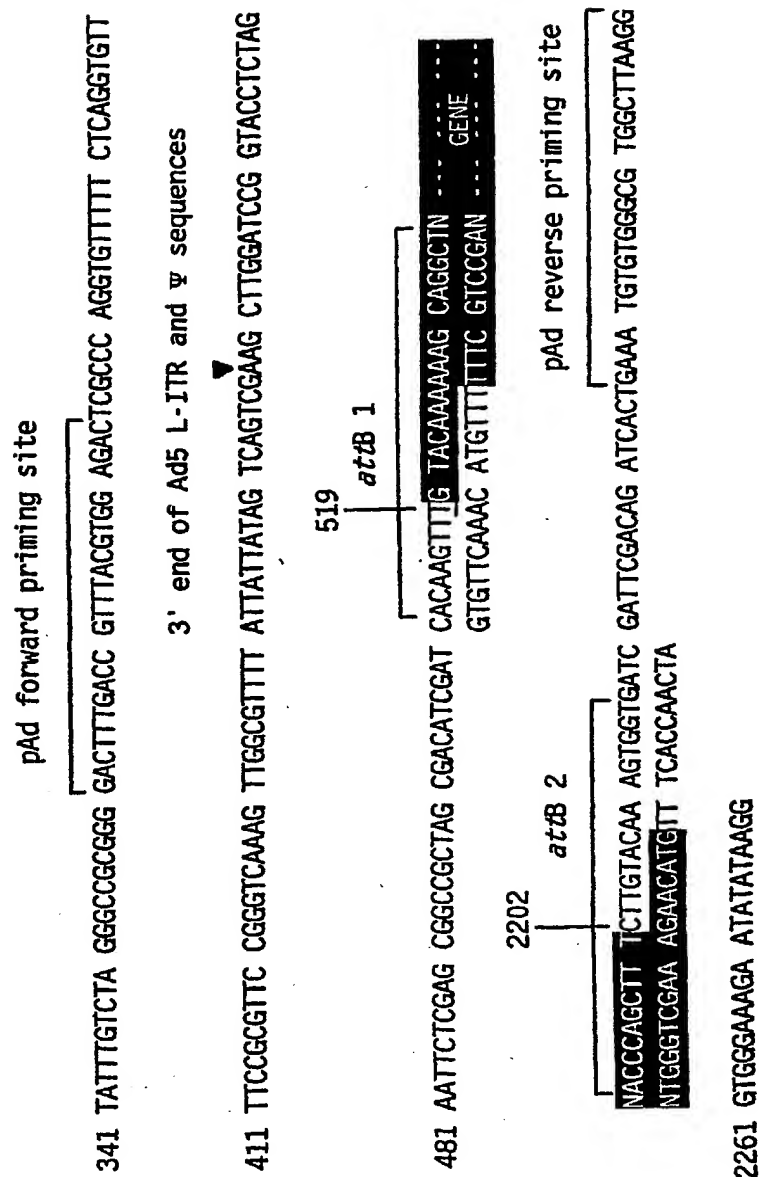


FIG.12

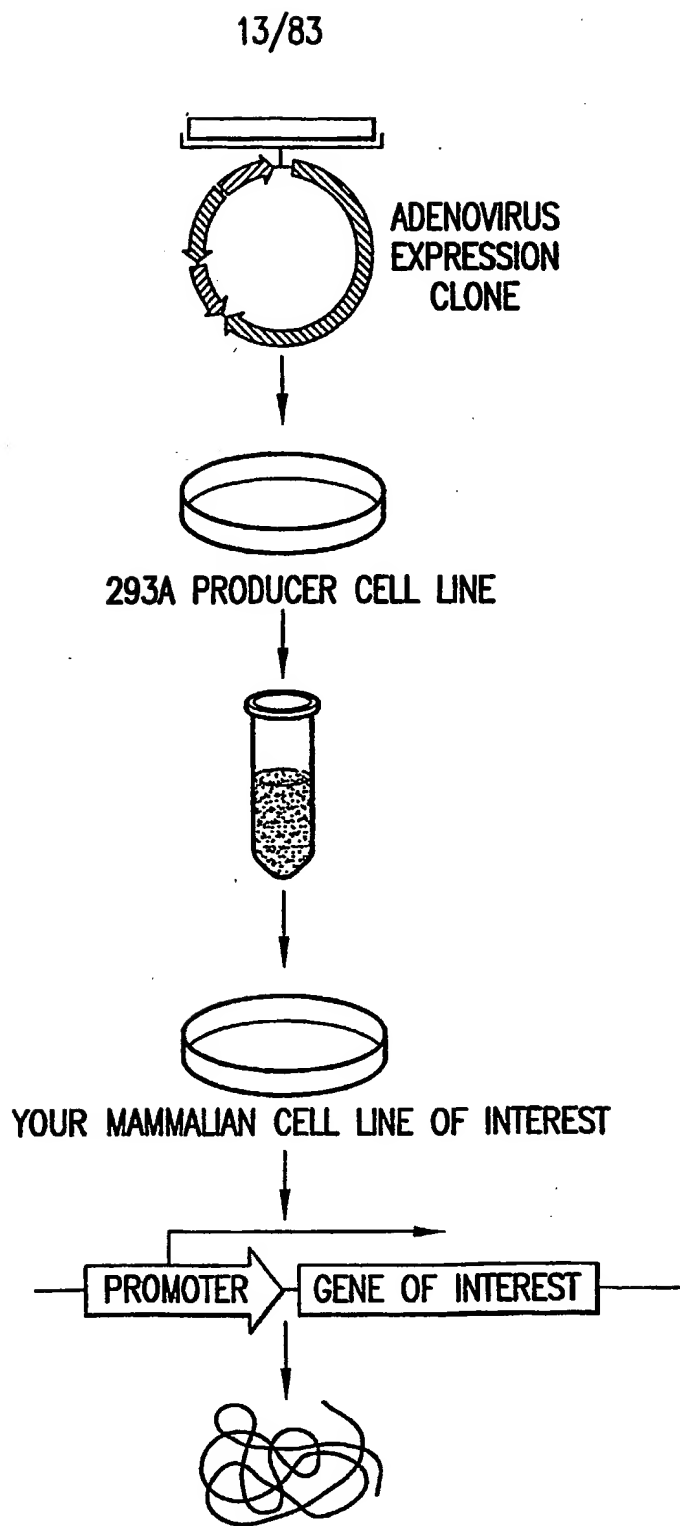


FIG.13

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FIG. 14A

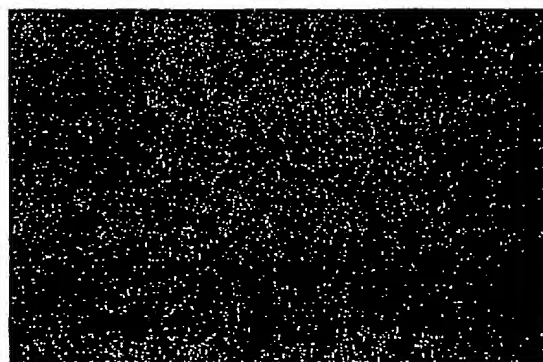


FIG. 14B



FIG. 14C

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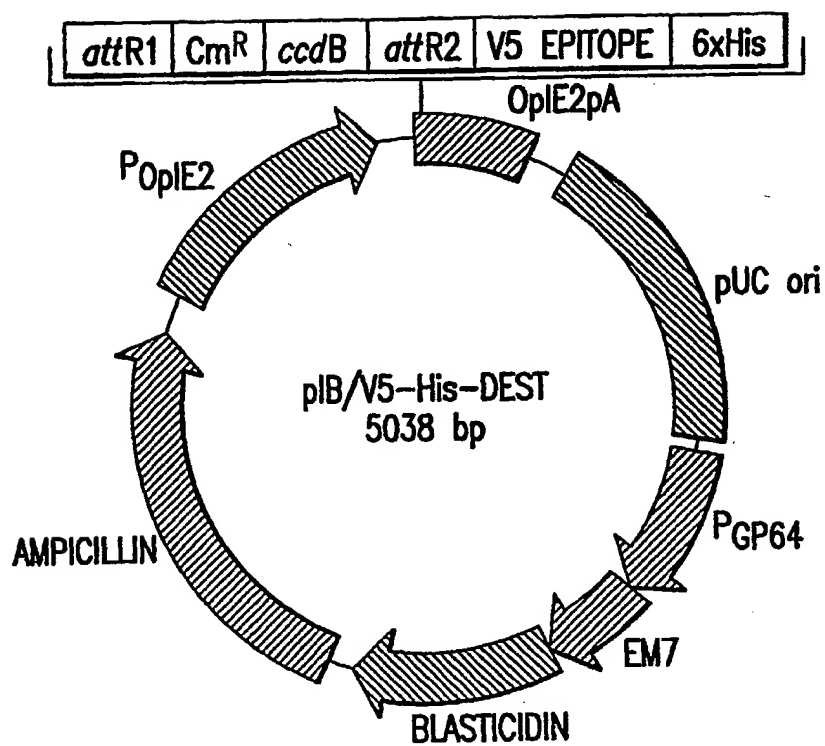


FIG.15

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1 GGATCATGAT GATAACAAT GTATGGTGCT AATGTTGCTT CAACAACAAT TCTGTTGAAC  
61 TGTGTTTTCA TGTTTGCCAA CAAGCACCTT TATACTCGGT GGCCTCCCA CCACCAACTT  
121 TTTTGCACTG CAAAAAACA CGCTTTTGCA CGCGGGCCCA TACATAGTAC AAACCTCTAGC  
181 TTTCTAGAC TATTTTACAT AAATAGTCTA CACGGTTGTA TAGCTCCAA ATACACTACC  
241 ACACATTGAA CCTTTTGGCA GTGCAAAAA GTACGTGTCG GCAGTCACGT AGGCCGGCCT  
301 TATCGGGTCG CGTCTGTGCA CGTACGAATC ACATTATCGG ACCGGACGAG TGTGTCTTA  
361 TCGTGACAGG ACGCCAGCTT CCTGTGTTGC TAACCGAGC CGGACGCAAC TCCTTATCGG  
421 AACAGGACGC GCCTCCATAT CAGCCGGCGG TTATCTCATG CGCGTGACCG GACACGAGGC  
481 GCCCGTCCCG CTTATCGGC CTATAATAC AGCCCGCAAC GATCTGGTAA ACACAGTTGA  
541 ACAGCATCTG TTCGAATTTA

TATA  
Start of Transcription

FIG.16

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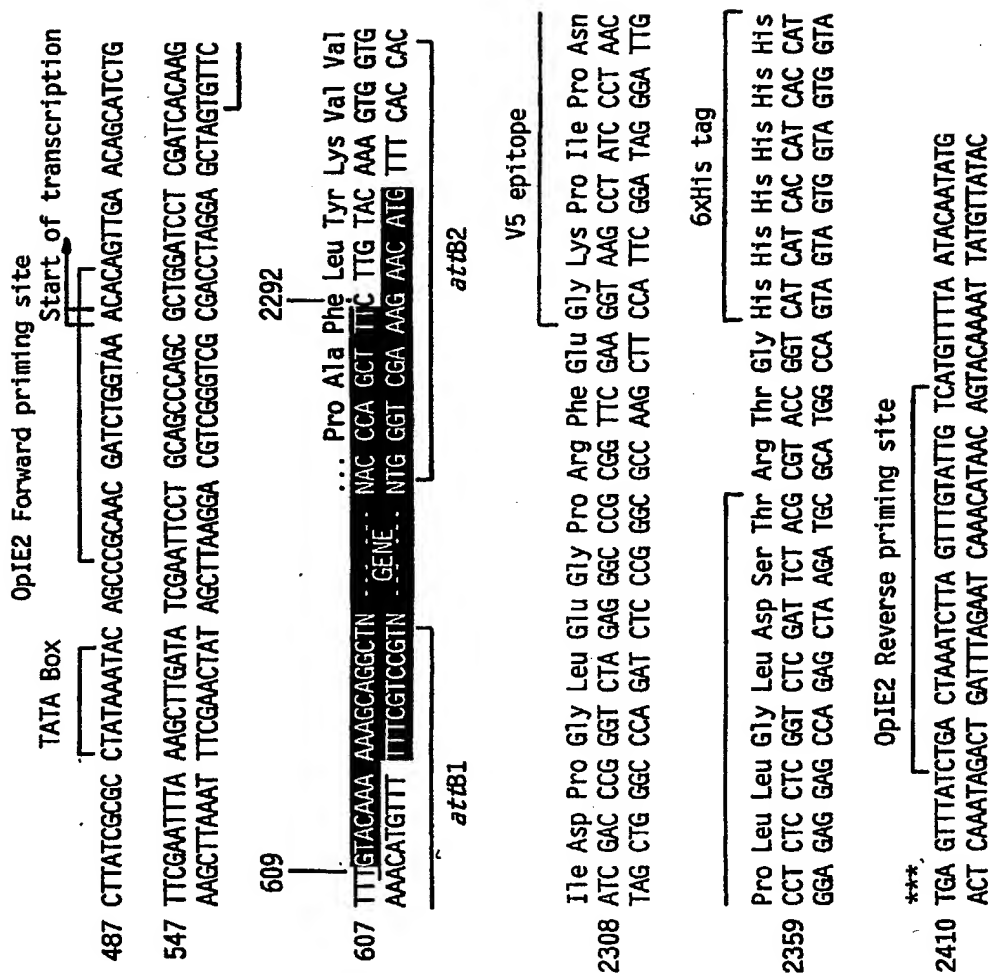


FIG. 17

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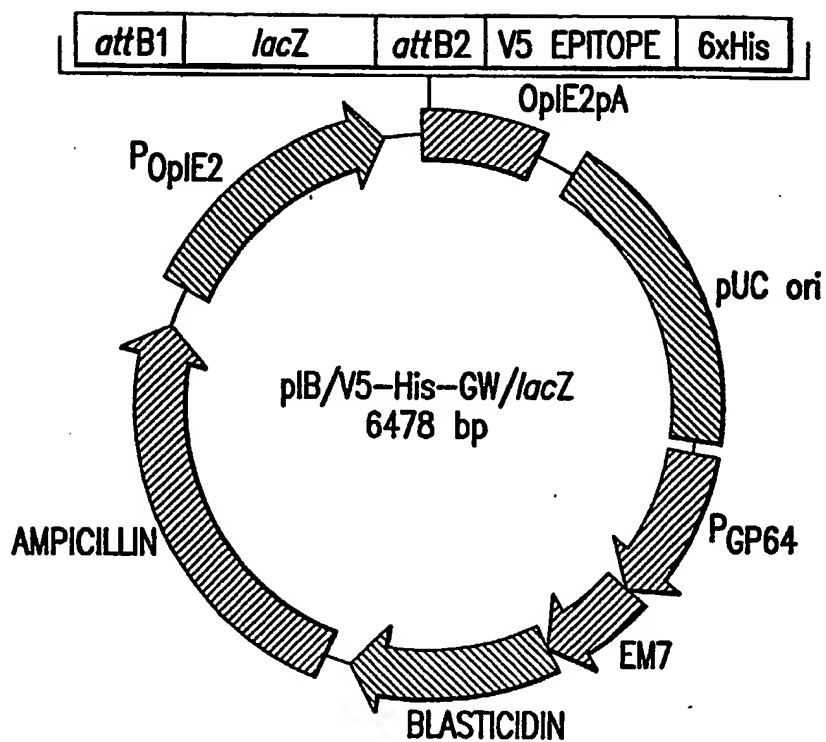


FIG.18



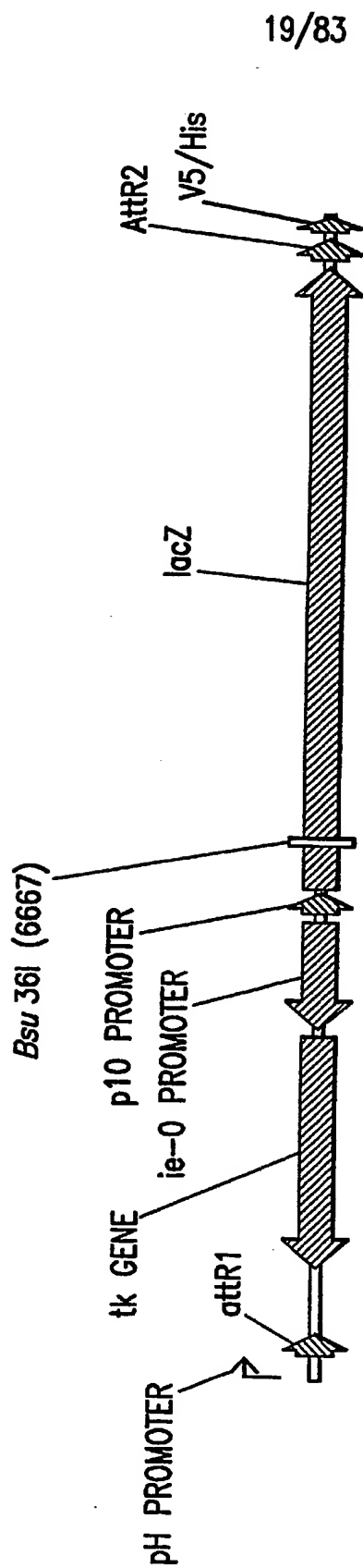


FIG. 19A

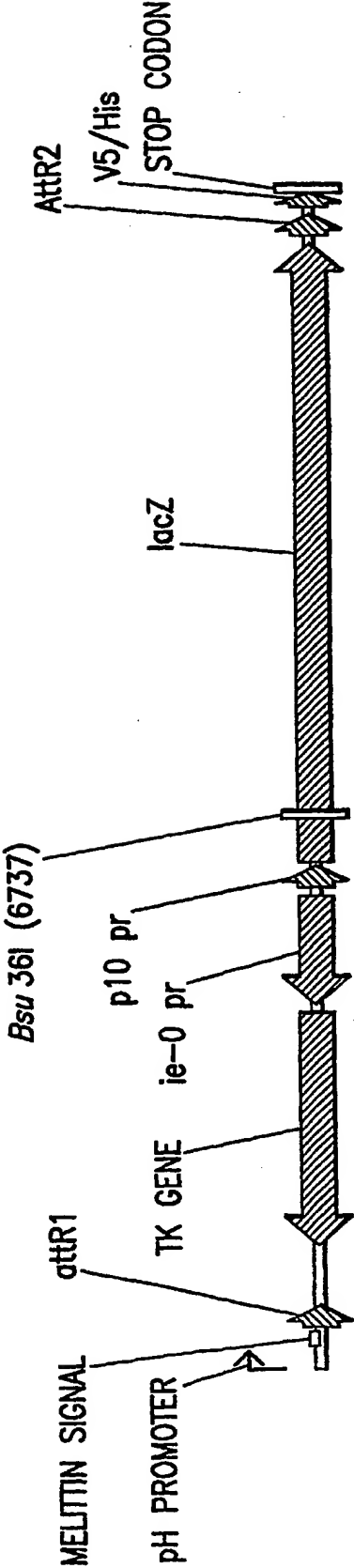


FIG. 19B

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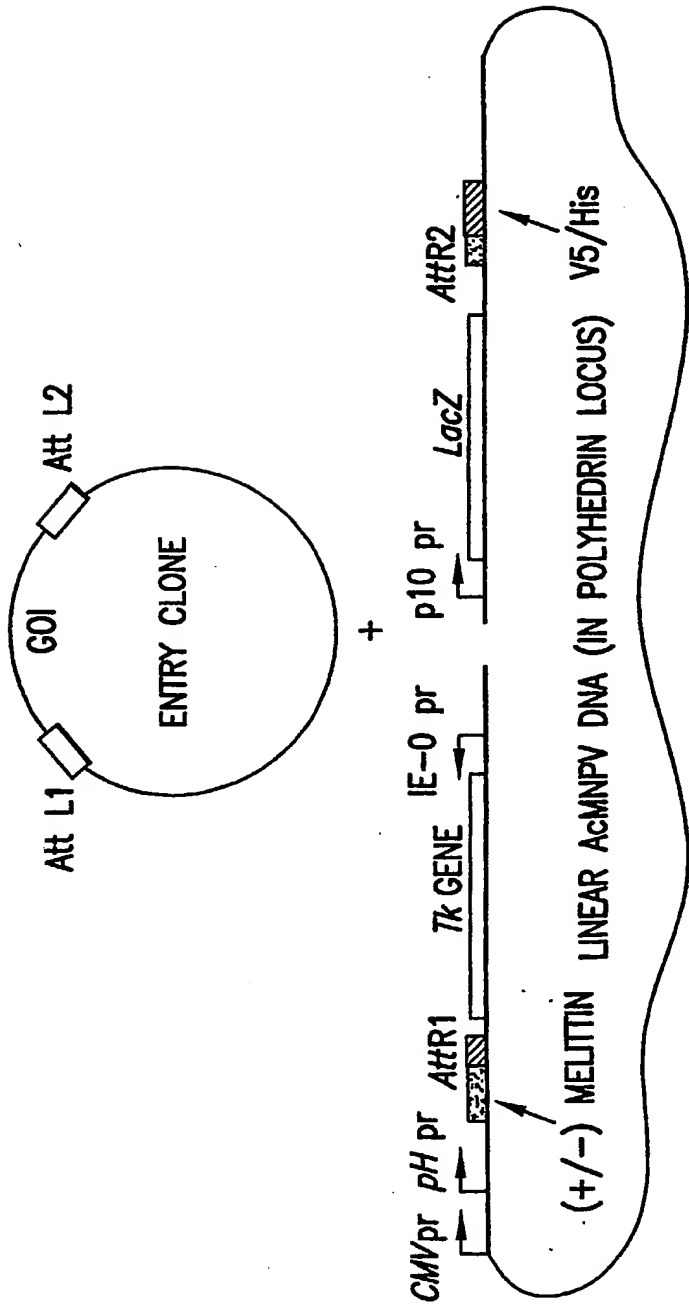
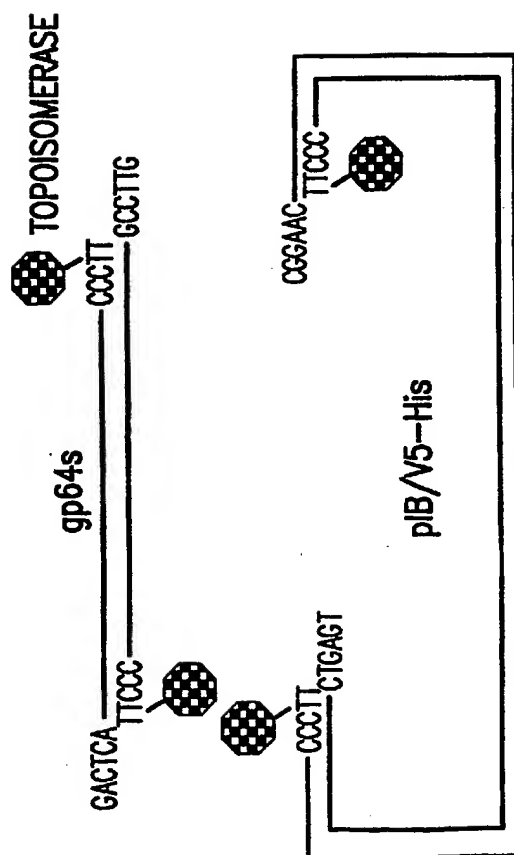


FIG.20



**FIG. 21**

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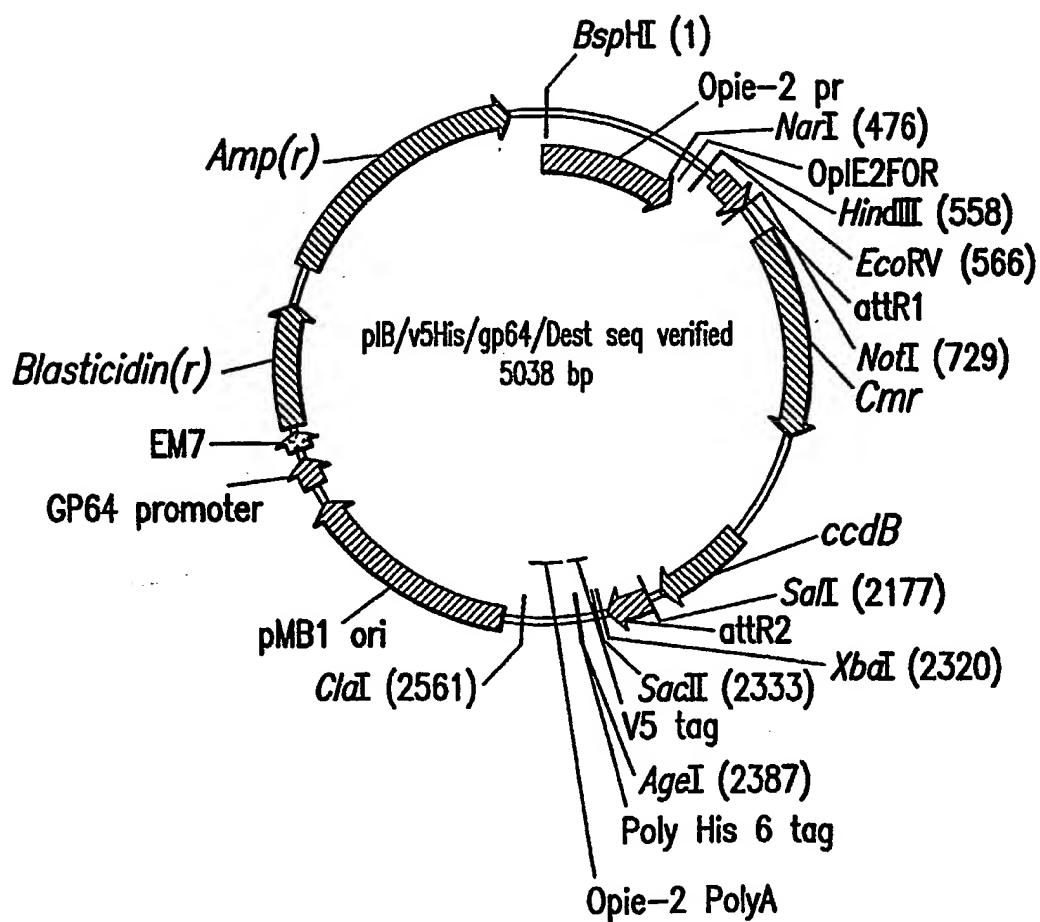


FIG.22

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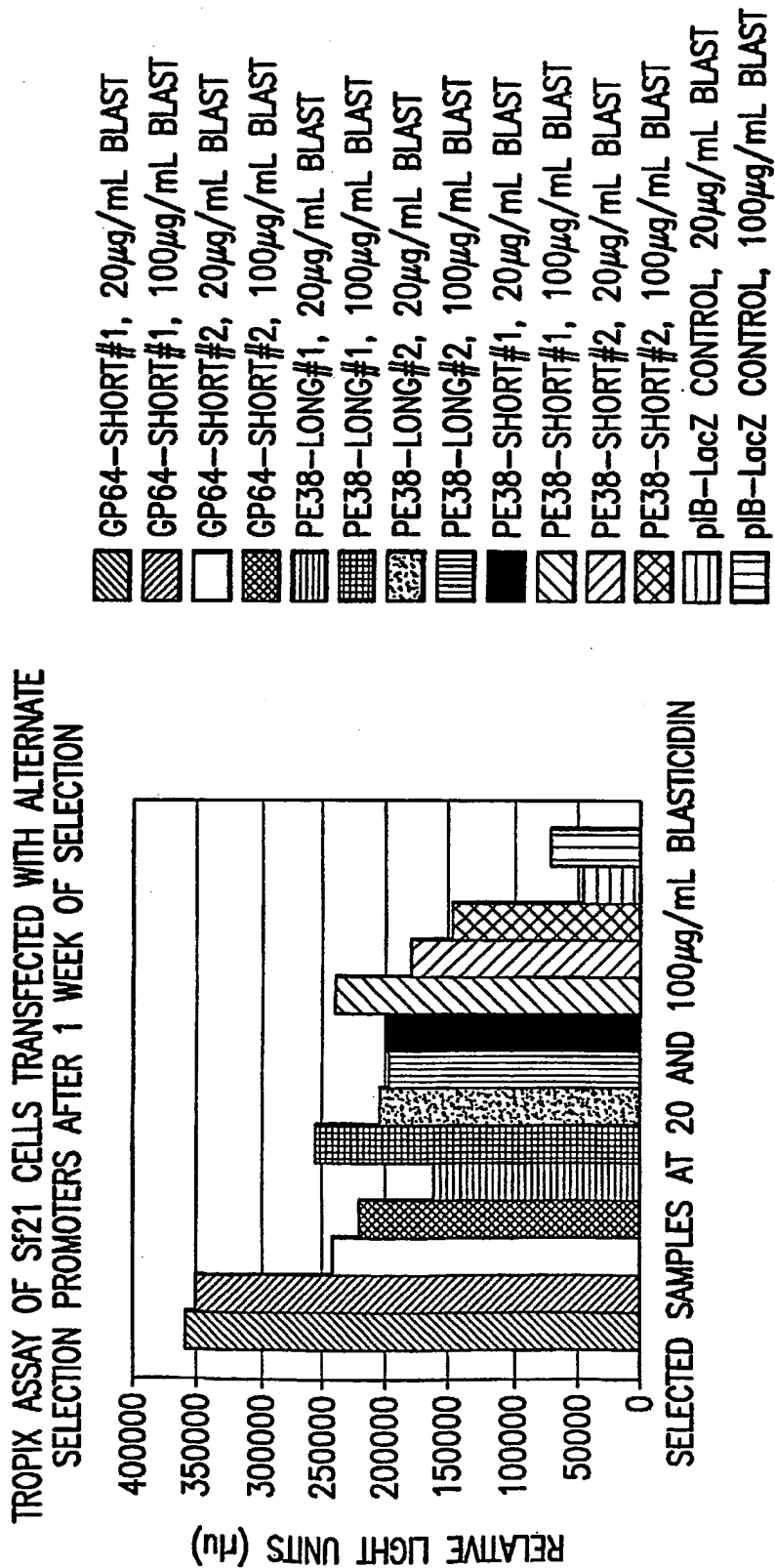


FIG.23

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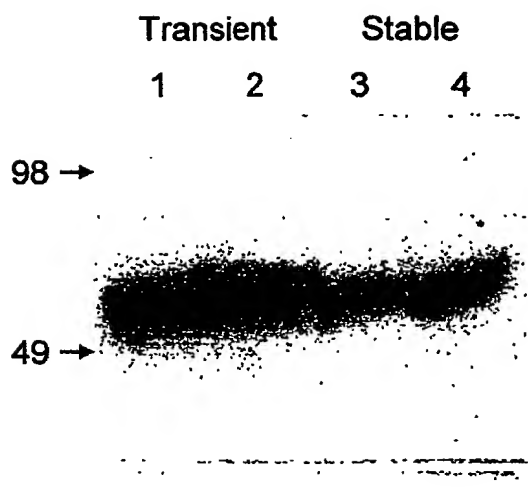


FIG.24

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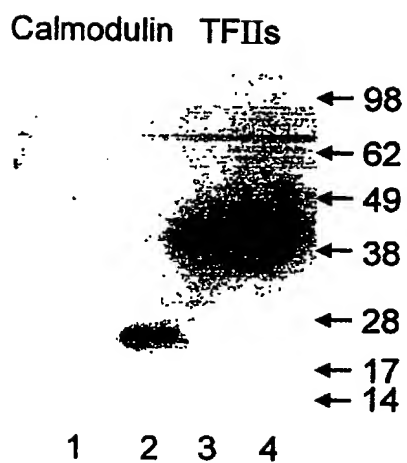


FIG.25A

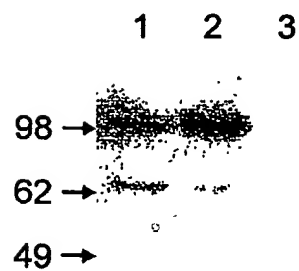


FIG.25B



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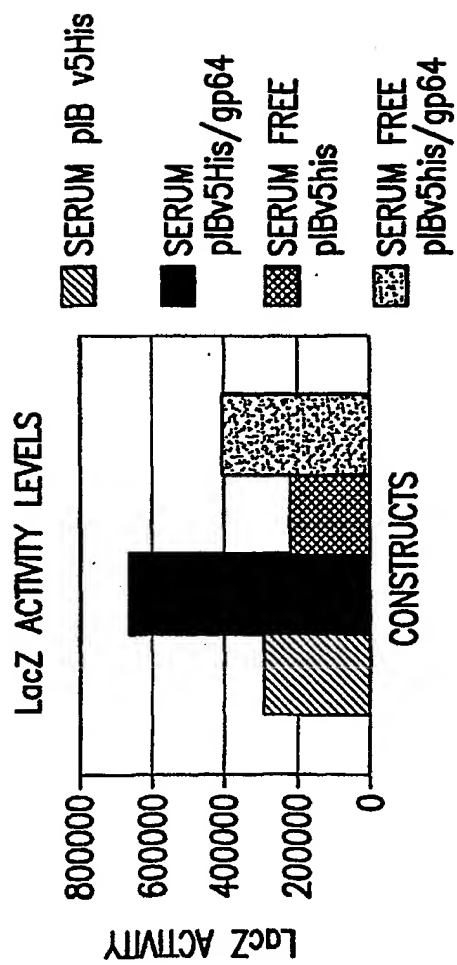
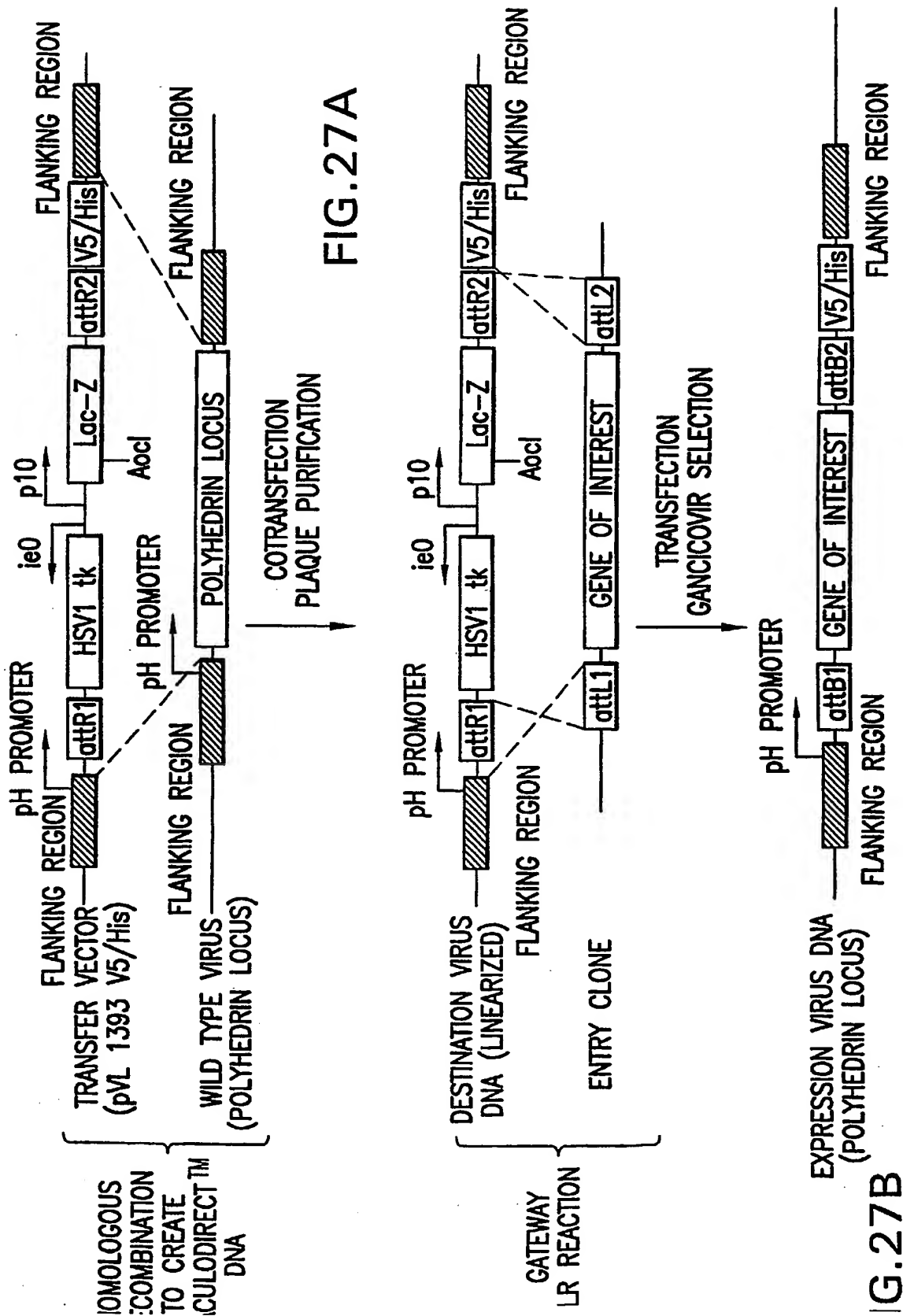


FIG.26

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# HTP PROTOCOL FOR USE WITH BACULODIRECT (TM)

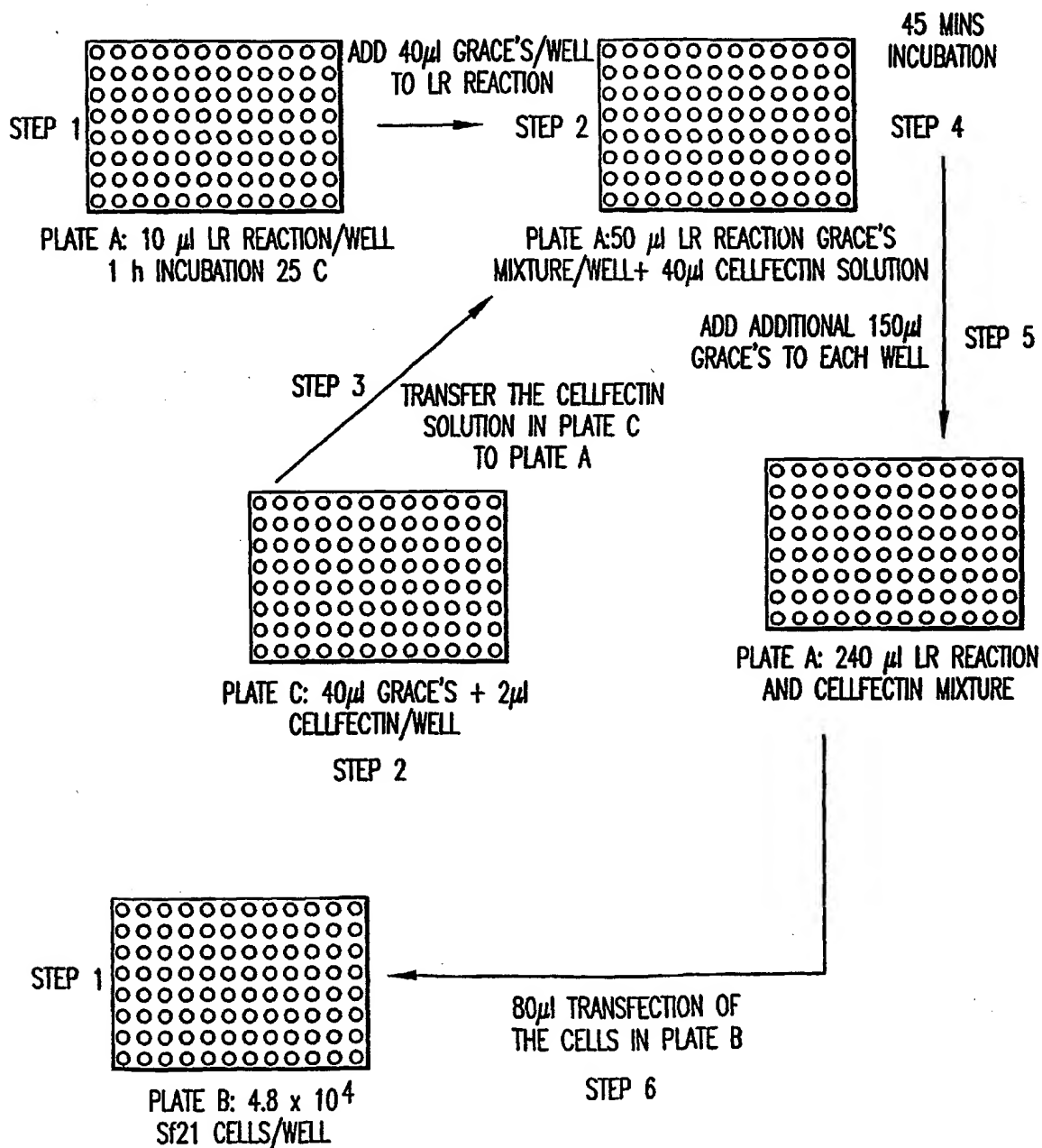


FIG.28

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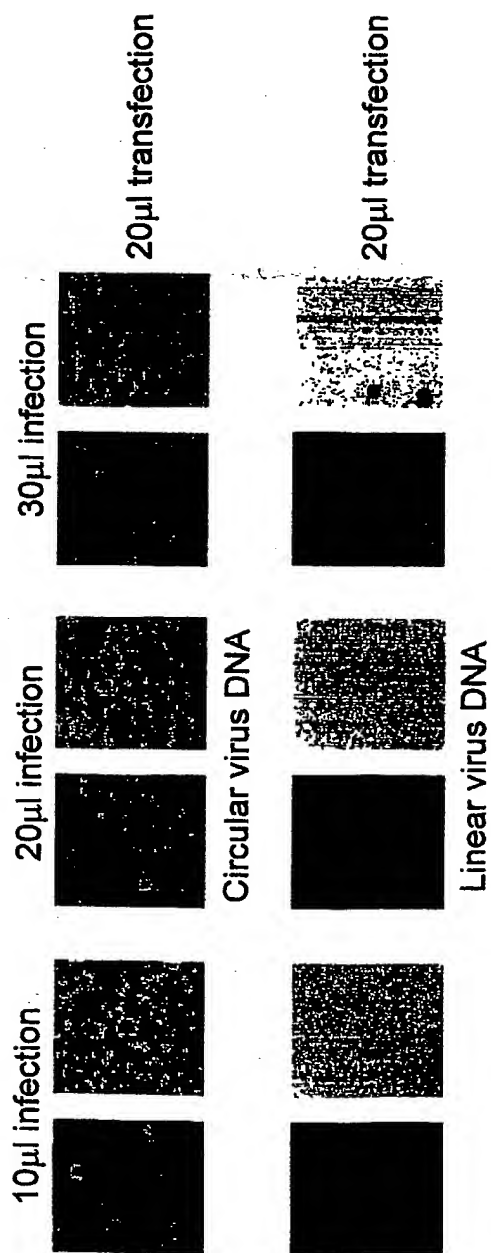


FIG.29

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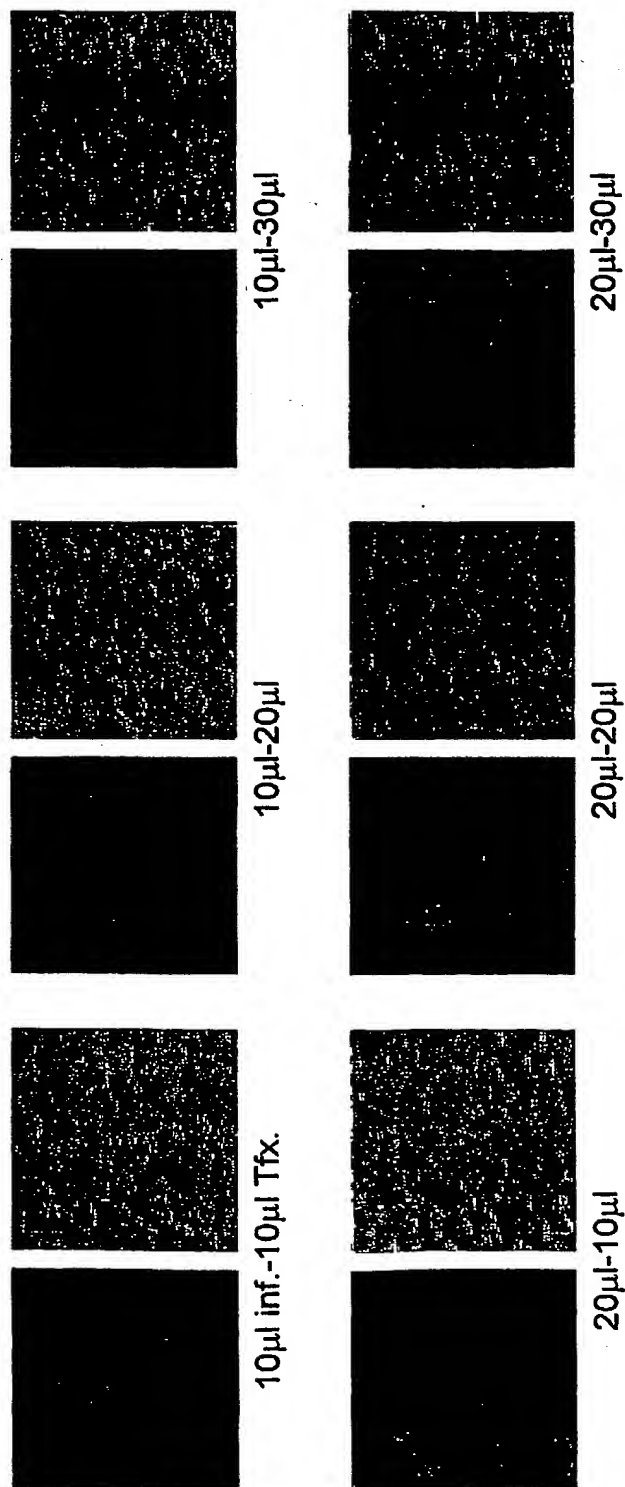


FIG. 30

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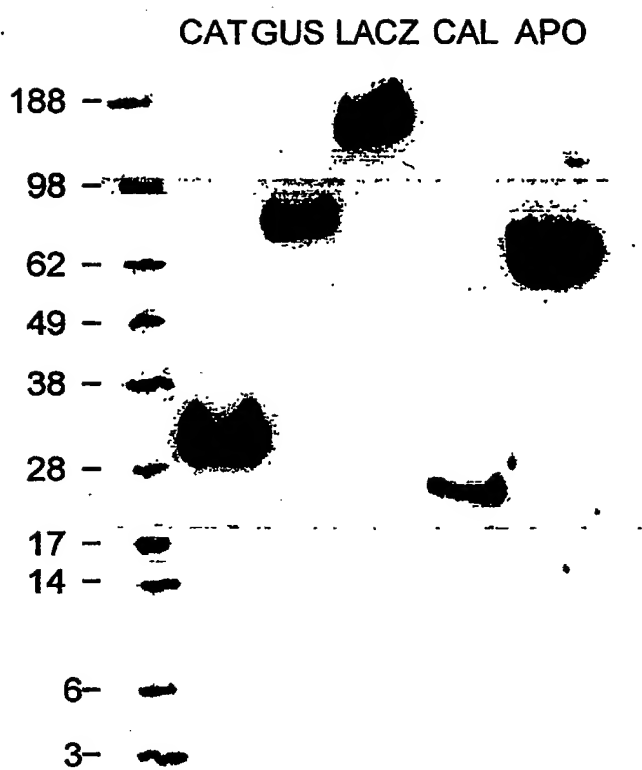


FIG.31

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| VIRUS \ TITER pfu/ml | TRANSFECT         | INFECT            |
|----------------------|-------------------|-------------------|
| BACULODIRECT™        | $9.8 \times 10^6$ | $6.9 \times 10^8$ |
| BAC TO BAC           | $6.9 \times 10^6$ | $7.2 \times 10^8$ |
| MAXBAC               | N/A               | $3.6 \times 10^8$ |

TITER COMPARISON OF THE THREE VIRUSES BY USING TCID<sub>50</sub>

| VIRUS \ TITER pfu/ml | TRANSFECT       | INFECT.         |
|----------------------|-----------------|-----------------|
| BACULODIRECT™        | $6 \times 10^6$ | $3 \times 10^8$ |
| BAC TO BAC           | $8 \times 10^6$ | $5 \times 10^8$ |
| MAXBAC               | N/A             | $3 \times 10^8$ |

TITER COMPARISON OF THE THREE VIRUSES BY USING PLAQUE ASSAY

FIG.32

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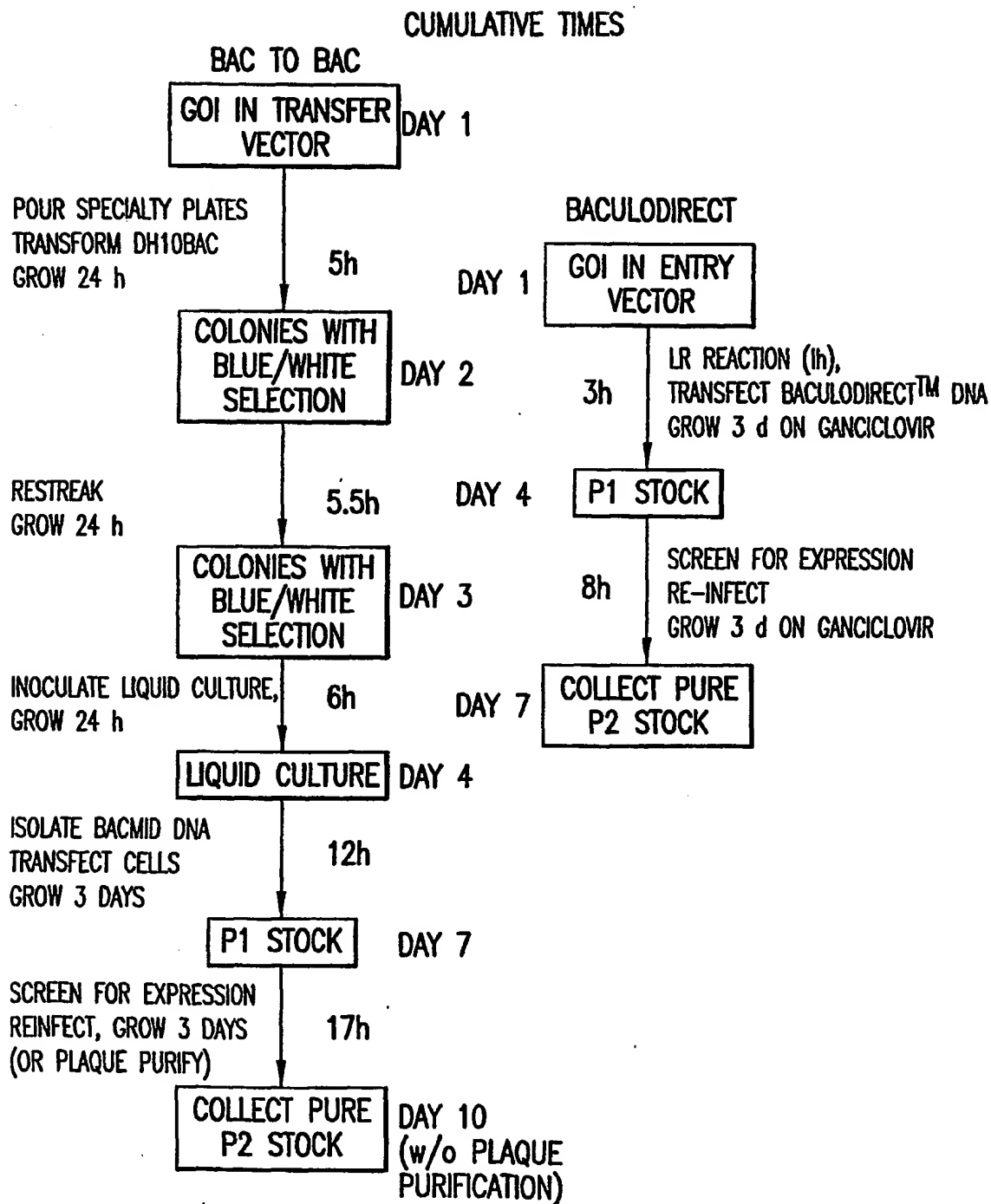


FIG.33



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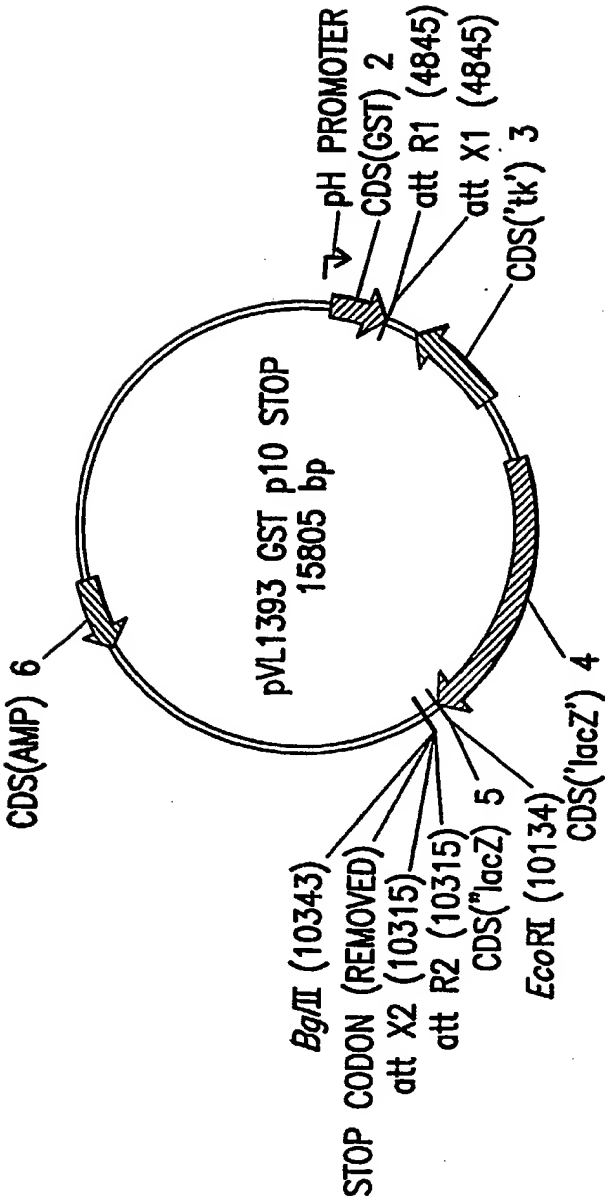


FIG.34

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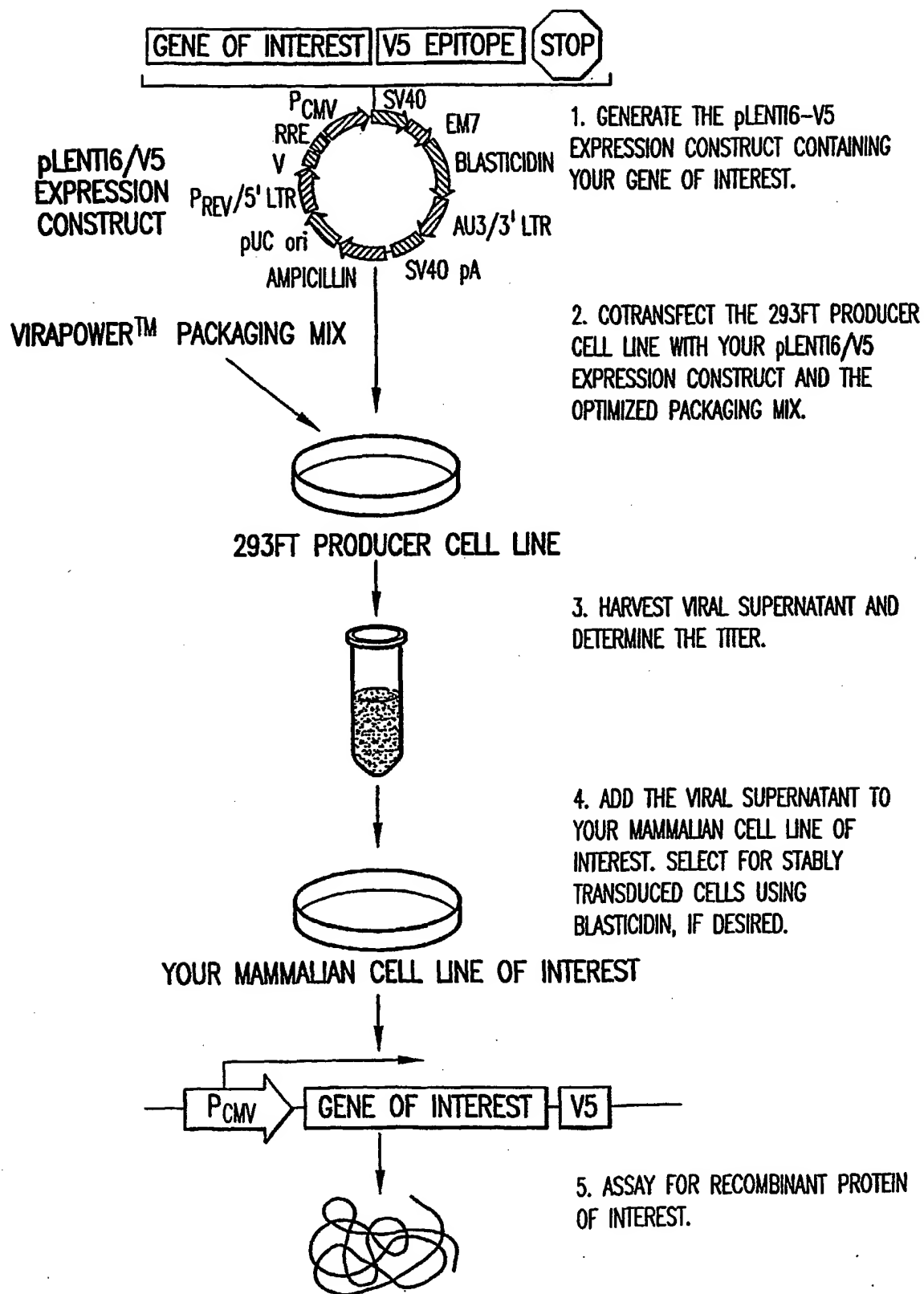


FIG 35

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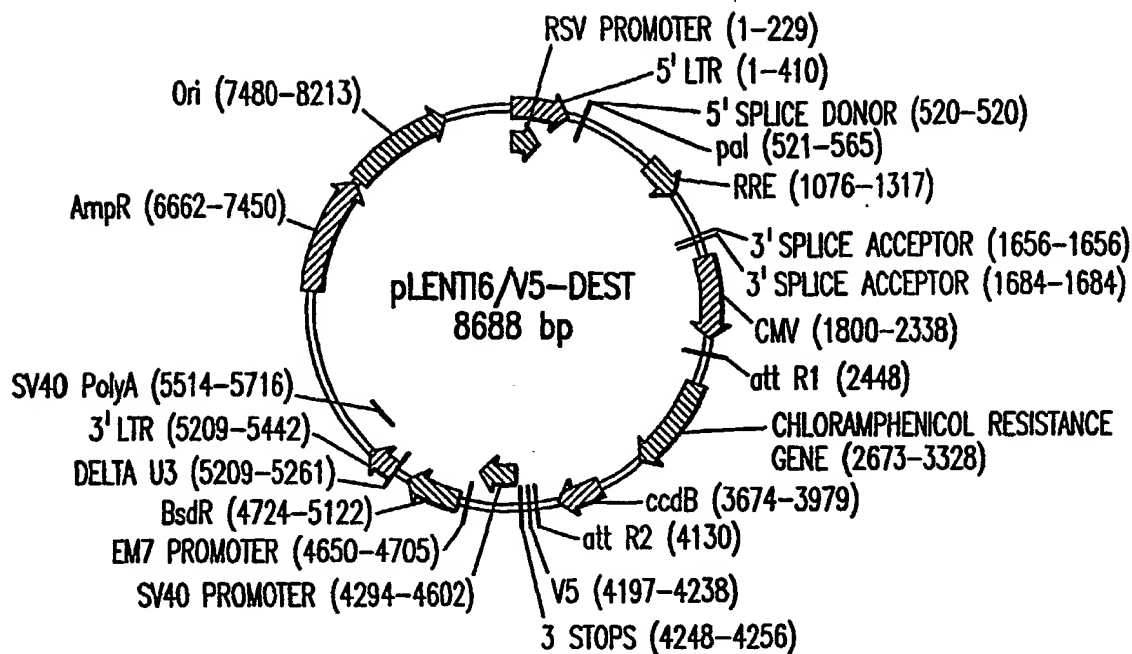


FIG.36A

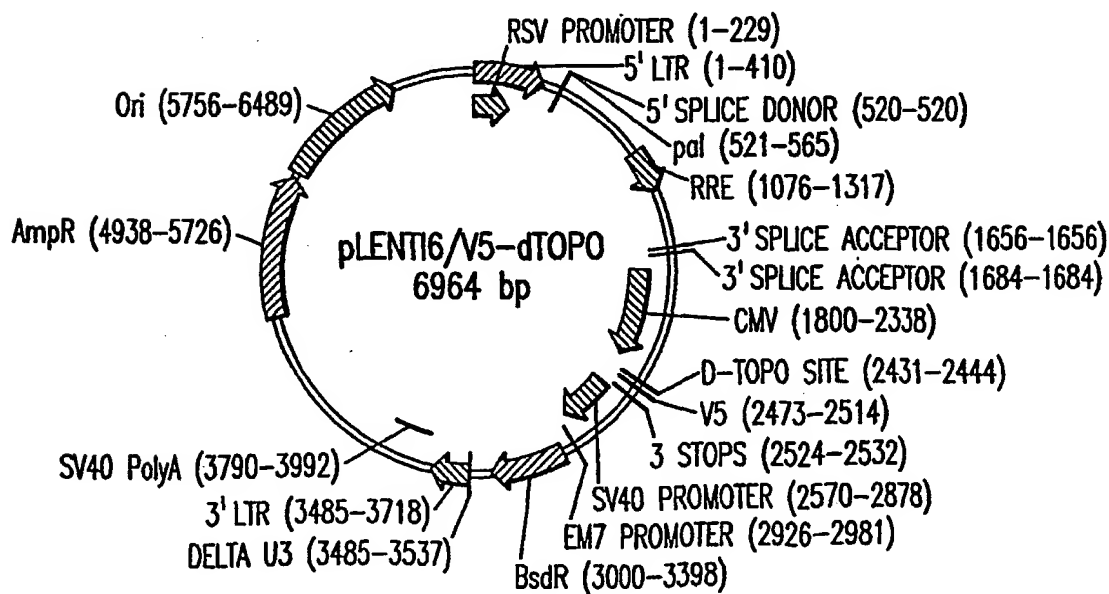


FIG 36B

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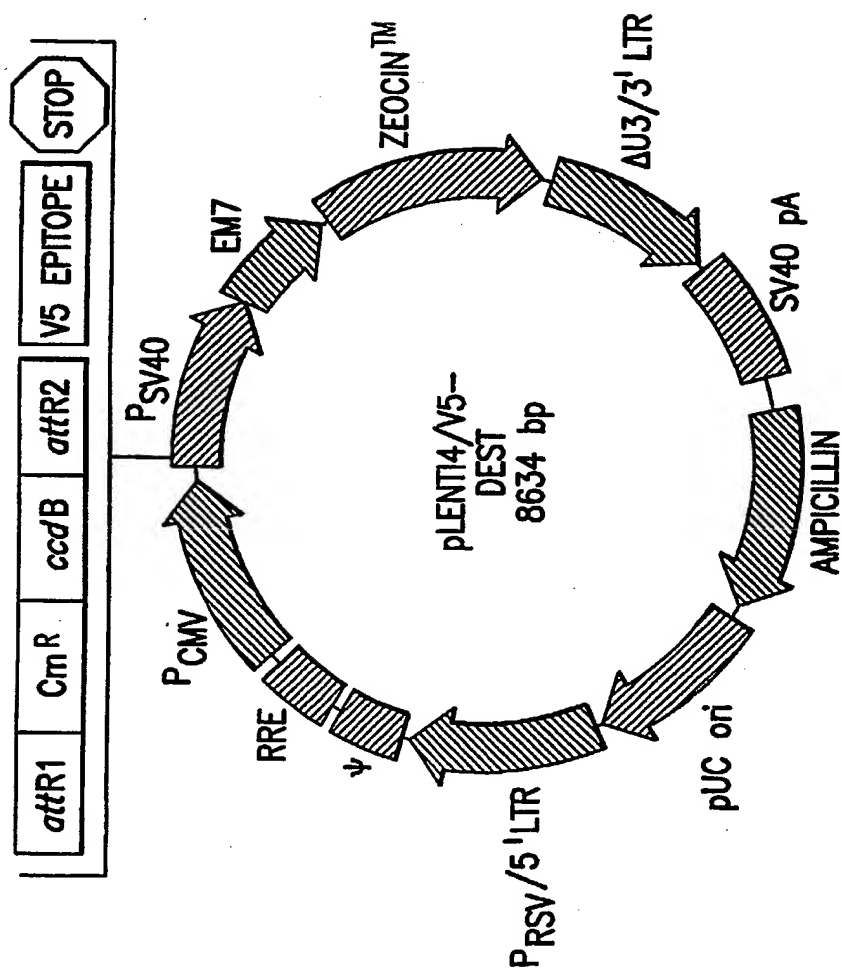


FIG.36C

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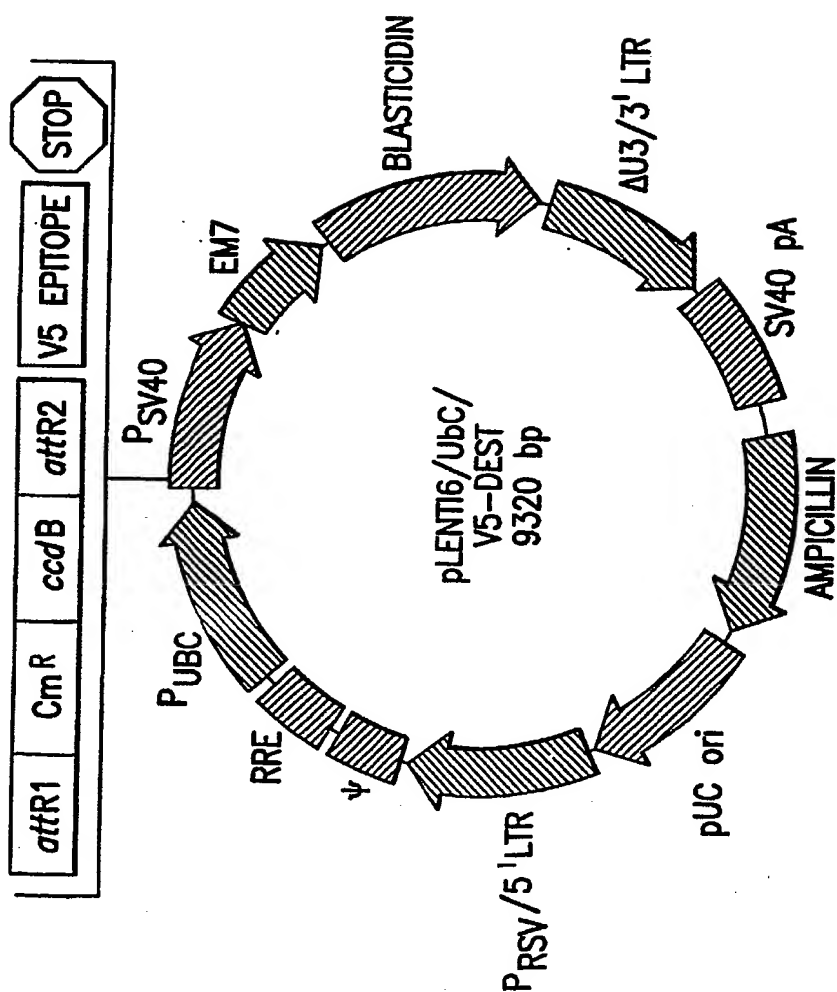


FIG.36D

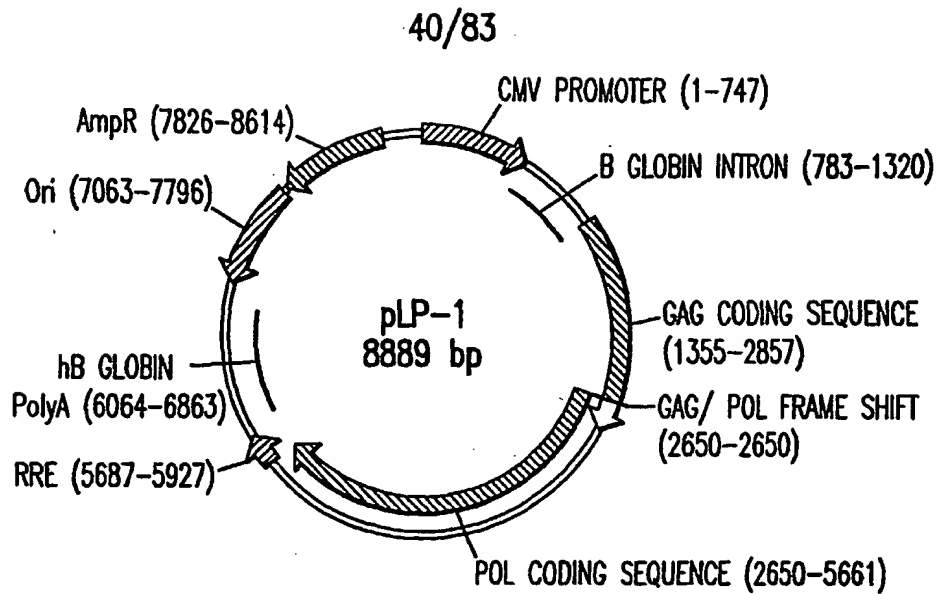


FIG.37A

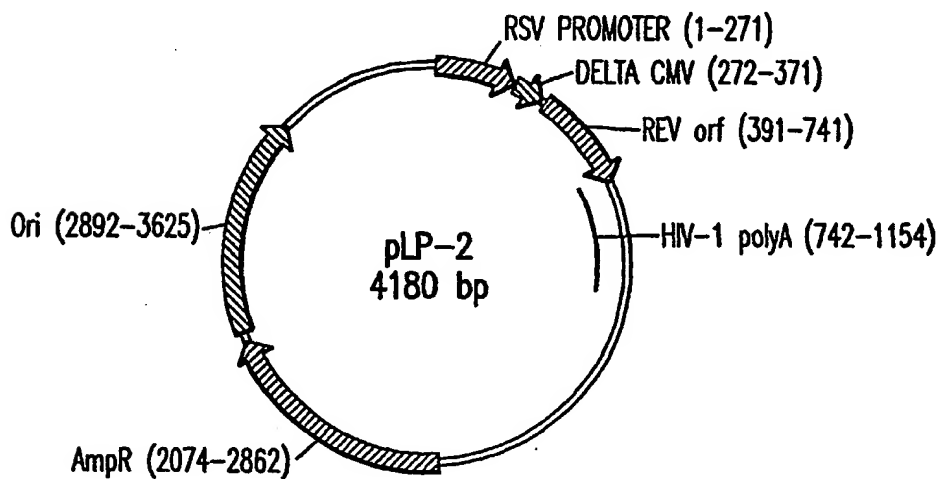


FIG.37B

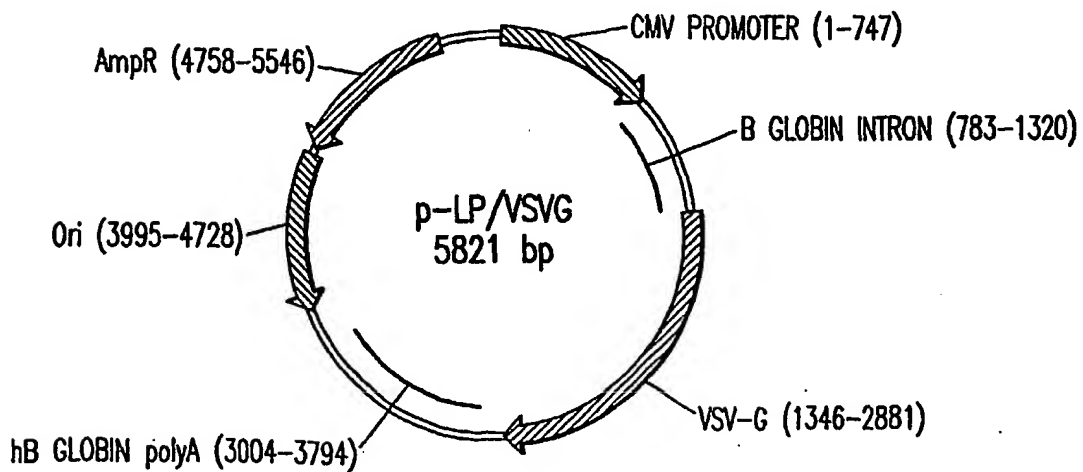


FIG. 37C

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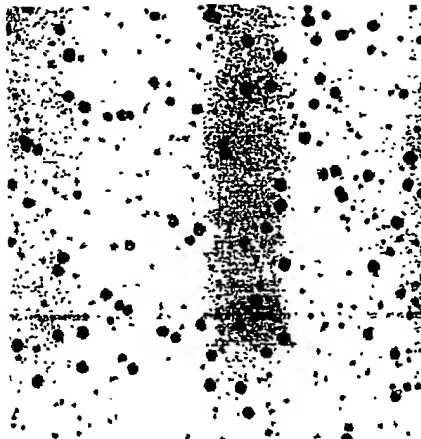


FIG.38A

| [Bsd] IN PLATE \ LR REACTION | DEST ALONE | DEST + CAT                    |
|------------------------------|------------|-------------------------------|
|                              |            |                               |
| NO Bsd                       | 24         | $320^1$<br>$(12/24 = 50\%)^2$ |
| 50 $\mu$ g/ml Bsd            | 0          | $162$<br>$(24/24 = 100\%)^2$  |

<sup>1</sup>SEE PHOTO ABOVE<sup>2</sup>PERCENTAGE OF CORRECT COLONIES

FIG.38B

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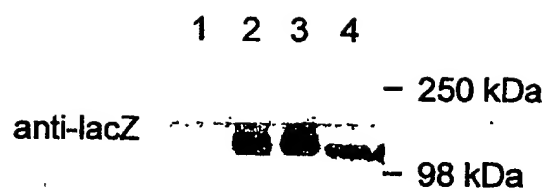


FIG.39A

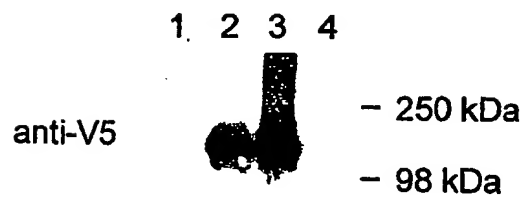


FIG.39B



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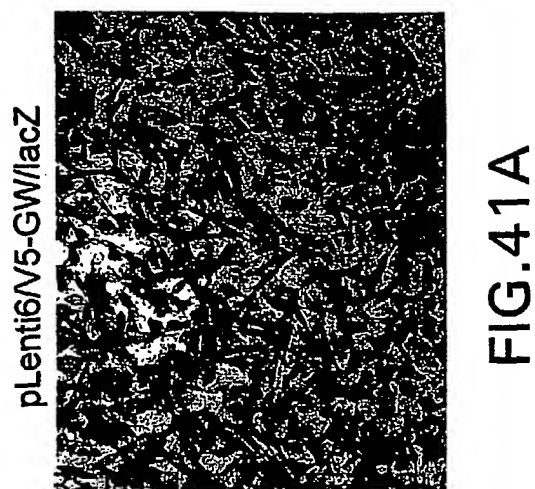
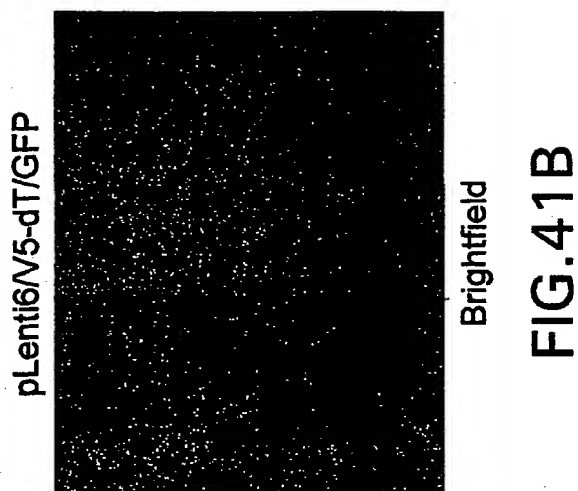
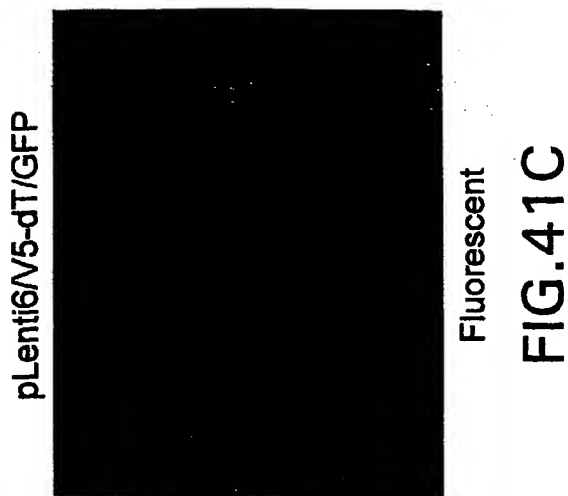
EXAMPLES OF PRODUCTION TITERS (Bsd<sup>R</sup> cfu/ml)

|       | EMPTY             | LacZ              | GFP             | CAT             | PKC             |
|-------|-------------------|-------------------|-----------------|-----------------|-----------------|
| EXP 1 | $6 \times 10^6$   | $5 \times 10^5$   | $4 \times 10^6$ | N.D.            | N.D.            |
| EXP 2 | $3 \times 10^7$   | $3 \times 10^5$   | $6 \times 10^6$ | $8 \times 10^6$ | N.D.            |
| EXP 3 | $7 \times 10^6$   | $6 \times 10^5$   | $2 \times 10^6$ | $1 \times 10^7$ | $3 \times 10^6$ |
| AVG   | $1.4 \times 10^7$ | $4.7 \times 10^5$ | $4 \times 10^6$ | $9 \times 10^6$ | $3 \times 10^6$ |

N.D. = NOT DETERMINED

FIG.40

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FIG.42A

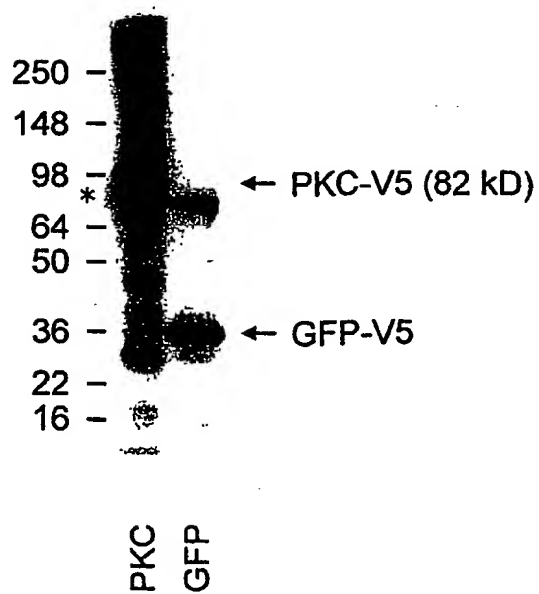


FIG.42B

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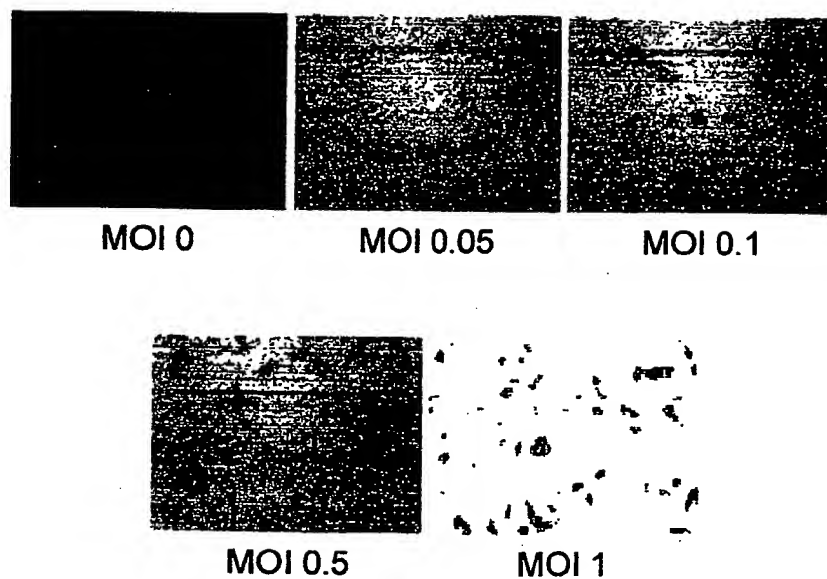


FIG.43A

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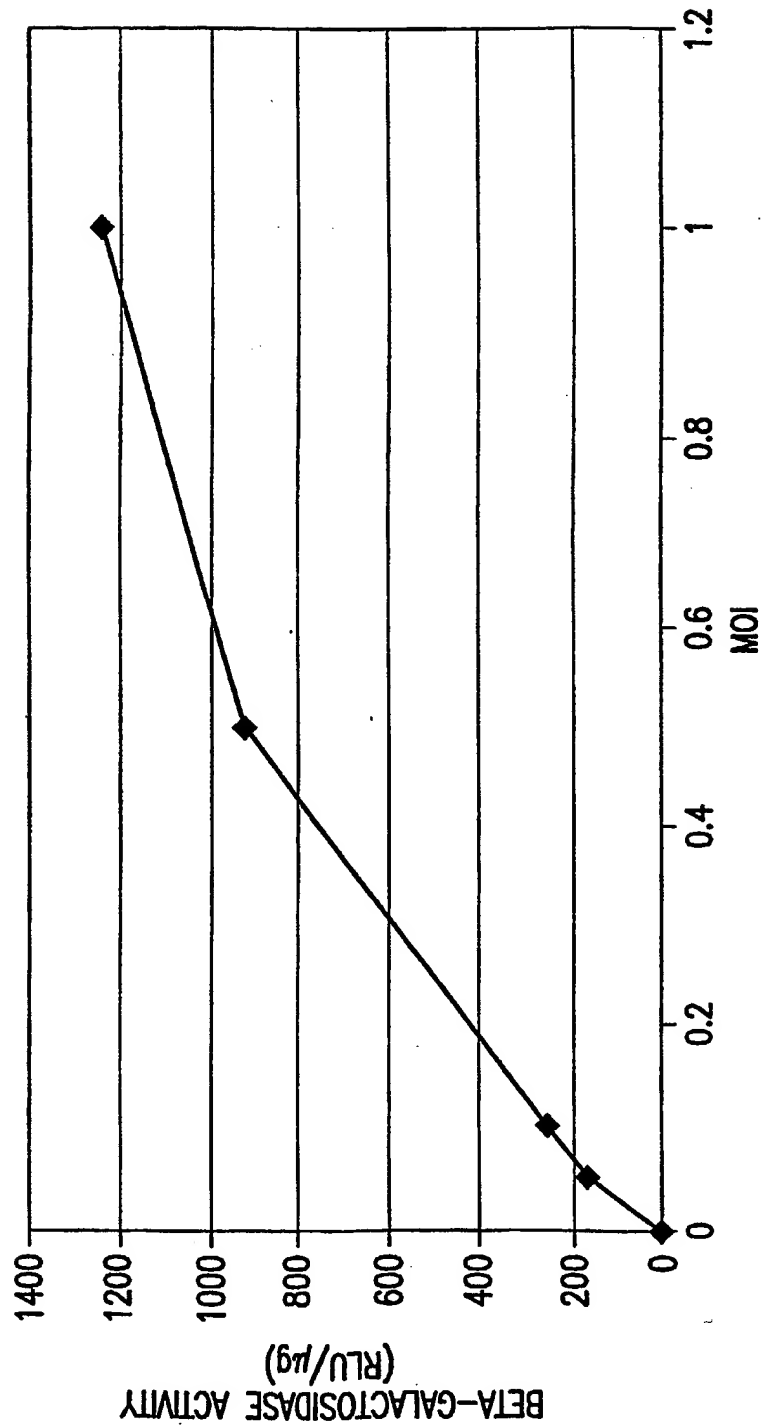
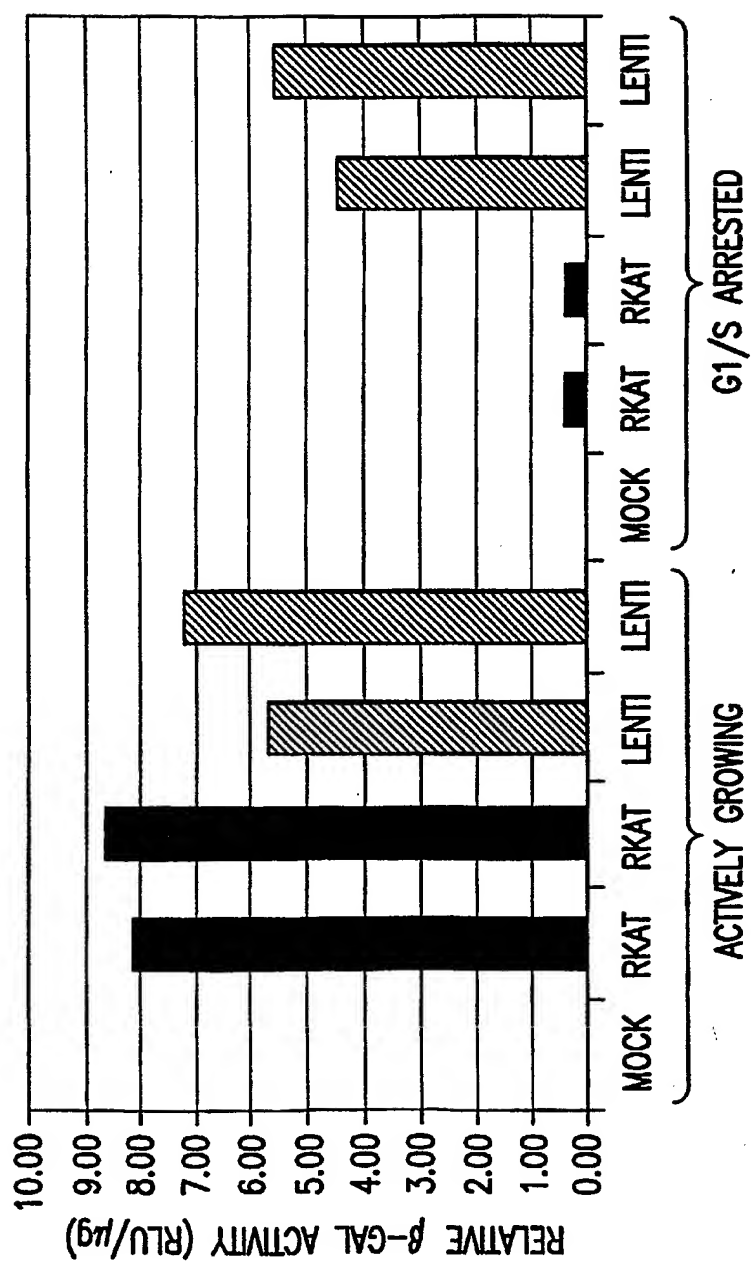


FIG. 43B

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**FIG. 44A**

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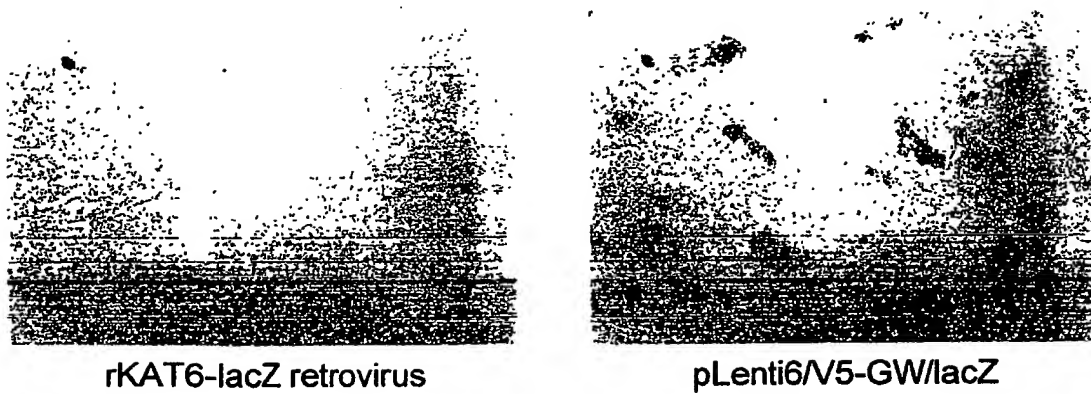


FIG.44B

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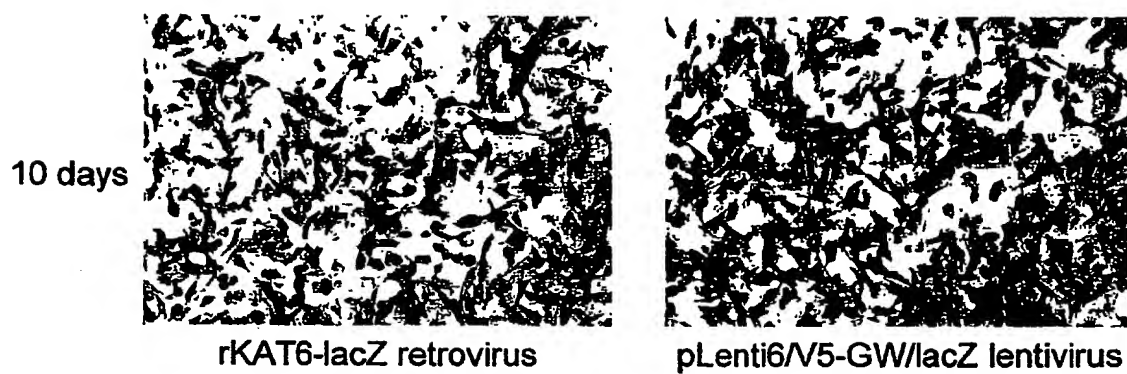


FIG.45A

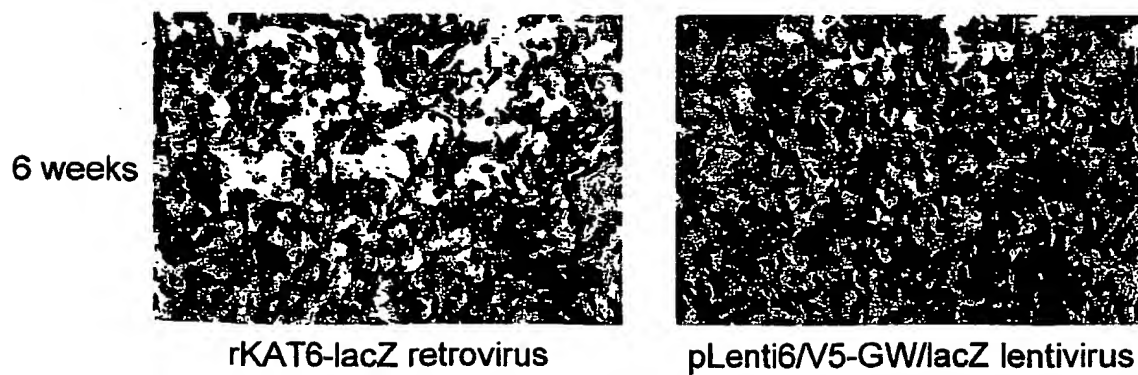


FIG.45B



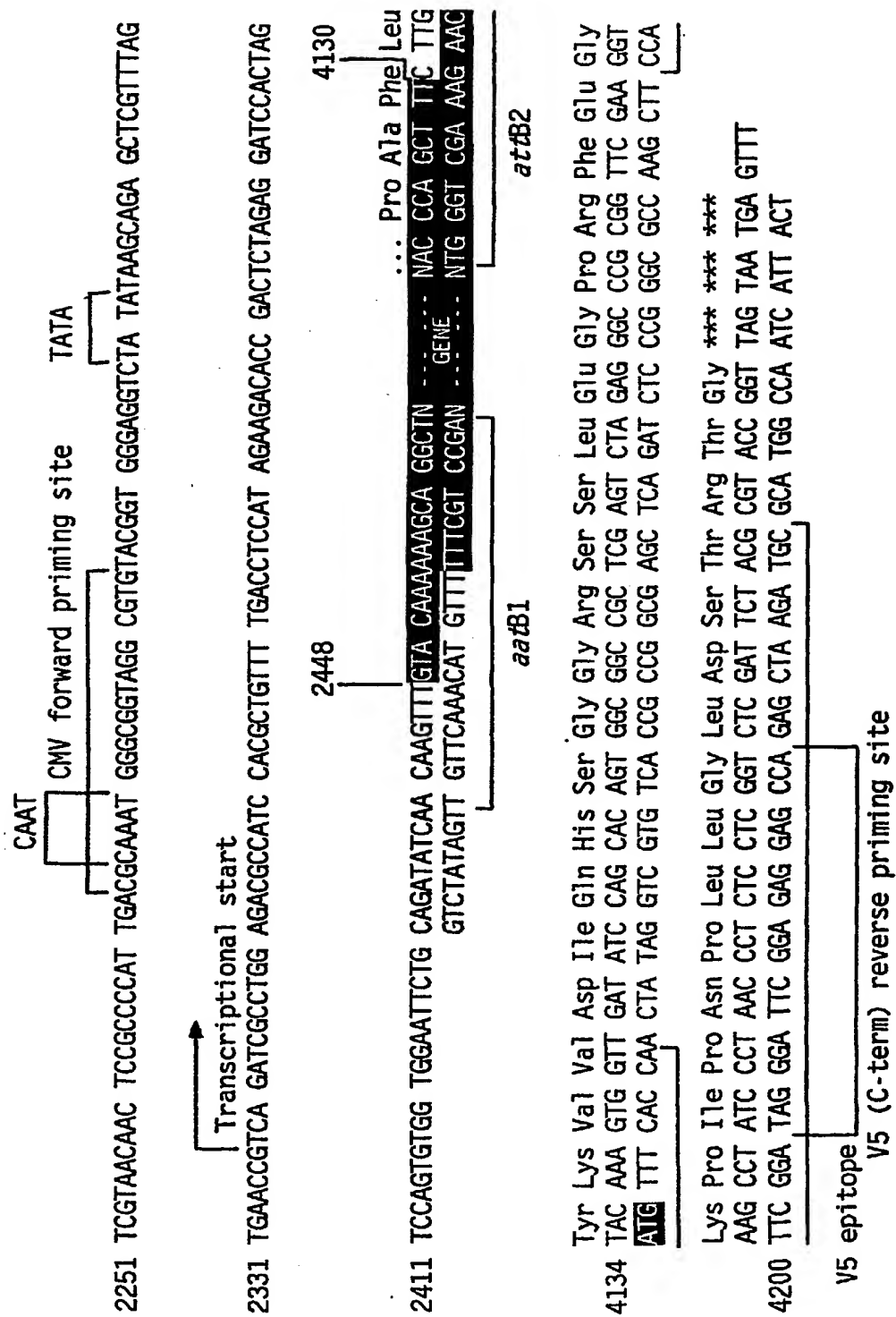


FIG.46A

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UB forward priming site

2881 TTGGCGAGTG TGTGTTGGA AGTTTTTTAG GCACCTTTTG AAATGTAATC ATTGGGTCA ATATGTAATT TTCAGTGTTA

2961 GACTAGTAAA TTGTCGCTA AATCTGGCC GTTTTGGCT TTTTGTAG ACGAAGCTTG GTACCGAGCT CGGATCCACT

3079 4762

3041 AGTCCAGTGT GGTGGAATTC TGCAGATATC AACAGTTTG TACAAAAAAG CAGGCTN ... Pro Ala Phe  
ACGTCTATAG TTGTTCAAAC ATGTTTTC GTCCGAN ... GENE ... NAC CCA GCT TTC  
NTG GGT CGA AAG

aaB1

atB2

Leu Tyr Lys Val Val Asp Ile Gln His Ser Gly Gly Arg Ser Ser Leu Glu Gly Pro Arg Phe Glu  
4763 TTG TAC AAA GTG GTT GAT ATC CAG CAC AGT GGC GGC CGC TCG AGT CTA GAG GGC CCG CGG TTC GAA  
AAC ATG TTT CAC CAA CTA TAG GTC GTG TCA CCG CCG GCG AGC TCA GAT CTC CCG GGC GCC AAG CTT

Gly Lys Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Arg Thr Gly \*\*\* \*\*\* \*\*\*

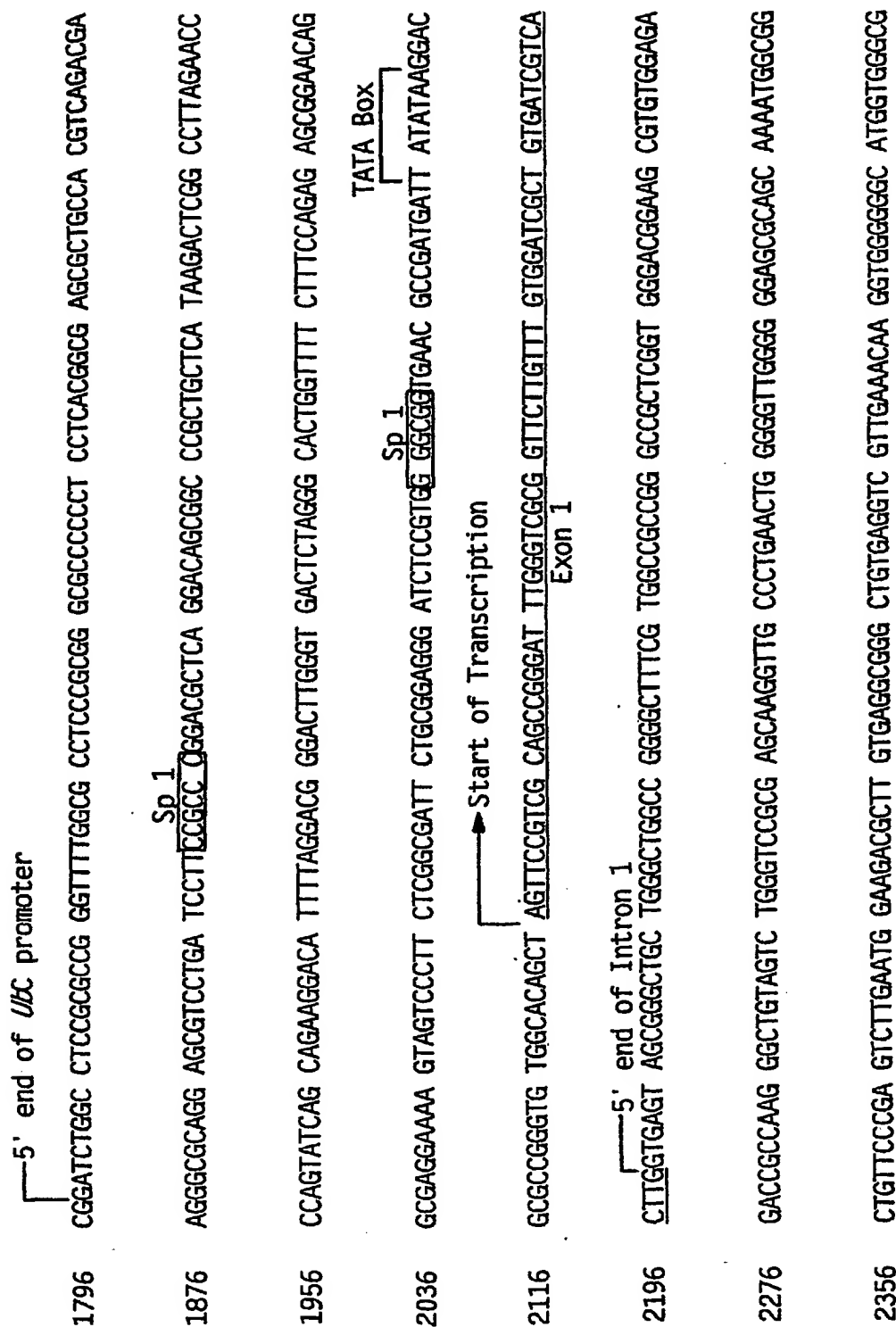
4829 GGT AAG CCT ATC CCT AAC CCT CTC CTC GGT CTC GAT TCT ACG CGT ACC GGT TAG TAA TGA GTTT  
CCA TTC GGA TAG GGA TTC GGA GAG CCA GAG CTA AGA TGC GCA TGG CCA

V5 epitope

V5 (C-term) reverse priming site

FIG.46B

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2436 GCAAGAACCC AAGGTCTTGA GGCCTTCGCT AATGCGGGA AGCTCTTATT CGGGTGAGAT GGGCTGGGGC ACCATCTGGG  
 2516 GACCCTGACG TGAAGTTTGT CACTGACTGG AGAACTCGGT TTGTCGTCTG TTGCGGGGGC GGCAGTTATG CCGTGCCCGTT  
 2596 GGGCAGTGCA CCGGTACCTT TGGGAGCGG CGCCCTCGTC GTGTCGTGAC GTCACCCGTT CTGTTGGCTT ATAATGCAGG  
 2676 GTGGGGCCAC CTGCCGGTAG GTGTGCGGTA GGCTTTTCTC CGTCGCAGGA CGCAGGGTTC GGGCCTAGGG TAGGCTCTCC  
 2756 TGAATCGACA GCGGCCGGAC CTCTGGTGAG GGGAGGATA AGTGAGGCGT CAGTTTCTTT GGTCGGTTTT ATGTACCTAT  
 2836 CTTCTTAAGT AGCTGAAGCT CCGGTTTGA ACTATCGCT CCGGGTTGGC GAGTGTGTTT TGTGAAGTTT TTTAGGCACC  
 2916 TTTTGAATG TAATCATTG GGTCAATATG TAATTTTCAG TGTTAGACTA GTAAATTGTC CGCTAAATTC TGGCCGTTTT

UB Forward priming site

3' end of Intron 1  
 2996 TGGCTTTTTT GTTAGACGAA GCTTGG....  
 5' end of Exon 2

FIG.46D

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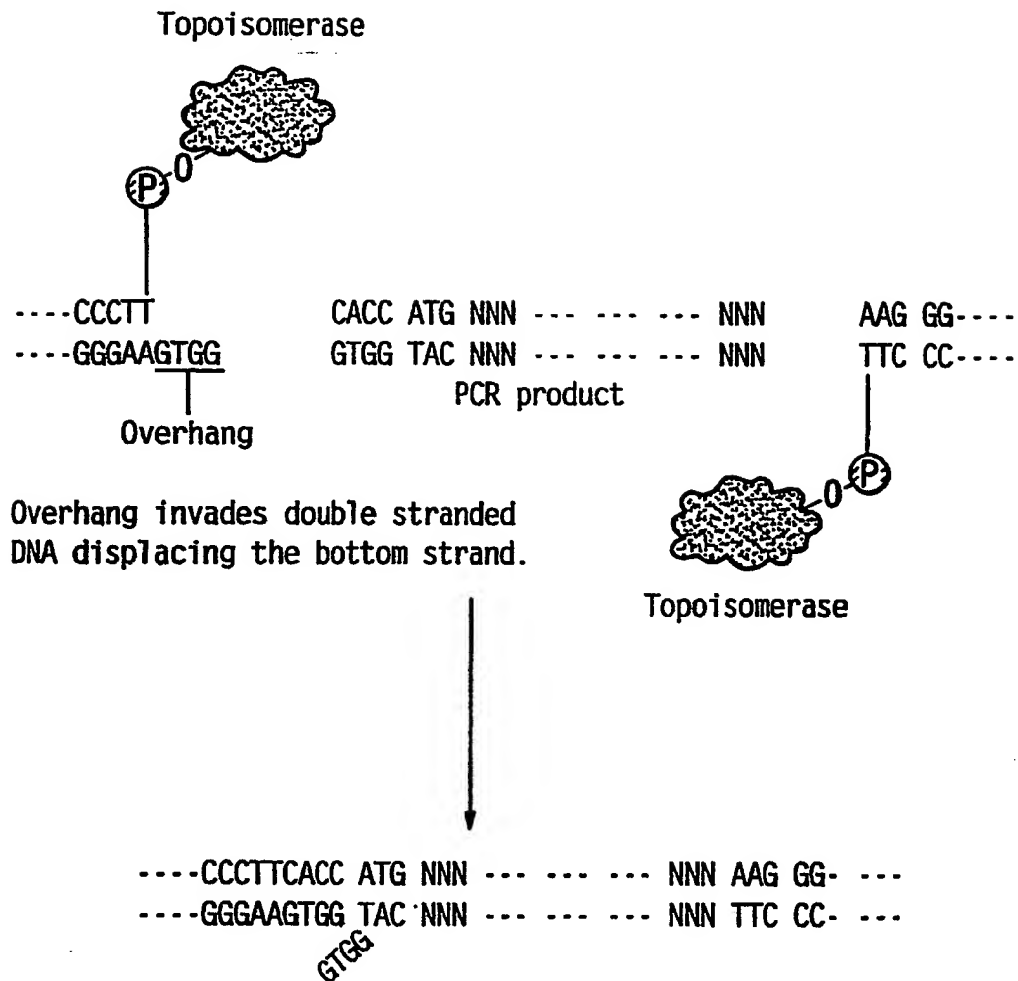


FIG.47

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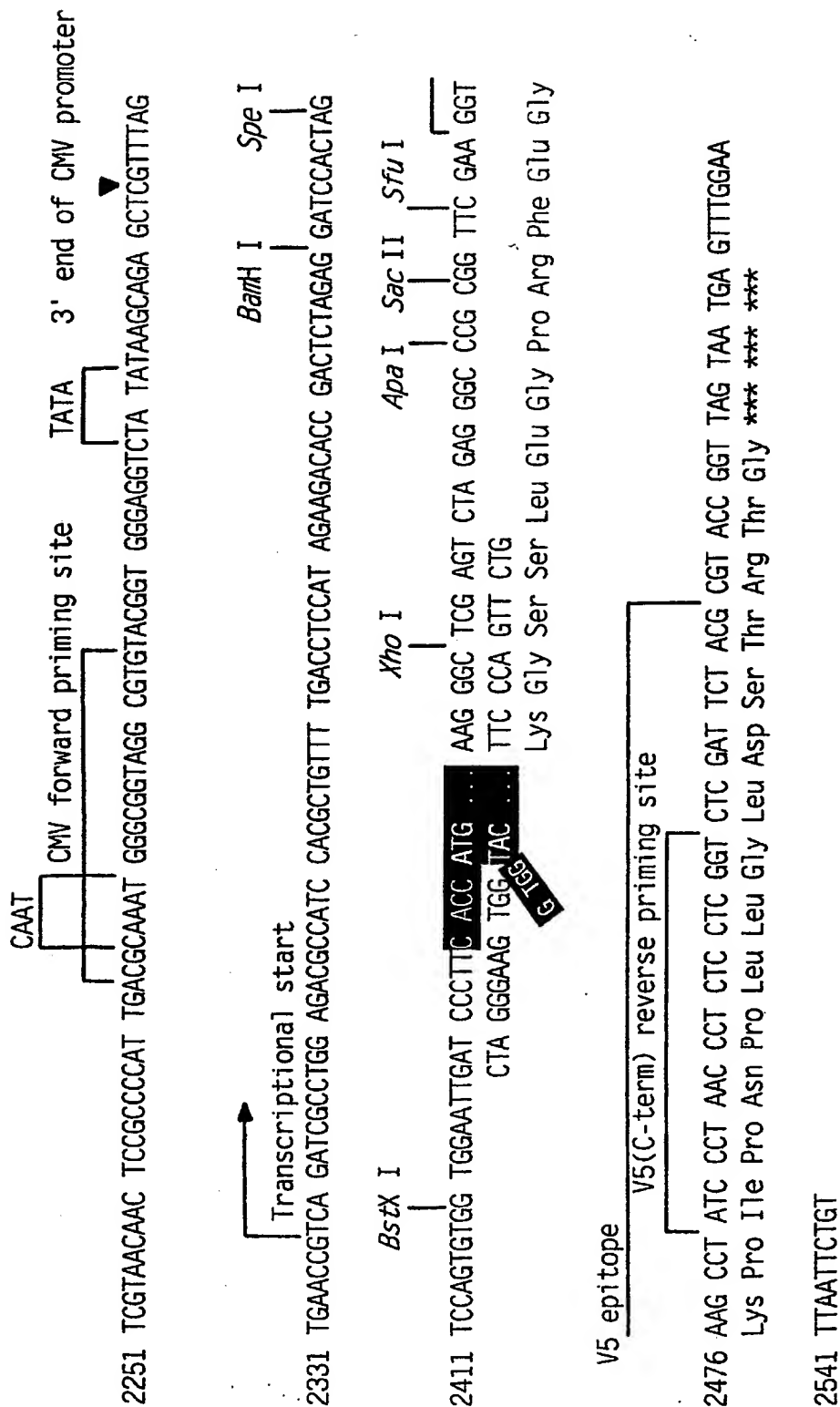


FIG.48

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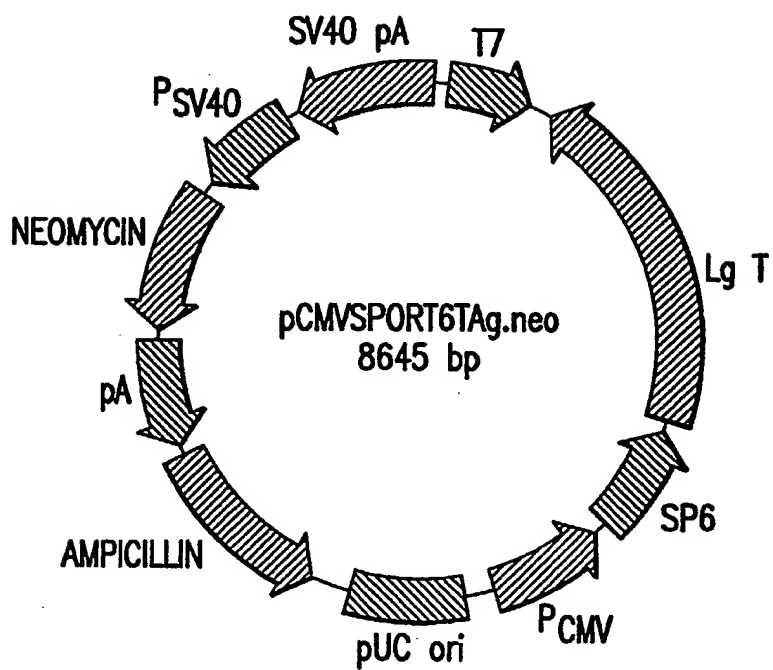


FIG.49

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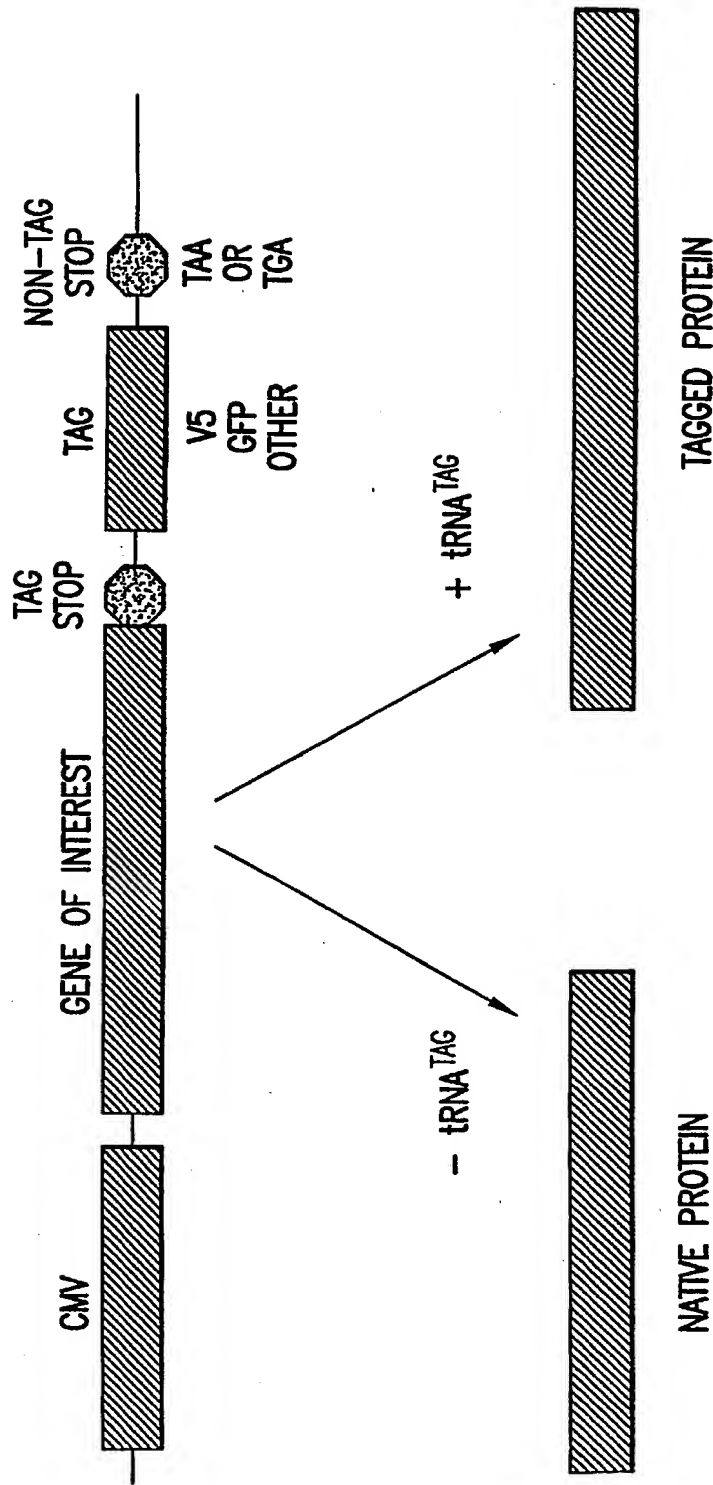


FIG.50



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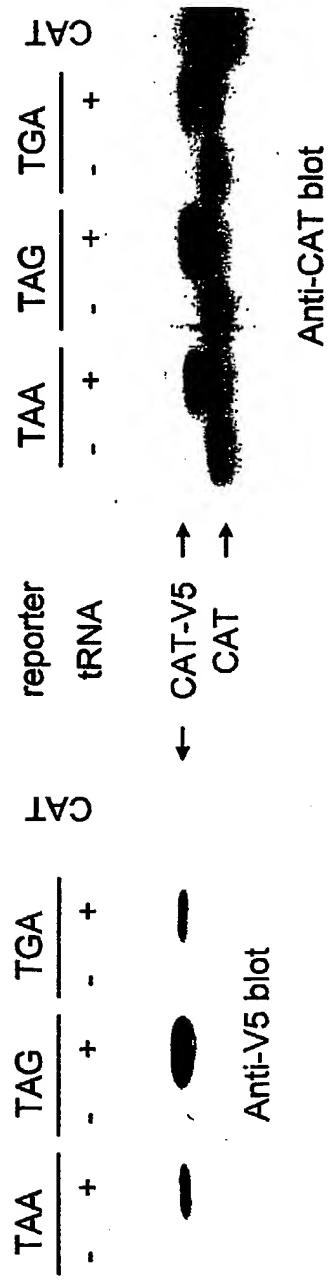


FIG.51A

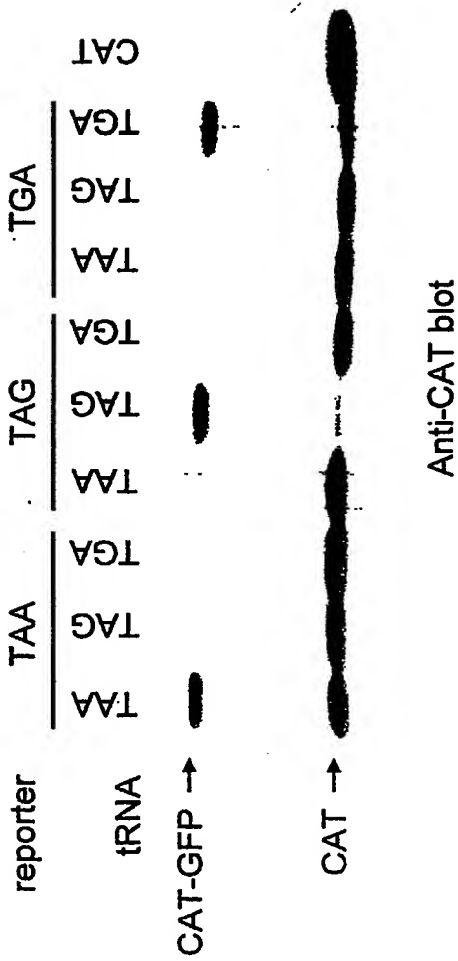


FIG.51B

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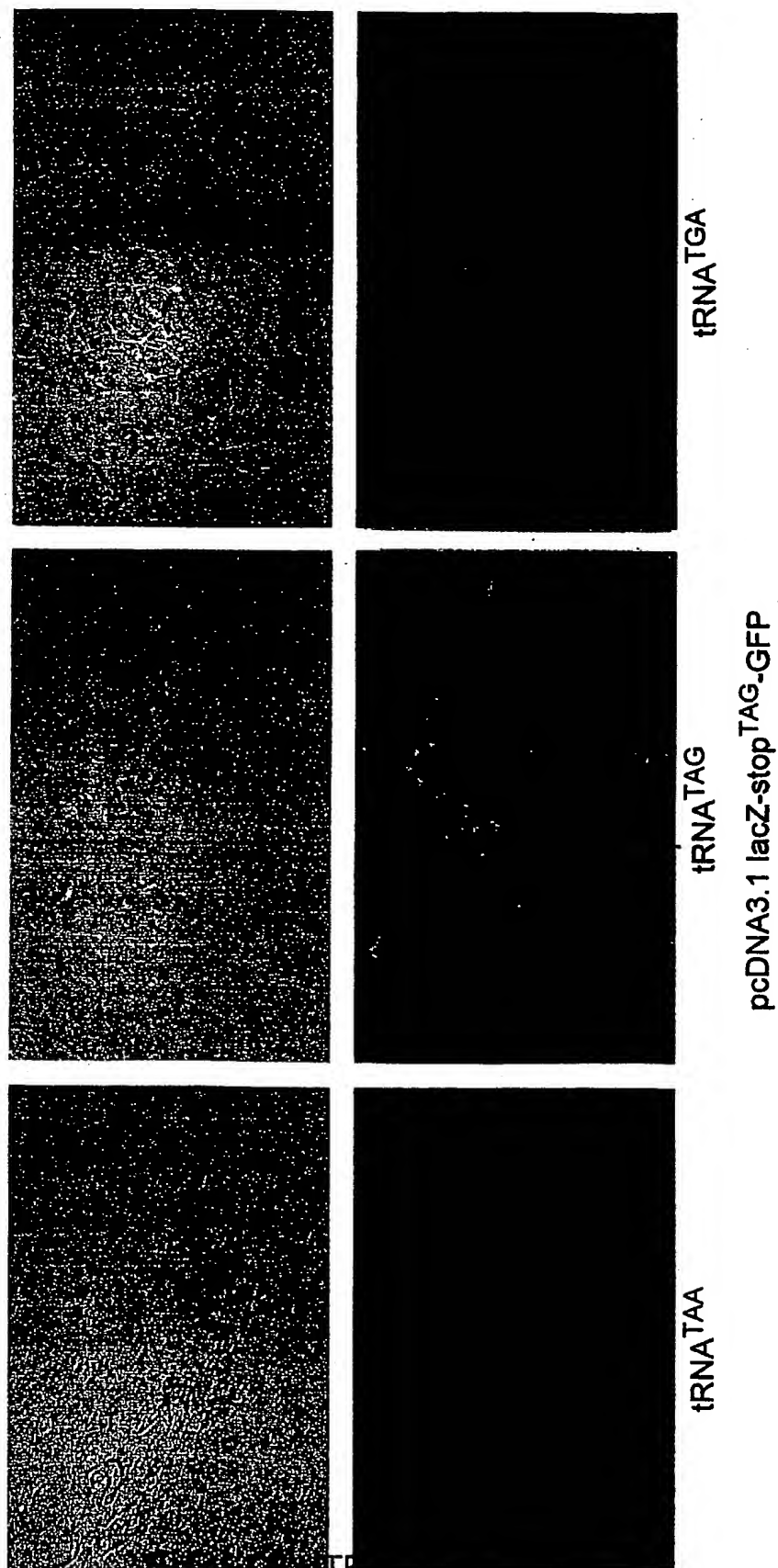


FIG. 52

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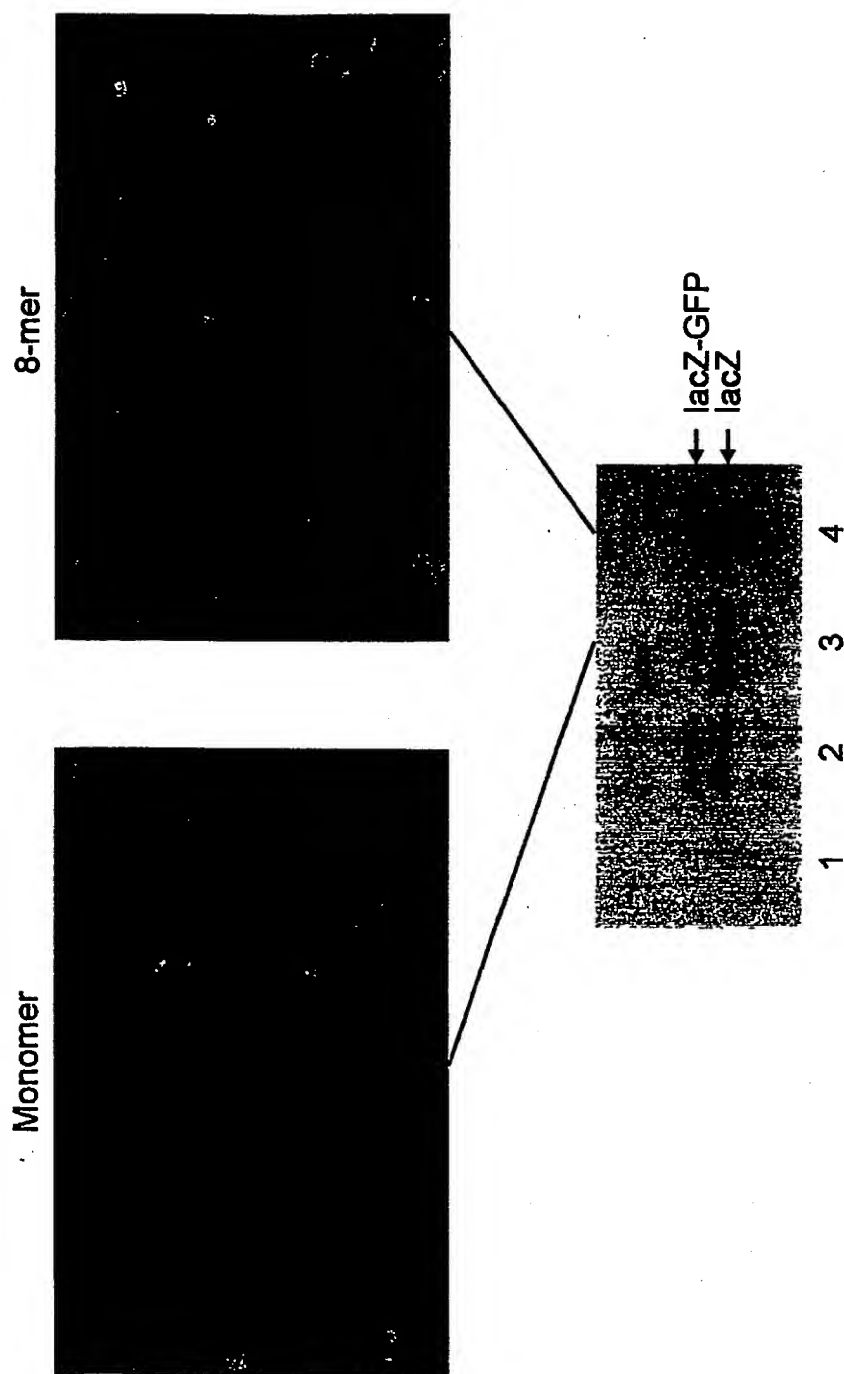


FIG. 53

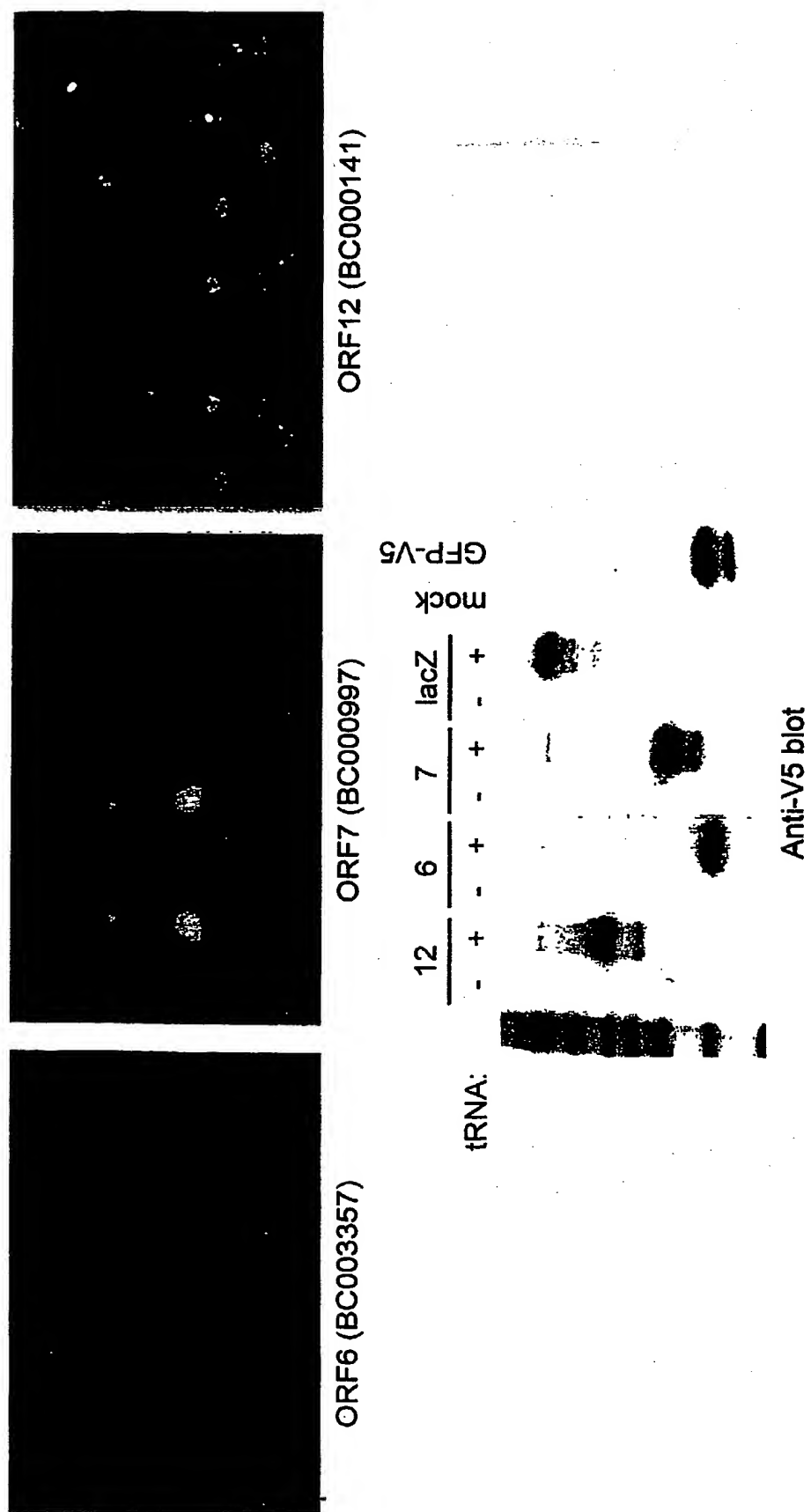


FIG. 54

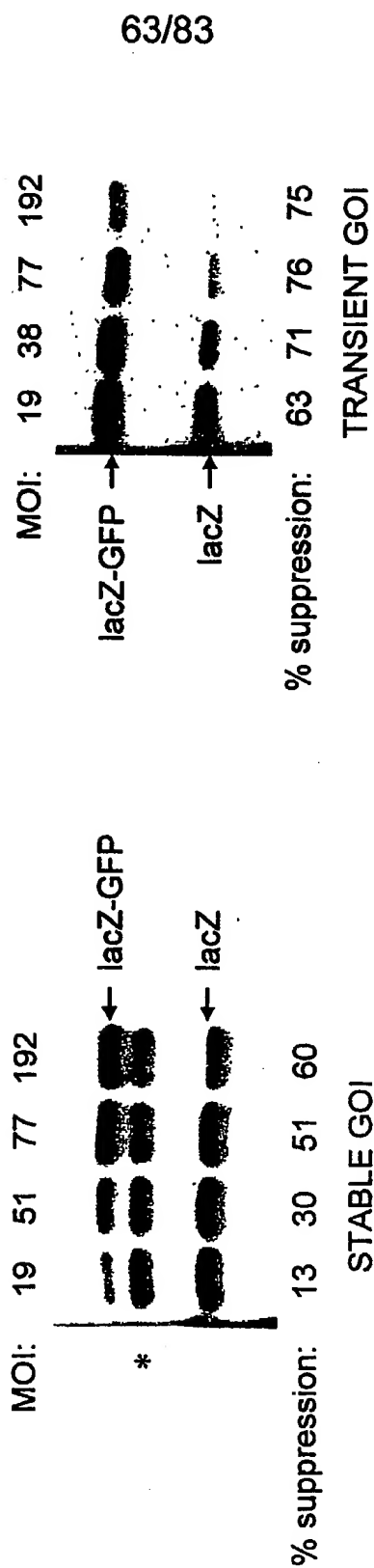


FIG.55B

FIG.55A

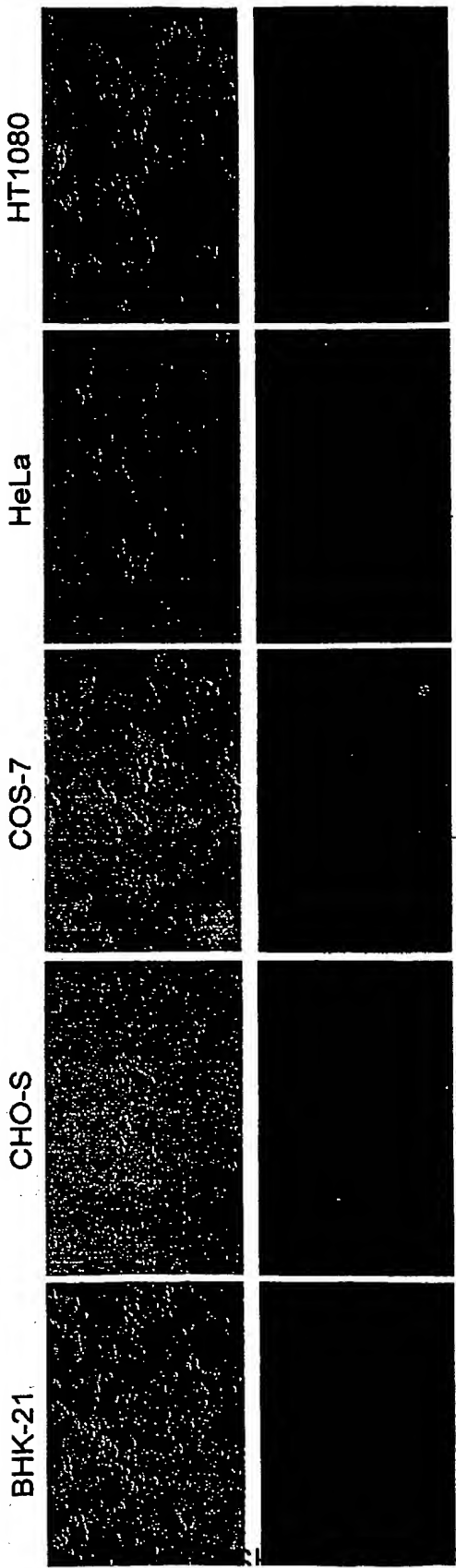


FIG. 56

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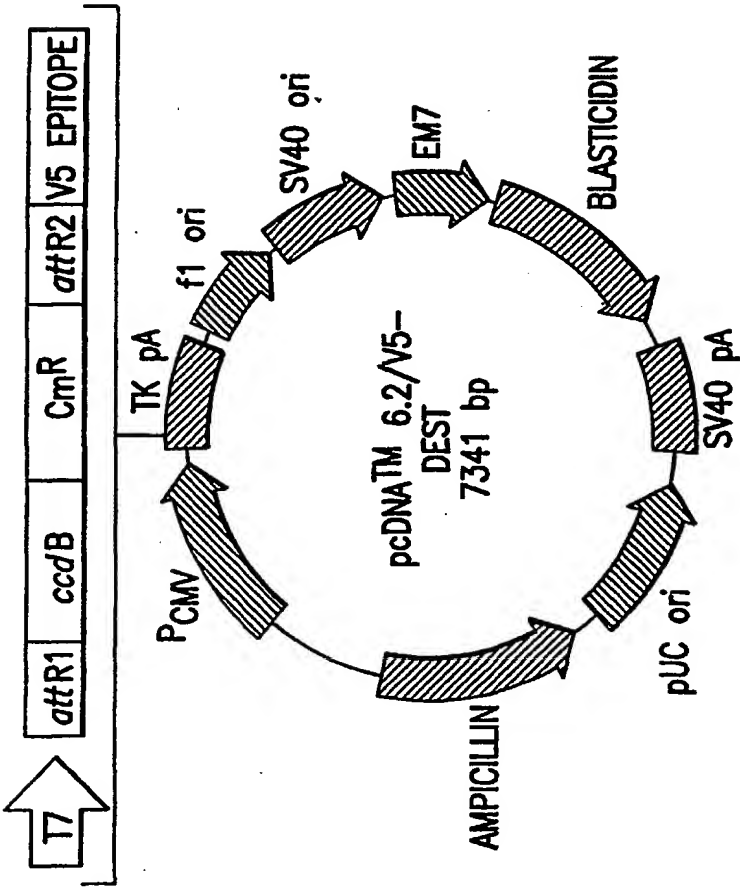


FIG.57

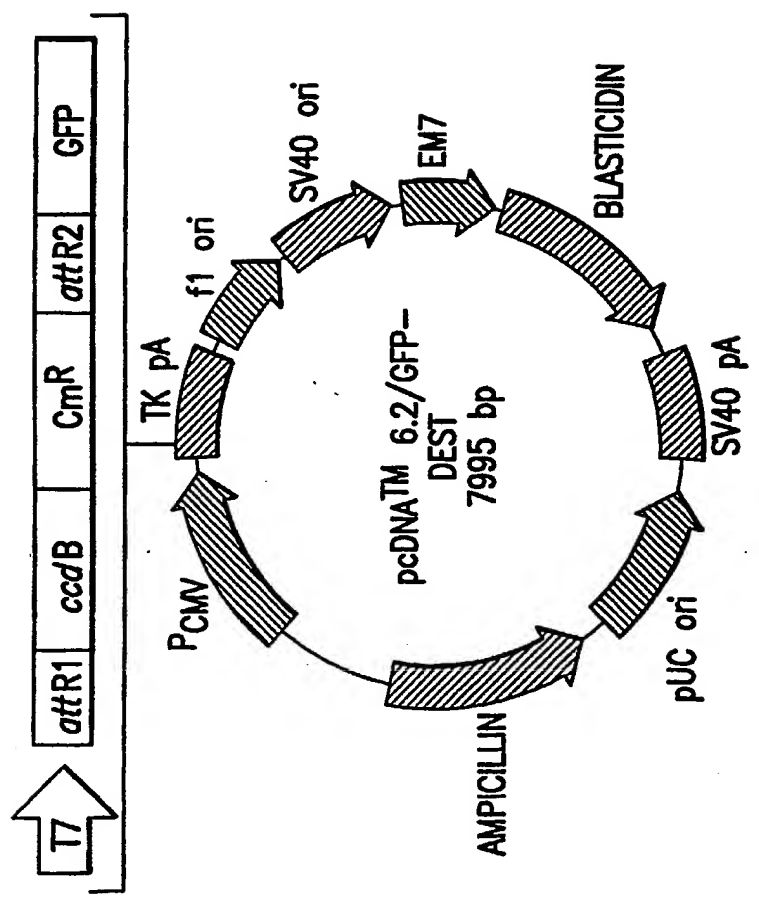


FIG.58



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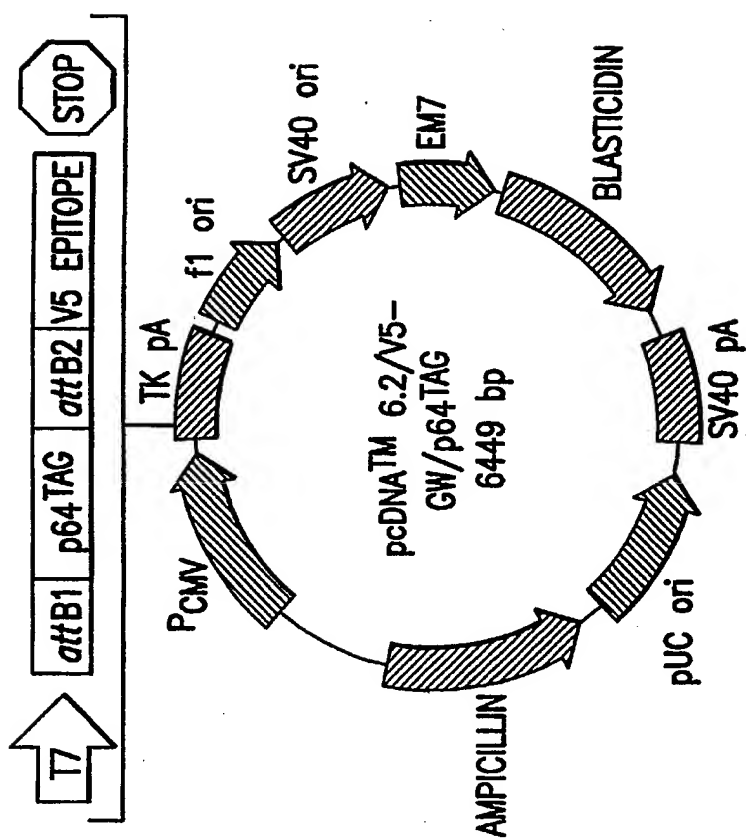


FIG.59

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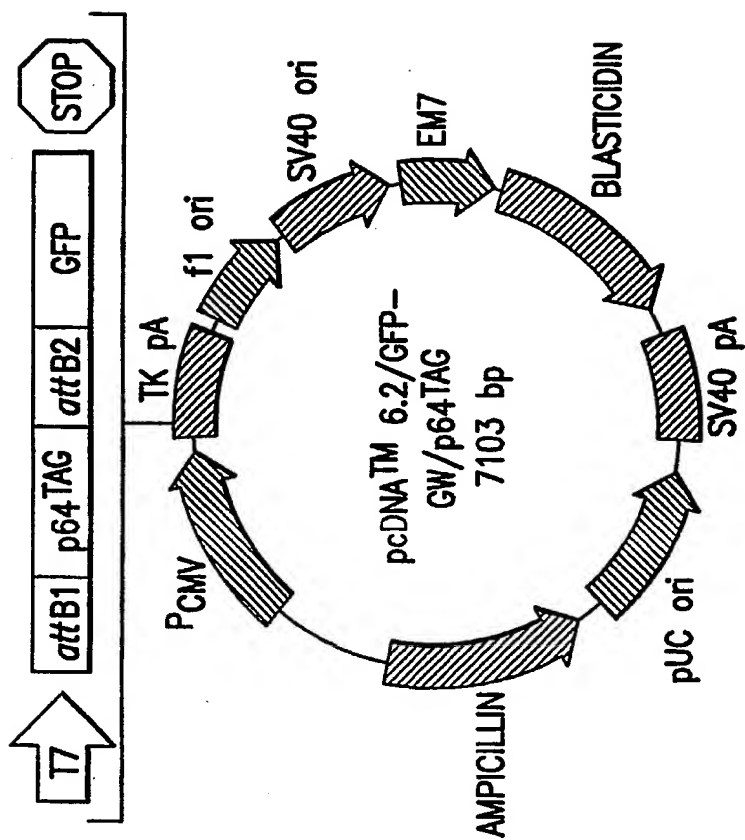


FIG.60

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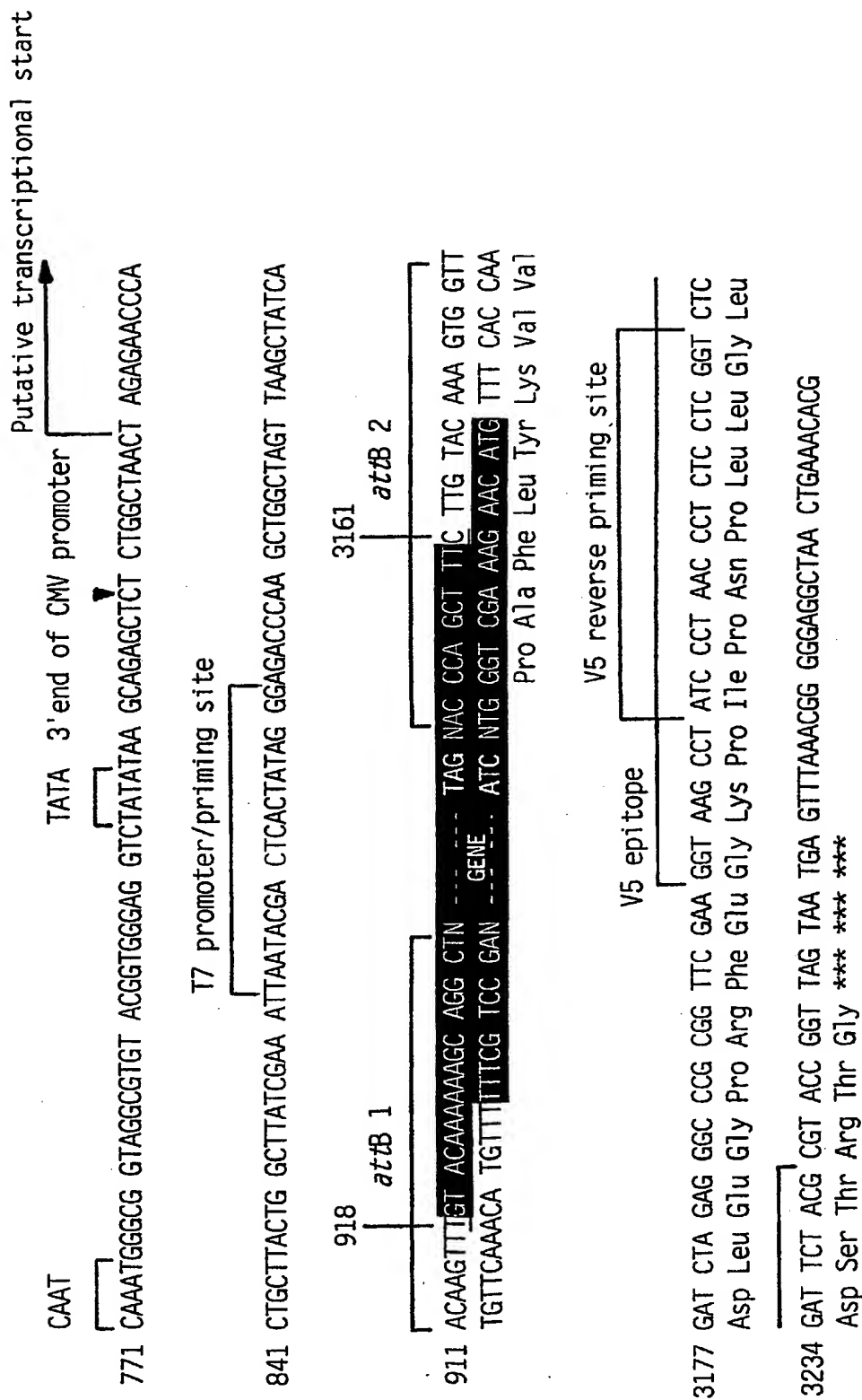


FIG.61A

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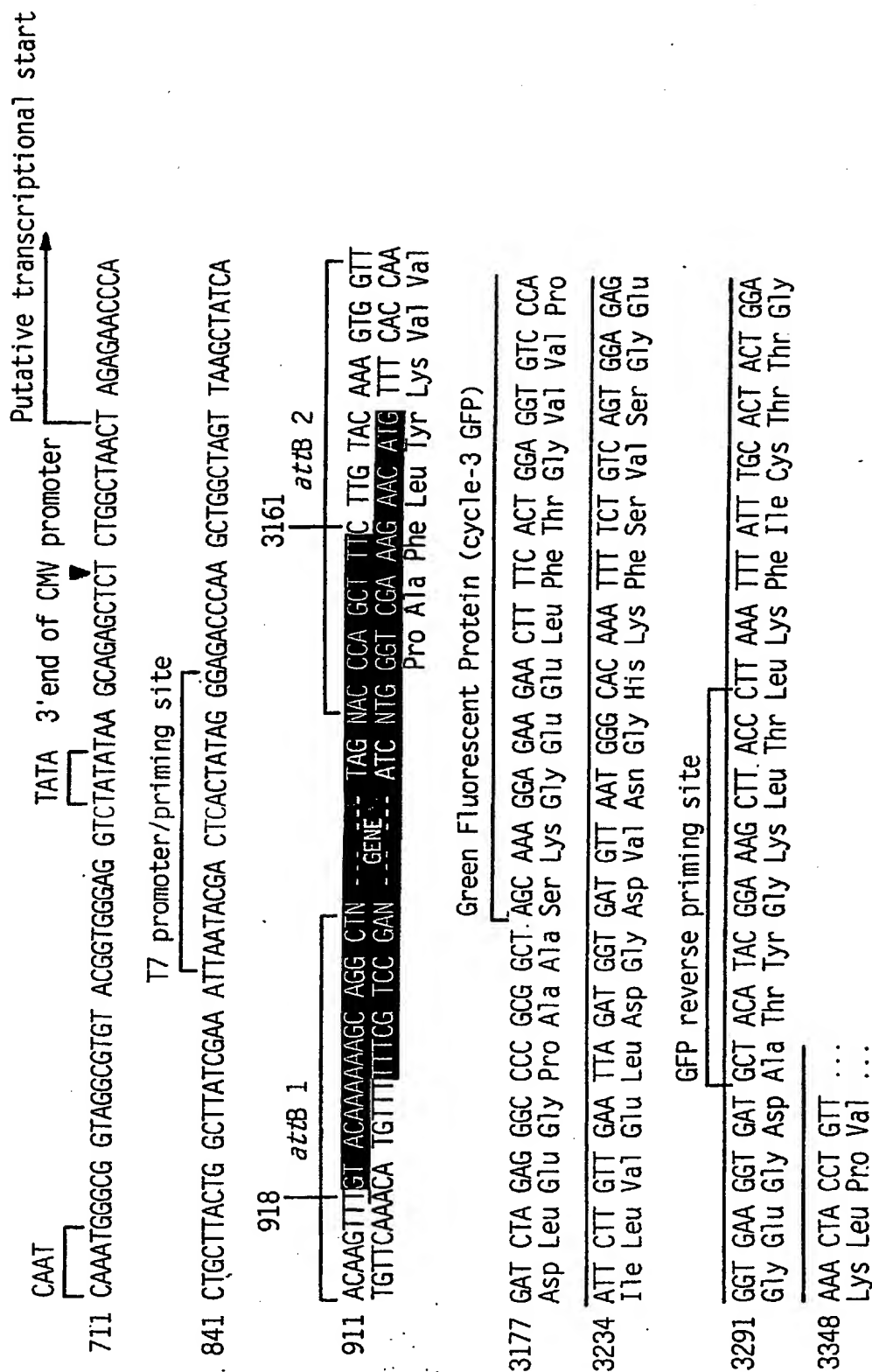
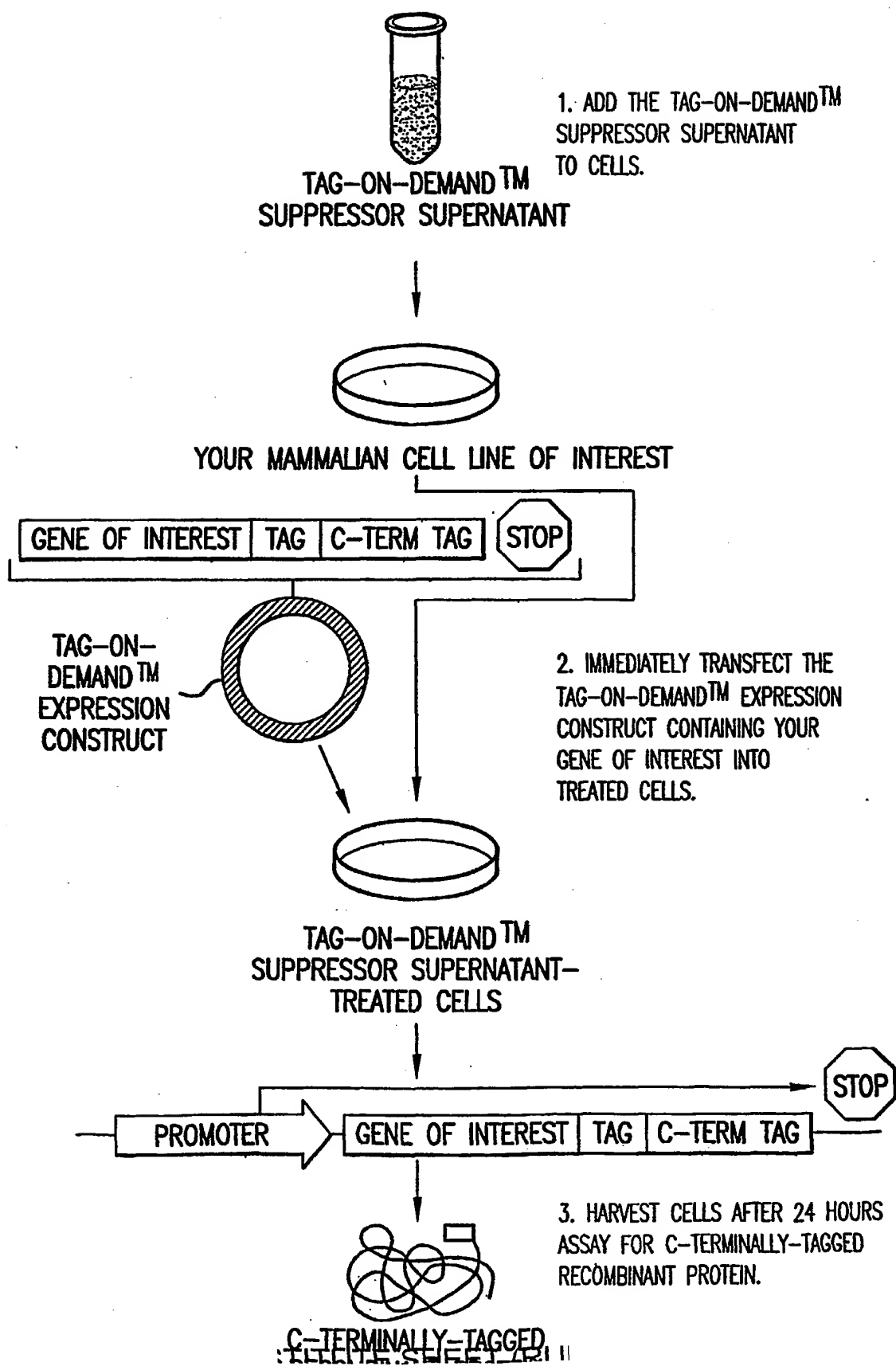


FIG. 61B

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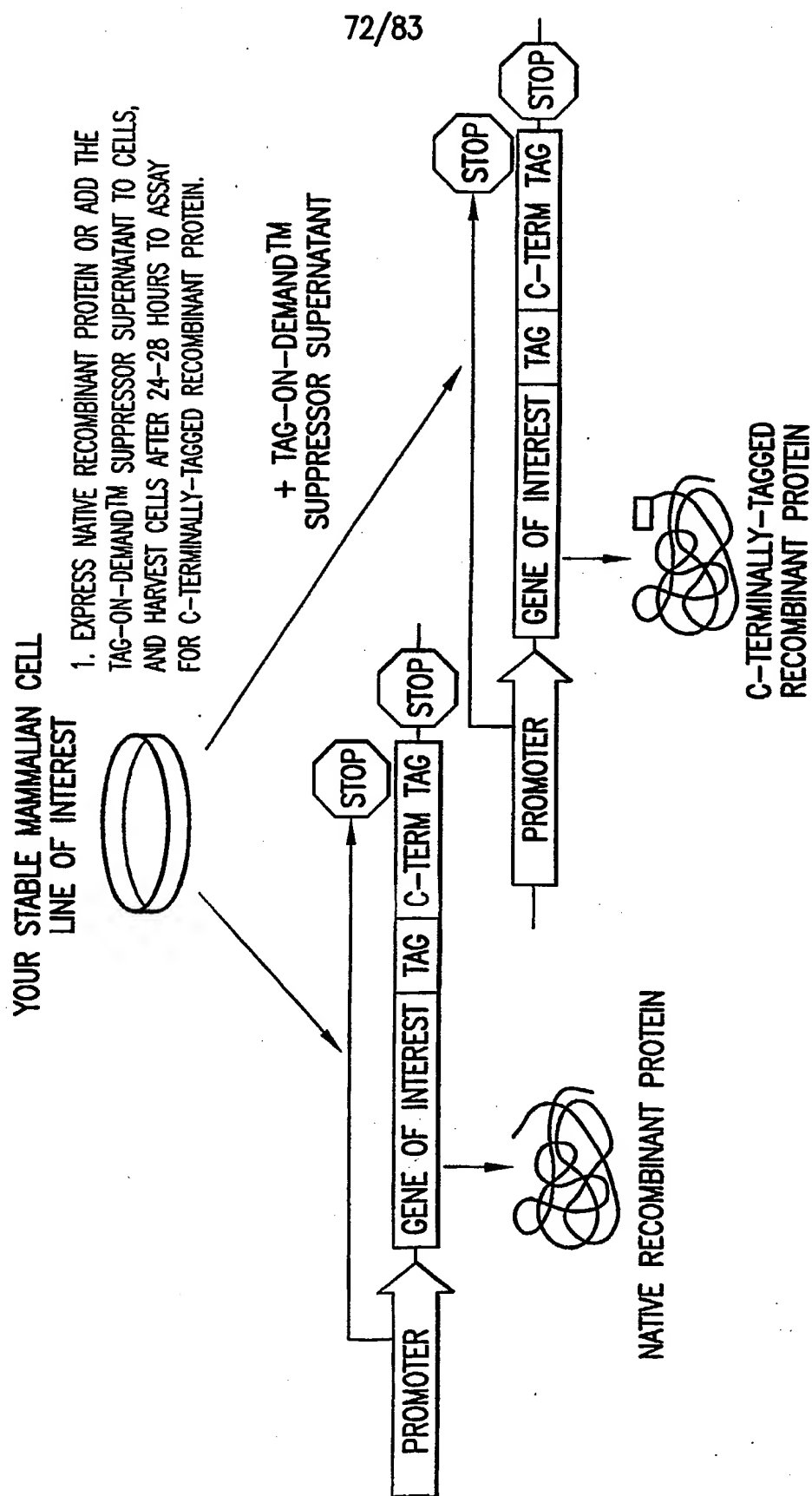


FIG.63

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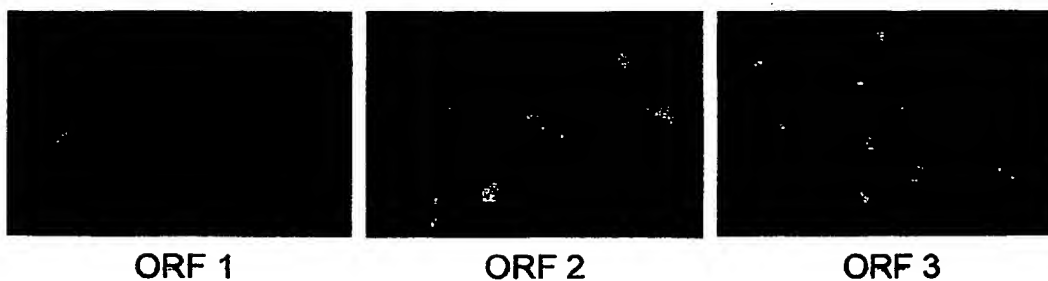


FIG.64

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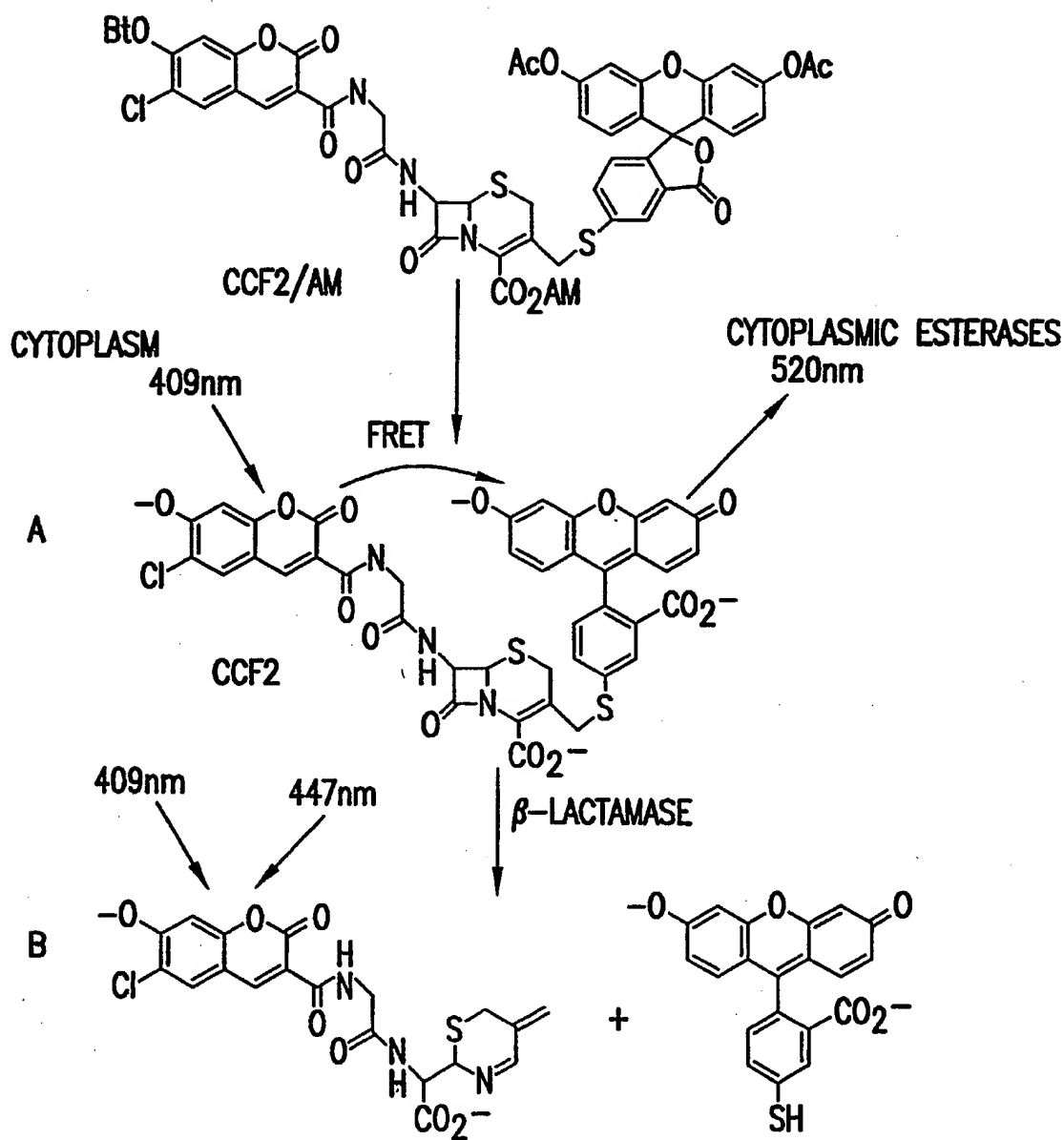


FIG.65



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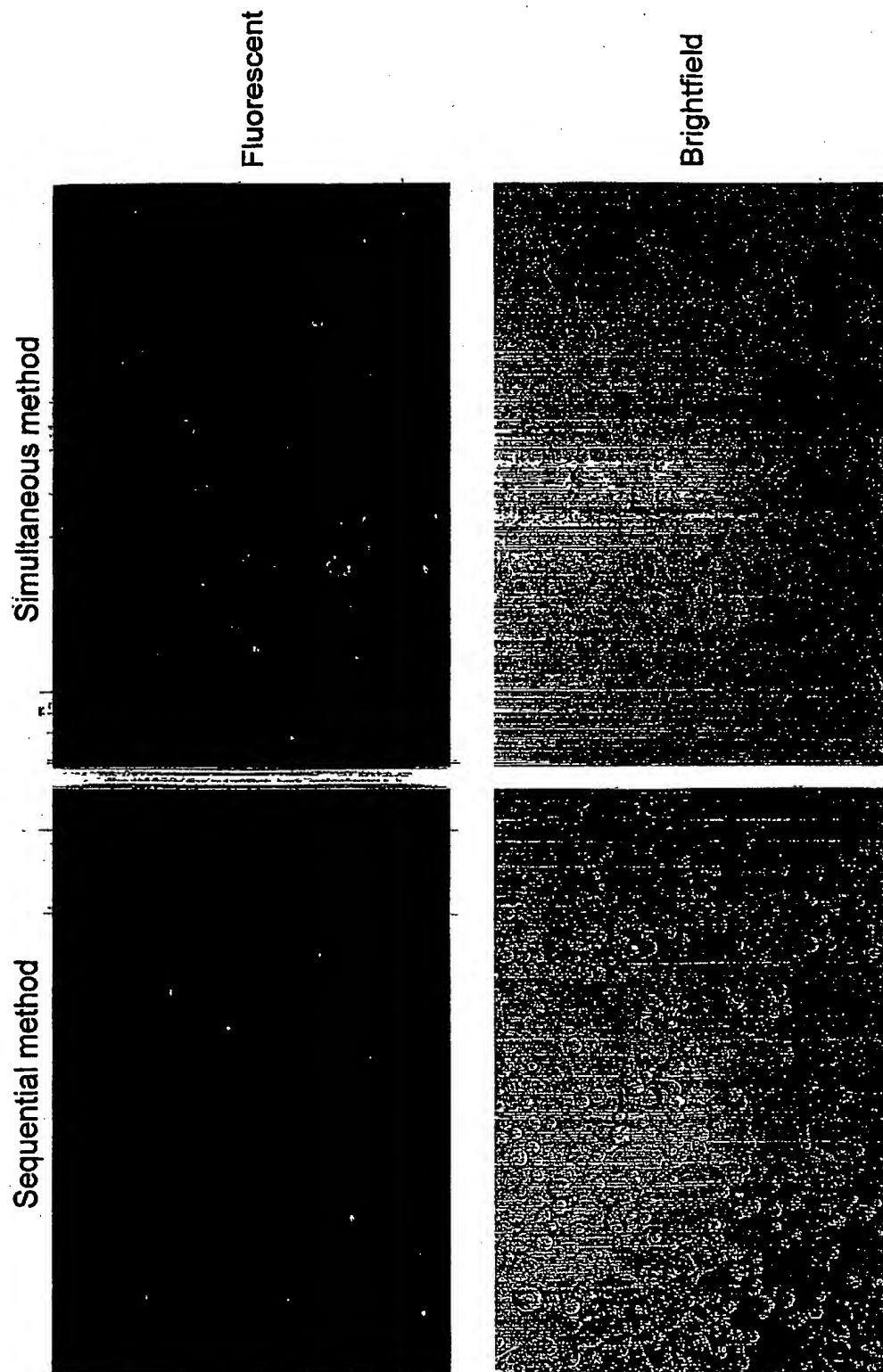


FIG.66

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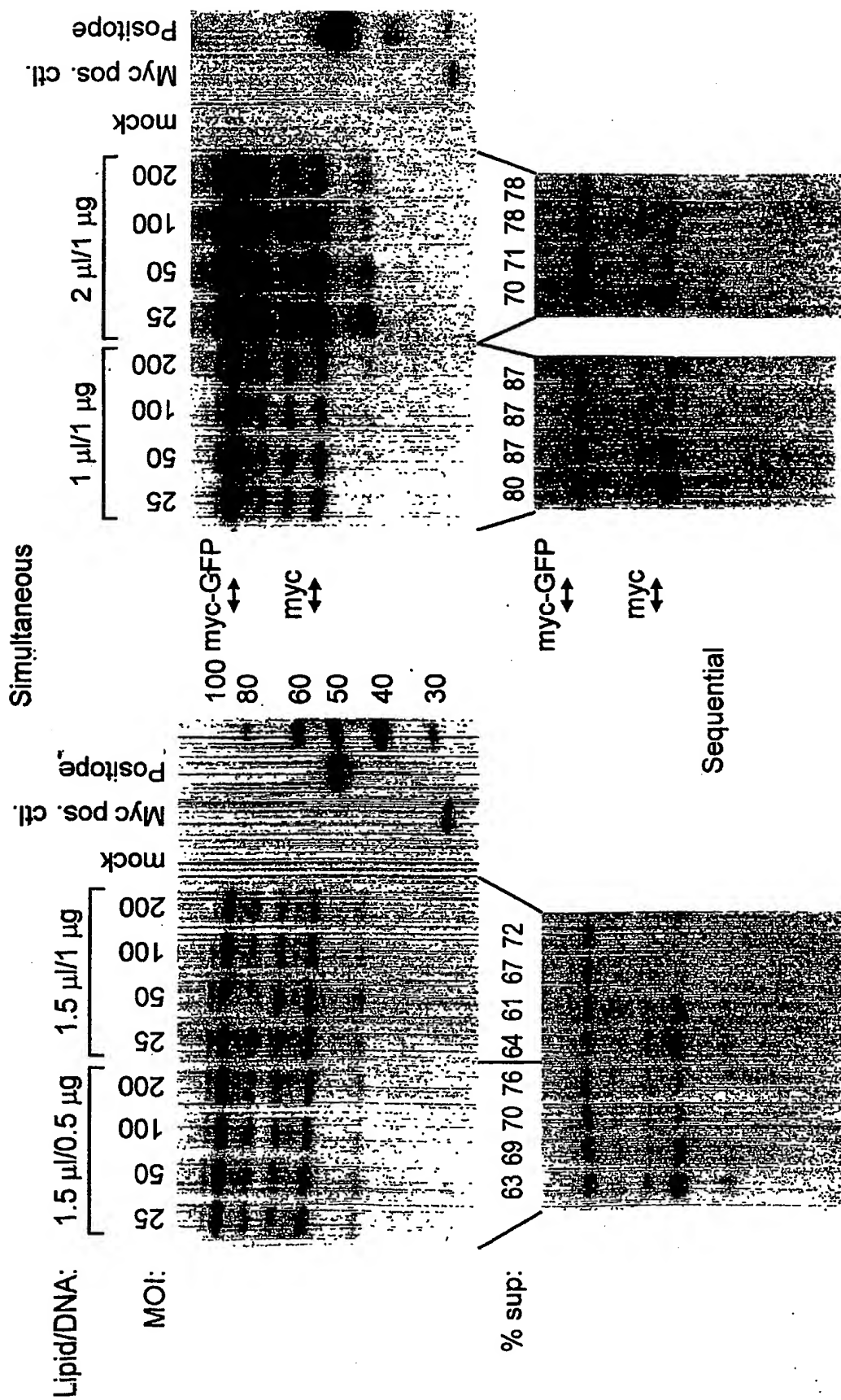


FIG. 67

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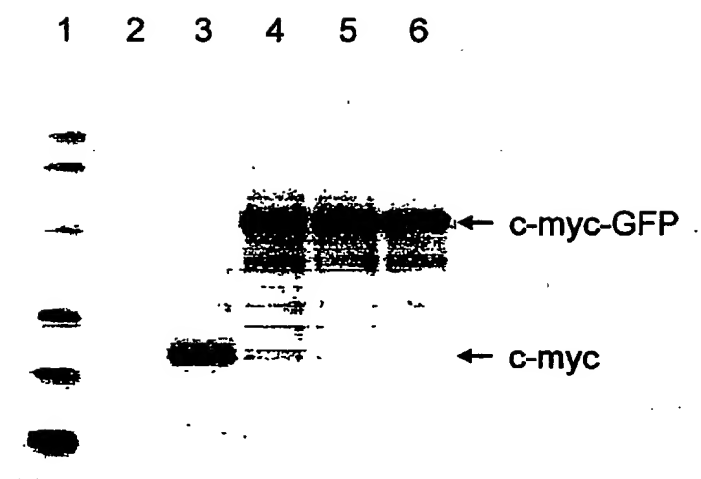


FIG.68

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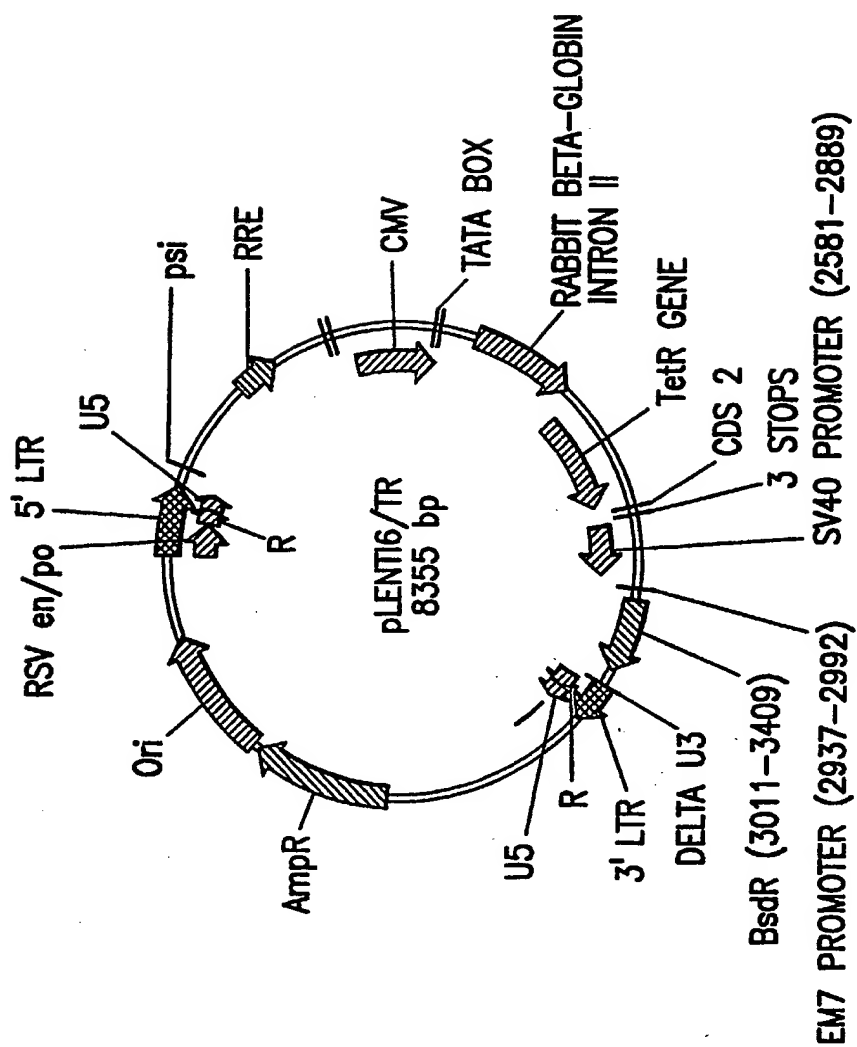


FIG.69

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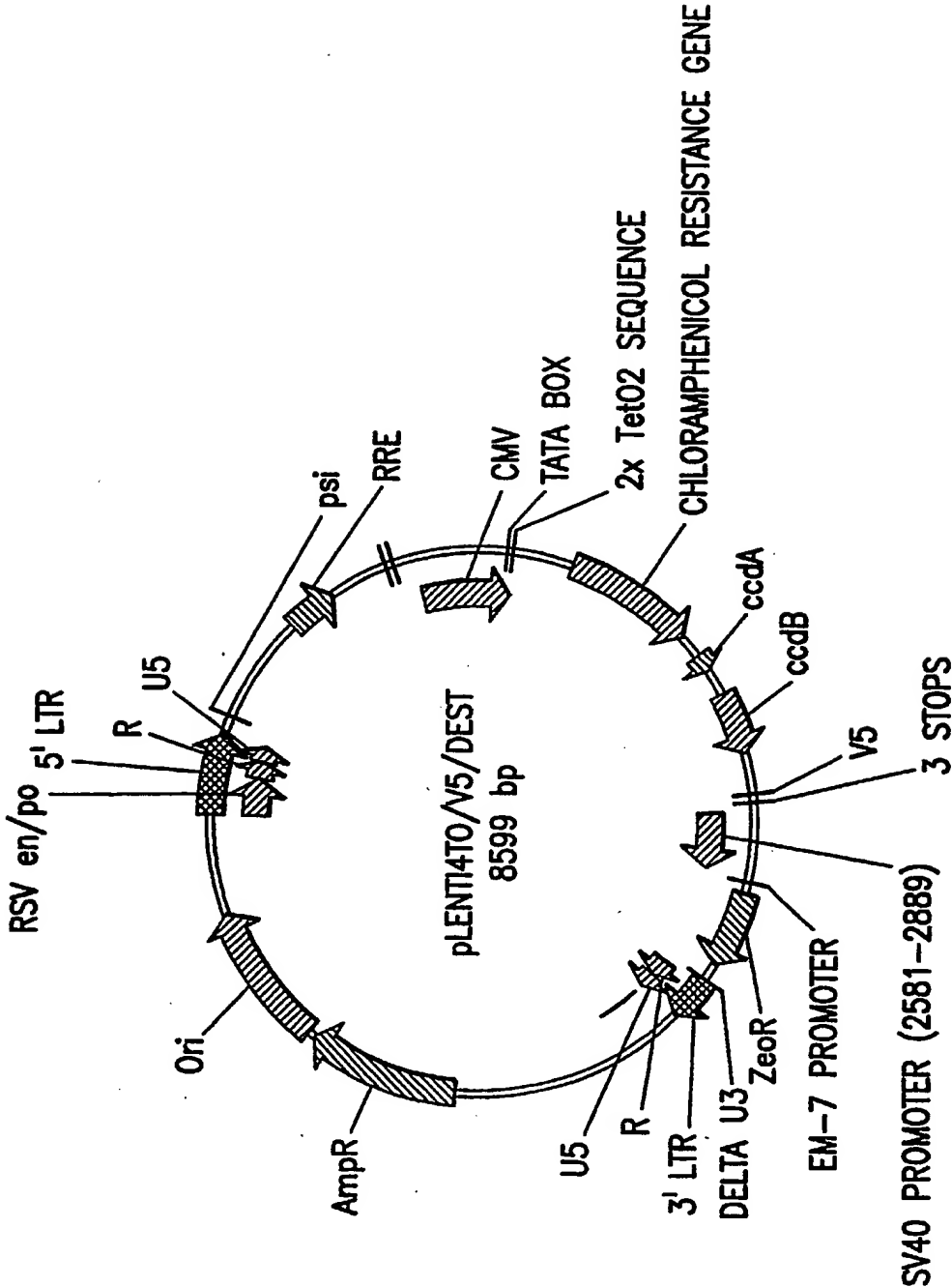


FIG.70A

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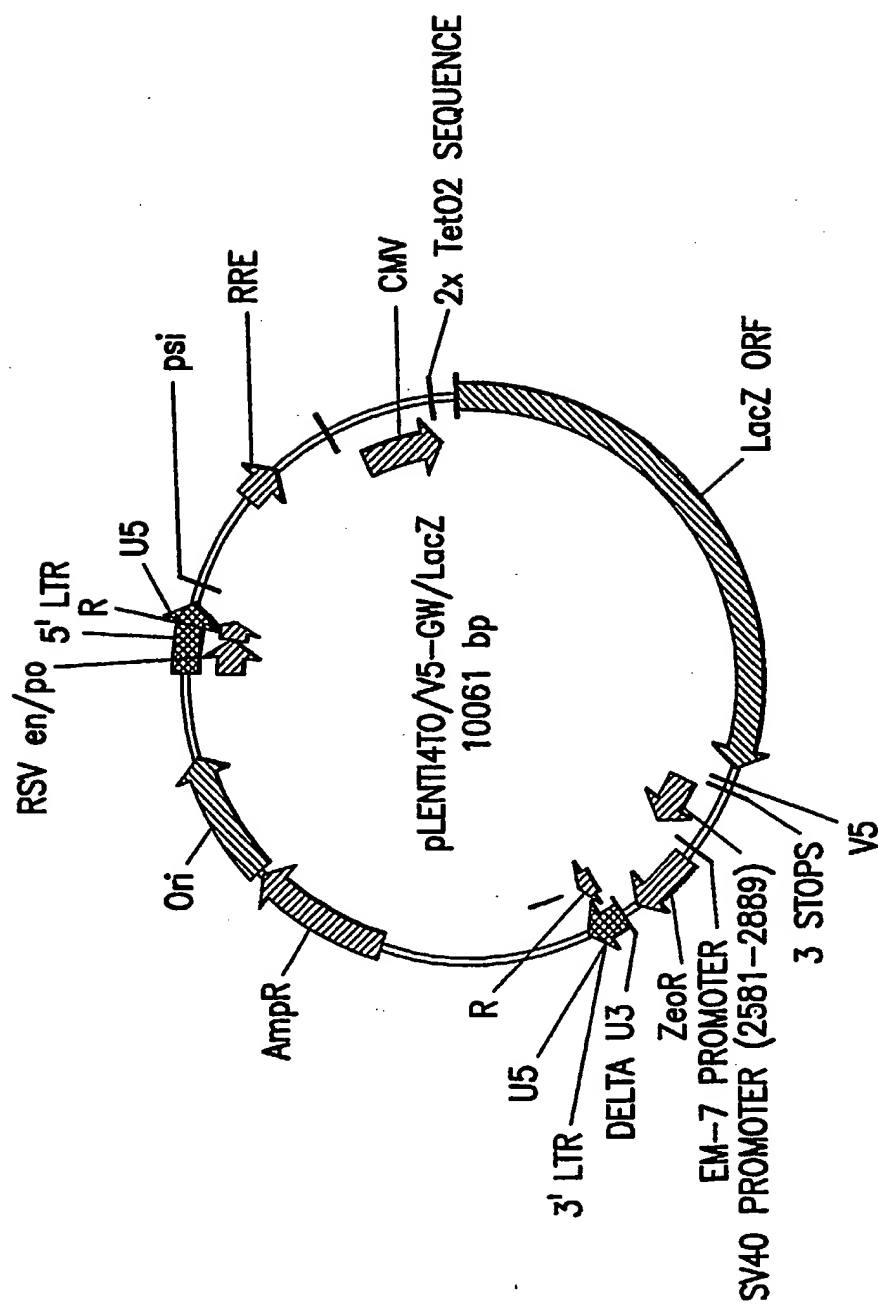


FIG. 70B

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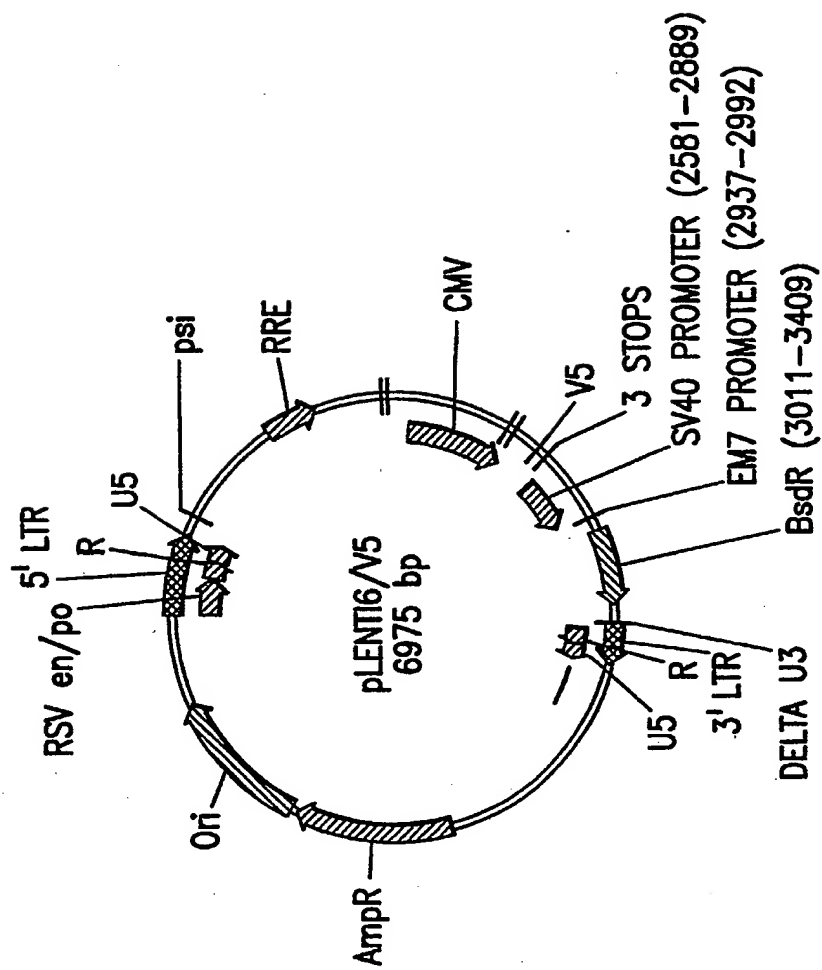


FIG. 71

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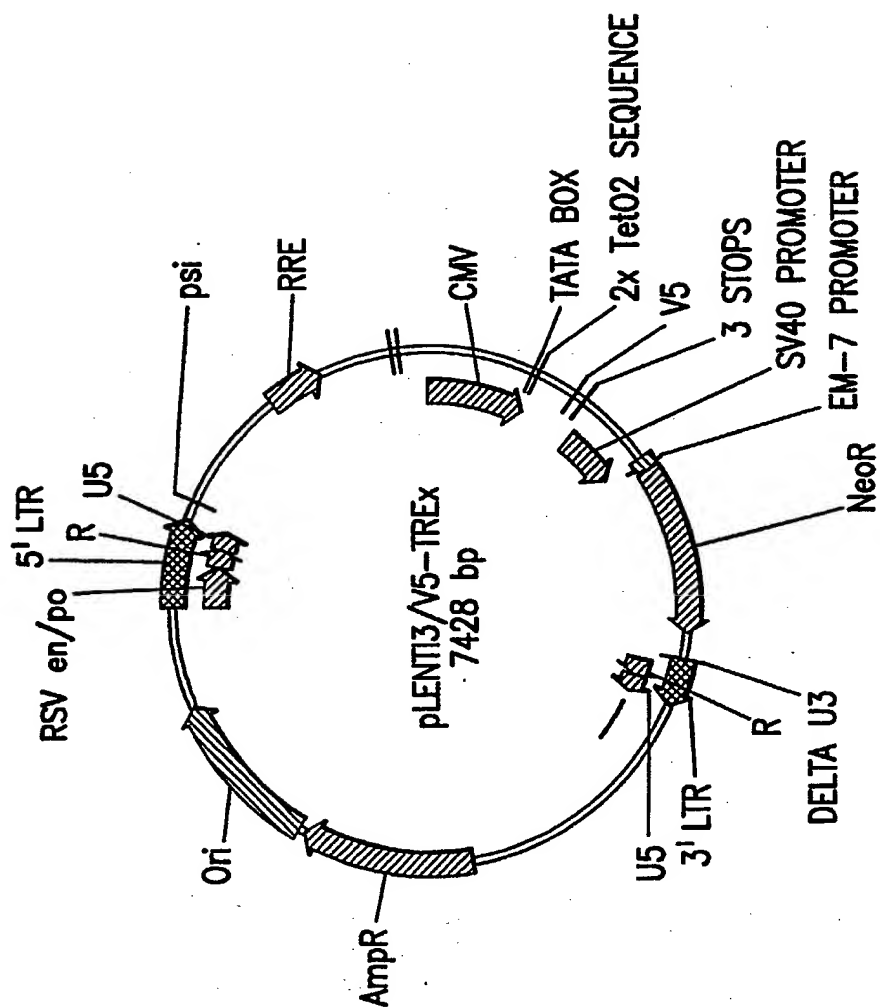


FIG.72



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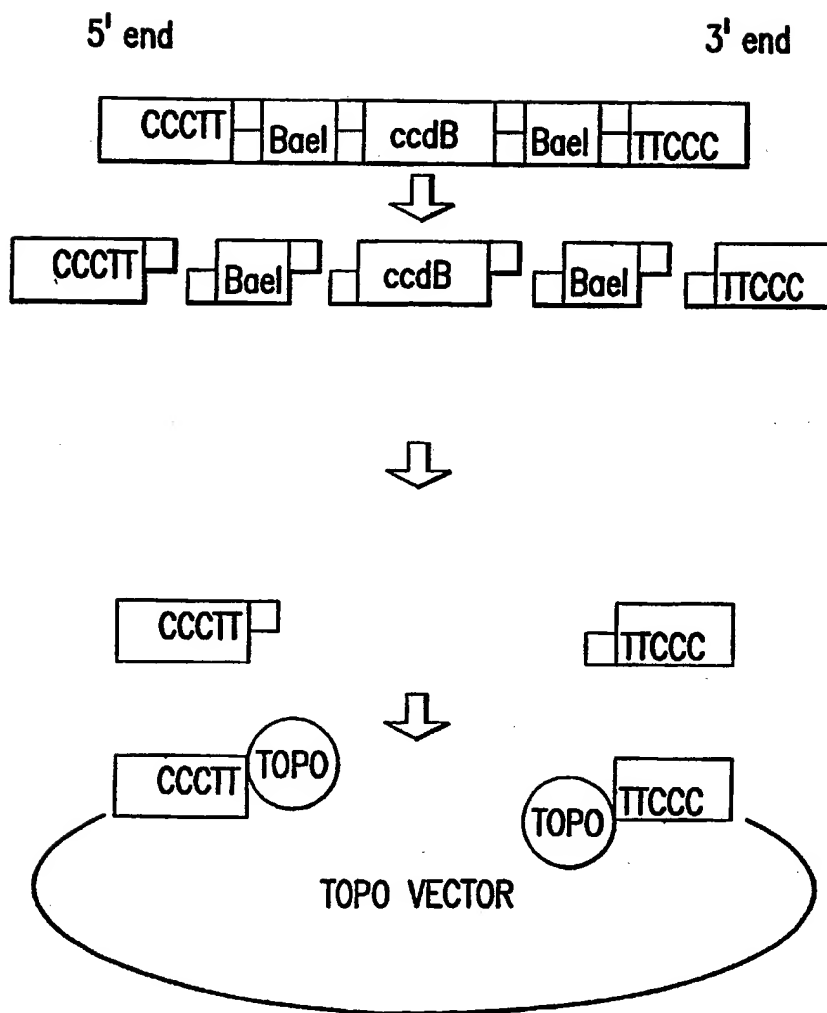


FIG.73